

ORIGINAL RESEARCH ARTICLE



Phenotypic Characterization of *EIF2AK4* Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically With Pulmonary Arterial Hypertension

Editorial, see p 2034

Charaka Hadinnapola,
MA, MB, BChir
et al

BACKGROUND: Pulmonary arterial hypertension (PAH) is a rare disease with an emerging genetic basis. Heterozygous mutations in the gene encoding the bone morphogenetic protein receptor type 2 (*BMPR2*) are the commonest genetic cause of PAH, whereas biallelic mutations in the eukaryotic translation initiation factor 2 alpha kinase 4 gene (*EIF2AK4*) are described in pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis. Here, we determine the frequency of these mutations and define the genotype-phenotype characteristics in a large cohort of patients diagnosed clinically with PAH.

METHODS: Whole-genome sequencing was performed on DNA from patients with idiopathic and heritable PAH and with pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis recruited to the National Institute of Health Research BioResource–Rare Diseases study. Heterozygous variants in *BMPR2* and biallelic *EIF2AK4* variants with a minor allele frequency of <1:10 000 in control data sets and predicted to be deleterious (by combined annotation-dependent depletion, PolyPhen-2, and *sorting intolerant from tolerant* predictions) were identified as potentially causal. Phenotype data from the time of diagnosis were also captured.

RESULTS: Eight hundred sixty-four patients with idiopathic or heritable PAH and 16 with pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis were recruited. Mutations in *BMPR2* were identified in 130 patients (14.8%). Biallelic mutations in *EIF2AK4* were identified in 5 patients with a clinical diagnosis of pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis. Furthermore, 9 patients with a clinical diagnosis of PAH carried biallelic *EIF2AK4* mutations. These patients had a reduced transfer coefficient for carbon monoxide (K_{CO} ; 33% [interquartile range, 30%–35%] predicted) and younger age at diagnosis (29 years; interquartile range, 23–38 years) and more interlobular septal thickening and mediastinal lymphadenopathy on computed tomography of the chest compared with patients with PAH without *EIF2AK4* mutations. However, radiological assessment alone could not accurately identify biallelic *EIF2AK4* mutation carriers. Patients with PAH with biallelic *EIF2AK4* mutations had a shorter survival.

CONCLUSIONS: Biallelic *EIF2AK4* mutations are found in patients classified clinically as having idiopathic and heritable PAH. These patients cannot be identified reliably by computed tomography, but a low K_{CO} and a young age at diagnosis suggests the underlying molecular diagnosis. Genetic testing can identify these misclassified patients, allowing appropriate management and early referral for lung transplantation.

The full author list is available on page 2031.

*Drs Gräf and Morrell contributed equally (see page 2031).

Correspondence to: Nicholas Morrell, MD, FRCP, FMedSci, Department of Medicine, University of Cambridge School of Clinical Medicine, Box 157, Addenbrooke's Hospital, Hills Rd, Cambridge, CB2 0QQ, UK. E-mail nmw23@cam.ac.uk

Sources of Funding, see page 2031

Key Words: genetics
■ hypertension, pulmonary
■ mutation ■ prognosis
■ pulmonary veno-occlusive disease

© 2017 The Authors. *Circulation* is published on behalf of the American Heart Association, Inc., by Wolters Kluwer Health, Inc. This is an open access article under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits use, distribution, and reproduction in any medium, provided that the original work is properly cited.

Clinical Perspective

What Is New?

- One percent of patients with a clinical diagnosis of pulmonary arterial hypertension (PAH) carry biallelic *EIF2AK4* mutations.
- Patients diagnosed clinically with PAH who had a transfer coefficient for carbon monoxide (Kco) <50% predicted and age of diagnosis <50 years were more likely to carry biallelic *EIF2AK4* mutations. The diagnostic yield for genetic testing in this group was 53%.
- Radiological assessment was unable to distinguish reliably between these patients and patients with idiopathic PAH.
- Histology from these patients may show predominantly pulmonary arteriopathy, with subtle involvement of the pulmonary veins and capillaries.
- Patients with PAH with biallelic *EIF2AK4* mutations had a worse prognosis compared with other patients with PAH.

What Are the Clinical Implications?

- Younger patients diagnosed with idiopathic PAH but with a low Kco have a high frequency of biallelic *EIF2AK4* mutations.
- Such patients should be reclassified as having pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis.
- Similar to patients with pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis, these patients have a poor prognosis compared with other patients with PAH.
- The spectrum of radiological and histological changes associated with biallelic *EIF2AK4* mutations is wider than previously assumed. The presence of only subtle or infrequent features associated with pulmonary veno-occlusive disease may lead to misclassification of these patients as having PAH.
- Genetic testing allows early identification of these patients, facilitating appropriate management.

