

IMRE TAAL

Causes of variation in littoral fish  
communities of the Eastern Baltic Sea:  
from community structure  
to individual life histories



DISSERTATIONES BIOLOGICAE UNIVERSITATIS TARTUENSIS

**327**

DISSERTATIONES BIOLOGICAE UNIVERSITATIS TARTUENSIS

**327**

**IMRE TAAL**

Causes of variation in littoral fish  
communities of the Eastern Baltic Sea:  
from community structure  
to individual life histories



UNIVERSITY OF TARTU  
Press

Estonian Marine Institute and Department of Zoology, Institute of Ecology and Earth Sciences, Faculty of Science and Technology, University of Tartu, Estonia.

This dissertation was accepted for commencement of the degree of *Doctor philosophiae* in Ichthyology and Fisheries Science from the University of Tartu on June 5<sup>th</sup> 2017, by the Scientific Council of the Institute of Ecology and Earth Sciences, University of Tartu.

Supervisors:      Markus Vetemaa, PhD  
                      Lauri Saks, PhD

Estonian Marine Institute  
University of Tartu  
Estonia

Opponent:        Johanna Mattila, PhD  
  
Swedish University of Agricultural Sciences  
Sweden

Commencement: Room 301, 46 Vanemuise Street, Tartu, on November 24, 2017, at 10.15 a.m.

Publication of this thesis is granted by the Estonian Marine Institute, University of Tartu.

ISSN 1024-6479  
ISBN 978-9949-77-590-3 (print)  
ISBN 978-9949-77-591-0 (pdf)

Autoriõigus Imre Taal, 2017

Tartu University Press  
[www.tyk.ee](http://www.tyk.ee)

## **CONTENTS**

LIST OF ORIGINAL PUBLICATIONS .....	6
DEFINITIONS AND ABBREVIATIONS .....	7
1. INTRODUCTION.....	8
2. STUDY SYSTEM.....	12
2.1. The study area.....	12
2.2. Field sampling and methods .....	14
3. RESULTS AND DISCUSSION .....	16
3.1. Diel changes in the fish assemblage of a sandy surf-zone area of the Baltic Sea .....	16
3.2. Diet composition of European smelt in a near-shore ecosystem .....	19
3.3. The littoral zone as a habitat of early out-migrating Atlantic salmon and anadromous brown trout juveniles in the Baltic Sea .....	21
4. CONCLUSIONS.....	28
SUMMARY IN ESTONIAN .....	29
ACKNOWLEDGEMENTS .....	33
REFERENCES.....	34
PUBLICATIONS .....	41
CURRICULUM VITAE .....	98
ELULOOKIRJELDUS.....	100

## LIST OF ORIGINAL PUBLICATIONS

The current thesis was based on the following papers, which are referred to in the text by the Roman numerals I–V:

- I. Taal I., Saks L., Kesler K., Verliin A., Jürgens K., Svirgsden R., Rohtla M., Hubel K., Albert A., Eschbaum R. & Vetemaa, M. 2017. Diel changes in the fish assemblage in a coastal surf-zone area in the eastern Baltic Sea. *Boreal Environment Research*, 22, 83–96.
- II. Taal I., Saks L., Nedolgova S., Verliin A., Kesler M., Jürgens K., Svirgsden R., Vetemaa M., & Saat T. 2014. Diet composition of smelt *Osmerus eperlanus* (Linnaeus) in brackish near-shore ecosystem (Eru Bay, Baltic Sea). *Ecology of Freshwater Fish*, 23, 121–128.
- III. Taal I., Kesler M., Saks L., Rohtla M., Verliin A., Svirgsden R., Jürgens K., Vetemaa M. & Saat T. 2014. Evidence for an autumn downstream migration of Atlantic salmon *Salmo salar* (Linnaeus) and brown trout *Salmo trutta* (Linnaeus) parr to the Baltic Sea. *Helgoland Marine Research*, 68, 373–377.
- IV. Taal, I., Rohtla, M., Saks, L., Kesler, M., Jürgens, K., Svirgsden, R., Matetski, L., Verliin, A., Paiste, P. & Vetemaa, M. 2017. Parr dispersal between streams *via* a marine environment: a novel mechanism behind straying for anadromous brown trout? *Ecology of Freshwater Fish*. doi: 10.1111/eff.12338.
- V. Taal, I., Rohtla, M., Saks, L., Svirgsden, R., Kesler, M., Matetski, L. & Vetemaa, M. 2017. Evidence of Atlantic salmon *Salmo salar* fry movement between fresh water and a brackish environment. *Journal of Fish Biology*, 91, 695–703.

### Author's contribution:

I was the lead author of all five publications. I generated the original research ideas and wrote the first drafts of the publications. I also took an active part in collecting field data and participated in data analysis.

Original papers are reproduced with the permission of John Wiley & Sons, Inc., Springer Science + Business Media, and the Boreal Environment Research Publishing Board.

## DEFINITIONS AND ABBREVIATIONS

**ANADROMOUS:** Fish that find a major part of their food in salt water, but spawn in freshwater (*e.g.* Atlantic salmon (*Salmo salar*), anadromous brown trout (*Salmo trutta*) (Jonsson & Jonsson 2011).

**ABT:** anadromous brown trout (sea trout).

**LITTORAL (ZONE):** There is no single definition of the concept of “littoral zone”, as it varies by context (*e.g.* oceans, Baltic Sea, rivers, lakes). In this dissertation, I define littoral (littoral zone) as the area ranging from the shoreline to the outer edge of the aquatic vegetation zone. In the habitats considered in this dissertation, the littoral reaches water depths of up to 10–15 metres (see also Hänninen *et al.* 2007).

**NEAR-SHORE ENVIRONMENT:** In this dissertation, I define near-shore as the marine environment up to 3 km from the coastline and less than 25 m deep.

**PARR:** Young fish (*e.g.* Atlantic salmon, anadromous brown trout) in freshwater, which are characterized by parr marks (a series of parallel vertical bars) along their flanks. Anadromous salmonids are considered parr from having consumed their yolk until smolting (Jonsson & Jonsson 2011). Post-independence of the yolk sac to the end of their first summer, young parr are also referred to as fry (*e.g.* Hendry & Cragg-Hine 2003).

**SMOLTING (SMOLTIFICATION):** The process involving morphological, physiological, biochemical, and behavioural changes, whereby salmonid parr pre-adapt to marine life (Jonsson & Jonsson 2011).

**SSME (Stream Shifting through the Marine Environment):** A previously undescribed life-history strategy of Atlantic salmon and anadromous brown trout. SSME involves salmon and ABT descending to the brackish waters of the Baltic Sea from their natal streams as parr or fry, and then migrating to non-natal streams before transforming into smolt (**IV**; **V**).

**STRAY:** Fish that enter a stream to spawn where they did not hatch (*e.g.* Atlantic salmon, anadromous brown trout) (Jonsson & Jonsson 2011).

**SURF-ZONE:** the shallow littoral area between the shoreline and the most seaward point of the breaker zone (surf line). The breaker zone is defined as the zone within which waves approaching the coastline break.

**YOY (young of the year):** Fish born within the past year, from transformation to juvenile until January 1<sup>st</sup> in the Northern Hemisphere and July 1<sup>st</sup> in the Southern Hemisphere (FishBase Glossary 2017).

## 1. INTRODUCTION

The shallow littoral is a highly productive environment characterized by the multitude of interactions within and between different trophic levels. It serves as an important reproduction, nursery, and foraging area to coastal marine biota (e.g. Thorman & Wiederholm 1986; Gibson & Robb 1996; Mustamäki *et al.* 2014; Lokko *et al.* 2016; Morkūnė *et al.* 2016). Littoral fish communities are highly variable, as species composition and abundance differ over both spatial and temporal scales (e.g. Thorman & Wiederholm 1986; Sundell 1994; Rajasilta *et al.* 1999; Wilber *et al.* 2003; Vahteri *et al.* 2009; Vasconcellos *et al.* 2011; Mustamäki *et al.* 2015). Earlier studies have indicated that the factors driving the occurrence and abundance of fish in near-shore areas can be divided as: (1) climatic events that affect year-class strength; (2) reproductive and feeding movements that impose seasonal variation; and (3) short-term physico-chemical conditions (e.g. turbidity, wave height, salinity, tidal stage, temperature, photoperiod), which may influence point abundance (reviewed by e.g. Ross *et al.* 1987; Wilber *et al.* 2003).

The Baltic Sea is very shallow (mean depth of 55 m, Myrberg & Andrejev 2003) and has a highly variable coastline (reviewed in Niemelä *et al.* 2015), ranging from soft-formed depositional shores to bedrock-dominated archipelagos. There are also considerable environmental constraints on abiotic and biotic parameters (e.g. salinity, temperature, nutrient status, and food base to fish) between different sub-basins of the Baltic Sea, and from estuaries and bays to the open sea (Neumann 1977; Uusitalo *et al.* 2005). In the brackish Baltic Sea, the fauna comprises a mixture of species of marine and freshwater origin, which have adapted to the low salinities of the environment. The salinity of the Baltic Sea decreases east- and north-wards, resulting in a decrease in the number of marine species and an increase of freshwater species (Järvekülg 1979; Voipio 1981; Ojaveer & Pihu 2003). The factors discussed above have a strong influence on fish community composition and abundances of different species in different areas of the Baltic Sea. Littoral habitats have often been considered important reproduction, nursery, and feeding areas for many fish species (e.g. Thorman & Wiederholm 1986; Vetemaa *et al.* 2006; Rohtla 2015). The shallow coastal zone is also inhabited by many small-sized fish species that to date have received minimal research attention (e.g. Sundell 1994; Rajasilta *et al.* 1999).

The overall economic importance (*sensu* landings biomass and revenue) of coastal fisheries in the Baltic Sea is small compared to pelagic fisheries regarding Baltic Sea herring (*Clupea harengus*), European sprat (*Sprattus sprattus*), and cod (*Gadus morhua*). However, littoral fish are important components in food chains of the whole Baltic ecosystem, transferring energy to upper trophic levels (e.g. Timola 1980; Lundström *et al.* 2007; Ustups *et al.* 2007; Mustamäki *et al.* 2014; Morkūnė *et al.* 2016). Some species, such as European perch (*Perca fluviatilis*) (e.g. Saat & Eschbaum 2002), European

smelt (*Osmerus eperlanus*) (e.g. Shpilev *et al.* 2005; Popov 2006; Shvagzhdy 2009), and European flounder (*Platichthys flesus*) (e.g. Ojaveer & Drevs 2003), are also important and highly valued species to local small-scale coastal fisheries. Due to the temporally highly variable biotic and abiotic conditions in the littoral areas, fish species composition and abundance may show high seasonal and inter-annual variation (e.g. Sundell 1994; Rajasilta *et al.* 1999; Vetemaa *et al.* 2006). Moreover, it has been recently shown that the seasonal locality-specific composition of the fish assemblage in shallow coastal areas changes significantly from early to late summer (Mustamäki *et al.* 2015).

Although littoral fish communities vary significantly in both space and time, there have been relatively few ichthyological studies carried out in the littoral zone compared to the amount of research concentrated on fish in other habitats of the Baltic Sea. Mustamäki (2015) stresses that fish monitoring interests are usually centred on long-term trends and the issue of seasonal changes is often circumvented by conducting such sampling at the same time each year. For this reason, descriptions of seasonal variation in fish communities of the Baltic Sea are scarce (reviewed by Mustamäki 2015). However, inter-annual, seasonal, and spatial dynamics of near-shore fish communities have still been relatively well studied (e.g. Vetemaa *et al.* 2006; Vahteri *et al.* 2009; Mustamäki *et al.* 2015). Nonetheless, there is only one study where diel variations in abundance of selected fish species inhabiting the shallow littoral zone of the Baltic Sea have been described (Thorman & Wiederholm 1986). Community level diel movements of littoral fish species in the Baltic Sea have not been described at all.

The behaviour of fish in the littoral zone can be strongly influenced by local habitat constraints, even when comparisons are performed between similar systems (Vasconcellos *et al.* 2011). Thus, it is possible that various coastal fish may show more diverse life history patterns (e.g. Landergren 2001; Limburg *et al.* 2001; Rohtla *et al.* 2012; Rohtla *et al.* 2014; Svärdsden *et al.* 2016) owing to adaption to the brackish conditions of the Baltic Sea, compared to conspecifics in strictly freshwater or more saline environments. It is therefore reasonable to speculate (in the context of the salinity gradient) that some coastal fish species inhabiting the Baltic Sea might even show sub-basin specific complex life history patterns. Therefore, new insight is needed into the functioning of littoral habitats as reproduction, nursery, and feeding areas, as well as local habitat spacing on short-term regular movement patterns of different fish species. Comprehensive evaluation of the ecological role of littoral habitats to mobile organisms such as fish should also include possible interactions with adjacent habitats (e.g. streams and the open sea).

Considering the gaps in our knowledge, my dissertation focuses on the dynamics of near-shore fish assemblages and alternative life-history patterns of fish species inhabiting the littoral zone in the north-eastern part of Baltic Sea. In particular, I address the topics of diel-scale variation in assemblage composition (**I**), seasonal foraging (**II**), and alternative nursery habitat use (**III, IV, V**), of anadromous fish in the littoral zone. All these factors are important in determining fish movements and elucidate a complex description of highly

dynamic Baltic Sea littoral ecosystems. The results of these studies can be applied in decision making related to the management and restoration of fish populations.

Temporal variations in surf-zone fish assemblage composition of the non-tidal Baltic Sea, with an emphasis on diel changes, were evaluated in Eru bay (**I**). In addition to diel behaviour, seasonal changes in fish assemblage composition are presented. To my knowledge, community level diel changes of the surf-zone fish assemblage have not been described in the Baltic Sea. Furthermore, it has been pointed out (Mustamäki 2015) that there is a trend towards community-level studies, along with recent developments in multivariate analysis methods. The need of such knowledge is furthered regarding ecosystem based management, which focuses on the system, rather than on a single species (reviewed by Mustamäki 2015). Understanding diel and seasonal variations of the littoral fish assemblage is also important to future studies where the aim is to evaluate the littoral fish assemblages of the Baltic Sea.

In this thesis, I describe the seasonal diet composition and feeding activity of the anadromous coastal fish species European smelt (*Osmerus eperlanus*) in a brackish littoral ecosystem (**II**). Shallow littoral areas are known as important feeding grounds to many fish species in the Baltic Sea (e.g. Thorman & Wiederholm 1986; Ustups *et al.* 2007; Mustamäki *et al.* 2014). Thus, temporal variations in fish assemblage composition in littoral areas are often related to feeding migrations (reviewed e.g. by Ross *et al.* 1987; Wilber *et al.* 2003). The knowledge of what and how much is consumed by fishes is central to understand food chains and webs, and the indirect importance of non-commercial marine biota to fisheries (Elliott *et al.* 2002). Hence, such studies may be essential to evaluate the ecological role of such species, as well as understanding the food web structure of the shallow littoral zone of the Baltic Sea. Moreover, diet composition and seasonal feeding patterns of certain species helps to understand the background mechanisms shaping diel and seasonal dynamics of the local littoral fish assemblage (**I**).

Unconventional types of juvenile salmonid migrations between streams and brackish littoral area of the Baltic Sea are described in the present thesis (**III**, **IV**, **V**). Those articles are in line with studies that document increasing complexity in the migration patterns of anadromous fish between freshwater and brackish littoral areas in the Baltic Sea (e.g. Limburg *et al.* 2001; Rohtla *et al.* 2012; Rohtla *et al.* 2014; Svirgsden *et al.* 2016). Regular movements of migratory fish species with relatively high commercial and recreational value, such as Atlantic salmon and brown trout, have been the subject of many studies (e.g. Klemetsen *et al.* 2003; Jonsson & Jonsson 2011). However, descriptions of alternative migrations of juvenile salmonids between freshwater and brackish environments are relatively scarce (e.g. Limburg *et al.* 2001; Ibbotson *et al.* 2013; Winter *et al.* 2016). Due to a lack of information, management decisions generally do not incorporate littoral habitats as provisional or permanent nursery, and wintering areas, for Atlantic salmon and sea trout parr and fry.

Thus, the results from studies **III**, **IV**, and **V** can be applied to future decision making related to the management and restoration of fish populations.

To conclude, the aims of this thesis were:

- to assess diel variations in the composition of the fish assemblage in the surf-zone of the non-tidal Baltic Sea (**I**).
- to estimate the predatory role of European smelt in a littoral habitat by describing seasonal variation of prey composition (**II**).
- to investigate whether littoral habitats of the Baltic Sea also function as a habitat for early out-migrating Atlantic salmon and ABT fry and parr (**III**, **IV**, **V**).

## 2. STUDY SYSTEM

### 2.1. The study area

The Baltic Sea is the second largest brackish waterbody in the world. Its main basin has very limited water exchange with the North Sea *via* the narrow and shallow Danish straits, and is characterized by a significant freshwater surplus due to large river runoffs, amounting to about 2% of the volume of the sea per annum (Myrberg & Andrejev 2003; reviewed by Ojaveer & Pihu 2003). Despite the relatively small surface area, the topography of the Baltic Sea is quite complex (*e.g.* Fig. 1), resulting in specific hydrographic characteristics in different sub-basins (Myrberg & Andrejev 2003; reviewed by Ojaveer & Pihu 2003). Moreover, crucial environmental factors (*e.g.* salinity, temperature, trophic status, effect of the photoperiod) also have considerable gradients from different littoral areas (*e.g.* the open coastal zone, estuaries, and bays) to the open sea (Neumann 1977; Thorman & Wiederholm 1983; Uusitalo *et al.* 2005). Due to variability in abiotic and biotic environmental parameters, near-shore fish communities of the Baltic Sea are highly variable. For instance, the occurrence and abundance of species may differ spatially and temporally over seasonal and diel scales (*e.g.* Thorman 1986a, b; Thorman & Wiederholm 1986; Sundell 1994; Vetemaa *et al.* 2006; Mustamäki *et al.* 2015), as near-shore littoral areas are known to constitute important spawning, nursery, and feeding areas (*e.g.* Sundell 1994; Ustups *et al.* 2003; Vetemaa *et al.* 2006; Rohtla 2015).

This thesis focuses on the north-eastern Baltic Sea as the study area (Fig. 1). Specifically, the field studies were conducted in the Gulf of Finland basin (Eru Bay (**I**, **II**, **III**) and the Toolse River (**V**)), Baltic Proper basin (Ermespu Stream (**IV**))), and Gulf of Riga basin (Loode and Lemmejõgi streams (**V**))). Eru Bay was used as a model area of a brackish ecosystem to describe: (1) temporal variations in the fish assemblage composition of the surf-zone, with an emphasis on diel changes of the relative proportions of different species by photoperiod (**I**); (2) seasonal diet composition and feeding activity of a small, but relatively abundant predator in the near-shore habitat, using European smelt as a model species (**II**); (3) alternative life-histories of Atlantic salmon and ABT parr in a littoral habitat (**III**). The Baltic Proper basin and Ermespu Stream were used to document whether juvenile anadromous brown trout descend to the brackish waters of the Baltic Sea from their natal streams as parr or fry, and then migrate into non-natal streams (SSME), before transforming into smolt (**IV**). The Gulf of Finland basin, Toolse River, and Loode and Lemmejõgi streams in the Gulf of Riga basin were used as models to describe the descent of Atlantic salmon fry from their natal streams to the brackish waters of the Baltic Sea, and their use of this environment as an alternative rearing habitat, before ascending back to freshwater streams (**V**)).

Eru Bay (**I**, **II**, **III**) is located in the centre of the southern part of the Gulf of Finland (Fig. 1). The Gulf of Finland is an elongated sub-basin at the north-

eastern extremity of the Baltic Sea, with a surface area of 29,571 km<sup>2</sup> and mean water depth of approximately 37 m. Salinity of the Gulf of Finland increases from east to west, and north to south. Surface salinity varies from 5–7 psu in the West, to 0–3 psu in the East (Alenius *et al.* 1998). Ice cover is a regular winter phenomenon in the Gulf of Finland (Alenius *et al.* 1998). The average number of ice days varies from 40 at the western end of the Gulf of Finland to 130 in some coastal bays in the north-eastern part of the Gulf of Finland (Seinä & Peltola 1991). The area is characterized by a strong seasonal thermocline at depths of 10–20 m (reviewed by Alenius *et al.* 1998), as well as by occasional coastal upwelling events (Myrberg & Andrejev 2003).

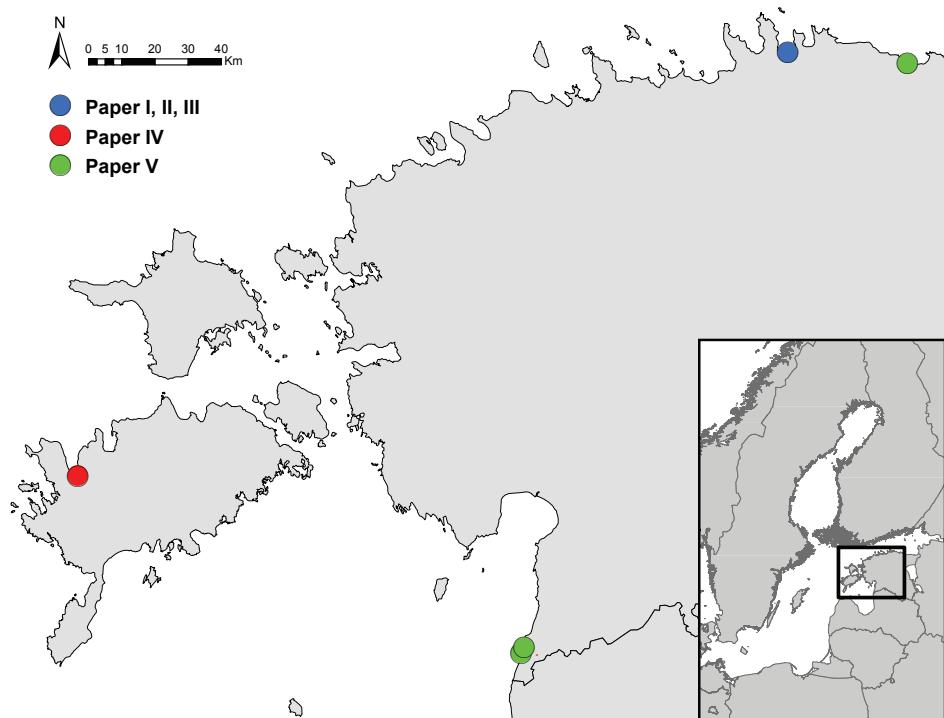
Although the Loobu River discharges into Eru Bay, the influence of freshwater on salinity levels in the area is very low (Martin *et al.* 2003). Surface water salinity of Eru Bay is usually 4.5–6.5 psu, which is typical for the central part of the Gulf of Finland. In the outer part of the bay, the bottom slope is relatively steep, with a depth of 10 m reached at 400–500 m from the shoreline (Martin *et al.* 2003), whereas the inner part of Eru Bay is characterized by a relatively gentle slope (a depth of 10 m is usually reached at >1000 m distance from the shoreline).

The local fish fauna comprises both marine and freshwater species. Coastal fisheries are mostly directed at salmonids and whitefish (Saat *et al.* 2003), as there are several important salmonid rivers in Northern Estonia (Kangur *et al.* 2003a, b). The coastline of the southern part of Eru Bay was suitable as a model area to study the dynamics of littoral fish assemblages because: (1) location, the area is sheltered from prevailing western winds and is only significantly influenced by winds from northern directions; (2) the gentle slope and flat bottom are suitable to use beach seining effectively (Lappalainen & Urho 2006); (3) human impact in this area is very low (Martin *et al.* 2003).

The Ermespu Stream (**IV**) is situated on Saaremaa Island and drains into Tagalaht Bay, which is part of the Baltic Proper (Fig. 1), where surface water salinity ranges between 5–7 ‰ (Martin *et al.* 2003). The influence of freshwater in this area is very low, as Tagalaht Bay receives marginal freshwater inflow from a few small streams (Järvekülg 2001). The Ermespu Stream is an appropriate system to document the SSME of anadromous brown trout, because: (1) no suitable spawning habitat is available to brown trout (*sensu* Ottaway *et al.* 1981; Louhi *et al.* 2008; Jonsson & Jonsson 2011), as the substrate consists entirely of fine sediment (Järvekülg *et al.* 2012); (2) of a neighbouring stream (Pidula Stream) with high densities of brown trout juveniles (Kesler *et al.* 2015).

The Toolse River (**V**) discharges into Kunda Bay (Gulf of Finland), where surface water salinity is usually < 5 ‰ (Järvekülg 1979). The Loode and Lemmejõgi streams (**V**) flow into the Gulf of Riga, where surface water salinity usually ranges between 4.5–6.0 ‰ (Leppäranta & Myrberg 2009). However, the high number of inflowing streams, and the relative isolation from the open Baltic Sea, may produce considerable temporary changes to the salinity gradient. Thus, salinity in the coastal zone of the Gulf of Riga can be variable

during periods of high discharge, and occasionally fluctuate only between 0.5–2.0 psu (reviewed by Ojaveer 1997). Those streams are suitable study systems to demonstrate SSME of Atlantic salmon, because the Lemmejõgi Stream (catchment area 55.7 km<sup>2</sup>), Loode Stream (catchment area 18.5 km<sup>2</sup>), and Toolse River (catchment area 84.3 km<sup>2</sup>), are not classified as historical or potential Atlantic salmon rivers (HELCOM 2011) due to their relatively small flow volume and size (Jonsson *et al.* 1991).



**Figure 1.** Study region and sampling sites of the present thesis.

## 2.2. Field sampling and methods

The fish samples from Eru Bay were collected with a beach (hand) seine (**I**, **II**, **III**) or gillnets (**II**, **III**). Detailed descriptions of the methods (*e.g.* beach seine dimensions, gillnet mesh sizes, weather conditions, fish sampling and processing, and otolith preparation and analysis) can be found in each specific publication (**I**, **II**, **III**).

Diel and seasonal changes in fish species composition and densities (**I**) in Eru Bay were assessed monthly during the ice-free period (April–December) in 2008. The beach seine was hauled perpendicular to the shore, covering a mean area of 964 m<sup>2</sup> per haul. The shallow ( $\leq 1$  m) littoral area of the sandy beach was divided into six seining stations. The sample sets were collected at dawn

(starting 30 minutes before sunrise), noon (starting 30 minutes before astronomical noon), dusk (starting 30 minutes before sunset), and midnight (starting 30 minutes before astronomical midnight). Fish were identified to the lowest possible taxonomic level and total length (TL) of each measured to the nearest 1 mm. Total biomass of each taxa was measured to the nearest 0.1 g.

Seasonal diet composition and feeding activity of adult and sub-adult European smelt in Eru Bay was studied by examining the stomach contents of 1397 individuals collected over the months of April to June and August to December 2009. Each prey item was identified to the lowest possible taxonomic level and counted (**II**).

Autumn migrant ABT and Atlantic salmon parr were collected in 2006, 2008, 2009, and 2013 from the shallow littoral zone of the Gulf of Finland. Age and migration pattern of ABT and Atlantic salmon were determined from sagittal otoliths (**III**).

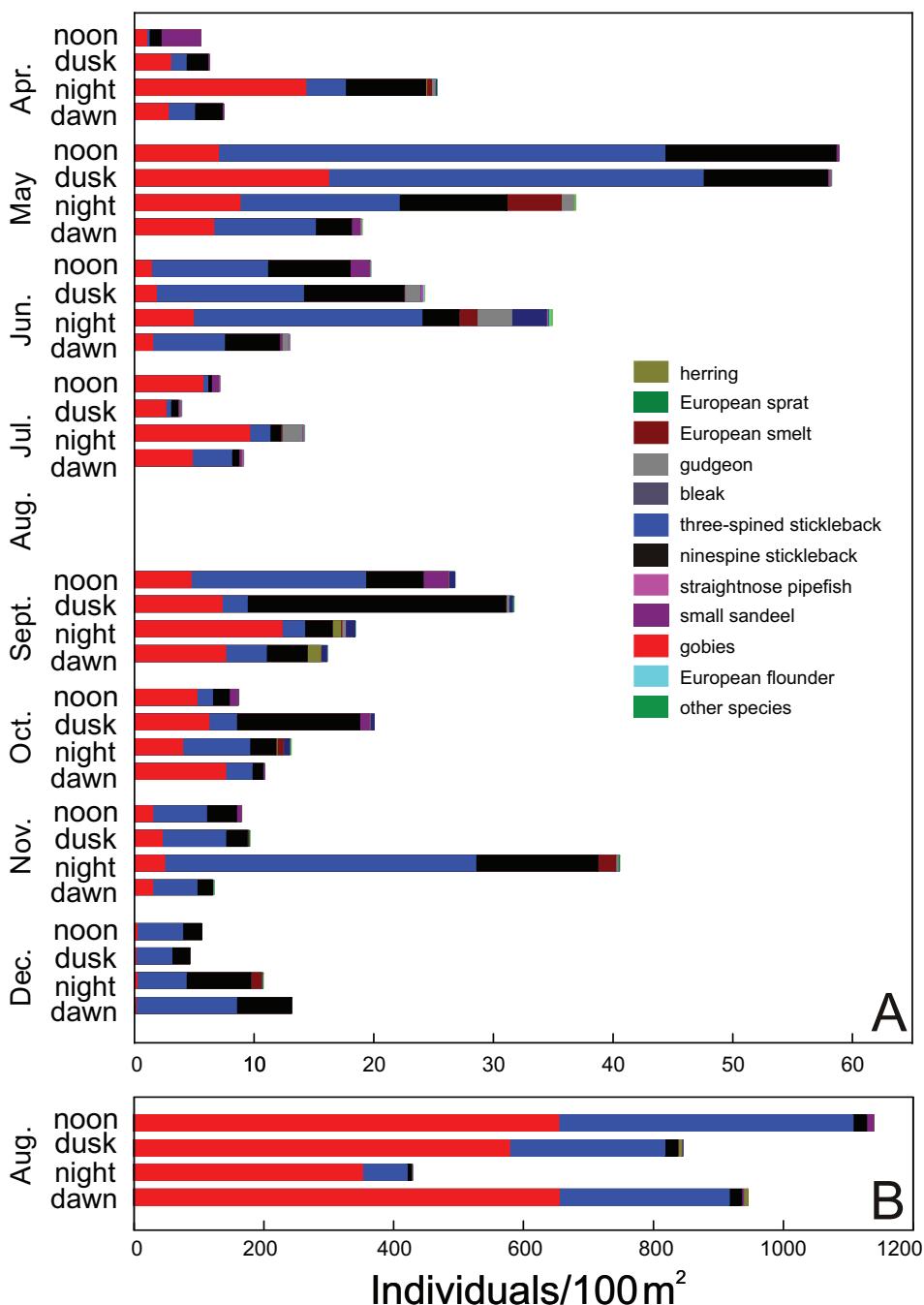
Early out-migrating ABT (**IV**) and Atlantic salmon parr (**V**) were electro-fished in the Ermespu Stream (**IV**), Lemmejõgi Stream, Loode Stream, and Toolse River (**V**). Age and use of alternative rearing habitats were determined from sagittal otoliths (**IV, V**). Detailed descriptions of these methods (*e.g.* fish sampling and processing, and otolith preparation and analysis) can be found in each specific publication (**IV, V**).

### **3. RESULTS AND DISCUSSION**

#### **3.1. Diel changes in the fish assemblage of a sandy surf-zone area of the Baltic Sea**

The composition of the fish assemblage in the surf-zone of Eru Bay, Gulf of Finland, was highly variable (Fig. 2) in relation to the time of day (dawn, noon, dusk, and midnight) during the ice-free period of 2008. Diel variation in fish assemblage composition of the surf-zone was also linked with seasonal changes (**I**). To my knowledge, this is the first documentation of diel variations in the fish assemblage of the surf-zone of the Baltic Sea, using a data set covering almost the entire year (the whole ice-free season). The surf-zone fish community consisted of 24 species and was dominated by three-spined stickleback (*Gasterosteus aculeatus*), ninespine stickleback (*Pungitius pungitius*), sand goby (*Pomatoschistus minutus*), and common goby (*Pomatoschistus microps*) (**I**). Species that exhibited the most variation during the diel cycle were three-spined stickleback, ninespine stickleback, gobies (*Pomatoschistus* spp.), European smelt, small sandeel (*Ammodytes tobianus*), gudgeon (*Gobio gobio*), and bleak (*Alburnus alburnus*). Some species (e.g. European smelt, gudgeon, bleak, small sandeel, ninespine-stickleback) showed distinguishable diel movement patterns throughout the study period, whereas others (e.g. three-spined stickleback, gobies) did not (Fig. 2). However, in the case of three-spined stickleback and gobies, high YOY abundance during August and September could have hampered finding overall distinguishable diel patterns, as the behaviour of juveniles can differ from that of adults (e.g. Thorman & Wiederholm 1983; Ustups *et al.* 2007; Mustamäki *et al.* 2015).

One of the main results of the present thesis was that the most conspicuous diel changes at fish assemblage level were induced by the presence of sub-adults and adults of relatively large species (e.g. European smelt, gudgeon, bleak, and small sandeel), which tended to have diel movement patterns, and only utilised the surf-zone during certain times of day (Fig. 2). Furthermore, combined results (**I**, **II**) hint that European smelt might even exhibit sub-basin specific diel foraging migration, probably induced by local abiotic and biotic environmental constraints. Midnight samples contained more species compared to those collected at dawn, noon, or dusk. However, not all species showed an increase in abundance at midnight. This is not in accordance with the single previous study of diel variations of the abundance of some surf-zone fish species in the Baltic Sea by Thorman & Wiederholm (1986). These authors concluded that all the fish species studied were more abundant at night than during daytime. Differences between the results of Thorman & Wiederholm (1986) and this thesis may have partly been caused by methodological differences, as well as habitat-specific and regional differences in the fish fauna. Possible sub-basin specific diel behavioural patterns of fish species also cannot be ruled out.



**Figure 2.** Mean densities of fish at different times of day during (A) April–July and September–December and (B) August in 2008. Note the scale difference between August and the other months. The common goby and sand goby were pooled as a species group termed “gobies” (Paper I, Figure 2).

This study complements the findings of Mustamäki *et al.* (2015), who showed that the seasonal composition of the fish assemblage in a similar shallow coastal area in the Baltic Sea changed significantly from early to late summer. In addition to supporting the findings of Mustamäki *et al.* (2015), the demonstrated diel migrations of fish between adjacent habitats (**I**) show that spatial changes in Baltic Sea littoral fish communities may take place over an even shorter period than described before. Specifically, the absence of some species during a particular time of day (*e.g.* European smelt, gudgeon, small sandeel), further stresses the effect of diel migrations on the composition of the fish assemblage of this ecosystem.

The results of this study also showed relatively low densities of surf-zone fish in July (compared to May, June, August, and September (Fig. 2)), which is in accordance with the mid-summer decline in fish abundance in the Baltic Sea (Sundell 1994) and the Kattegat (Thorman 1986a). The observed decline in fish abundances during July could be caused by several factors. Adult sticklebacks and gobies either die after spawning or move to deeper areas during mid-summer (*e.g.* Sundell 1994). The beach seine used may not have been efficient at catching small YOY specimens, which were present in the surf-zone. It is also possible that warmer water in the surf-zone during mid-summer may not be optimal to some species during most of the diel period, as in deeper, cooler waters, utilisation of energy from consumed food is more efficient (discussed by Thorman & Wiederholm 1986). Hence, the results of this present thesis and those of previous studies (Thorman 1986a; Sundell 1994), indicate that mid-summer sampling of surf-zone fish in the Baltic Sea may result in skewed results with regard to the composition of the fish assemblage if – due to the sampling method used – smaller species are underrepresented.

The observed diel variations in the composition of the fish assemblage of the surf-zone indicate the importance of considering the time of the day in sampling designs. The results of the present study demonstrate that the composition of a fish assemblage observed at a certain time of day cannot be directly extrapolated to another time of day, or regarded as representative of the whole astronomical day. To fully describe the fish assemblage of the surf-zone of the Baltic Sea, samples covering several different times of day (dawn, noon, dusk, and midnight) should be obtained. For example, estimates of the significance of this habitat to particular species could be very misleading if sampling is limited to only a specific diel period.

Diel migration of fish assemblages in the various littoral habitats of the Baltic Sea is not comprehensively known, as the present results (**I**) describe only the easily accessible sandy surf-zone. The sheltered location and flat bottom habitat without physical obstructions, which is highly suitable to beach sein (reviewed by Lappalainen & Urho 2006), made our study site (**I**) an ideal model area to describe diel and seasonal changes in surf-zone fish assemblages of the Baltic Sea. Furthermore, the wide range of littoral habitats in the Baltic Basin, as well as possible sub-basin dependent diel habitat utilization (**I, II**) and alternative life history patterns of fish (*e.g.* Landergren & Vallin 1998;

Landergren 2001, 2004; Limburg *et al.* 2001; Rohtla *et al.* 2012, 2014; **III**, **IV**, **V**) in the Baltic Sea makes this a compelling topic for further ecological studies.

### **3.2. Diet composition of European smelt in a near-shore ecosystem**

Prey composition and feeding activity of European smelt varied monthly and depended on the size of the individual in Eru Bay (Table 1). Gammarid amphipods (*Gammarus* spp., *Corophium volutator*) and mysids (*Mysis* spp.) were the most common prey, followed by isopods (*Idotea* spp., *Saduria entomon*), cladocerans (*Cercopagis pengoi*) decapods (*Palaemon adspersus*) and fish (e.g. three-spined stickleback, nine-spined stickleback, sand goby, common goby). Smelt also preyed also on molluscs, insects, insect larvae, and polychaete annelids. This study also revealed some previously unrecorded prey species of European smelt, e.g. bullhead (*Cottus gobio*), small sandeel, bay mussel (*Mytilus trossulus*), Baltic macoma (*Macoma baltica*), and ragworm (*Hediste diversicolor*) (see Nedolgová 2010 for details).

Whether adult smelt are a significant predator of larvae and young fish, has for a long time been the subject of discussion (reviewed by Nellbring 1989). Results of the present thesis (**II**) suggest that sub-adult and adult smelt are indeed an important predator in shallow littoral areas of the eastern Baltic Sea, as seven fish species were recorded in stomach contents. Even the smaller length classes of sub-adult and adult smelt tended to feed on fish, mostly sticklebacks and gobies.

The results of the present study suggest that European smelt inhabiting littoral areas of the Baltic Sea occupy a different ecological niche compared to smelt in offshore areas (e.g. Kostrichkina 1968, 1974; Shestakov 1970; Popov 1978, 2006; Urtans 1990; Ojaveer *et al.* 1997), because fish and benthic prey tend to be more important regarding the diet of coastal inhabiting smelt (**II**). Moreover, it was shown (**I**) that European smelt exhibited a distinct migration pattern to the surf-zone of Eru Bay, utilizing the habitat mostly, and during some months exclusively, at night (Fig. 2). This data conflicts with earlier studies conducted in the surf-zone of the Baltic Proper (Ustups *et al.* 2003, 2007), which indicated that adult European smelt were abundant at depths < 2m only during spawning migration in spring. This controversy can be partly explained by foraging migration induced by local environmental characteristics (**I**, **II**), and by differences in the sampling methodology (**II**). In Ustups *et al.* (2003, 2007), samples were taken in May, July, and September during daylight.

**Table 1.** Frequency of empty stomachs (%FES), and occurrence (%FO, abbreviated to %O in the table) and numerical (%N) indices of prey in European smelt stomachs ( $L_s$ , size class; N, number of analysed smelt; H, div, *Hediste diversicolor*; G, *Gastropoda*; S, ent, *Saduria entomon*; Gam, *Gammaridae*; C, vol, *Coryphium volutator*). An empty stomach was defined as the absence of any measurable organic material (Arrington *et al.* 2002). The percentages of frequency of occurrence (%F.O.) and numbers (%N) were calculated for each prey category according to the following equations (Hyslop 1980): %F.O. =  $100 \times F_i/n$ , where  $F_i$  is the number of stomachs containing a particular prey taxon; n is the total number of stomachs with any prey taxon; %N =  $100 \times N_i/N$ , where  $N_i$  is the total number of prey of a particular taxon; N is the total number of all prey identified (Paper II, Table 1).

Diel variation in the fish assemblages of shallow coastal areas are often related to feeding migrations (e.g. Thorman & Wiederholm 1986; Castillo-Rivera *et al.* 2010; Vasconcellos *et al.* 2011). Significant temporal variation in near-shore fish assemblages (**I**) could thus also be associated with the feeding behaviour of predatory fish in these habitats. Overall, based on current knowledge of European smelt diet (e.g. Popov 2006; Ustups *et al.* 2007; **II**), it can be speculated that European smelt may be an important predatory fish species that links shallow coastal areas to the upper trophic levels and deeper areas of the Baltic Sea. European smelt are an important part of the diet of grey seals *Halichoerus grypus* (e.g. Lundström *et al.* 2007) and various piscivorous birds, such as common guillemot (*Uria aalge*), red-throated diver (*Gavia stellata*), and great crested grebe (*Podiceps cristatus*) (e.g. Morkūnė *et al.* 2016) in the Baltic basin. Moreover, Morkūnė *et al.* (2016) pointed out that the fatty acid composition of anadromous smelt from the coastal zone is isotopically unique within the fish community. Morkūnė *et al.* (2016) hypothesized that this phenomenon is probably caused by an interaction of anadromous migration patterns, predatory and cannibalistic feeding, starvation during the spawning period, and allocation of nutrients into the reproductive tissues during winter. According to coastal fish monitoring data (Eschbaum *et al.* 2015), the European smelt is the most common predatory fish after Eurasian perch in near-shore areas of the southern Gulf of Finland. Therefore, it seems that at least in the case of the Eastern Gulf of Finland, the impact of European smelt on the food web of the surf-zone *sensu stricto* (**I**, **II**), as well as the coastal littoral zone *sensu lato* (**III**), has been underestimated.