Pulmonary arterial hypertension (PAH) is a heterogeneous and rare disorder that can be classified into idiopathic and heritable forms, associated with an underlying condition such as connective tissue disease or congenital heart disease or related to specific drugs and toxins.^{1,2} In addition, pulmonary veno-occlusive disease (PVOD) and pulmonary capillary hemangiomatosis (PCH) are even rarer forms of pulmonary hypertension that are grouped together with PAH under the current classification system.²

Clinical features described in patients with PVOD/PCH include a low transfer coefficient for carbon monoxide (Kco) and oxygen desaturation on exertion, as well as the presence of centrilobular ground glass opacification, interlobular septal thickening, and mediastinal lymph-

adenopathy on high-resolution computed tomography (CT) of the lung parenchyma.^{3,4} However, these clinical and radiological features have also been reported in idiopathic PAH.^{5–7} Consequently, the clinical distinction between PVOD/PCH and idiopathic PAH can be challenging. It has been estimated that 10% of patients with PVOD/PCH are misdiagnosed as having idiopathic PAH.^{8,9} The diagnosis of PVOD/PCH is often confirmed only postmortem or from explanted lungs by histology.

The histological features of PVOD/PCH typically include pulmonary venous obstructions and pulmonary capillary proliferation, although the distribution of these changes within the lung can be heterogeneous.^{10,11} Pulmonary artery smooth muscle hypertrophy and intimal hyperplasia, similar to the changes observed in other forms of PAH, may also be present. Furthermore, pulmonary venous changes have been reported in cases of idiopathic PAH, patients with scleroderma-associated PAH, and those with *BMPR2* mutations to varying extents.^{12,13}

A major advance in the molecular diagnosis of PVOD/PCH was the finding of biallelic mutations in the gene encoding the eukaryotic translation initiation factor 2 alpha kinase 4 (*EIF2AK4*) in both familial (100%) and sporadic (20% to 25%) cases of PVOD/PCH.^{14,15} *EIF2AK4* is an activator of the integrated stress response pathway and responds to environmental stresses, including amino acid deprivation, by phosphorylating the α subunit of eukaryotic translation initiation factor 2.^{11,16,17} These discoveries suggest that *EIF2AK4* mutations are specific to PVOD/PCH and that finding biallelic *EIF2AK4* mutations in a patient with pulmonary hypertension would be diagnostic of PVOD/PCH. Patients with PVOD/PCH have a poor prognosis and risk fatal pulmonary edema with the use of pulmonary artery vasodilator therapies.^{4,18–20} Consequently, early and accurate diagnosis is vital to guide clinical management.

Heterozygous mutations in the gene encoding the bone morphogenetic protein type 2 receptor (*BMPR2*) are the most common genetic cause of PAH. They are found in \approx 17% of individuals with idiopathic PAH and 82% with a family history of the disease.²¹ However, mutations in *BMPR2* have also been reported in patients with histologically proven PVOD.^{4,22–24} Thus, considerable uncertainty remains as to what extent the finding of *EIF2AK4* or *BMPR2* mutations reliably predicts the clinical phenotype and response to therapy in a population of patients with PAH.

Here, we report the genetic and phenotypic characteristics of patients assessed for *BMPR2* and *EIF2AK4* mutations through whole-genome sequencing within a large cohort (n=880) of patients with PAH recruited to the National Institute of Health Research (NIHR) BRIDGE study (BioResource–Rare Diseases) (Table 1 in the online-only Data Supplement). The frequency of mutations in other previously reported genes associated with PAH will be reported in a future publication. In this study, we identified and characterized patients with a clinical

and radiological diagnosis of idiopathic PAH who were found to possess biallelic *EIF2AK4* mutations. These patients had a low Kco and were diagnosed at a younger age compared with patients with idiopathic PAH without mutations in these genes. We show that, in common with patients diagnosed clinically with PVOD/PCH, patients with PAH with biallelic *EIF2AK4* mutations have a shorter survival. We conclude that clinical assessment alone is inadequate for the accurate diagnosis of PVOD/PCH. Clinical genetic testing in younger patients presenting clinically with PAH but with a low Kco will allow appropriate classification, leading to better risk stratification and management of these patients.

METHODS

Ethics Approval and Consent

UK patients (621 [70.6%]) were recruited prospectively to the BRIDGE study and provided written informed consent for genetic analysis and the capture of clinical data (BRIDGE study 13/EE/0325). In addition, the study included patients recruited retrospectively from non-UK centers (191 [21.7%]) and deceased UK patients (68 [7.7%]) if they had signed local tissue bank consent forms allowing genetic sequencing.

Explanted lung tissue from an individual undergoing lung transplantation for end-stage PAH was collected under Papworth Hospital Research Tissue Bank ethics (08/H0304/56).