### **3.3. The littoral zone as a habitat of early out-migrating Atlantic salmon and anadromous brown trout juveniles in the Baltic Sea**

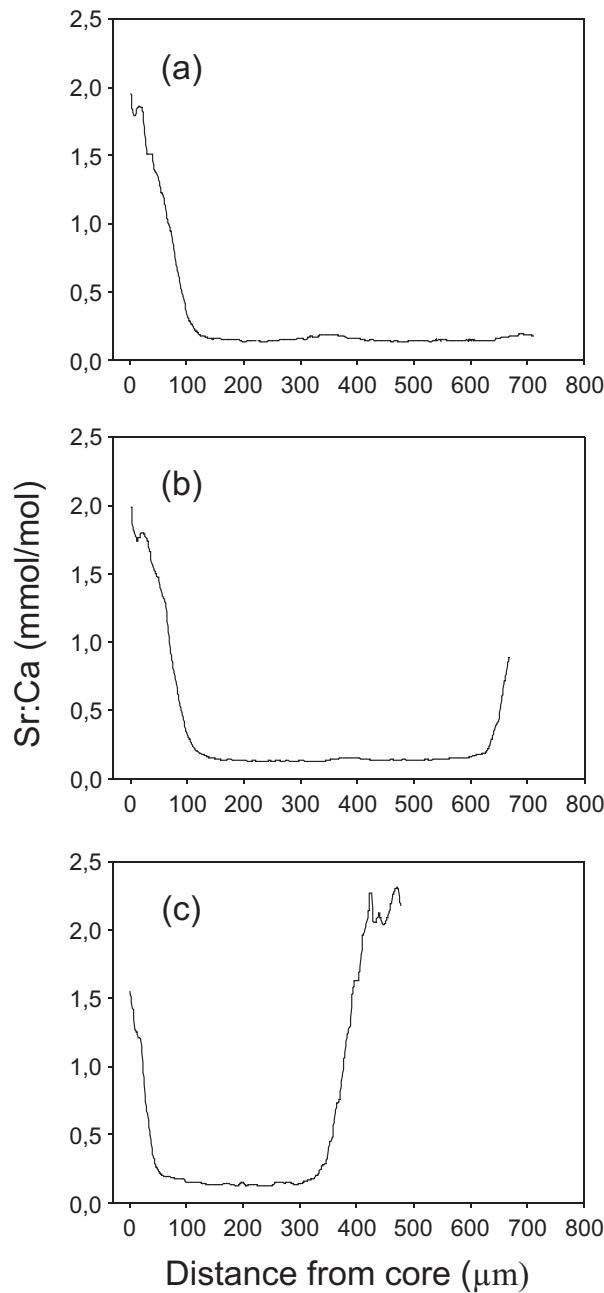
This study presents the first records of autumn descent of Atlantic salmon and ABT to the brackish littoral habitats of the Baltic Sea Basin (Figure 3; see also Table 1 in **III**). The prevalence of this behaviour in these species is still unknown in the Baltic Sea. However, our relatively wide temporal range of observations (2006, 2008, 2009, and 2013) indicates that autumn descent of anadromous salmonid parr is not a random event (**III**). As a novel contribution to science, stream shifting through the marine environment (SSME) of early out-migrating fry and parr of ABT (Figure 4; see also Table 1 in paper **IV**) and Atlantic salmon (Figure 5; see also Table 1 in paper **V**) is presented in the current study for the first time. Moreover, residency in a saline environment has also not been previously demonstrated in the case of Atlantic salmon fry (Figure 5; see also Table 1 in paper **V**).

Shallow littoral habitats are important feeding areas for juvenile ABT (Klemetsen *et al.* 2003; Jonsson & Jonsson 2011), whereas Atlantic salmon

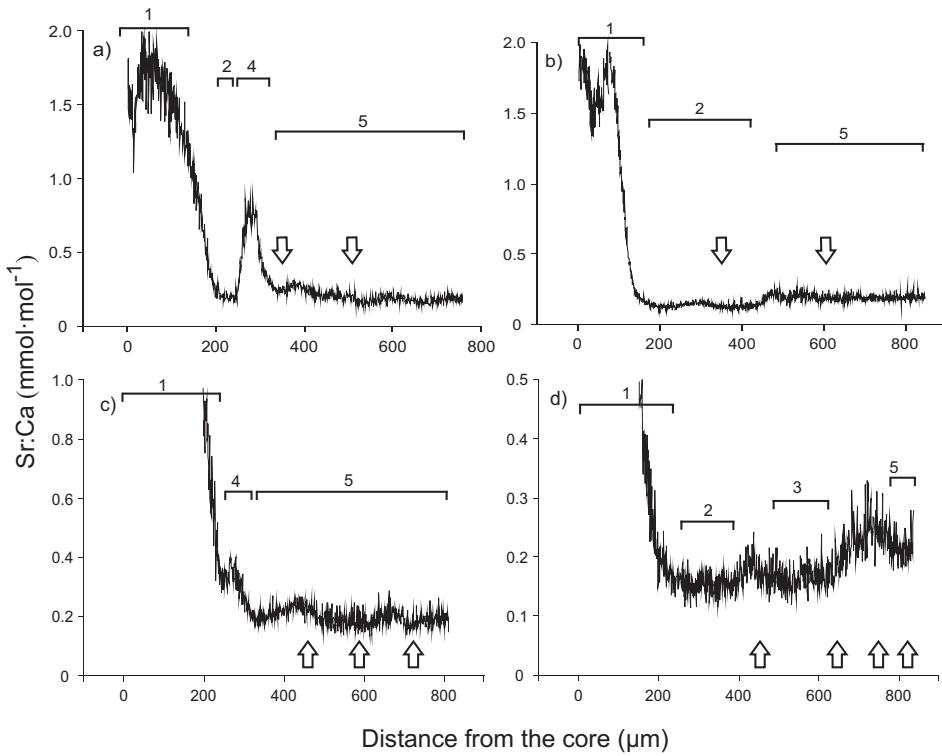
smolt usually inhabit littoral areas during a brief period at the beginning of their sea sojourn, before moving into the open sea (reviewed by Jonsson & Jonsson 2011). During smolting, salmonid parr undergo extensive physiological and morphological transformations to adapt to the saline environment (e.g. Klemetsen *et al.* 2003; Jonsson & Jonsson 2011). Juvenile salmon and ABT are traditionally considered to stay in their native stream from at least one to several years as parr, prior to their descent to the sea as smolts in spring. However, salmonids are known to have diverse life histories, which differ in the duration of freshwater and or marine rearing, age at maturation, and incidence of repeat spawning (e.g. Klemetsen *et al.* 2003; Koski 2009; Jonsson & Jonsson 2011; Roni *et al.* 2012; Bond *et al.* 2014; Shrimpton *et al.* 2014; Walsworth *et al.* 2015). It is therefore not surprising that seaward movements of Atlantic salmon and ABT have also been documented during times of the year not associated with the spring parr-smolt transformation (e.g. Cunjak *et al.* 1989; Järvi *et al.* 1996; Limburg *et al.* 2001; Winter *et al.* 2016).

Autumn descent of Atlantic salmon parr has been observed in Canada (Cunjak *et al.* 1989), Great Britain (e.g. Youngson *et al.* 1983; Riley *et al.* 2002; Pinder *et al.* 2007; Ibbotson *et al.* 2013), and Norway (Jensen *et al.* 2012). This behaviour has been recorded in the case of ABT populations in Ireland, Norway (Jonsson & Jonsson 2009), New Zealand (Holmes *et al.* 2014), Great Britain, and Denmark (Winter *et al.* 2016). However, besides autumn descent, ABT can follow other alternative migration patterns in coastal areas of the Baltic Sea. It has also been reported that fry and parr of ABT may descend to the sea soon after hatching in spring and early summer (Järvi *et al.* 1996; Limburg *et al.*, 2001; Landergren 2004), and use the littoral as an alternative nursery habitat.

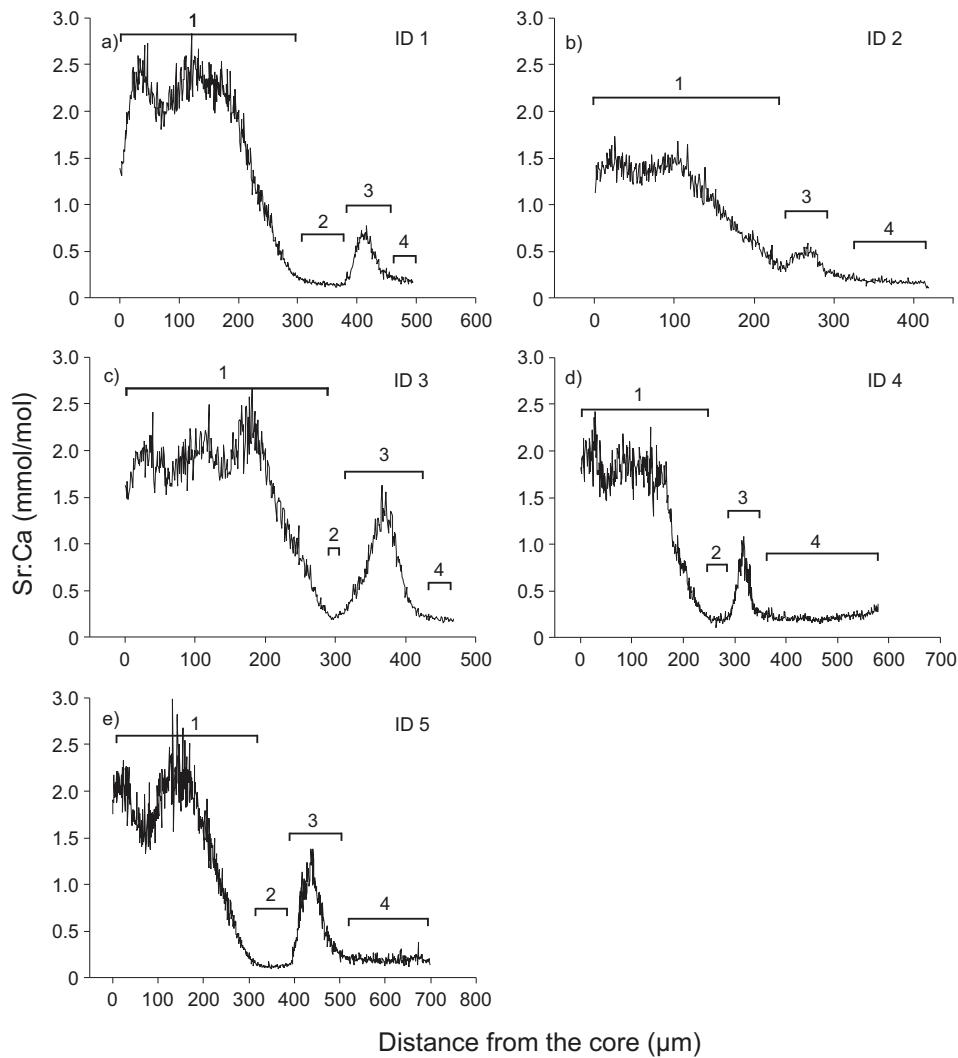
The exact ecological and physiological mechanisms behind early out-migration and SSME of Atlantic salmon and ABT fry and parr (**III**, **IV**, **V**), are still unknown. The autumn descent of Atlantic salmon parr has been associated with high volume water flows (e.g. Youngson *et al.* 1983). Ibbotson *et al.* (2013) reviewed a number of potential mechanisms that could trigger autumn migration of Atlantic salmon. The causes behind autumn migration could include intrinsic factors such as size and position of the individual in the dominance hierarchy, and extrinsic factors associated with density and habitat, related to cover (shelter) availability, supply of food, and the presence of predators (Ibbotson *et al.* 2013). The phenomenon of early (spring-summer) out-migrating ABT fry from small Gotland streams (with high number of spawners and decreasing water levels during spring and summer) may be due to competition (e.g. for food, space) and or the result of variable hydrological conditions (e.g. decreasing water levels during dry periods in summer and early autumn), and small distance of spawning grounds from the sea (e.g. Limburg *et al.* 2001; Landergren 2004). Observed springtime out-migration of YOY Atlantic salmon in this study (**V**) hint that fry or young parr may also disperse downstream from their natal habitat during high volume water flows. Such an effect of high flows could be further supported by the spawning site preferences of Atlantic salmon (Jonsson *et al.* 1991).



**Figure 3.** Examples of Sr:Ca profiles. (a) Anadromous brown trout collected from the sea on the 14<sup>th</sup> November 2009, with no seawater signal; (b) salmon collected from the sea on the 28<sup>th</sup> December 2013, with a moderate seawater signal; (c) salmon collected from the sea on the 7<sup>th</sup> December 2006, with a stabilized seawater signal. The peaks at the beginning of the Sr:Ca profiles represent the maternally derived seawater signal (Kalish 1990) (Paper III, Figure 2).



**Figure 4.** Representative otolith profiles (Sr:Ca) of brown trout from the Ermespu Stream: (a) parr (ID 9) with a clear Sr:Ca marine signal (note that the pre-SSME natal freshwater Sr:Ca value is not stabilized); (b) parr (ID 15) with a clear shift in Sr:Ca values; (c) parr (ID 14) that descended/drifted to the sea immediately after yolk sac reabsorption or even during it, and subsequently entered the Ermespu Stream; (d) parr (ID 1) that possibly migrated between tributaries within its natal watershed, and subsequently migrated to the Ermespu Stream *via* the marine environment, as evidenced by the increased Sr:Ca values. Numbered brackets represent distinct stages in the chemical profiles: (1) maternally influenced region (*sensu* Kalish 1990); (2) natal freshwater habitat; (3) possible migration between tributaries in the natal watershed; (4) brackish water experience; and (5) non-natal freshwater habitat (the Ermespu Stream). The Sr:Ca freshwater threshold is  $< 0.5 \text{ mmol}\cdot\text{mol}^{-1}$  in the freshwaters of Estonia (Matetski 2014). Note the different scales of the Sr:Ca profiles. Arrows denote the annuli (Paper IV, Figure 2).



**Figure 5.** Otolith Sr:Ca profiles of Atlantic salmon (a, d, e) parr with distinct natal values, followed by a clear marine signal, and subsequent decrease and stabilization of values to levels different from natal ones; (b, c) parr that descended/drifted to the sea immediately after yolk sac resorption or even during it, and subsequently entered a non-natal freshwater habitat. Numbered brackets represent distinct stages in the chemical profiles: (1) maternally influenced region (*sensu* Kalish 1990); (2) natal freshwater habitat; (3) brackish water experience; and (4) non-natal freshwater habitat. The Sr:Ca freshwater threshold is  $< 0.5 \text{ mmol} \cdot \text{mol}^{-1}$  in Estonian freshwaters (Matetski 2014) (Paper V, Figure 2).

According to Jonsson *et al.* (1991), Atlantic salmon prefer to spawn in relatively large rivers in terms of water volume and catchment area, compared to e.g. ABT. All the alternative migrations of Atlantic salmon and ABT not associated with the spring parr-smolt transformation (e.g. Youngson *et al.* 1983; Limburg *et al.* 2001; Jonsson & Jonsson 2009; **III**, **IV**, **V**), parallel various potamodromous movements of salmonids from inlet or outlet streams (spawning and nursery grounds) into freshwater lakes or from tributaries into the main river (Klemetsen *et al.* 2003; Jonsson & Jonsson 2011). It is also known that juveniles of anadromous Dolly Varden trout (*Salvelinus malma*) may use the marine environment to reach non-natal streams in which they overwinter (e.g. Armstrong 1974; Bond *et al.* 2014).

The autumn descent of Atlantic salmon (e.g. Cunjak *et al.* 1989; Pinder *et al.* 2007) and ABT (Jonsson & Jonsson 2009) to estuarine habitats has been previously reported for the Atlantic Ocean. However, Riley *et al.* (2008) demonstrated that autumn descending Atlantic salmon parr are physiologically not sufficiently adapted to full-strength seawater (35 %), and thus their expected survival is very low. Nonetheless, a low proportion of Atlantic salmon and ABT autumn migrants are reported to return as adults during the spawning run (e.g. Jonsson & Jonsson 2009; Riley *et al.* 2009). However, in the case of ABT it has been experimentally demonstrated (Landergren 2001) that the low saline environment of the Baltic Sea (6–7 %) is not a physiological obstacle to the survival of early out-migrating parr. This may also be relevant for Atlantic salmon, as phylogeny and life history of this species is similar to brown trout (Klemetsen *et al.* 2003; Jonsson & Jonsson 2011). Hence, the results of **III**, **IV**, **V** and previous studies (Järvi *et al.* 1996; Limburg *et al.*, 2001; Landergren 2004), suggest that the shallow littoral areas of the Baltic Sea might act as alternative nursery areas to Atlantic salmon and ABT fry and parr.

In the Baltic Sea, investigation of the origin of adult sea-caught ABT from the Gotland area revealed that some fish showed no evidence of freshwater history, thereby raising the possibility that a proportion of this population may not depend on riverine spawning at all (Limburg *et al.* 2001). However, this hypothesis is contradicted by the conclusions of Landergren & Vallin (1998), who suggested that spawning grounds located at stream outlets could, due to the low salinity (<4 %), contribute to ABT recruitment. However, eggs deposited in such areas face the risk of being adversely effected by storms, thus making successful recruitment questionable (Landergren & Vallin 1998). The conclusions of Landergren & Vallin (1998) are also supported by the results of **IV** and **V**. In particular, I propose that Atlantic salmon (**IV**) and ABT (**V**) fry may emigrate from natal streams before environmental chemical signals are incorporated into the otolith. This hypothesis seems to be supported by an alternative explanation of the origin of brackish water born brown trout suggested by Limburg *et al.* (2001). These authors discussed whether the maternally-donated otolith Sr may obscure evidence of early residency of the egg in freshwater. In Estonia, sheltered stream outlets are mostly characterized by no suitable spawning habitat for salmonids. Stream outlets with suitable

spawning substrate (gravel), however, are exposed to the sea and ambient surface salinity probably does not stay below a 4 ‰ level throughout the incubation period. Thus, in the Estonian part of the Baltic Sea, spawning of brown trout (and Atlantic salmon) in brackish water is highly unlikely.

It is widely accepted that salmonids imprint their natal stream during smoltification, and return there to spawn as adults (“homing”) (e.g. Quinn 1993; Hansen & Jonsson 1994). Thus, ABT and Atlantic salmon parr that undergo SSME may become smolts in non-natal streams and thus fixate their homing to these new waterbodies. Furthermore, SSME (**IV**, **V**) could be a possible factor behind the adult straying behaviour documented during the spawning run of Atlantic salmon and ABT (Jonsson *et al.* 2003; Degerman *et al.* 2012). However, it is unclear whether these early out-migrating juveniles (permanently inhabiting brackish littoral areas, Limburg *et al.* 2001) return to their natal rivers as adults or become strayers. Similar, unresolved questions on early out-migrating ABT were raised by Landergren (2004). Understanding the mechanisms behind these phenomena may be important, as existing salinity values are mostly projected to drop further due to increases in rainfall over the upcoming decades (e.g. Kjellström & Ruosteenoja 2007; Meier *et al.* 2012). Thus, the issue of alternative life-histories of Atlantic salmon (**III**, **V**) and ABT (Landergren & Vallin 1998; Landergren 2001; Limburg *et al.* 2001; Landergren 2004; **III**, **IV**) in the brackish environment of the east Baltic Sea may become even more relevant in the future, especially in the context of population abundance assessment and conservation.

## 4. CONCLUSIONS

1. The fish assemblage inhabiting the surf-zone of the Baltic Sea can vary significantly on a diel and seasonal basis. Fine-scale assemblage level variations of littoral fish are more complex and may take place over an even shorter time frame than previously known. To accurately quantitatively describe the surf-zone fish community, samples covering different photoperiods should be obtained. This is important, because crucial aspects, such as the significance of the scrutable littoral habitat to certain species, could be missed when sampling only during a limited time-frame (**I**).
2. Even the smaller length classes of sub-adult and adult European smelt (*Osmerus eperlanus*) tend to feed on fish. Thus, smelt is an important predator in shallow littoral areas, being the second most common predatory fish in the near-shore areas of the southern Gulf of Finland. European smelt inhabiting littoral areas of the Baltic Sea occupy a different ecological niche than smelt in offshore areas (**II**). It is reasonable to speculate that at least in the case of the Eastern Gulf of Finland, the impact of European smelt is seriously underestimated in the food webs of the surf-zone *sensu stricto*, as well as in coastal littoral zone *sensu lato*.
3. Early descent, as well as stream shifting through the marine environment, of Atlantic salmon (*Salmo salar*) and anadromous brown trout (*Salmo trutta*) fry (**V**) and parr (**III**, **IV**), indicate that the shallow littoral zone of the Baltic Sea may play a significant role as a permanent or provisional nursery area to these species. Therefore, the possible life history traits of salmonids, as well as the importance of littoral areas as habitats to these species in less-saline parts of the Baltic Sea, should be taken into account with reference to stock management and the assessment of salmonid populations.
4. It is possible that Atlantic salmon and ABT parr that become smolts in non-natal streams may fixate their homing to these new waterbodies (**IV**, **V**). Thus, the phenomenon of stream shifting through the marine environment may constitute at least one possible mechanism behind the straying behaviour documented during the spawning runs of Atlantic salmon and ABT in the Baltic Sea basin.
5. It is likely that some coastal fish species inhabiting the Baltic Sea might show sub-basin specific behaviour in terms of their regular diel movements (e.g. European smelt (**I**, **II**)) and alternative life history patterns (e.g. Atlantic salmon, sea trout (**III**, **IV**, **V**)).

## SUMMARY IN ESTONIAN

### Läänemere idaosa litoraali kalakoosluste varieeruvus ja selle põhjused

Litoraal on veekogude (ookeanid, mered, järvad jne) bentali ökoloogiline sügavusvöönd, mis hõlmab rannikupiirkonna, kus kasvab põhjataimestik. Ookeanite ja merede (mereliste ökosüsteemide) madalaveeliseid litoraalialasid iseloomustavad muutlikud keskkonnatingimused (nt soolsus, veetemperatuur), mitmekesine elustik ja suur produktiivsus. Litoraalialad on vee-elustikule oluliseks sigimis-, kasvu- ja toitumisalaks. Muutlike abiootiliste ja biootiliste keskkonnategurite koosmõjul varieerub litoraalialade kalakoosluste liigiline koosseis väga suurel määral nii ajas (ööpäevane, aastaringne, aastatevaheline muutlikkus) kui ka ruumis (erinevad litoraalialade elupaigad). Riimveelise Läänemere erinevatele piirkondadele ning ka sama piirkonna erinevatele elupaikadele – näiteks Eru lahe siseosa vörrelduna lahe suudmeala või Soome lahe avaosaga – on iseloomulik vee soolsuse, temperatuuri ja toitelisuse kiire muutumine. Läänemere kalastik koosneb nii mageveelist kui ka merelist päritolu liikidest. Merelist päritolu liikide arvukus väheneb ning mageveeliste liikide osakaal suureneb koos soolsuse vähenemisega Läänemere ida ja põhjaosas. Lisaks rannakalanduse jaoks olulistele sihtliikidele, näiteks räim (*Clupea harengus*), meritint (*Osmerus eperlanus*), ahven (*Perca fluviatilis*) ja lest (*Platichthys flesus*) on Läänemere litoraalialad elupaigaks ka mitmetele kalanduslikult ebaolulistele liikidele: rünt (*Gobio gobio*), viidikas (*Alburnus alburnus*), ogalik (*Gasterosteus aculeatus*), luukarits (*Pungitius pungitius*), väike tobias (*Ammodytes tobianus*), madunõel (*Nerophis ophidion*), pisimudi-lake (*Pomatoschistus microps*) jne. Need pigem väheldaste mõõtmetega kalaliigid on toiduobjektideks Läänemere rannikulähedasi alasid asustavatele loomadele (putukad, kalad, linnud, imetajad), mistõttu moodustavad olulise lüli ökosüsteemi toiduahelas.

Vaatamata litoraalialade suurele tähtsusele mereliste ökosüsteemide funktsioneerimisel on Läänemere litoraali kalakoosluseid siiski suhteliselt vähe uuritud. Seni sel teemal avaldatud tööd on enamjaolt keskendunud aastate või aastaaegade vahelise abiootilise ja biootilise komponendi mõjule kalakooslusele või mõnele üksikule liigile. Seevastu Läänemere litoraali kalakoosluste öö-päevalringsetest muutustest on äärmiselt vähe teada. Kalakoosluste ööpäevalringset käitumist Läänemeres on minu teada analüüsitud vaid ühes avaldatud teadusartiklis ja sealgi on käsitletud vaid vähesid kalaliike. Teema päevalkohasusele osutavad ka viimaste teadusuuringute tulemused, mis näitavad, et mitme Läänemere litoraali asustava kalaliigi väikesemastaabilised vee sügavusest sõltuvad elupaiga eelistused muutuvad suve jooksul oluliselt. Lähtudes eelnevast ning merelistes ökosüsteemides kirjeldatud kalakoosluste ööpäevalringse dünaamika mitmetahulisusest, võiski eeldada, et Läänemere litoraali asustava kalakoosluse valgustingimustest sõltuv ümberpaiknemine erinevate

elupaikade vahel võib olla keerukam kui seni arvatud. Valgustingimustest sõltuv kalastiku ööpäevaringne dünaamika on otseselt seotud ka liikidevaheliste vastasmõjudega nii troofiliste tasemete piires kui ka nende vahel. Mingi konkreetse liigi käitumine on seotud tema toidukonkurentide, saakobjektide ja/või kiskjate elupaigaspetsiifilise arvukuse ja käitumisega uuritavas keskkonnas. Seega võib isegi väliselt sarnastel litoraalialadel kalade käitumine kohalike biootiliste ja abiootiliste tegurite koosmõjul märkimisväärtselt erineda.

Kuna Läänemere alampiirkondade keskkonnatingimused on väga erinevad, on tõenäoline, et ka samade kalaliikide käitumismustrid võivad oluliselt erineda nii alampiirkondade (nt Soome laht võrrelduna Läänemere avaosaga), merealade (nt litoraalala võrrelduna avamerega) kui ka erinevate mikroelupaikade (nt erinev põhjasubstraat, põhjataimestiku katvus) vahel. Hiljutised uurimused on näidanud, et mitmetele fakultatiivselt anadroomsetele liikidele (nt meriforell (*Salmo trutta*), luts (*Lota lota*), säinas (*Leuciscus idus*), haug (*Esox lucius*) on Läänemere litoraalalad oluliseks kasvualaks. Lisaks on näidatud, et kalade ränded litoraalialade ja vooluveekogude vahel on palju keerukamat kui senini arvatud; seni kirjeldamata rändestrateegiaid on Läänemere piirkonnas tähdatud nii meriforellil, haugil, säinal, lepamaimul (*Phoxinus phoxinus*) kui ka lutsul. Piltlikumateks näideteks võib pidada osaliselt anadroomse lepamaimu populatsiooni esinemist Saaremaal ning Gotlandi saarel vaadeldavat meriforelli noorjärkude väga varajast laskumist merre (mõne kuu jooksul pärast koorumist). Kõik eelnev viitab võimalusele, et suure hulga eripalgeliste keskkonnatingimuste mõjul võivad kalad lisaks lühiajalisele (ööpäevaringsele) käitumisele omada mitmeid elupaiga- ja piirkonnaspetsiifilisi elukäigumustreid. Seega moodustavad nii Läänemere litoraalala, sinna suubuvad vooluveekogud kui ka avameri väga kompleksse ning mitmekesise ökosüsteemi.

Väitekirja eesmärk on selgitada Läänemere litoraali asustavate kalaliikide ööpäevaringseid ja aastasiseseid käitumismustreid. Kalakoosluse valgustingimustest ja aastaajast sõltuva dünaamika uurimiseks kasutati madalat litoraalala (vee sügavus  $\leq 1$  m) Eru lahes (**I**). Litoraali kalastiku sesoonsete toitumisökoloogiliste eripärade kirjeldamiseks (võrrelduna nt avamerealadega) kasutati mudelobjektina meritinti (**II**). Anadroomsete kalade noorjärkude alternatiivseid rändestrateegiaid jõgede ja Läänemere litoraalialade vahel uuriti lõhe (*Salmo salar*) (**III, V**) ja meriforelli (**III, IV**) näitel.

Saadud tulemused kinnitavad, et Läänemere madalate litoraalialade kalastiku liigiline koosseis muutub oluliselt nii aasta kui ka ööpäeva jooksul (**I**). Selgus, et Läänemere idaosa litoraali kalakoosluse struktuuri mõjutavad ööpäeva lõikes suurel määral suhteliselt suuremakasvuliste liikide (nt meritint, rünt, viidikas, väike tobias) täiskasvanud isendid. Mitmetel püükides võrdlemisi arvukalt esinevatel liikidel (nt meritint, rünt, viidikas, luukarits, väike tobias) kujunesid kogu püügiperiodi lõikes välja kindlad valgustingimustest (hommik, keskpäev, õhtu, keskõö) sõltuvad käitumismustrid. Näiteks meritint esines uurimisalal peamiselt öösel ning väike tobias päeval (**I**). Meritindi puhul võivad käesolevas töös saadud uurimustulemused viidata kõnealuse liigi seni kirjeldamata elupaigaspetsiifilisele toitumisrändele (**I, II**). Läänemere litoraalialade

kalakooslustes domineerivate liikide, nagu ogaliku ja mudilate (väike mudilake (*Pomatoschistus minutus*) ja pisimudilake) arvukuse muutused uurimisalal sõltusid küll oluliselt valgustingimustest, kuid kogu vaadeldud perioodi (aprill kuni detsember) kattev lihtsalt selgitatav ööpäevane esinemismuster puudus (**I**). See tulemus on mõnevõrra vastuolus ainukese varasema Lääänemere madalas litoralis (<1,2m) elavate kalade ööpäevast dünaamikat käsitleva uuringuga, mis kirjeldas kõikide vaadeldud liikide (sh ogaliku ja pisimudilakese) arvukuse regulaarset tõusu öistes proovipüükides vörrelduna valge ajaga. Käesoleva uuringu tulemustest järeltub, et erinevatel ajahetkedel (nt päev ja öö) kogutud proovid pole omavahel üks ühele vörreldavad (**I**). Teiste sõnadega, mingi konkreetse litoraali piirkonna ööpäevas muutuva liigilise koosseisu täielikuks kirjeldamiseks on vajalik proovipüükide tegemine nii hommikul, keskpäeval, öhtul kui ka öösel. Lisaks viitavad käesoleva uuringu tulemused, et suve keskosa (juuli) on kaldanoodaga tehtavaks (vee sügavus  $\leq 1$  m) litoraali asustava kalastiku seireks ebasobiv periood, kuna siis ei ole nimetatud püügivahendiga võimalik kalakoosluse struktuuri adekvaatselt hinnata. Näiteks ogaliku, väikese mudilakese ja pisimudilakese samasuvised noorjärgud on suve keskosas kaldanoodaga tabamiseks liialt väiksed (**I**). Lisaks hoiduvad mitmed parema ujumisvõimega täiskasvanud kalad väga soojaveelisel perioodil tõenäoliselt kaldast kaugemale või on sigimispingutuse tagajärvel (nt ogalik, väike mudilake, pisimudilake) hukkunud.

Meritint, mitmetes Lääänemere alampiirkondades suhteliselt arvukas kalaliik, on sealsetes toiduvõrgustikes oluliseks lüliksi. Töö tulemused näitasid, et Lääänemere idaosa madalat litoraaliala asustava meritindi toitumiskäitumine muutub aasta jooksul oluliselt ning erineb avamere elupaikades kirjeldatust (**II**). Samuti sõltub meritindi saakobjektide valik Eru lahes kala pikkusest (**II**). Kõige arvukamateks meritindi saakobjektideks Eru lahes olid erinevad kirpvähilised ja kuulmiksabat (**II**). Ihtüoloogilistes teadustekstides korduma kippuvale küsimusele – Kas tint on röövkala – võib uuringu tulemuste põhjal vastata jaatavalta. Nimelt esines kõikide analüüsitud pikkusrühmade saagis kalu, eelkõige ogalikke, luukaritsaid, väike mudilakesi ja pisimudilakesi. Lisaks eelpoolmainitule olid meritindi saakobjektideks Eru lahes veel kakandilised, vesikirbulised, kümnejalalised, limused, putukad ja hulkharjasussid. Mitmeid uurimuses kirjeldatud saakobjekte (nt voldas (*Cottus gobio*), väike tobias, sööday rannakarp (*Mytilus trossulus*), munajas punntigu (*Macoma baltica*), harjasliimukas (*Hediste diversicolor*) jne) pole varasemates kirjandusallikates tindi toiduobjektidena nimetatud. Käesoleva uuringu tulemused näitavad, et tindil võib litoraalivööndis olla teistsugune ökoloogiline nišš kui avamerealadel (**II**). Meritinti võib Soome lahe rannikumeres pidada ahvena kõrval üheks olulisemaks röövkalaks ning on alust arvata, et liigi tähtsust litoraalialade toiduvõrgustiku osana on seni alahinnatud.

Traditsioniliselt on soolaseveeline litoraal lõhe ja meriforelli noorjärkudele oluliseks elupaigaks alles pärast kevadist hõbestumist (smoltifitseerumist, mille käigus kalade füsioloogia kohandub eluks merevees). Lõhilased on samas tuntud oma plastiliste elukäigustrateegiate poolest, mis varieeruvad liigi siseselt

nii rändemustriteilt (nt noorjärkude aastajast sõltuv elupaiga valik, kuderänne jne) kui ka näiteks suguküpse saabumise vanuse ja suuruse poolest. Üksikutes lõhe ja meriforelli populatsioonides on täheldatud ka vastsete ja tähnikute varajast (enne smoltifitseerumist) rännet riimveelistesse elupaikadesse. Reeglinä toimub lõhe ja forelli tähnikute varajane laskumine merre sügisel. Seestust Gotlandi saare ojadest laskuvad meriforelli vastsed ja tähnikud Läänemerre juba paari kuu jooksul pärast koorumist. Kirjeldatud alternatiivsete rändemustrite põhjuseks on pakutud näiteks jõe väga väkest või väga suurt vooluhulka, suurt asustustihedust või talvitumiseks sobivate elupaikade vähesust.

Käesolevas töös näidatakse esmakordsest nii lõhe kui ka meriforelli sügisel merre laskunud tähnikute esinemist Läänemeres (**III**). Samuti näidati mõlema liigi vastsete ning tähnikute rändamist sünnejõest Läänemere litoraali ning seal omakorda uude vooluveekogusse (**IV, V**). Selline läbi riimveelise litoraali toimuv mageveelise elupaiga vahetus on nii lõhe kui ka meriforelli seni kirjeldamata rändestrategia. Lõhi puhul pole kunagi varem dokumenteeritud isegi vastsete laskumist jõgedest merre.

Seega võib nii lõhe kui ka forelli noorjärkude rändestrategia magevee ning riimveelise litoraaliala vahel olla seni teada olnust oluliselt keerukam. Katseiliselt on näidatud, et 6–7 % soolsusega riimvesi ei oma meriforelli tähnikute elutegevusele negatiivset mõju. Lõhe puhul on Läänemerest oluliselt soolasesmas Lõuna-Inglismaal (Inglise kanali vesikond) dokumenteeritud üksikute sügisel tähnikunaookeanisse laskunud kalade kuderännet jõgedesse. Seega võivad Läänemere riimveelised litoraalialad olla seni arvatust olulisemaks kasvualaks alternatiivseid rändestrategiaid kasutavatele anadroomsetele lõhilastele noorjärkudele. Kirjanduse andmetel jätabad lõhilased oma sünnejõe „lõhna“ meelde smoltifitseerumise käigus, et tulla täiskasvanuna samasse vee-kogusse kudema. Läbi riimvee vooluveekogu vahetavad noorkalad võivad osaliselt olla ka anadroomsete lõhilaste populatsioonides esinevate „eksijate“ (st kalad, kes koevad oma kodujõe asemel mujal) varem kirjeldamata tekkemehhanismiks (**IV, V**). Vastse või tähnikuna kodujõge vahetades vermitakse lõhilastele smoltifitseerumise käigus uue elupaiga „muster“ ning vastav isend on seetõttu nii-öelda „eksinud“ juba enne täiskasvanuks saamist.

Kokkuvõtteks, käesoleva väitekirja tulemused näitavad et Läänemere litoraali asustavate kalaliikide ööpäevaringne ja aastajast sõltuv käitumine (sh toitumiskäitumine) on seni kirjeldatust oluliselt mitmetahulised (**I, II**). Selgus, et litoraali ja sellega piirnevate veeökosüsteemide (nt vooluveekogud, avameri) kalastike vahelised seosed on samuti seni arvatust oluliselt keerukamat (**III, IV, V**). Ülalpool kirjeldatud tulemused aitavad täita vajakajäämisi varasematest bioloogilistest ja metodoloogilistest teadmistest, ning on seega kasutatavad nii edaspidistes teadustöödes kui kalavarude majandamisel.

## **ACKNOWLEDGEMENTS**

My deepest gratitude goes to my supervisors Dr. Markus Vetemaa and Dr. Lauri Saks for their support, guidance, and constructive criticism during my studies. Preparation of this thesis would have been impossible without the help of my co-authors, colleagues, and friends who helped me during long fieldworks, provided literature, or advice. I hereby thank them all for their contribution (in alphabetical order): Anu Albert, Margit Eero, Redik Eschbaum, Kalvi Hubel, Kristiina Jürgens, Katri Kalda, Risto Kalda, Martin Kesler, Kairi Kullasepp, Maie Leirost, Lagle Matetski, Snežana Nedolgová, Rein Nellis, Maarja Nõmm, James Phillips, Mari-Liis Põlme, Ulvi Päädam, Mehis Rohtla, Katerina Rumvolt, Toomas Saat, Silver Sirp, Roland Svirgson, Toomas Tammaru, Andro “Mesi” Truuverk, Tiiu Tõrra, Aare Verliin and Erki Ōunap. Also, many thanks to the several referees for their valuable criticism on the manuscripts of the papers and this thesis. Last but not least, I wish to thank my family for their patience and understanding during my slow academic progress.

This study was funded by the Estonian Science Foundation’s grant 8281 and target financed project SF0180005s10.

## REFERENCES

- Alenius, P., Myrberg, K. & Nekrasov, A. 1998. The physical oceanography of the Gulf of Finland: a review. *Boreal Environment Research* 3: 97–125.
- Armstrong, R.H. 1974. Migrations of anadromous Dolly Varden (*Salvelinus malma*) in southeastern Alaska. *Journal of Fisheries Research Board of Canada* 31: 435–444.
- Arrington, D.A., Winemiller, K.O., Loftus, W.F. & Akin, S. 2002. How often do fishes „run on empty”? *Ecology* 83: 2145–2151.
- Bond, M.H., Crane, P.E., Larson, W.A. & Quinn, T.P. 2014. Is isolation by adaptation driving genetic divergence among proximate Dolly Varden char populations? *Ecology and Evolution* 4: 2515–2532.
- Castillo-Rivera, M., Zárate-Hernández, R., Ortiz-Burgos, S. & Zavala-Hurtado J. 2010. Diel and seasonal variability in the fish community structure of a mud-bottom estuarine habitat in the Gulf of Mexico. *Marine Ecology* 31: 633–642.
- Cunjak, R.A., Chadwick, E.M.P. & Shears, M. 1989. Downstream movements and estuarine residence by Atlantic salmon parr (*Salmo salar*). *Canadian Journal of Fisheries and Aquatic Sciences* 46: 1466–1471.
- Degerman, E., Leonardsson, K. & Lundqvist, H. 2012. Coastal migrations, temporary use of neighbouring rivers, and growth of Sea trout (*Salmo trutta*) from nine northern Baltic Sea rivers. *ICES Journal of Marine Science* 69: 971–980.
- Elliott, M., Hemingway, K.L., Costello, M.J., Duhamel, S., Hostens, K., Labropoulou, M., Marshall, S. & Winkler, H. 2002. Links between Fish and Other Trophic Levels. In: Elliott, M. & Hemingway, K.L. (eds.) *Fishes in Estuaries*. Blackwell Science Ltd, pp. 124–216.
- Eschbaum, R., Hubel, K., Jürgens, K., Rohtla, M., Špilev, H. & Talvik, Ü. 2015. *Kalandise riikliku andmekogumise programmi täitmine, andmete analüüs ning soovitused kalavarude haldamiseks 2015. aastal*. Töövõtulepingu nr 4–1.1/14/92 II vahearuanne. 199 pp. (in Estonian). [online] Available from [http://www.envir.ee/sites/default/files/akp\\_2014\\_rannikumere\\_kalad\\_aruanne.pdf](http://www.envir.ee/sites/default/files/akp_2014_rannikumere_kalad_aruanne.pdf) [accessed 22 May 2017]
- FishBase Glossary. 2017. System glossary. [online]. Available from <http://www.fishbase.org/glossary/Glossary.php?q=young-of-the-year> [accessed 17 April 2017]
- Gibson, R.N. & Robb, L. 1996. Piscine predation on juvenile fishes on a Scottish sandy beach. *Journal of Fish Biology* 49: 120–138.
- Hansen, L.P. & Jonsson, B. 1994. Homing in Atlantic salmon: Effects of juvenile learning on transplanted post-spawners. *Animal Behaviour* 47: 220–222.
- HELCOM. 2011. Salmon and Sea Trout Populations and Rivers in the Baltic Sea – Helcom assessment of salmon (*Salmo salar*) and sea trout (*Salmo trutta*) populations and habitats in rivers flowing to the Baltic Sea. Baltic Sea Environment Proceedings No. 126A. Available at <http://www.helcom.fi/Lists/Publications/BSEP126A.pdf> [accessed 8 March 2017].
- Hendry, K. & Cragg-Hine, D. 2003. Ecology of the Atlantic Salmon. Conserving Natura 2000 Rivers Ecology Series No. 7. English Nature, Peterborough. 32 pp.
- Holmes, R., Hayes, J.W., Jiang, W., Quarterman, A. & Davey, L.N. 2014. Emigration and mortality of juvenile brown trout in a New Zealand headwater tributary. *Ecology of Freshwater Fish* 23: 631–643.
- Hyslop, E.J. 1980. Stomach contents analysis – a review of methods and their application. *Journal of Fish Biology* 17: 411–429.