Recruitment and Patients

The BRIDGE study is a prospective study recruiting both prevalent and incident patients with selected rare diseases. Recruitment to the BRIDGE PAH study started in January 2013, and the last patient included in this analysis was recruited on June 15, 2016. Patients with idiopathic PAH, heritable PAH, PVOD, and PCH, diagnosed according to international guidelines at specialist pulmonary hypertension centers in the United Kingdom, the Netherlands, and France, were recruited (Figure 1 and Table II in the online-only Data Supplement).² This included 14 patients with confirmed mutations in *BMPR2*.

Throughout this article, we classify patients recruited to the study as having idiopathic PAH or familial PAH on the

basis of the absence or presence of a family history of the disease. The term heritable PAH does not distinguish between patients with sporadic PAH with a mutation and patients with a mutation who have a family history. Therefore, the term heritable PAH is used only when referring to previous publications and guidelines.

Patients with other rare diseases and their unaffected relatives recruited to the BRIDGE study (Table III in the online-only Data Supplement) acted as control subjects without PAH for the genetic analysis.

Whole-Genome Sequencing and Variant Calling

Next-generation sequencing with 100- to 150-base pair (bp) paired-end sequencing was performed on DNA libraries created from genomic DNA with Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, CA).

Reads were aligned against the Genome Reference Consortium human genome (build 37), and variants were called with the Isaac Aligner and Variant Caller (version 2, Illumina Inc). Variants in *BMPR2* and *EIF2AK4* were extracted and annotated with the Ensembl Variant Effect Predictor version 84.²⁵ Deletions (resulting in the loss of >50 bp) were identified by applying Isaac Copy Number Variant Caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina). Further information is provided in the online-only Data Supplement.

Likely causal variants were identified on the basis of minor allele frequency and predicted deleteriousness. Variants were considered further if they had a minor allele frequency of <1 in 10 000 in unrelated BRIDGE control subjects without PAH and the ExAC database.²⁶ The rare variants that passed the minor allele frequency filtering were then assessed for deleteriousness. Variants were considered pathogenic on the basis of a combined annotation-dependent depletion score of ≥ 15 and PolyPhen-2 or *sorting intolerant from tolerant* predictions not classified as benign or tolerated, respectively.^{27–29}

Overrepresentation Analyses

For comparison of variant frequencies between disease and control groups, only variants from unrelated individuals were used. The PRIMUS software package was used to identify

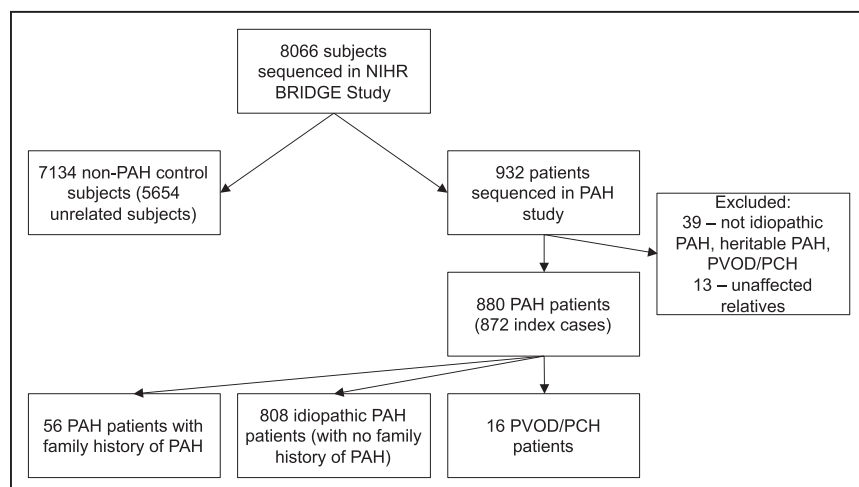


Figure 1. Subjects recruited to the National Institute of Health Research (NIHR) BRIDGE study (BioResource–Rare Diseases) and the clinical diagnostic categories of patients with pulmonary arterial hypertension (PAH) included in this study.

PVOD/PCH indicates pulmonary veno-occlusive disease/pulmonary capillary hemangiomatosis.

nonrelated individuals among both BRIDGE control subjects without PAH and patients with PAH.³⁰ The number of unrelated control subjects was maximized by including either patients with other rare diseases or their unaffected relatives. The frequency of rare and predicted deleterious heterozygous *EIF2AK4* variants in PAH index cases was also compared with publically available information in the ExAC database (<http://exac.broadinstitute.org>).²⁶ This analysis provides the maximum estimate of the frequency of heterozygous *EIF2AK4* variants in the ExAC database because variants in ExAC were assumed not to be in a compound heterozygous state.

Phenotypic Data Capture and CT Assessment

Paper and electronic patient records of patients with PAH were reviewed to capture demographic and phenotypic variables from the time of diagnosis and follow-up. Survival data for UK patients were obtained from recruiting centers through the NHS National Spine and local databases. Anonymized information was captured securely online with the free OpenClinica software, adapted for data capture specific to PAH.