- Hänninen, J., Toivonen, R., Vahteri, P., Vuorinen, I. & Helminen, H. 2007. Environmental Factors Shaping the Littoral Biodiversity in the Finnish Archipelago, northern Baltic, and the Value of Low Biodiversity. Turku: SEILI Archipelago Research Institute Publications 4. 19 pp.
- Ibbotson, A.T., Riley, W.D., Beaumont, W.R.C., Cook, A.C., Ives, M.J., Pinder, A.C. & Scott, L.J. 2013. The source of autumn and spring downstream migration juvenile Atlantic salmon in a small lowland river. *Ecology of Freshwater Fish* 22: 73–81.
- Jensen, A.J., Finstad, B., Fiske, P., Hvidsten, N.A., Rikardsen, A.H. & Saksgård, L. 2012. Timing of smolt migration in sympatric populations of Atlantic salmon (*Salmo salar*), brown trout (*Salmo trutta*), and Arctic char (*Salvelinus alpinus*). *Canadian Journal of Fisheries and Aquatic Sciences* 69: 711–723.
- Jonsson, B. & Jonsson, N. 2009. Migratory timing, marine survival and growth of anadromous brown trout *Salmo trutta* in the River Imsa, Norway. *Journal of Fish Biology* 74: 621–638.
- Jonsson, B. & Jonsson, N. 2011. Ecology of Atlantic salmon and brown trout. Habitat as a template for life histories. Dordrecht, Springer. 708 pp.
- Jonsson, N., Hansen, L.P. & Jonsson, B. 1991. Variation in age, size and repeat spawning of adult Atlantic salmon in relation to river discharge. *Journal of Animal Ecology* 60: 937–947.
- Jonsson, B., Jonsson, N. & Hansen, L. P. 2003. Atlantic salmon straying from the River Imsa. *Journal of Fish Biology* 62: 641–657.
- Järvekülg, A. [Ярвекюльг, А.] 1979. The bottom fauna of the eastern part of the Baltic Sea. Tallinn: Valgus. 382 pp. In Russian.
- Järvekülg, A. 2001. Eesti jõed (in Estonian). University of Tartu Press, Tartu. 750 pp.
- Järvekülg, R., Kesler, M., Pihu, R. & Lauringson, G. 2012. Meriforelli kudejõgede taastootmispotentsiaali hindamine 2011. EMÜ PKI Limnoloogiakeskus / TÜ Eesti Mere-instituut / MTÜ Trulling. Tartu. 122 pp (in Estonian). [online] Available from <http://www.envir.ee/sites/default/files/mfaruanne2011lisadeta.pdf> [accessed 22 May 2017].
- Järvi, T., Holmgren, K., Rubin, J.F., Petersson, E., Lundberg, S. & Glimsäter, C. 1996. Newly-emerged *Salmo trutta* fry that migrate to the sea – an alternative choice of feeding habitat? *Nordic Journal of Freshwater Research* 72: 52–62.
- Kalish, J.M. 1990. Use of otolith microchemistry to distinguish the progeny of sympatric anadromous and non-anadromous salmonids. *Fishery Bulletin* 88: 657–666.
- Kangur, M., Paaver, T. & Drevs, T. 2003a. Salmon, *Salmo salar* L. In: Ojaveer, E., Saat, T. & Pihu, E. (eds.) *Fishes of Estonia*. Tallinn: Estonian Academy Publishers, pp. 91–97.
- Kangur, M., Paaver, T., Drevs, T. & Turovski, A. 2003b. Sea trout, *Salmo trutta* L. In: Ojaveer, E., Saat, T. & Pihu, E. (eds.) *Fishes of Estonia*. Tallinn: Estonian Academy Publishers, pp. 97–104.
- Kesler, M., Taal, I. & Svigrsden, R. 2015. Kalanduse riikliku andmekogumise programmi täitmine, vaalaliste juhusliku kaaspüügi hindamine Läänemerel ning soovitused kalavarude haldamiseks 2015. aastal. Töövõtulepingu nr 4–1.1/14/92 II vaheuaruanne. Osa: Lõhe ja meriforell. 74 pp (in Estonian). [online]. Available from [http://www.envir.ee/sites/default/files/akp\\_2014\\_lohe\\_meriforelli\\_aruanne.pdf](http://www.envir.ee/sites/default/files/akp_2014_lohe_meriforelli_aruanne.pdf) [accessed 22 May 2017].
- Kjellström, E. & Ruosteenoja, K. 2007. Present-day and future precipitation in the Baltic Sea region as simulated in a suite of regional climate models. *Climatic Change* 81: 281–291.

- Klemetsen, A., Amundsen, P.-A., Dempson, J.B., Jonsson, B., Jonsson, N., O'Connell, M.F. & Mortensen, E. 2003. Atlantic salmon *Salmo salar* L., brown trout *Salmo trutta* L. and Arctic charr *Salvelinus alpinus* (L.): a review of aspects of their life histories. *Ecology of Freshwater Fish* 12: 1–59.
- Koski, K.V. 2009. The fate of coho salmon nomads: the story of an estuarine-rearing strategy promoting resilience. *Ecology and Society* 14(1): 4. [online] Available from <http://www.ecologyandsociety.org/vol14/iss1/art4/> [accessed 8 March 2017].
- Kostrichkina, E.M. [Костричкина, Е.М.] 1968. Feeding relations of fishes in the Gulf of Riga. *Rybokhozyajstvennye issledovaniya v bassejne Baltijskogo morya* 4: 109–135. In Russian.
- Kostrichkina, E.M. [Костричкина, Е.М.] 1974. Dial rhythm of feeding and food composition of some benthophagous fish of the Gulf of Riga. *Rybokhozyajstvennye issledovaniya v bassejne Baltijskogo morya* 10: 28–37. In Russian.
- Landergren, P. 2001. Survival and growth of sea trout parr in fresh and brackish water. *Journal of Fish Biology* 58: 591–593.
- Landergren, P. 2004. Factors affecting early migration of sea trout *Salmo trutta* parr to brackish water. *Fisheries Research* 67: 283–294.
- Landergren, P. & Vallin, L. 1998. Spawning of sea trout, *Salmo trutta* L., in brackish waters—lost effort or successful strategy? *Fisheries Research* 35: 229–236.
- Lappalainen, A. & Urho, L. 2006. Young-of-the-year fish species composition in small coastal bays in the northern Baltic Sea, surveyed with beach seine and small underwater detonations. *Boreal Environment Research* 11: 431–440.
- Leppäranta, M. & Myrberg, K. 2009. Physical Oceanography of the Baltic Sea. Springer Berlin Heidelberg. 378 pp.
- Limburg, K.E., Landergren, P., Westin, L., Elfman, M. & Kristiansson, P. 2001. Flexible modes of anadromy in Baltic sea trout: making the most of marginal spawning streams. *Journal of Fish Biology* 59: 682–695.
- Lokko, K., Virro, T. & Kotta, J. 2016. Seasonal variability in the structure and functional diversity of psammic rotifer communities: role of environmental parameters. *Hydrobiologia* 796: 287–307.
- Louhi, P., Mäki-Petäys, A. & Erkinaro, J. 2008. Spawning habitat of Atlantic salmon and brown trout: general criteria and intragravel factors. *River Research and Applications* 24: 330–339.
- Lundström, K., Hjerne, O., Alexandersson, A. & Karlsson, O. 2007. Estimation of grey seal (*Halichoerus grypus*) diet composition in the Baltic Sea. *NAMMCO Scientific Publications* 6: 177–196.
- Martin, G., Torn, K., Kotta, J. & Orav-Kotta, H. 2003. Estonian marine phytobenthos monitoring programme: Preliminary results and future perspectives. *Proceedings of the Estonian Academy of Sciences. Biology, Ecology* 52: 112–124.
- Matetski, L. 2014. Distinguishing juvenile sea trout (*Salmo trutta*) from different natal streams based on otolith elemental fingerprints. Master's Thesis. University of Tartu. 65 pp (in Estonian with English summary).
- Meier, H.E.M., Andersson, H.C., Arheimer, B., Blenckner, T., Chubarenko, B., Donnelly, C., Eilola, K., Gustafsson, B.G., Hansson, A., Havenhand, J., Höglund, A., Kuznetsov, I., MacKenzie, B.R., Müller-Karulis, B., Neumann, T., Niiranen, S., Piwowarczyk, J., Raudsepp, U., Reckermann, M., Ruoho-Airola, T., Savchuk, O.P., Schenk, F., Schimanke, S., Väli, G., Weslawski, J-M. & Zorita, E. 2012. Comparing reconstructed past variations and future projections of the Baltic Sea ecosystem—first results from multi-model ensemble simulations. *Environmental Research Letters* 7: 1–8.

- Morkūnė, R., Lesutienė, J., Barisevičiūtė, R., Morkūnas, J. & Gasiūnaitė, Z.R. 2016. Food Sources of Wintering Piscivorous Waterbirds in Coastal Waters: a Triple Stable Isotope Approach for the Southeastern Baltic Sea. *Estuarine, Coastal and Shelf Science* 171: 41–50.
- Mustamäki, N. 2015. Spatial and temporal variation in fish populations and assemblages in coastal waters of the northern Baltic Proper. Doctoral Thesis. Environmental and Marine Biology, Husö biological station, Faculty of Science and Engineering, Åbo Akademi University. 41 pp.
- Mustamäki, N., Cederberg, T. & Mattila, J. 2014. Diet, stable isotopes and morphology of Eurasian perch (*Perca fluviatilis*) in littoral and pelagic habitats in the northern Baltic Proper. *Environmental Biology of Fishes* 97: 675–689.
- Mustamäki, N., Jokinen, H., Scheinin, M., Bonsdorff, E. & Mattila, J. 2015. Seasonal small-scale variation in distribution among depth zones in a coastal Baltic Sea fish assemblage. *ICES Journal of Marine Science* 72: 2374–2384.
- Myrberg, K. & Andrejev, O. 2003. Main upwelling regions in the Baltic Sea – a statistical analysis based on three-dimensional modelling. *Boreal Environment Research* 8: 97–112.
- Nedolgova, S. 2010. Feeding of smelt in Eru Bay. Master's Thesis. University of Tartu. 47 pp. (In Estonian with English summary).
- Nellbring, S. 1989. The ecology of smelts (genus *Osmerus*): a literature review. *Nordic Journal of Freshwater Research* 65: 116–145.
- Neumann, E. 1977. Activity and distribution of benthic Fish in some Baltic archipelagos with special reference to temperatuure. *Ambio Special Report* 5: 47–55.
- Niemelä, P., Tolvanen, H., Rönkä, M., Kellomäki, S., Krug, J., Schurgers, G., Lehtikoinen, E. & Kalliola, R. 2015. Environmental Impacts—Coastal Ecosystems, Birds and forests. In: Bolle, H-J., Menenti, M. & Rasool, S.I. (eds.) *Second Assessment of Climate Change for the Baltic Sea Basin, Regional Climate Studies*. Springer Science, pp. 291–306.
- Ojaveer, E. & Drevs, T. 2003. Flounder, *Platichthys flesus trachurus* (Duncker). In: Ojaveer, E., Saat, T. & Pihu, E. (eds.) *Fishes of Estonia*. Tallinn: Estonian Academy Publishers, pp. 362–370.
- Ojaveer, E. & Pihu, E. 2003. Estonian natural fish waters. In: Ojaveer, E., Pihu, E. & Saat, T. (eds) *Fishes of Estonia*. Tallinn: Estonian Academy Publishers, pp. 15–27.
- Ojaveer, H. 1997. Composition and dynamics of fish stocks in the Gulf of Riga ecosystem. *Dissertationes Biologicae Universitatis Tartuensis* 31. Tartu University Press. 33 pp.
- Ojaveer, H., Lankov, A., Lumberg, A. & Turovski, A. 1997. Forage fishes in the Brackish Gulf of Riga Ecosystem. In: Forage Fishes in Marine Ecosystems. Alaska Sea Grant College Program. University of Alaska Fairbanks, AK-SG-97-01, pp. 293–309.
- Ottaway, E.M., Carling, P.A., Clarke, A. & Reader, N.A. 1981. Observations on the structure of brown trout, *Salmo trutta* Linnaeus, redds. *Journal of Fish Biology* 19: 593–607.
- Pinder, A.C., Riley, W.D., Ibbotson, A.T. & Beaumont, W.R.C. 2007. Evidence for an autumn downstream migration and the subsequent estuarine residence of 0+ year juvenile Atlantic salmon *Salmo salar* L., in England. *Journal of fish Biology* 71: 260–264.
- Popov, A.N. [Попов, А.Н.] 1978. Feeding of smelt in the eastern part of Gulf of Finland. *Izvestiya GOSNIORH* 129: 53–63. In Russian.

- Popov, A.N. [Попов, А.Н.] 2006. Biology and reproduction of smelt (*Osmerus eperlanus* L.) in the eastern Gulf of Finland. In: Lavrentieva, G.M. & Susloparova, O.N. (eds.) Ecological aspects of hydroconstruction impact upon biota of the eastern Gulf of Finland. *Proceeding of FGNU GOSNIORH* 331: 92–118. In Russian.
- Quinn, T.P. 1993. A review of homing and straying of wild and hatchery-produced salmon. *Fisheries Research* 18: 29–44.
- Rajasilta, M., Mankki, J., Ranta-Aho, K. & Vuorinen, I. 1999. Littoral fish communities in the Archipelago Sea, SW Finland: a preliminary study of changes over 20 years. *Hydrobiologia* 393: 253–260.
- Riley, W.D., Eagle, M.O. & Ives, S.J. 2002. The onset of downstream movement of juvenile Atlantic salmon, *Salmo salar* L., in a chalk stream. *Fisheries Management and Ecology* 9: 87–94.
- Riley, W.D., Ibbotson, A.T., Lower, N., Cook, A.C., Moore, A., Mizuno, S., Pinder, A.C., Beaumont, W.R.C. & Privitera, L. 2008. Physiological seawater adaptation in juvenile Atlantic salmon (*Salmo salar*) autumn migrants. *Freshwater Biology* 53: 754–755.
- Riley, W.D., Ibbotson, A.T. & Beaumont, W.R.C. 2009. Adult returns from Atlantic salmon, *Salmo salar*, parr autumn migrants. *Fisheries management and Ecology* 16: 75–76.
- Rohtla, M. 2015. Otolith sclerochronological studies on migrations, spawning habitat preferences and age of freshwater fishes inhabiting the Baltic Sea. *Dissertationes Biologicae Universitatis Tartuensis* 271. University of Tartu Press. 49 pp.
- Rohtla, M., Vetemaa, M., Urtson, K. & Soesoo, A. 2012. Early life migration patterns of Baltic Sea pike *Esox lucius*. *Journal of Fish Biology* 80: 886–893.
- Rohtla, M., Vetemaa, M., Taal, I., Svirmsden, R., Urtson, K., Saks, L., Verliin, A., Kesler, M. & Saat, T. 2014. Life history of anadromous burbot (*Lota lota*, Linneaus) in the brackish Baltic Sea inferred from otolith microchemistry. *Ecology of Freshwater Fish* 23: 141–148.
- Roni, P., Bennett, T., Holland, R., Pess, G., Hanson, K., Moses, R., McHenry, M., Ehinger, W. & Walter, J. 2012. Factors Affecting Migration Timing, Growth, and Survival of Juvenile Coho Salmon in Two Coastal Washington Watersheds. *Transactions of the American Fisheries Society* 141(4): 890–906.
- Ross, S.T., McMichael R.H.Jr. & Ruple, D.L. 1987. Seasonal and diel variation in the standing crop of fishes and macroinvertebrates from a Gulf of Mexico surf zone. *Estuarine, Coastal and Shelf Science* 25: 391–412.
- Saat, T. & Eschbaum, R. 2002. Väinamere kalastik ja selle muutused viimastel aastakümnetel (Fishes of the Väinameri; changes during recent decades). In: Saat, T. (ed.) *Väinamere kalastik ja kalandus*. Tartu, Tartu University Publishers, pp. 9–45. In Estonian with English summary.
- Saat, T., Eschbaum, R., Vetemaa, M. & Verliin, A. 2003. Ten years of coastal fish monitoring in Estonia: dynamics of fish assemblages and populations. ICES CM 2003/R: 14. 17 pp.
- Seinä, A. & Peltola, J. 1991. Duration of the ice season and statistics of fast ice thickness along the Finnish coast 1961–1990. *Finnish Marine Research* 258: 3–46.
- Shestakov, V.G. [Шестаков, В.Г.] 1970. Feeding of smelt in Pärnu Bay. *Trudy Baltijskogo Nautsno-issledovatel'skogo Instituta Rybnogo Khoziaistva* 4: 349–360. In Russian.
- Shpilev, H., Ojaveer, E. & Lankov, A. 2005. Smelt (*Osmerus eperlanus* L.) in the Baltic Sea. *Proceedings of the Estonian Academy of Sciences: Biology, Ecology* 54: 230–241.

- Shrimpton, J.M., Warren, K.D., Todd, N.L., McRae, C.J., Glova, G.J., Telmer, K.H. & Clarke, A.D. 2014. Freshwater movement patterns by juvenile Pacific salmon *Oncorhynchus* spp. before they migrate to the ocean: Oh the places you'll go! *Journal of Fish Biology* 85: 987–1004.
- Shvagzhdys, A. 2009. Impact of environmental conditions on smelt catch fluctuations in the Nemunas River and the Curonian Lagoon. *Ekologija* 55: 204–214.
- Sundell, J. 1994. Dynamics and composition of littoral fish fauna along the coast of SW-Finland. *Aqua Fennica* 24 (1): 37–49.
- Svirgsten, R., Rohtla, M., Albert, A., Taal, I., Saks, L., Verliin, A. & Vetemaa, M. 2016. Do Eurasian minnows (*Phoxinus phoxinus* L.) inhabiting brackish water enter fresh water to reproduce: Evidence from a study on otolith microchemistry. *Ecology of Freshwater Fish*. doi: 10.1111/eff.12326
- Thorman, S. 1986a. Seasonal colonisation and effects of salinity and temperature on species richness and abundance of fish of some brackish and estuarine shallow waters in Sweden. *Holarctic Ecology* 9: 126–132.
- Thorman, S. 1986b. Physical factors affecting the abundance and species richness of fishes in the shallow waters of the southern Bothnian Sea, Sweden. *Estuarine, Coastal and Shelf Science* 22: 357–369.
- Thorman, S. & Wiederholm, A-M. 1983. Seasonal occurrence and food resource use of an assemblage of near-shore fish in the Bothnian Sea, Sweden. *Marine Ecology – Progress Series* 10: 223–229.
- Thorman, S. & Wiederholm, A-M. 1986. Food, habitat and time niches in a coastal fish species assemblage in a brackish water bay in the Bothnian Sea, Sweden. *Journal of Experimental Marine Biology and Ecology* 95:67–86.
- Timola, O. 1980. The diet of smelt, *Osmerus eperlanus eperlanus* (L.), in the northeastern Bothnian Bay. *Bothnian Bay Reports* 2: 9–16.
- Urtans, E. [Уртанс, Э.] 1990. Feeding characteristics of smelt (*Osmerus eperlanus eperlanus* L.) and eelpout (*Zoarces viviparus* L.) in the Gulf of Riga. *Fischereiforschung* 28: 34–38. In Russian.
- Ustups, D., Urtans, E., Minde, A. & Uzars, D. 2003. The structure and dynamics of fish communities in the Latvian coastal zone (Pape-Pērkone), Baltic Sea. *Acta Universitatis Latviensis* 662: 33–44.
- Ustups, D., Uzars, D. & Müller-Karulis, B. 2007. Structure and feeding ecology of the fish community in the surf zone of the eastern Baltic Latvian coast. *Proceedings of the Latvian Academy of Sciences, Section B* 61: 20–30.
- Uusitalo, L., Vehkalahti, K., Kuikka, S. & Söderkultalahti, P. 2005. Studying species associations from commercial catch data: a Baltic Sea application. *Fisheries Research* 72: 301–310.
- Vahteri, P., O'Brien, K. & Vuorinen, I. 2009. Zonation and spatial distribution of littoral fish communities from the southwestern Finnish coast (Archipelago and Bothnian Sea, Northern Baltic Sea). *Estuarine, Coastal and Shelf Science* 82: 35–40.
- Vasconcellos, R.M., Araújo, F.G., Santos, J.N.S. & Silva, M.A. 2011. Diel seasonality in fish biodiversity in a sandy beach in south-eastern Brazil. *Journal of the Marine Biological Association of the United Kingdom* 91(6): 1337–1344.
- Vetemaa, M., Eschbaum, R., Verliin, A., Albert, A., Eero, M., Lillemägi, R., Pihlak, M. & Saat, T. 2006. Annual and seasonal dynamics of fish in the brackish-water Matsalu Bay, Estonia. *Ecology of Freshwater Fish* 15: 211–220.
- Voipio, A. 1981. *The Baltic Sea*. Amsterdam: Elsevier. 418 pp.

- Walsworth, T.E., Schindler, D.E., Griffiths, J.R. & Zimmerman, C.E. 2015. Diverse juvenile life-history behaviours contribute to the spawning stock of an anadromous fish population. *Ecology of Freshwater Fish* 24: 204–213.
- Wilber, D.H., Clarke, D.G., Burlas, M.H., Ruben, H. & Will, R.J. 2003. Spatial and temporal variability in surf zone fish assemblages on the coast of northern New Jersey. *Estuarine, Coastal and Shelf Science* 56: 291–304.
- Winter, E.R., Tummers, J.S., Aarestrup, K., Baktoft, H. & Lucas, M.C. 2016. Investigating the phenology of seaward migration of juvenile brown trout (*Salmo trutta*) in two European populations. *Hydrobiologia* 775: 139–151.
- Youngson, A.F., Buck, R.J.G., Simpson, T.H. & Hay, D.W. 1983. The autumn and spring emigrations of juvenile Atlantic salmon, *Salmo salar* L., from the Girnock Burn, Aberdeenshire, Scotland: environmental release of migration. *Journal of Fish Biology* 23: 625–639.

## **PUBLICATIONS**

# CURRICULUM VITAE

**Name:** Imre Taal  
**Date of birth:** 12.11.1982  
**Citizenship:** Estonian  
**Address:** Estonian Marine Institute, University of Tartu,  
Vanemuise 46a, Tartu 51014, Estonia  
**E-mail:** imre.taal@ut.ee

**Education history:**

2008–... University of Tartu, doctoral studies in zoology and hydrobiology  
2001–2007 University of Tartu, *baccalaureus scientiarum* in hydrobiology

**Language skills:** Estonian, English, Russian

**Professional employment:**

2013–... Junior Researcher, University of Tartu, Estonian Marine Institute  
2004–2013 Laboratory Assistant, University of Tartu, Estonian Marine Institute

**Research interests:** Migration and feeding ecology of fish in the Baltic Sea region

**Publications:**

- Verliin, A., Saks, L., Svirgsden, R., Vetemaa, M., Rohtla, M., **Taal, I.** & Saat, T. 2013. Whitefish (*Coregonus lavaretus* (L.)) landings in the Baltic Sea during the past 100 years: combining official datasets and grey literature. *Advances in Limnology* 64: 133–152.
- Rohtla, M., Vetemaa, M., **Taal, I.**, Svirgsden, R., Urtson, K., Saks, L., Verliin, A., Kesler, M. & Saat, T. 2014. Life history of anadromous burbot (*Lota lota*, Linneaus) in the brackish Baltic Sea inferred from otolith microchemistry. *Ecology of Freshwater Fish* 23: 141–148.
- Rohtla, M., Vetemaa, M., Svirgsden, R., **Taal, I.**, Saks, L., Kesler, M., Verliin, A. & Saat, T. 2014. Using otolith 87Sr:86Sr as a natal chemical tag in the progeny of anadromous Baltic Sea pike (*Esox lucius*) – a pilot study. *Boreal Environment Research* 19: 379–386.
- Taal, I.**, Saks, L., Nedolgová, S., Verliin, A., Kesler, M., Jürgens, K., Svirgsden, R., Vetemaa, M. & Saat, T. 2014. Diet composition of smelt *Osmerus eperlanus* (Linnaeus) in brackish near-shore ecosystem (Eru Bay, Baltic Sea). *Ecology of Freshwater Fish* 23: 121–128.
- Taal, I.**, Kesler, K., Saks, L., Rohtla, M., Verliin, A., Svirgsden, R., Jürgens, K., Vetemaa, M. & Saat, T. 2014. Evidence for an autumn downstream

- migration of Atlantic salmon *Salmo salar* (Linnaeus) and brown trout *Salmo trutta* (Linnaeus) parr to the Baltic Sea. *Helgoland Marine Research* 68: 373–377.
- Rohtla, M., **Taal, I.**, Svirgsden, R. & Vetemaa, M. 2015. Old timers from the Baltic Sea: revisiting the population structure and maximum recorded age of ide *Leuciscus idus*. *Fisheries Research* 165: 74–78.
- Rohtla, M., Svirgsden, R., **Taal, I.**, Saks, L., Eschbaum, R. & Vetemaa, M. 2015. Life history characteristics of ide *Leuciscus idus* in the Eastern Baltic Sea. *Fisheries Management and Ecology* 22: 239–248.
- Svirgsden, R., Albert, A., Rohtla, M., **Taal, I.**, Saks, L., Verliin, A., Kesler, M., Hubel, K., Vetemaa, M. & Saat, T. 2015. Variations in egg characteristics of ruffe *Gymnocephalus cernua* inhabiting brackish and freshwater environments. *Helgoland Marine Research* 69: 273–283.
- Svirgsden, R., Rohtla, M., Albert, A., **Taal, I.**, Saks, L., Verliin, A. & Vetemaa, M. 2016. Do Eurasian minnows (*Phoxinus phoxinus* L.) inhabiting brackish water enter fresh water to reproduce: Evidence from a study on otolith microchemistry. *Ecology of Freshwater Fish*. doi: 10.1111/eff.12326
- Verliin, A., Kesler, M., Svirgsden, R., **Taal, I.**, Saks, L., Rohtla, M., Hubel, K., Eschbaum, R., Vetemaa, M. & Saat, T. 2017. Invasion of round goby to the temperate salmonid streams in the Baltic Sea. *Ichthyological Research* 64: 155–158.
- Taal, I.**, Saks, L., Kesler, K., Verliin, A., Jürgens, K., Svirgsden, R., Rohtla, M., Hubel, K., Albert, A., & Vetemaa, M. 2017. Diel changes in the fish assemblage in a coastal surf-zone area in the eastern Baltic Sea. *Boreal Environment Research* 22: 83–96.
- Taal, I.**, Rohtla, M., Saks, L., Kesler, M., Jürgens, K., Svirgsden, R., Matetski, L., Verliin, A., Paiste, P. & Vetemaa, M. 2017. Parr dispersal between streams via a marine environment: a novel mechanism behind straying for anadromous brown trout? *Ecology of Freshwater Fish*. doi: 10.1111/eff.12338
- Rohtla, M., Matetski, L., Svirgsden, R., Kesler, M., **Taal, I.**, Saura, A., Vaittinen, M. & Vetemaa, M. 2017. Do sea trout *Salmo trutta* parr surveys monitor the densities of anadromous or resident maternal origin parr, or both? *Fisheries Management and Ecology* 24: 156–162.
- Taal, I.**, Rohtla, M., Saks, L., Svirgsden, R., Kesler, M., Matetski, L. & Vetemaa, M. 2017. Evidence of Atlantic salmon *Salmo salar* fry movement between fresh water and a brackish environment. *Journal of Fish Biology* 91: 695–703.

#### Dissertations supervised:

- Snežana Nedolgová, Master's Degree, 2010, (sup) Imre Taal, Toomas Saat, Feeding of smelt in Eru Bay, University of Tartu. In Estonian.
- Katri Kalda, Master's Degree, 2015, (sup) Imre Taal, Changes in body sizes of fishes after preservation in ethanol and formalin, University of Tartu. In Estonian.

## ELULOOKIRJELDUS

**Nimi:** Imre Taal  
**Sünniaeg:** 12.11.1982  
**Kodakondus:** Eesti  
**Kontaktaadress:** Eesti Mereinstituut, Tartu Ülikool, Vanemuise 46a, Tartu 51014, Eesti  
**E-post:** imre.taal@ut.ee

**Haridus:**  
2008–... Tartu Ülikool, doktoriõpe zooloogia ja hüdrobioloogia erialal  
2001–2007 Tartu Ülikool, *baccalaureus scientiarum* hüdrobioloogia erialal

**Keelteoskus:** eesti, inglise, vene

**Teenistuskäik:**  
2013–... Nooremteadur, Eesti Mereinstituut, Tartu Ülikool  
2004–2013 Laborant, Eesti Mereinstituut, Tartu Ülikool

**Teadustöö põhisuunad:** Kalade ränded ja toitumine Läänemeres

**Publikatsioonide loetelu:**

- Verliin, A., Saks, L., Svirgsden, R., Vetemaa, M., Rohtla, M., **Taal, I.** & Saat, T. 2013. Whitefish (*Coregonus lavaretus* (L.)) landings in the Baltic Sea during the past 100 years: combining official datasets and grey literature. *Advances in Limnology* 64: 133–152.
- Rohtla, M., Vetemaa, M., **Taal, I.**, Svirgsden, R., Urtson, K., Saks, L., Verliin, A., Kesler, M. & Saat, T. 2014. Life history of anadromous burbot (*Lota lota*, Linneaus) in the brackish Baltic Sea inferred from otolith micro-chemistry. *Ecology of Freshwater Fish* 23: 141–148.
- Rohtla, M., Vetemaa, M., Svirgsden, R., **Taal, I.**, Saks, L., Kesler, M., Verliin, A. & Saat, T. 2014. Using otolith  $^{87}\text{Sr}$ : $^{86}\text{Sr}$  as a natal chemical tag in the progeny of anadromous Baltic Sea pike (*Esox lucius*) – a pilot study. *Boreal Environment Research* 19: 379–386.
- Taal, I.**, Saks, L., Nedolgov, S., Verliin, A., Kesler, M., Jürgens, K., Svirgsden, R., Vetemaa, M. & Saat, T. 2014. Diet composition of smelt *Osmerus eperlanus* (Linnaeus) in brackish near-shore ecosystem (Eru Bay, Baltic Sea). *Ecology of Freshwater Fish* 23: 121–128.
- Taal, I.**, Kesler, K., Saks, L., Rohtla, M., Verliin, A., Svirgsden, R., Jürgens, K., Vetemaa, M. & Saat, T. 2014. Evidence for an autumn downstream migration of Atlantic salmon *Salmo salar* (Linnaeus) and brown trout *Salmo trutta* (Linnaeus) parr to the Baltic Sea. *Helgoland Marine Research* 68: 373–377.

- Rohtla, M., **Taal, I.**, Svirgsden, R. & Vetemaa, M. 2015. Old timers from the Baltic Sea: revisiting the population structure and maximum recorded age of ide *Leuciscus idus*. *Fisheries Research* 165: 74–78.
- Rohtla, M., Svirgsden, R., **Taal, I.**, Saks, L., Eschbaum, R. & Vetemaa, M. 2015. Life history characteristics of ide *Leuciscus idus* in the Eastern Baltic Sea. *Fisheries Management and Ecology* 22: 239–248.
- Svirgsden, R., Albert, A., Rohtla, M., **Taal, I.**, Saks, L., Verliin, A., Kesler, M., Hubel, K., Vetemaa, M. & Saat, T. 2015. Variations in egg characteristics of ruffe *Gymnocephalus cernua* inhabiting brackish and freshwater environments. *Helgoland Marine Research* 69: 273–283.
- Svirgsden, R., Rohtla, M., Albert, A., **Taal, I.**, Saks, L., Verliin, A & Vetemaa, M. 2016. Do Eurasian minnows (*Phoxinus phoxinus* L.) inhabiting brackish water enter fresh water to reproduce: Evidence from a study on otolith microchemistry. *Ecology of Freshwater Fish*. doi: 10.1111/eff.12326
- Verliin, A., Kesler, M., Svirgsden, R., **Taal, I.**, Saks, L., Rohtla, M., Hubel, K., Eschbaum, R., Vetemaa, M. & Saat, T. 2017. Invasion of round goby to the temperate salmonid streams in the Baltic Sea. *Ichthyological Research* 64: 155–158.
- Taal, I.**, Saks, L., Kesler, K., Verliin, A., Jürgens, K., Svirgsden, R., Rohtla, M., Hubel, K., Albert, A., Eschbaum, R. & Vetemaa, M. 2017. Diel changes in the fish assemblage in a coastal surf-zone area in the eastern Baltic Sea. *Boreal Environment Research* 22: 83–96.
- Taal, I.**, Rohtla, M., Saks, L., Kesler, M., Jürgens, K., Svirgsden, R., Matetski, L., Verliin, A., Paiste, P. & Vetemaa, M. 2017. Parr dispersal between streams via a marine environment: a novel mechanism behind straying for anadromous brown trout? *Ecology of Freshwater Fish*. doi: 10.1111/eff.12338
- Rohtla, M., Matetski, L., Svirgsden, R., Kesler, M., **Taal, I.**, Saura, A., Vaittinen, M. & Vetemaa, M. 2017. Do sea trout *Salmo trutta* parr surveys monitor the densities of anadromous or resident maternal origin parr, or both? *Fisheries Management and Ecology* 24: 156–162.
- Taal, I.**, Rohtla, M., Saks, L., Svirgsden, R., Kesler, M., Matetski, L. & Vetemaa, M. 2017. Evidence of Atlantic salmon *Salmo salar* fry movement between fresh water and a brackish environment. *Journal of Fish Biology* 91: 695–703.

#### **Juhendatud väitekirjad:**

- Snežana Nedolgova, magistrikraad, 2010, (juh) Imre Taal, Toomas Saat, Meritindi toitumine Eru lahes, Tartu Ülikool.
- Katri Kalda, magistrikraad, 2015, (juh) Imre Taal, Formaliini ja etanooliga fikseerimise mõju kalade kehamõõtmetele, Tartu Ülikool.

# DISSERTATIONES BIOLOGICAE UNIVERSITATIS TARTUENSIS

1. **Toivo Maimets.** Studies of human oncoprotein p53. Tartu, 1991, 96 p.
2. **Enn K. Seppet.** Thyroid state control over energy metabolism, ion transport and contractile functions in rat heart. Tartu, 1991, 135 p.
3. **Kristjan Zobel.** Epifüütsete makrosamblike värtus õhu saastuse indikaatoritena Hamar-Dobani boreaalsetes mägimetsades. Tartu, 1992, 131 lk.
4. **Andres Mäe.** Conjugal mobilization of catabolic plasmids by transposable elements in helper plasmids. Tartu, 1992, 91 p.
5. **Maia Kivisaar.** Studies on phenol degradation genes of *Pseudomonas* sp. strain EST 1001. Tartu, 1992, 61 p.
6. **Allan Nurk.** Nucleotide sequences of phenol degradative genes from *Pseudomonas* sp. strain EST 1001 and their transcriptional activation in *Pseudomonas putida*. Tartu, 1992, 72 p.
7. **Ülo Tamm.** The genus *Populus* L. in Estonia: variation of the species biology and introduction. Tartu, 1993, 91 p.
8. **Jaanus Remme.** Studies on the peptidyltransferase centre of the *E.coli* ribosome. Tartu, 1993, 68 p.
9. **Ülo Langel.** Galanin and galanin antagonists. Tartu, 1993, 97 p.
10. **Arvo Käärd.** The development of an automatic online dynamic fluorescence-based pH-dependent fiber optic penicillin flowthrough biosensor for the control of the benzylpenicillin hydrolysis. Tartu, 1993, 117 p.
11. **Lilian Järvekülg.** Antigenic analysis and development of sensitive immunoassay for potato viruses. Tartu, 1993, 147 p.
12. **Jaak Palumets.** Analysis of phytomass partition in Norway spruce. Tartu, 1993, 47 p.
13. **Arne Sellin.** Variation in hydraulic architecture of *Picea abies* (L.) Karst. trees grown under different environmental conditions. Tartu, 1994, 119 p.
13. **Mati Reeben.** Regulation of light neurofilament gene expression. Tartu, 1994, 108 p.
14. **Urmas Tartes.** Respiration rhythms in insects. Tartu, 1995, 109 p.
15. **Ülo Puurand.** The complete nucleotide sequence and infections *in vitro* transcripts from cloned cDNA of a potato A potyvirus. Tartu, 1995, 96 p.
16. **Peeter Hõrak.** Pathways of selection in avian reproduction: a functional framework and its application in the population study of the great tit (*Parus major*). Tartu, 1995, 118 p.
17. **Erkki Truve.** Studies on specific and broad spectrum virus resistance in transgenic plants. Tartu, 1996, 158 p.
18. **Illar Pata.** Cloning and characterization of human and mouse ribosomal protein S6-encoding genes. Tartu, 1996, 60 p.
19. **Ülo Niinemets.** Importance of structural features of leaves and canopy in determining species shade-tolerance in temperature deciduous woody taxa. Tartu, 1996, 150 p.

20. **Ants Kurg.** Bovine leukemia virus: molecular studies on the packaging region and DNA diagnostics in cattle. Tartu, 1996, 104 p.
21. **Ene Ustav.** E2 as the modulator of the BPV1 DNA replication. Tartu, 1996, 100 p.
22. **Aksel Soosaar.** Role of helix-loop-helix and nuclear hormone receptor transcription factors in neurogenesis. Tartu, 1996, 109 p.
23. **Maido Remm.** Human papillomavirus type 18: replication, transformation and gene expression. Tartu, 1997, 117 p.
24. **Tiiu Kull.** Population dynamics in *Cypripedium calceolus* L. Tartu, 1997, 124 p.
25. **Kalle Olli.** Evolutionary life-strategies of autotrophic planktonic micro-organisms in the Baltic Sea. Tartu, 1997, 180 p.
26. **Meelis Pärtel.** Species diversity and community dynamics in calcareous grassland communities in Western Estonia. Tartu, 1997, 124 p.
27. **Malle Leht.** The Genus *Potentilla* L. in Estonia, Latvia and Lithuania: distribution, morphology and taxonomy. Tartu, 1997, 186 p.
28. **Tanel Tenson.** Ribosomes, peptides and antibiotic resistance. Tartu, 1997, 80 p.
29. **Arvo Tuvikene.** Assessment of inland water pollution using biomarker responses in fish *in vivo* and *in vitro*. Tartu, 1997, 160 p.
30. **Urmas Saarma.** Tuning ribosomal elongation cycle by mutagenesis of 23S rRNA. Tartu, 1997, 134 p.
31. **Henn Ojaveer.** Composition and dynamics of fish stocks in the gulf of Riga ecosystem. Tartu, 1997, 138 p.
32. **Lembi Lõugas.** Post-glacial development of vertebrate fauna in Estonian water bodies. Tartu, 1997, 138 p.
33. **Margus Pooga.** Cell penetrating peptide, transportan, and its predecessors, galanin-based chimeric peptides. Tartu, 1998, 110 p.
34. **Andres Saag.** Evolutionary relationships in some cetrarioid genera (Lichenized Ascomycota). Tartu, 1998, 196 p.
35. **Aivar Liiv.** Ribosomal large subunit assembly *in vivo*. Tartu, 1998, 158 p.
36. **Tatjana Oja.** Isoenzyme diversity and phylogenetic affinities among the eurasian annual bromes (*Bromus* L., Poaceae). Tartu, 1998, 92 p.
37. **Mari Moora.** The influence of arbuscular mycorrhizal (AM) symbiosis on the competition and coexistence of calcareous grassland plant species. Tartu, 1998, 78 p.
38. **Olavi Kurina.** Fungus gnats in Estonia (*Diptera: Bolitophilidae, Keroplatidae, Macroceridae, Ditomyiidae, Diadocidiidae, Mycetophilidae*). Tartu, 1998, 200 p.
39. **Andrus Tasa.** Biological leaching of shales: black shale and oil shale. Tartu, 1998, 98 p.
40. **Arnold Kristjuhan.** Studies on transcriptional activator properties of tumor suppressor protein p53. Tartu, 1998, 86 p.
41. **Sulev Ingerpuu.** Characterization of some human myeloid cell surface and nuclear differentiation antigens. Tartu, 1998, 163 p.