CT images of the chest, when available, were reviewed independently by 2 cardiothoracic radiologists (A.S. and N.S.) with specialist imaging experience in pulmonary hypertension who were blinded to the underlying diagnoses with a customized proforma. Further information is provided in the supplemental materials and [Tables IV and V in the online-only Data Supplement](#).

Statistical Analysis

Statistical analysis was performed in R (www.r-project.org). Further information is provided in the [online-only Data Supplement](#).

Semiparametric Cox proportional hazard models were used to assess survival between groups with the survival package in R. Time from diagnosis to both death and death or transplantation was assessed. Age at diagnosis and sex were used as covariates in the models. To avoid immortal time bias arising from the inclusion of retrospectively recruited patients and prevalent patients, a sensitivity analysis was conducted. In this analysis, only prospectively recruited patients from the UK were included, and patients entered the risk set only from the time they consented to the study. Further information is provided in the [online-only Data Supplement](#).

RESULTS

Study Patients

Whole-genome sequencing was performed on 932 patients recruited to the NIHR BRIDGE PAH study and 7134 control subjects without PAH recruited to other NIHR BRIDGE study cohorts. Fifty-two patients were excluded from further analysis because they did not have a clinical diagnosis of idiopathic PAH, heritable PAH, PVOD, or PCH (Figure 1). The remaining 880 patients (of whom 872 were defined as unrelated index cases)

consisted of 16 patients (1.8%) with a clinical diagnosis of PVOD/PCH, 56 (6.4%) with PAH and a family history of the disease (referred to as familial PAH), and 808 (91.8%) with idiopathic PAH and no known family history. One of the 16 patients with a clinical diagnosis of PVOD/PCH had an affected sister, whereas the remainder had the sporadic form of the disease.

BMPR2 Mutations in the PAH Cohort

Rare and predicted deleterious *BMPR2* mutations (single-nucleotide variants, indels, and larger deletions) were found in 41 patients (73.2%) with familial PAH and 89 patients (11.0%) with idiopathic PAH. No *BMPR2* mutations were found in patients with a clinical diagnosis of PVOD/PCH.

Rare and Predicted Deleterious *EIF2AK4* Variants in the PAH Cohort

Sixty-nine rare and predicted deleterious *EIF2AK4* single-nucleotide variants and indels were present in the NIHR BRIDGE study. No large deletions were found that affected the *EIF2AK4* gene locus. The variants are summarized in [Table VI in the online-only Data Supplement](#). Five of the 16 patients (31.3%) with clinically diagnosed PVOD/PCH carried biallelic *EIF2AK4* mutations (2 homozygotes and 3 compound heterozygotes).

Twenty-five *EIF2AK4* variants were also found in 19 patients (2.2%) diagnosed clinically with PAH, in whom there was no clinical suspicion of PVOD/PCH (5 homozygotes, 4 compound heterozygotes, and 10 heterozygotes; [Table VII in the online-only Data Supplement](#)). One of these patients with a homozygous *EIF2AK4* mutation (c.3097C>T creating a premature stop codon) had a sister who had died of PAH. There was no reported family history of PVOD/PCH.

The remaining rare *EIF2AK4* variants were found in a heterozygous state in 36 control subjects (0.5%). Four of these variants appeared in >1 control subject without PAH, and none were shared with patients with PAH.

Overrepresentation of Rare Heterozygous *EIF2AK4* Variants in Patients With Idiopathic PAH Compared With Control Subjects

The proportion of patients with a clinical diagnosis of idiopathic PAH carrying heterozygous rare *EIF2AK4* variants (1.2%) was significantly greater than the percentage of control subjects without PAH (0.5%; $P=0.030$). A similar overrepresentation in patients with idiopathic PAH was observed compared with allele frequencies in the ExAC database (0.6%; $P=0.042$). Two patients with idiopathic PAH with heterozygous rare

EIF2AK4 variants also carried a rare and predicted deleterious *BMPR2* mutation.

Phenotype of Patients With a Clinical Diagnosis of PAH and Biallelic *EIF2AK4* Mutations

Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations presented at a younger age (median, 29 years; interquartile range, 23–38 years) compared with patients without mutations in the PAH associated genes (51 years; IQR, 37–65 years; $P=0.024$; Table 1). Mean pulmonary artery pressure, cardiac output, and pulmonary vascular resistance were not significantly different between patients with PAH with biallelic *EIF2AK4* mutations and the other groups. As previously reported, hemodynamic variables were significantly worse in patients with *BMPR2* mutations compared with those without any mutations in these genes.

The patients with PAH with biallelic *EIF2AK4* mutations exhibited a reduced Kco (33% [IQR, 30%–35%] predicted) compared with *BMPR2* mutation carriers (81% [IQR, 73%–92%] predicted; $P<0.001$) and patients with PAH with no identified mutation (71% [IQR, 51%–85%] predicted; $P=0.001$). Patients with PAH with biallelic *EIF2AK4* mutations had no obstructive or restrictive deficit on spirometry. These differences remained after the exclusion of patients with abnormal

spirometry in the other groups (forced expiratory volume in 1 second of expiration [FEV_1] $<80\%$ or forced vital capacity [FVC] $<80\%$; Table VIII in the online-only Data Supplement).

Digital clubbing was overrepresented among patients with biallelic *EIF2AK4* mutations diagnosed clinically with PAH (42%; $P=0.002$). Eleven percent of patients with a clinical diagnosis of PVOD were clubbed.

Only 1 patient with a heterozygous rare and predicted deleterious *EIF2AK4* variant (c.2516T>C) had a reduced Kco (54% predicted) with normal spirometry (FEV_1 , 102% predicted; FVC, 98% predicted; and total lung capacity, 100% predicted). There was mild paraseptal emphysema on thoracic CT ($<5\%$ of the lung parenchyma affected). This patient, a 44-year-old white man diagnosed with idiopathic PAH, also carried a rare and deleterious *BMPR2* splice acceptor mutation (c.853-2A>G).

We questioned whether Kco was a predictor of biallelic *EIF2AK4* mutations in the wider cohort. However, among patients with PAH with no mutations and normal spirometry ($n=255$), a reduced Kco ($<50\%$ predicted) was present in 65 patients (25.5%). In these patients with a reduced Kco and preserved spirometry, 90.8% were >50 years old at diagnosis, and 69.2% had a history of coronary artery disease, left ventricular dysfunction, or cardiovascular risk factors (diabetes mellitus, systemic hypertension, or hyperlipidemia).

Table 1. Phenotypic Summary of *EIF2AK4* Variant Carriers: Patients With a Clinical Diagnosis of PAH and Biallelic *EIF2AK4* Mutations Were Younger at Diagnosis and Had a Significantly Reduced Kco Compared With Other Groups

	PAH Patients With <i>BMPR2</i> Mutations*	PAH Patients With No Mutations in PAH-Associated Genes	PAH Patients With <i>EIF2AK4</i> Heterozygous Variants	PAH Patients With Biallelic <i>EIF2AK4</i> Mutations	Patients With PVOD/PCH	P Value
n	130	704	8	9	16	
Age, y	39 (31–52)	51 (37–65)	49 (36–67)	29 (23–38)	57 (41–69)	<0.001
Female, n (%)	85 (65.4)	494 (70.2)	7 (87.5)	4 (44.4)	9 (56.2)	0.180
White, n (%)	108 (83.1)	551 (78.5)	5 (62.5)	2 (22.2)	13 (81.2)	0.002
Digital clubbing, n (%)	6 (9.7)	10 (3.4)	0 (0)	3 (42.9)	1 (11.1)	0.002
BMI, kg/m ²	28 (24–33)	28 (24–33)	26 (23–28)	24 (20–27)	27 (24–31)	0.216
mPAP, mm Hg	57 (51–69)	52 (44–61)	44 (42–52)	52 (46–65)	48 (40–58)	<0.001
CO, L/min	3 (3–4)	4 (3–5)	3 (3–5)	5 (3–6)	4 (3–4)	<0.001
PVR, WU	15 (11–20)	10 (7–14)	9 (6–10)	9 (8–13)	10 (9–12)	<0.001
Vasoresponders, n (%)	0 (0)	28 (17.5)	0 (0)	0 (0)	0 (0)	0.011
FEV_1 , % predicted	90 (78–99)	84 (72–95)	83 (71–94)	94 (85–100)	85 (70–95)	0.031
FVC, % predicted	97 (86–109)	95 (82–106)	96 (75–98)	100 (86–119)	97 (81–103)	0.310
KCO, % predicted	81 (73–92)	71 (51–85)	81 (72–95)	33 (30–35)	37 (32–47)	<0.001
Resting S_{aO_2} , %	96 (94–97)	96 (93–97)	98 (98–98)	91 (90–94)	94 (91–95)	0.010
S_{aO_2} after walk test, %	94 (90–97)	92 (85–96)	94 (84–96)	78 (75–82)	88 (85–89)	<0.001

BMI indicates body mass index; CO, cardiac output; FEV_1 , forced expiratory volume in 1 second; FVC, forced vital capacity; Kco, transfer coefficient for carbon monoxide; mPAP, mean pulmonary artery pressure; PAH, pulmonary arterial hypertension; PCH, pulmonary capillary hemangiomas; PVOD, pulmonary veno-occlusive disease; PVR, pulmonary vascular resistance; and S_{aO_2} , arterial oxygen saturation.

*Also includes the 2 patients with a heterozygous *EIF2AK4* variant and a *BMPR2* variant. Data presented as median (interquartile range) unless indicated. Percentages were calculated from the number of patients for whom data were available as the denominator.