42. **Veljo Kisand.** Responses of planktonic bacteria to the abiotic and biotic factors in the shallow lake Võrtsjärv. Tartu, 1998, 118 p.
43. **Kadri Põldmaa.** Studies in the systematics of hypomyces and allied genera (Hypocreales, Ascomycota). Tartu, 1998, 178 p.
44. **Markus Vetemaa.** Reproduction parameters of fish as indicators in environmental monitoring. Tartu, 1998, 117 p.
45. **Heli Talvik.** Prepatent periods and species composition of different *Oesophagostomum* spp. populations in Estonia and Denmark. Tartu, 1998, 104 p.
46. **Katrin Heinsoo.** Cuticular and stomatal antechamber conductance to water vapour diffusion in *Picea abies* (L.) karst. Tartu, 1999, 133 p.
47. **Tarmo Annilo.** Studies on mammalian ribosomal protein S7. Tartu, 1998, 77 p.
48. **Indrek Ots.** Health state indicies of reproducing great tits (*Parus major*): sources of variation and connections with life-history traits. Tartu, 1999, 117 p.
49. **Juan Jose Cantero.** Plant community diversity and habitat relationships in central Argentina grasslands. Tartu, 1999, 161 p.
50. **Rein Kalamees.** Seed bank, seed rain and community regeneration in Estonian calcareous grasslands. Tartu, 1999, 107 p.
51. **Sulev Kõks.** Cholecystokinin (CCK) – induced anxiety in rats: influence of environmental stimuli and involvement of endopioid mechanisms and serotonin. Tartu, 1999, 123 p.
52. **Ebe Sild.** Impact of increasing concentrations of O<sub>3</sub> and CO<sub>2</sub> on wheat, clover and pasture. Tartu, 1999, 123 p.
53. **Ljudmilla Timofejeva.** Electron microscopical analysis of the synaptoneuronal complex formation in cereals. Tartu, 1999, 99 p.
54. **Andres Valkna.** Interactions of galanin receptor with ligands and G-proteins: studies with synthetic peptides. Tartu, 1999, 103 p.
55. **Taavi Virro.** Life cycles of planktonic rotifers in lake Peipsi. Tartu, 1999, 101 p.
56. **Ana Rebane.** Mammalian ribosomal protein S3a genes and intron-encoded small nucleolar RNAs U73 and U82. Tartu, 1999, 85 p.
57. **Tiina Tamm.** Cocksfoot mottle virus: the genome organisation and translational strategies. Tartu, 2000, 101 p.
58. **Reet Kurg.** Structure-function relationship of the bovine papilloma virus E2 protein. Tartu, 2000, 89 p.
59. **Toomas Kivilild.** The origins of Southern and Western Eurasian populations: an mtDNA study. Tartu, 2000, 121 p.
60. **Niilo Kaldalu.** Studies of the TOL plasmid transcription factor XylS. Tartu, 2000, 88 p.
61. **Dina Lepik.** Modulation of viral DNA replication by tumor suppressor protein p53. Tartu, 2000, 106 p.

62. **Kai Vellak.** Influence of different factors on the diversity of the bryophyte vegetation in forest and wooded meadow communities. Tartu, 2000, 122 p.
63. **Jonne Kotta.** Impact of eutrophication and biological invasionas on the structure and functions of benthic macrofauna. Tartu, 2000, 160 p.
64. **Georg Martin.** Phytobenthic communities of the Gulf of Riga and the inner sea the West-Estonian archipelago. Tartu, 2000, 139 p.
65. **Silvia Sepp.** Morphological and genetical variation of *Alchemilla L.* in Estonia. Tartu, 2000. 124 p.
66. **Jaan Liira.** On the determinants of structure and diversity in herbaceous plant communities. Tartu, 2000, 96 p.
67. **Priit Zingel.** The role of planktonic ciliates in lake ecosystems. Tartu, 2001, 111 p.
68. **Tiit Teder.** Direct and indirect effects in Host-parasitoid interactions: ecological and evolutionary consequences. Tartu, 2001, 122 p.
69. **Hannes Kollist.** Leaf apoplastic ascorbate as ozone scavenger and its transport across the plasma membrane. Tartu, 2001, 80 p.
70. **Reet Marits.** Role of two-component regulator system PehR-PehS and extracellular protease PrtW in virulence of *Erwinia Carotovora* subsp. *Carotovora*. Tartu, 2001, 112 p.
71. **Vallo Tilgar.** Effect of calcium supplementation on reproductive performance of the pied flycatcher *Ficedula hypoleuca* and the great tit *Parus major*, breeding in Nothern temperate forests. Tartu, 2002, 126 p.
72. **Rita Hõrak.** Regulation of transposition of transposon Tn4652 in *Pseudomonas putida*. Tartu, 2002, 108 p.
73. **Liina Eek-Piirssoo.** The effect of fertilization, mowing and additional illumination on the structure of a species-rich grassland community. Tartu, 2002, 74 p.
74. **Krõõt Aasamaa.** Shoot hydraulic conductance and stomatal conductance of six temperate deciduous tree species. Tartu, 2002, 110 p.
75. **Nele Ingerpuu.** Bryophyte diversity and vascular plants. Tartu, 2002, 112 p.
76. **Neeme Tõnisson.** Mutation detection by primer extension on oligonucleotide microarrays. Tartu, 2002, 124 p.
77. **Margus Pensa.** Variation in needle retention of Scots pine in relation to leaf morphology, nitrogen conservation and tree age. Tartu, 2003, 110 p.
78. **Asko Lõhmus.** Habitat preferences and quality for birds of prey: from principles to applications. Tartu, 2003, 168 p.
79. **Viljar Jaks.** p53 – a switch in cellular circuit. Tartu, 2003, 160 p.
80. **Jaana Männik.** Characterization and genetic studies of four ATP-binding cassette (ABC) transporters. Tartu, 2003, 140 p.
81. **Marek Sammul.** Competition and coexistence of clonal plants in relation to productivity. Tartu, 2003, 159 p
82. **Ivar Ilves.** Virus-cell interactions in the replication cycle of bovine papillomavirus type 1. Tartu, 2003, 89 p.

83. **Andres Männik.** Design and characterization of a novel vector system based on the stable replicator of bovine papillomavirus type 1. Tartu, 2003, 109 p.
84. **Ivika Ostonen.** Fine root structure, dynamics and proportion in net primary production of Norway spruce forest ecosystem in relation to site conditions. Tartu, 2003, 158 p.
85. **Gudrun Veldre.** Somatic status of 12–15-year-old Tartu schoolchildren. Tartu, 2003, 199 p.
86. **Ülo Väli.** The greater spotted eagle *Aquila clanga* and the lesser spotted eagle *A. pomarina*: taxonomy, phylogeography and ecology. Tartu, 2004, 159 p.
87. **Aare Abroi.** The determinants for the native activities of the bovine papillomavirus type 1 E2 protein are separable. Tartu, 2004, 135 p.
88. **Tiina Kahre.** Cystic fibrosis in Estonia. Tartu, 2004, 116 p.
89. **Helen Orav-Kotta.** Habitat choice and feeding activity of benthic suspension feeders and mesograzers in the northern Baltic Sea. Tartu, 2004, 117 p.
90. **Maarja Öpik.** Diversity of arbuscular mycorrhizal fungi in the roots of perennial plants and their effect on plant performance. Tartu, 2004, 175 p.
91. **Kadri Tali.** Species structure of *Neotinea ustulata*. Tartu, 2004, 109 p.
92. **Kristiina Tambets.** Towards the understanding of post-glacial spread of human mitochondrial DNA haplogroups in Europe and beyond: a phylogeographic approach. Tartu, 2004, 163 p.
93. **Arvi Jõers.** Regulation of p53-dependent transcription. Tartu, 2004, 103 p.
94. **Lilian Kadaja.** Studies on modulation of the activity of tumor suppressor protein p53. Tartu, 2004, 103 p.
95. **Jaak Truu.** Oil shale industry wastewater: impact on river microbial community and possibilities for bioremediation. Tartu, 2004, 128 p.
96. **Maire Peters.** Natural horizontal transfer of the *pheBA* operon. Tartu, 2004, 105 p.
97. **Ülo Maiväli.** Studies on the structure-function relationship of the bacterial ribosome. Tartu, 2004, 130 p.
98. **Merit Otsus.** Plant community regeneration and species diversity in dry calcareous grasslands. Tartu, 2004, 103 p.
99. **Mikk Heidemaa.** Systematic studies on sawflies of the genera *Dolerus*, *Empria*, and *Caliroa* (Hymenoptera: Tenthredinidae). Tartu, 2004, 167 p.
100. **Ilmar Tõnno.** The impact of nitrogen and phosphorus concentration and N/P ratio on cyanobacterial dominance and N<sub>2</sub> fixation in some Estonian lakes. Tartu, 2004, 111 p.
101. **Lauri Saks.** Immune function, parasites, and carotenoid-based ornaments in greenfinches. Tartu, 2004, 144 p.
102. **Siiri Roots.** Human Y-chromosomal variation in European populations. Tartu, 2004, 142 p.
103. **Eve Vedler.** Structure of the 2,4-dichloro-phenoxyacetic acid-degradative plasmid pEST4011. Tartu, 2005. 106 p.

104. **Andres Tover.** Regulation of transcription of the phenol degradation *pheBA* operon in *Pseudomonas putida*. Tartu, 2005, 126 p.
105. **Helen Udras.** Hexose kinases and glucose transport in the yeast *Hansenula polymorpha*. Tartu, 2005, 100 p.
106. **Ave Suija.** Lichens and lichenicolous fungi in Estonia: diversity, distribution patterns, taxonomy. Tartu, 2005, 162 p.
107. **Piret Lõhmus.** Forest lichens and their substrata in Estonia. Tartu, 2005, 162 p.
108. **Inga Lips.** Abiotic factors controlling the cyanobacterial bloom occurrence in the Gulf of Finland. Tartu, 2005, 156 p.
109. **Kaasik, Krista.** Circadian clock genes in mammalian clockwork, metabolism and behaviour. Tartu, 2005, 121 p.
110. **Juhan Javoš.** The effects of experience on host acceptance in ovipositing moths. Tartu, 2005, 112 p.
111. **Tiina Sedman.** Characterization of the yeast *Saccharomyces cerevisiae* mitochondrial DNA helicase Hml1. Tartu, 2005, 103 p.
112. **Ruth Aguraiuja.** Hawaiian endemic fern lineage *Diellia* (Aspleniaceae): distribution, population structure and ecology. Tartu, 2005, 112 p.
113. **Riho Teras.** Regulation of transcription from the fusion promoters generated by transposition of Tn4652 into the upstream region of *pheBA* operon in *Pseudomonas putida*. Tartu, 2005, 106 p.
114. **Mait Metspalu.** Through the course of prehistory in india: tracing the mtDNA trail. Tartu, 2005, 138 p.
115. **Elin Lõhmussaar.** The comparative patterns of linkage disequilibrium in European populations and its implication for genetic association studies. Tartu, 2006, 124 p.
116. **Priit Kupper.** Hydraulic and environmental limitations to leaf water relations in trees with respect to canopy position. Tartu, 2006, 126 p.
117. **Heili Ilves.** Stress-induced transposition of Tn4652 in *Pseudomonas Putida*. Tartu, 2006, 120 p.
118. **Silja Kuusk.** Biochemical properties of Hml1p, a DNA helicase from *Saccharomyces cerevisiae* mitochondria. Tartu, 2006, 126 p.
119. **Kersti Püssa.** Forest edges on medium resolution landsat thematic mapper satellite images. Tartu, 2006, 90 p.
120. **Lea Tummeleht.** Physiological condition and immune function in great tits (*Parus major* l.): Sources of variation and trade-offs in relation to growth. Tartu, 2006, 94 p.
121. **Toomas Esperk.** Larval instar as a key element of insect growth schedules. Tartu, 2006, 186 p.
122. **Harri Valdmann.** Lynx (*Lynx lynx*) and wolf (*Canis lupus*) in the Baltic region: Diets, helminth parasites and genetic variation. Tartu, 2006, 102 p.
123. **Priit Jõers.** Studies of the mitochondrial helicase Hml1p in *Candida albicans* and *Saccharomyces cerevisiae*. Tartu, 2006, 113 p.
124. **Kersti Lilleväli.** Gata3 and Gata2 in inner ear development. Tartu, 2007, 123 p.

125. **Kai Rünk.** Comparative ecology of three fern species: *Dryopteris carthusiana* (Vill.) H.P. Fuchs, *D. expansa* (C. Presl) Fraser-Jenkins & Jermy and *D. dilatata* (Hoffm.) A. Gray (Dryopteridaceae). Tartu, 2007, 143 p.
126. **Aveliina Helm.** Formation and persistence of dry grassland diversity: role of human history and landscape structure. Tartu, 2007, 89 p.
127. **Leho Tedersoo.** Ectomycorrhizal fungi: diversity and community structure in Estonia, Seychelles and Australia. Tartu, 2007, 233 p.
128. **Marko Mägi.** The habitat-related variation of reproductive performance of great tits in a deciduous-coniferous forest mosaic: looking for causes and consequences. Tartu, 2007, 135 p.
129. **Valeria Lulla.** Replication strategies and applications of Semliki Forest virus. Tartu, 2007, 109 p.
130. **Ülle Reier.** Estonian threatened vascular plant species: causes of rarity and conservation. Tartu, 2007, 79 p.
131. **Inga Jüriado.** Diversity of lichen species in Estonia: influence of regional and local factors. Tartu, 2007, 171 p.
132. **Tatjana Krama.** Mobbing behaviour in birds: costs and reciprocity based cooperation. Tartu, 2007, 112 p.
133. **Signe Saumaa.** The role of DNA mismatch repair and oxidative DNA damage defense systems in avoidance of stationary phase mutations in *Pseudomonas putida*. Tartu, 2007, 172 p.
134. **Reedik Mägi.** The linkage disequilibrium and the selection of genetic markers for association studies in european populations. Tartu, 2007, 96 p.
135. **Priit Kilgas.** Blood parameters as indicators of physiological condition and skeletal development in great tits (*Parus major*): natural variation and application in the reproductive ecology of birds. Tartu, 2007, 129 p.
136. **Anu Albert.** The role of water salinity in structuring eastern Baltic coastal fish communities. Tartu, 2007, 95 p.
137. **Kärt Padari.** Protein transduction mechanisms of transportans. Tartu, 2008, 128 p.
138. **Siiri-Lii Sandre.** Selective forces on larval colouration in a moth. Tartu, 2008, 125 p.
139. **Ülle Jõgar.** Conservation and restoration of semi-natural floodplain meadows and their rare plant species. Tartu, 2008, 99 p.
140. **Lauri Laanisto.** Macroecological approach in vegetation science: generality of ecological relationships at the global scale. Tartu, 2008, 133 p.
141. **Reidar Andreson.** Methods and software for predicting PCR failure rate in large genomes. Tartu, 2008, 105 p.
142. **Birgot Paavel.** Bio-optical properties of turbid lakes. Tartu, 2008, 175 p.
143. **Kaire Torn.** Distribution and ecology of charophytes in the Baltic Sea. Tartu, 2008, 98 p.
144. **Vladimir Vimberg.** Peptide mediated macrolide resistance. Tartu, 2008, 190 p.
145. **Daima Örd.** Studies on the stress-inducible pseudokinase TRB3, a novel inhibitor of transcription factor ATF4. Tartu, 2008, 108 p.

146. **Lauri Saag.** Taxonomic and ecologic problems in the genus *Lepraria* (*Stereocaulaceae*, lichenised *Ascomycota*). Tartu, 2008, 175 p.
147. **Ulvi Karu.** Antioxidant protection, carotenoids and coccidians in green-finches – assessment of the costs of immune activation and mechanisms of parasite resistance in a passerine with carotenoid-based ornaments. Tartu, 2008, 124 p.
148. **Jaanus Remm.** Tree-cavities in forests: density, characteristics and occupancy by animals. Tartu, 2008, 128 p.
149. **Epp Moks.** Tapeworm parasites *Echinococcus multilocularis* and *E. granulosus* in Estonia: phylogenetic relationships and occurrence in wild carnivores and ungulates. Tartu, 2008, 82 p.
150. **Eve Eensalu.** Acclimation of stomatal structure and function in tree canopy: effect of light and CO<sub>2</sub> concentration. Tartu, 2008, 108 p.
151. **Janne Pullat.** Design, functionlization and application of an *in situ* synthesized oligonucleotide microarray. Tartu, 2008, 108 p.
152. **Marta Putrinš.** Responses of *Pseudomonas putida* to phenol-induced metabolic and stress signals. Tartu, 2008, 142 p.
153. **Marina Semtšenko.** Plant root behaviour: responses to neighbours and physical obstructions. Tartu, 2008, 106 p.
154. **Marge Starast.** Influence of cultivation techniques on productivity and fruit quality of some *Vaccinium* and *Rubus* taxa. Tartu, 2008, 154 p.
155. **Age Tats.** Sequence motifs influencing the efficiency of translation. Tartu, 2009, 104 p.
156. **Radi Tegova.** The role of specialized DNA polymerases in mutagenesis in *Pseudomonas putida*. Tartu, 2009, 124 p.
157. **Tsipe Aavik.** Plant species richness, composition and functional trait pattern in agricultural landscapes – the role of land use intensity and landscape structure. Tartu, 2009, 112 p.
158. **Kaja Kiiver.** Semliki forest virus based vectors and cell lines for studying the replication and interactions of alphaviruses and hepaciviruses. Tartu, 2009, 104 p.
159. **Meelis Kadaja.** Papillomavirus Replication Machinery Induces Genomic Instability in its Host Cell. Tartu, 2009, 126 p.
160. **Pille Hallast.** Human and chimpanzee Luteinizing hormone/Chorionic Gonadotropin beta (*LHB/CGB*) gene clusters: diversity and divergence of young duplicated genes. Tartu, 2009, 168 p.
161. **Ain Vellak.** Spatial and temporal aspects of plant species conservation. Tartu, 2009, 86 p.
162. **Triinu Remmel.** Body size evolution in insects with different colouration strategies: the role of predation risk. Tartu, 2009, 168 p.
163. **Jaana Salujõe.** Zooplankton as the indicator of ecological quality and fish predation in lake ecosystems. Tartu, 2009, 129 p.
164. **Ele Vahtmäe.** Mapping benthic habitat with remote sensing in optically complex coastal environments. Tartu, 2009, 109 p.

165. **Liisa Metsamaa.** Model-based assessment to improve the use of remote sensing in recognition and quantitative mapping of cyanobacteria. Tartu, 2009, 114 p.
166. **Pille Säälik.** The role of endocytosis in the protein transduction by cell-penetrating peptides. Tartu, 2009, 155 p.
167. **Lauri Peil.** Ribosome assembly factors in *Escherichia coli*. Tartu, 2009, 147 p.
168. **Lea Hallik.** Generality and specificity in light harvesting, carbon gain capacity and shade tolerance among plant functional groups. Tartu, 2009, 99 p.
169. **Mariliis Tark.** Mutagenic potential of DNA damage repair and tolerance mechanisms under starvation stress. Tartu, 2009, 191 p.
170. **Riinu Rannap.** Impacts of habitat loss and restoration on amphibian populations. Tartu, 2009, 117 p.
171. **Maarja Adojaan.** Molecular variation of HIV-1 and the use of this knowledge in vaccine development. Tartu, 2009, 95 p.
172. **Signe Altmäe.** Genomics and transcriptomics of human induced ovarian folliculogenesis. Tartu, 2010, 179 p.
173. **Triin Suvi.** Mycorrhizal fungi of native and introduced trees in the Seychelles Islands. Tartu, 2010, 107 p.
174. **Velda Lauringson.** Role of suspension feeding in a brackish-water coastal sea. Tartu, 2010, 123 p.
175. **Eero Talts.** Photosynthetic cyclic electron transport – measurement and variably proton-coupled mechanism. Tartu, 2010, 121 p.
176. **Mari Nelis.** Genetic structure of the Estonian population and genetic distance from other populations of European descent. Tartu, 2010, 97 p.
177. **Kaarel Krjutškov.** Arrayed Primer Extension-2 as a multiplex PCR-based method for nucleic acid variation analysis: method and applications. Tartu, 2010, 129 p.
178. **Egle Köster.** Morphological and genetical variation within species complexes: *Anthyllis vulneraria* s. l. and *Alchemilla vulgaris* (coll.). Tartu, 2010, 101 p.
179. **Erki Öunap.** Systematic studies on the subfamily Sterrhinae (Lepidoptera: Geometridae). Tartu, 2010, 111 p.
180. **Merike Jõesaar.** Diversity of key catabolic genes at degradation of phenol and *p*-cresol in pseudomonads. Tartu, 2010, 125 p.
181. **Kristjan Herkül.** Effects of physical disturbance and habitat-modifying species on sediment properties and benthic communities in the northern Baltic Sea. Tartu, 2010, 123 p.
182. **Arto Pulk.** Studies on bacterial ribosomes by chemical modification approaches. Tartu, 2010, 161 p.
183. **Maria Pöllupüü.** Ecological relations of cladocerans in a brackish-water ecosystem. Tartu, 2010, 126 p.
184. **Toomas Silla.** Study of the segregation mechanism of the Bovine Papillomavirus Type 1. Tartu, 2010, 188 p.