Given the high prevalence of a low Kco with preserved spirometry in the wider cohort, we restricted an analysis to patients <50 years of age who at the time of diagnosis had normal spirometry (n=164). Even in this group, a significant proportion (n=15, 9.1%) had a Kco <50% predicted (Figure 2). Eight of these 15 patients carried biallelic *EIF2AK4* mutations. One patient with biallelic *EIF2AK4* mutations was 70 years of age at diagnosis and subsequently did not meet this cutoff.

Among patients with normal spirometry, the presence of a Kco <50% predicted and age at diagnosis <50 years had a high sensitivity (0.889) and specificity (0.977) for identifying patients who carry biallelic *EIF2AK4* mutations; the positive predictive value was low (0.533). Nevertheless, in terms of the diagnostic yield, although genetic testing for biallelic *EIF2AK4* mutations in the entire cohort of patients diagnosed clinically with PAH yielded a 1% detection rate, the presence of biallelic *EIF2AK4* mutations in patients with PAH with a Kco <50% predicted with normal spirometry and <50 years of age at diagnosis was 53%.

CT Features of *EIF2AK4* Mutation Carriers

Centrilobular ground glass opacification extent, mediastinal lymphadenopathy, and interlobular septal thickening are considered suggestive of PVOD/PCH. However, we found subtle or gross centrilobular ground glass opacification in 38% of patients diagnosed clinically with PAH and carrying no mutations (n=21) and 67% of patients with PAH with *BMPR2* mutations (n=21). This was not significantly different compared with patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations (86%, n=7) and patients with a clinical diagnosis

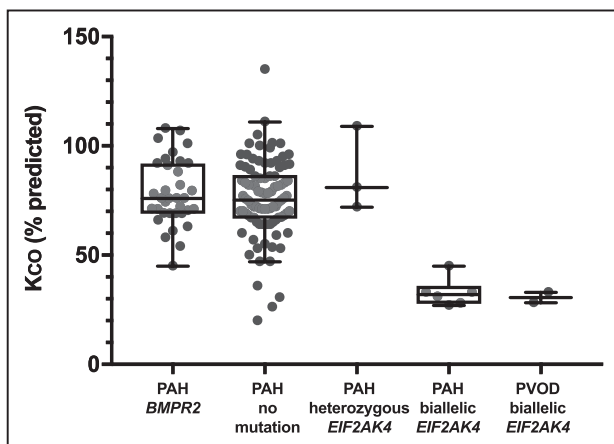


Figure 2. The transfer coefficient for carbon monoxide (Kco) is influenced by genotype in pulmonary arterial hypertension (PAH).

Patients with forced expiratory volume in 1 second of expiration <80% predicted and forced vital capacity <80% predicted and diagnosed with PAH or pulmonary veno-occlusive disease (PVOD)/pulmonary capillary hemangiomatosis after 50 years of age were excluded from the plot.

nosis of PVOD (50%, n=14). Gross interlobular septal thickening and mediastinal lymphadenopathy were significantly more frequent among patients with PAH and biallelic *EIF2AK4* mutations (29% and 57%, respectively) and those with PVOD (64% and 79%) compared with patients with PAH and no mutation (5% and 0%) or *BMPR2* mutations (5% and 10%). A radiological suspicion of PVOD/PCH was raised in 71% of those with PVOD, 57% of patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations, 14% of patients with PAH with no mutation, and 5% of those with *BMPR2* mutations (Table 2).

A further CT analysis comparing patients with biallelic *EIF2AK4* mutations (with a clinical diagnosis of PVOD/PCH or PAH; n=11) and those with a clinical diagnosis of PVOD but not carrying biallelic *EIF2AK4* mutations (n=10) was made (Table IX in the online-only Data Supplement). Patients with biallelic *EIF2AK4* mutations were younger at diagnosis (27 years; IQR, 23–34 years) compared with those with PVOD and no *EIF2AK4* mutations (68 years; IQR, 64–72 years; $P=0.001$). The patients with biallelic *EIF2AK4* mutations also had a lower Kco (32% [IQR, 29%–33%] predicted) compared with patients with PVOD and no *EIF2AK4* mutations (41.4% [IQR, 37%–54%] predicted; $P=0.013$). Centrilobular ground glass opacification appeared more extensive in those with biallelic *EIF2AK4* mutations (82%) compared with those without a mutation (10%; $P=0.012$). However, pleural effusions were more common among those without a mutation (40%) compared with patients with biallelic *EIF2AK4* mutations (0%; $P=0.035$). This may suggest that patients with biallelic *EIF2AK4* mutations have a distinct radiological phenotype compared with patients with PVOD and no biallelic *EIF2AK4* mutations.