185. **Gyaneshwer Chaubey.** The demographic history of India: A perspective based on genetic evidence. Tartu, 2010, 184 p.
186. **Katrin Kepp.** Genes involved in cardiovascular traits: detection of genetic variation in Estonian and Czech populations. Tartu, 2010, 164 p.
187. **Virve Sõber.** The role of biotic interactions in plant reproductive performance. Tartu, 2010, 92 p.
188. **Kersti Kangro.** The response of phytoplankton community to the changes in nutrient loading. Tartu, 2010, 144 p.
189. **Joachim M. Gerhold.** Replication and Recombination of mitochondrial DNA in Yeast. Tartu, 2010, 120 p.
190. **Helen Tammert.** Ecological role of physiological and phylogenetic diversity in aquatic bacterial communities. Tartu, 2010, 140 p.
191. **Elle Rajandu.** Factors determining plant and lichen species diversity and composition in Estonian *Calamagrostis* and *Hepatica* site type forests. Tartu, 2010, 123 p.
192. **Paula Ann Kivistik.** ColR-ColS signalling system and transposition of Tn4652 in the adaptation of *Pseudomonas putida*. Tartu, 2010, 118 p.
193. **Siim Sõber.** Blood pressure genetics: from candidate genes to genome-wide association studies. Tartu, 2011, 120 p.
194. **Kalle Kipper.** Studies on the role of helix 69 of 23S rRNA in the factor-dependent stages of translation initiation, elongation, and termination. Tartu, 2011, 178 p.
195. **Triinu Siibak.** Effect of antibiotics on ribosome assembly is indirect. Tartu, 2011, 134 p.
196. **Tambet Tõnissoo.** Identification and molecular analysis of the role of guanine nucleotide exchange factor RIC-8 in mouse development and neural function. Tartu, 2011, 110 p.
197. **Helin Räägel.** Multiple faces of cell-penetrating peptides – their intracellular trafficking, stability and endosomal escape during protein transduction. Tartu, 2011, 161 p.
198. **Andres Jaanus.** Phytoplankton in Estonian coastal waters – variability, trends and response to environmental pressures. Tartu, 2011, 157 p.
199. **Tiit Nikopensius.** Genetic predisposition to nonsyndromic orofacial clefts. Tartu, 2011, 152 p.
200. **Signe Värv.** Studies on the mechanisms of RNA polymerase II-dependent transcription elongation. Tartu, 2011, 108 p.
201. **Kristjan Välk.** Gene expression profiling and genome-wide association studies of non-small cell lung cancer. Tartu, 2011, 98 p.
202. **Arno Pöllumäe.** Spatio-temporal patterns of native and invasive zooplankton species under changing climate and eutrophication conditions. Tartu, 2011, 153 p.
203. **Egle Tammeleht.** Brown bear (*Ursus arctos*) population structure, demographic processes and variations in diet in northern Eurasia. Tartu, 2011, 143 p.

205. **Teele Jairus.** Species composition and host preference among ectomycorrhizal fungi in Australian and African ecosystems. Tartu, 2011, 106 p.
206. **Kessy Abarenkov.** PlutoF – cloud database and computing services supporting biological research. Tartu, 2011, 125 p.
207. **Marina Grigorova.** Fine-scale genetic variation of follicle-stimulating hormone beta-subunit coding gene (*FSHB*) and its association with reproductive health. Tartu, 2011, 184 p.
208. **Anu Tiitsaar.** The effects of predation risk and habitat history on butterfly communities. Tartu, 2011, 97 p.
209. **Elin Sild.** Oxidative defences in immunoecological context: validation and application of assays for nitric oxide production and oxidative burst in a wild passerine. Tartu, 2011, 105 p.
210. **Irja Saar.** The taxonomy and phylogeny of the genera *Cystoderma* and *Cystodermella* (Agaricales, Fungi). Tartu, 2012, 167 p.
211. **Pauli Saag.** Natural variation in plumage bacterial assemblages in two wild breeding passserines. Tartu, 2012, 113 p.
212. **Aleksei Lulla.** Alphaviral nonstructural protease and its polyprotein substrate: arrangements for the perfect marriage. Tartu, 2012, 143 p.
213. **Mari Järve.** Different genetic perspectives on human history in Europe and the Caucasus: the stories told by uniparental and autosomal markers. Tartu, 2012, 119 p.
214. **Ott Scheler.** The application of tmRNA as a marker molecule in bacterial diagnostics using microarray and biosensor technology. Tartu, 2012, 93 p.
215. **Anna Balikova.** Studies on the functions of tumor-associated mucin-like leukosialin (CD43) in human cancer cells. Tartu, 2012, 129 p.
216. **Triinu Kõressaar.** Improvement of PCR primer design for detection of prokaryotic species. Tartu, 2012, 83 p.
217. **Tuul Sepp.** Hematological health state indices of greenfinches: sources of individual variation and responses to immune system manipulation. Tartu, 2012, 117 p.
218. **Rya Ero.** Modifier view of the bacterial ribosome. Tartu, 2012, 146 p.
219. **Mohammad Bahram.** Biogeography of ectomycorrhizal fungi across different spatial scales. Tartu, 2012, 165 p.
220. **Annely Lorents.** Overcoming the plasma membrane barrier: uptake of amphipathic cell-penetrating peptides induces influx of calcium ions and downstream responses. Tartu, 2012, 113 p.
221. **Katrin Männik.** Exploring the genomics of cognitive impairment: whole-genome SNP genotyping experience in Estonian patients and general population. Tartu, 2012, 171 p.
222. **Marko Prouš.** Taxonomy and phylogeny of the sawfly genus *Empria* (Hymenoptera, Tenthredinidae). Tartu, 2012, 192 p.
223. **Triinu Visnapuu.** Levansucrases encoded in the genome of *Pseudomonas syringae* pv. tomato DC3000: heterologous expression, biochemical characterization, mutational analysis and spectrum of polymerization products. Tartu, 2012, 160 p.

224. **Nele Tamberg.** Studies on Semliki Forest virus replication and pathogenesis. Tartu, 2012, 109 p.
225. **Tõnu Esko.** Novel applications of SNP array data in the analysis of the genetic structure of Europeans and in genetic association studies. Tartu, 2012, 149 p.
226. **Timo Arula.** Ecology of early life-history stages of herring *Clupea harengus membras* in the northeastern Baltic Sea. Tartu, 2012, 143 p.
227. **Inga Hiiesalu.** Belowground plant diversity and coexistence patterns in grassland ecosystems. Tartu, 2012, 130 p.
228. **Kadri Koorem.** The influence of abiotic and biotic factors on small-scale plant community patterns and regeneration in boreonemoral forest. Tartu, 2012, 114 p.
229. **Liis Andresen.** Regulation of virulence in plant-pathogenic pectobacteria. Tartu, 2012, 122 p.
230. **Kaupo Kohv.** The direct and indirect effects of management on boreal forest structure and field layer vegetation. Tartu, 2012, 124 p.
231. **Mart Jüssi.** Living on an edge: landlocked seals in changing climate. Tartu, 2012, 114 p.
232. **Riina Klais.** Phytoplankton trends in the Baltic Sea. Tartu, 2012, 136 p.
233. **Rauno Veeroja.** Effects of winter weather, population density and timing of reproduction on life-history traits and population dynamics of moose (*Alces alces*) in Estonia. Tartu, 2012, 92 p.
234. **Marju Keis.** Brown bear (*Ursus arctos*) phylogeography in northern Eurasia. Tartu, 2013, 142 p.
235. **Sergei Põlme.** Biogeography and ecology of *alnus-* associated ectomycorrhizal fungi – from regional to global scale. Tartu, 2013, 90 p.
236. **Liis Uusküla.** Placental gene expression in normal and complicated pregnancy. Tartu, 2013, 173 p.
237. **Marko Lõoke.** Studies on DNA replication initiation in *Saccharomyces cerevisiae*. Tartu, 2013, 112 p.
238. **Anne Aan.** Light- and nitrogen-use and biomass allocation along productivity gradients in multilayer plant communities. Tartu, 2013, 127 p.
239. **Heidi Tamm.** Comprehending phylogenetic diversity – case studies in three groups of ascomycetes. Tartu, 2013, 136 p.
240. **Liina Kangur.** High-Pressure Spectroscopy Study of Chromophore-Binding Hydrogen Bonds in Light-Harvesting Complexes of Photosynthetic Bacteria. Tartu, 2013, 150 p.
241. **Margus Leppik.** Substrate specificity of the multisite specific pseudouridine synthase RluD. Tartu, 2013, 111 p.
242. **Lauris Kaplinski.** The application of oligonucleotide hybridization model for PCR and microarray optimization. Tartu, 2013, 103 p.
243. **Merli Pärnoja.** Patterns of macrophyte distribution and productivity in coastal ecosystems: effect of abiotic and biotic forcing. Tartu, 2013, 155 p.
244. **Tõnu Margus.** Distribution and phylogeny of the bacterial translational GTPases and the MqsR/YgiT regulatory system. Tartu, 2013, 126 p.

245. **Pille Mänd.** Light use capacity and carbon and nitrogen budget of plants: remote assessment and physiological determinants. Tartu, 2013, 128 p.
246. **Mario Plaas.** Animal model of Wolfram Syndrome in mice: behavioural, biochemical and psychopharmacological characterization. Tartu, 2013, 144 p.
247. **Georgi Hudjašov.** Maps of mitochondrial DNA, Y-chromosome and tyrosinase variation in Eurasian and Oceanian populations. Tartu, 2013, 115 p.
248. **Mari Lepik.** Plasticity to light in herbaceous plants and its importance for community structure and diversity. Tartu, 2013, 102 p.
249. **Ede Leppik.** Diversity of lichens in semi-natural habitats of Estonia. Tartu, 2013, 151 p.
250. **Ülle Saks.** Arbuscular mycorrhizal fungal diversity patterns in boreo-nemoral forest ecosystems. Tartu, 2013, 151 p.
251. **Eneli Oitmaa.** Development of arrayed primer extension microarray assays for molecular diagnostic applications. Tartu, 2013, 147 p.
252. **Jekaterina Jutkina.** The horizontal gene pool for aromatics degradation: bacterial catabolic plasmids of the Baltic Sea aquatic system. Tartu, 2013, 121 p.
253. **Helen Vellau.** Reaction norms for size and age at maturity in insects: rules and exceptions. Tartu, 2014, 132 p.
254. **Randel Kreitsberg.** Using biomarkers in assessment of environmental contamination in fish – new perspectives. Tartu, 2014, 107 p.
255. **Krista Takkis.** Changes in plant species richness and population performance in response to habitat loss and fragmentation. Tartu, 2014, 141 p.
256. **Liina Nagirnaja.** Global and fine-scale genetic determinants of recurrent pregnancy loss. Tartu, 2014, 211 p.
257. **Triin Triisberg.** Factors influencing the re-vegetation of abandoned extracted peatlands in Estonia. Tartu, 2014, 133 p.
258. **Villu Soon.** A phylogenetic revision of the *Chrysis ignita* species group (Hymenoptera: Chrysidae) with emphasis on the northern European fauna. Tartu, 2014, 211 p.
259. **Andrei Nikonov.** RNA-Dependent RNA Polymerase Activity as a Basis for the Detection of Positive-Strand RNA Viruses by Vertebrate Host Cells. Tartu, 2014, 207 p.
260. **Eele Ōunapuu-Pikas.** Spatio-temporal variability of leaf hydraulic conductance in woody plants: ecophysiological consequences. Tartu, 2014, 135 p.
261. **Marju Männiste.** Physiological ecology of greenfinches: information content of feathers in relation to immune function and behavior. Tartu, 2014, 121 p.
262. **Katre Kets.** Effects of elevated concentrations of CO<sub>2</sub> and O<sub>3</sub> on leaf photosynthetic parameters in *Populus tremuloides*: diurnal, seasonal and interannual patterns. Tartu, 2014, 115 p.

263. **Külli Lokko**. Seasonal and spatial variability of zooplankton communities in relation to environmental parameters. Tartu, 2014, 129 p.
264. **Olga Žilina**. Chromosomal microarray analysis as diagnostic tool: Estonian experience. Tartu, 2014, 152 p.
265. **Kertu Lõhmus**. Colonisation ecology of forest-dwelling vascular plants and the conservation value of rural manor parks. Tartu, 2014, 111 p.
266. **Anu Aun**. Mitochondria as integral modulators of cellular signaling. Tartu, 2014, 167 p.
267. **Chandana Basu Mallick**. Genetics of adaptive traits and gender-specific demographic processes in South Asian populations. Tartu, 2014, 160 p.
268. **Riin Tamme**. The relationship between small-scale environmental heterogeneity and plant species diversity. Tartu, 2014, 130 p.
269. **Liina Remm**. Impacts of forest drainage on biodiversity and habitat quality: implications for sustainable management and conservation. Tartu, 2015, 126 p.
270. **Tiina Talve**. Genetic diversity and taxonomy within the genus *Rhinanthus*. Tartu, 2015, 106 p.
271. **Mehis Rohtla**. Otolith sclerochronological studies on migrations, spawning habitat preferences and age of freshwater fishes inhabiting the Baltic Sea. Tartu, 2015, 137 p.
272. **Alexey Reschikov**. The world fauna of the genus *Lathrolestes* (Hymenoptera, Ichneumonidae). Tartu, 2015, 247 p.
273. **Martin Pook**. Studies on artificial and extracellular matrix protein-rich surfaces as regulators of cell growth and differentiation. Tartu, 2015, 142 p.
274. **Mai Kukumägi**. Factors affecting soil respiration and its components in silver birch and Norway spruce stands. Tartu, 2015, 155 p.
275. **Helen Karu**. Development of ecosystems under human activity in the North-East Estonian industrial region: forests on post-mining sites and bogs. Tartu, 2015, 152 p.
276. **Hedi Peterson**. Exploiting high-throughput data for establishing relationships between genes. Tartu, 2015, 186 p.
277. **Priit Adler**. Analysis and visualisation of large scale microarray data. Tartu, 2015, 126 p.
278. **Aigar Niglas**. Effects of environmental factors on gas exchange in deciduous trees: focus on photosynthetic water-use efficiency. Tartu, 2015, 152 p.
279. **Silja Laht**. Classification and identification of conopeptides using profile hidden Markov models and position-specific scoring matrices. Tartu, 2015, 100 p.
280. **Martin Kesler**. Biological characteristics and restoration of Atlantic salmon *Salmo salar* populations in the Rivers of Northern Estonia. Tartu, 2015, 97 p.
281. **Pratyush Kumar Das**. Biochemical perspective on alphaviral nonstructural protein 2: a tale from multiple domains to enzymatic profiling. Tartu, 2015, 205 p

282. **Priit Palta.** Computational methods for DNA copy number detection. Tartu, 2015, 130 p.
283. **Julia Sidorenko.** Combating DNA damage and maintenance of genome integrity in pseudomonads. Tartu, 2015, 174 p.
284. **Anastasiia Kovtun-Kante.** Charophytes of Estonian inland and coastal waters: distribution and environmental preferences. Tartu, 2015, 97 p.
285. **Ly Lindman.** The ecology of protected butterfly species in Estonia. Tartu, 2015, 171 p.
286. **Jaanis Lodjak.** Association of Insulin-like Growth Factor I and Corticosterone with Nestling Growth and Fledging Success in Wild Passerines. Tartu, 2016, 113 p.
287. **Ann Kraut.** Conservation of Wood-Inhabiting Biodiversity – Semi-Natural Forests as an Opportunity. Tartu, 2016, 141 p.
288. **Tiit Örd.** Functions and regulation of the mammalian pseudokinase TRIB3. Tartu, 2016, 182. p.
289. **Kairi Käiro.** Biological Quality According to Macroinvertebrates in Streams of Estonia (Baltic Ecoregion of Europe): Effects of Human-induced Hydromorphological Changes. Tartu, 2016, 126 p.
290. **Leidi Laurimaa.** *Echinococcus multilocularis* and other zoonotic parasites in Estonian canids. Tartu, 2016, 144 p.
291. **Helerin Margus.** Characterization of cell-penetrating peptide/nucleic acid nanocomplexes and their cell-entry mechanisms. Tartu, 2016, 173 p.
292. **Kadri Runnel.** Fungal targets and tools for forest conservation. Tartu, 2016, 157 p.
293. **Urmo Võsa.** MicroRNAs in disease and health: aberrant regulation in lung cancer and association with genomic variation. Tartu, 2016, 163 p.
294. **Kristina Mäemets-Allas.** Studies on cell growth promoting AKT signalling pathway – a promising anti-cancer drug target. Tartu, 2016, 146 p.
295. **Janeli Viil.** Studies on cellular and molecular mechanisms that drive normal and regenerative processes in the liver and pathological processes in Dupuytren's contracture. Tartu, 2016, 175 p.
296. **Ene Kook.** Genetic diversity and evolution of *Pulmonaria angustifolia* L. and *Myosotis laxa sensu lato* (Boraginaceae). Tartu, 2016, 106 p.
297. **Kadri Peil.** RNA polymerase II-dependent transcription elongation in *Saccharomyces cerevisiae*. Tartu, 2016, 113 p.
298. **Katrin Ruisu.** The role of RIC8A in mouse development and its function in cell-matrix adhesion and actin cytoskeletal organisation. Tartu, 2016, 129 p.
299. **Janely Pae.** Translocation of cell-penetrating peptides across biological membranes and interactions with plasma membrane constituents. Tartu, 2016, 126 p.
300. **Argo Ronk.** Plant diversity patterns across Europe: observed and dark diversity. Tartu, 2016, 153 p.

301. **Kristiina Mark.** Diversification and species delimitation of lichenized fungi in selected groups of the family Parmeliaceae (Ascomycota). Tartu, 2016, 181 p.
302. **Jaak-Albert Metsoja.** Vegetation dynamics in floodplain meadows: influence of mowing and sediment application. Tartu, 2016, 140 p.
303. **Hedvig Tamman.** The GraTA toxin-antitoxin system of *Pseudomonas putida*: regulation and role in stress tolerance. Tartu, 2016, 154 p.
304. **Kadri Pärtel.** Application of ultrastructural and molecular data in the taxonomy of helotialean fungi. Tartu, 2016, 183 p.
305. **Maris Hindrikson.** Grey wolf (*Canis lupus*) populations in Estonia and Europe: genetic diversity, population structure and -processes, and hybridization between wolves and dogs. Tartu, 2016, 121 p.
306. **Polina Degtjarenko.** Impacts of alkaline dust pollution on biodiversity of plants and lichens: from communities to genetic diversity. Tartu, 2016, 126 p.
307. **Liina Pajusalu.** The effect of CO<sub>2</sub> enrichment on net photosynthesis of macrophytes in a brackish water environment. Tartu, 2016, 126 p.
308. **Stoyan Tankov.** Random walks in the stringent response. Tartu, 2016, 94 p.
309. **Liis Leitsalu.** Communicating genomic research results to population-based biobank participants. Tartu, 2016, 158 p.
310. **Richard Meitern.** Redox physiology of wild birds: validation and application of techniques for detecting oxidative stress. Tartu, 2016, 134 p.
311. **Kaie Lokk.** Comparative genome-wide DNA methylation studies of healthy human tissues and non-small cell lung cancer tissue. Tartu, 2016, 127 p.
312. **Mikhail Kurašin.** Processivity of cellulases and chitinases. Tartu, 2017, 132 p.
313. **Carmen Tali.** Scavenger receptors as a target for nucleic acid delivery with peptide vectors. Tartu, 2017, 155 p.
314. **Katarina Oganjan.** Distribution, feeding and habitat of benthic suspension feeders in a shallow coastal sea. Tartu, 2017, 132 p.
315. **Taavi Paal.** Immigration limitation of forest plants into wooded landscape corridors. Tartu, 2017, 145 p.
316. **Kadri Ōunap.** The Williams-Beuren syndrome chromosome region protein WBSCR22 is a ribosome biogenesis factor. Tartu, 2017, 135 p.
317. **Riin Tamm.** In-depth analysis of factors affecting variability in thiopurine methyltransferase activity. Tartu, 2017, 170 p.
318. **Keiu Kask.** The role of RIC8A in the development and regulation of mouse nervous system. Tartu, 2017, 184 p.
319. **Tiiia Möller.** Mapping and modelling of the spatial distribution of benthic macrovegetation in the NE Baltic Sea with a special focus on the eelgrass *Zostera marina* Linnaeus, 1753. Tartu, 2017, 162 p.
320. **Silva Kasela.** Genetic regulation of gene expression: detection of tissue- and cell type-specific effects. Tartu, 2017, 150 p.

321. **Karmen Süld.** Food habits, parasites and space use of the raccoon dog *Nyctereutes procyonoides*: the role of an alien species as a predator and vector of zoonotic diseases in Estonia. Tartu, 2017, p.
322. **Ragne Oja.** Consequences of supplementary feeding of wild boar – concern for ground-nesting birds and endoparasite infection. Tartu, 2017, 141 p.
323. **Riin Kont.** The acquisition of cellulose chain by a processive cellobiohydrolase. Tartu, 2017, 117 p.
324. **Liis Kasari.** Plant diversity of semi-natural grasslands: drivers, current status and conservation challenges. Tartu, 2017, 141 p.
325. **Sirgi Saar.** Belowground interactions: the roles of plant genetic relatedness, root exudation and soil legacies. Tartu, 2017, 113 p.
326. **Sten Anslan.** Molecular identification of Collembola and their fungal associates. Tartu, 2017, 125 p.