Response to Pulmonary Artery Vasodilator Therapies

The response to pulmonary artery vasodilator therapies at 1 and 3 years was assessed for patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations and the other patients with PAH included in the CT analysis. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations did not improve their functional class at either 1 or 3 years after diagnosis, unlike the other PAH groups (Table X in the online-only Data Supplement).

Histological Features of a Biallelic *EIF2AK4* Mutation Carrier

The explanted lungs of 1 patient diagnosed with idiopathic PAH but found to have a homozygous *EIF2AK4* missense mutation (c.1795G>C, p.G599R) were as-

Table 2. Radiological Features and Consensus Radiological Diagnosis of Patients With PAH in the CT Substudy

	Group	Patients With PAH With <i>BMPR2</i> Mutations (n=21), n (%)	Patients With PAH With No Mutations in the Previously Reported PAH Genes (n=21), n (%)	Patients With PAH With Heterozygous <i>EIF2AK4</i> Variants (n=4), n (%)	Patients With PAH With Biallelic <i>EIF2AK4</i> Mutations (n=7), n (%)	Patients With PVOD (n=14), n (%)	P Value
Centrilobular ground glass opacification density	None	7 (33.3)	13 (61.9)	2 (50.0)	1 (14.3)	7 (50.0)	0.122
	Subtle	12 (57.1)	5 (23.8)	0 (0.0)	2 (28.6)	3 (21.4)	
	Present	2 (9.5)	3 (14.3)	2 (50.0)	4 (57.1)	4 (28.6)	
Centrilobular ground glass opacification extent	None	8 (38.1)	13 (61.9)	2 (50.0)	1 (14.3)	8 (57.1)	0.077
	<5%	0 (0.0)	3 (14.3)	0 (0.0)	1 (14.3)	1 (7.1)	
	5%–25%	2 (9.5)	0 (0.0)	1 (25.0)	2 (28.6)	1 (7.1)	
	25%–50%	2 (9.5)	4 (19.0)	0 (0.0)	0 (0.0)	2 (14.3)	
	50%–75%	5 (23.8)	1 (4.8)	0 (0.0)	2 (28.6)	0 (0.0)	
	75%–100%	4 (19.0)	0 (0.0)	1 (25.0)	1 (14.3)	2 (14.3)	
Interlobular septal thickening	None	17 (81.0)	18 (85.7)	4 (100.0)	5 (71.4)	4 (28.6)	0.001
	Subtle	3 (14.3)	2 (9.5)	0 (0.0)	0 (0.0)	1 (7.1)	
	Present	1 (4.8)	1 (4.8)	0 (0.0)	2 (28.6)	9 (64.3)	
Mediastinal lymphadenopathy	None	19 (90.5)	21 (100.0)	4 (100.0)	3 (42.9)	3 (21.4)	<0.001
	Present	2 (9.5)	0 (0.0)	0 (0.0)	4 (57.1)	11 (78.6)	
Pleural effusion	None	17 (81.0)	21 (100.0)	3 (75.0)	7 (100.0)	10 (71.4)	0.048
	Small	4 (19.0)	0 (0.0)	1 (25.0)	0 (0.0)	4 (28.6)	
Neovascularity	None	12 (57.1)	18 (85.7)	4 (100.0)	6 (85.7)	13 (92.9)	0.077
	Present	9 (42.9)	3 (14.3)	0 (0.0)	1 (14.3)	1 (7.1)	
CT diagnosis	PAH	20 (95.2)	18 (85.7)	3 (75.0)	3 (42.9)	4 (28.6)	
	Possible PVOD/PCH	1 (4.8)	3 (14.3)	1 (25.0)	4 (57.1)	10 (71.4)	

CT indicates computed tomography; PAH, pulmonary arterial hypertension; and PVOD, pulmonary veno-occlusive disease.

sessed. The predominant histological feature was pulmonary arterial vasculopathy. The pulmonary arteries predominantly showed concentric and eccentric intimal fibrosis. No plexiform lesions were observed. Although infrequent, there was some fibrosis of the septal veins and venules, some of which were nearly completely occluded. Although there was evidence of capillary congestion, no capillary hemangiomatosis was observed (Figure 3). The missense variant carried by this patient was not reported in the ExAC database, occurs in a conserved area of the genome (Genomic Evolutionary Rate Profiling score, 5.5), and was predicted to be deleterious (combined annotation-dependent depletion score, 32; PolyPhen-2 prediction of “probably damaging [1],” *sorting intolerant from tolerant* prediction of “deleterious [0]”). The same homozygous mutation was also found in a second unrelated patient with a clinical diagnosis of idiopathic PAH.

Impact of Genotype on Survival

Eight hundred fifty-eight patients were included in the Cox proportional hazards model (Table XI and Figure I in the online-only Data Supplement). Patients diagnosed clinically as having PAH with biallelic *EIF2AK4*

mutations had a shorter survival time from diagnosis compared with the *BMPR2* mutation carriers ($P<0.001$) and those without any variants in PAH-associated genes ($P<0.001$). Age ($P<0.001$) and sex ($P=0.001$) also had a significant effect on survival, with male sex and older age at diagnosis associated with shorter survival in the model. Similar results were obtained in the assessment of time to death or transplantation (Table XII in the online-only Data Supplement). In the sensitivity analysis, including only prospectively recruited UK patients, only 2 events occurred in the biallelic *EIF2AK4* group. Thus, no significant difference was observed in mortality between patients diagnosed clinically as having PAH with biallelic *EIF2AK4* mutations and patients with *BMPR2* mutations ($P=0.215$) or patients without any variants in PAH-associated genes ($P=0.282$; Table XIII in the online-only Data Supplement).

DISCUSSION

This is the first study to analyze the frequency of *EIF2AK4* rare variation in a large cohort of patients with PAH and to make detailed phenotypic and radiological assessments. Previously, the presence of biallelic *EIF2AK4* mutations was reported in patients with a clear

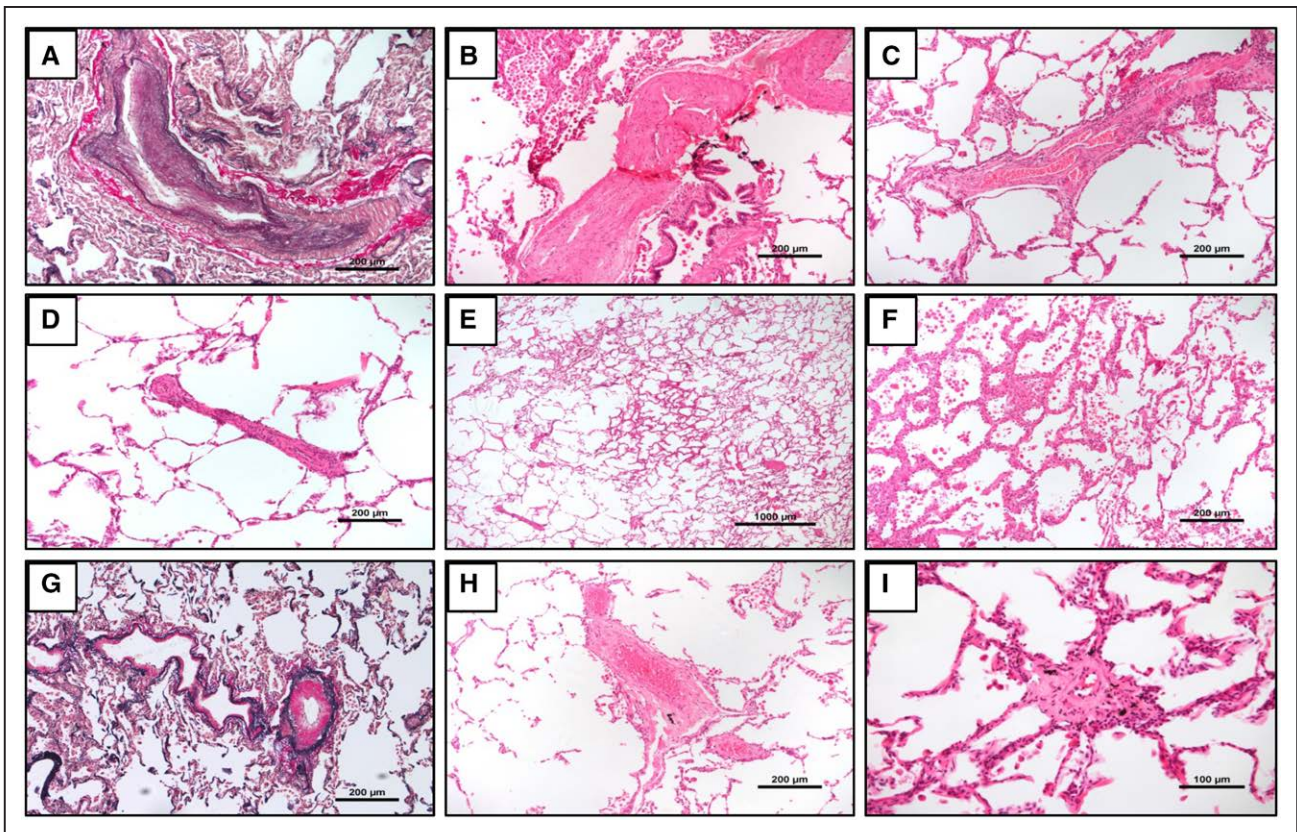


Figure 3. Representative histopathologic images from 1 patient with clinically diagnosed idiopathic pulmonary arterial hypertension (PAH) but found to have a rare (not reported in the ExAC database) and predicted deleterious (combined annotation-dependent depletion score, 32) homozygous *EIF2AK4* missense variant (c.1795G>C).

The patient was of Pakistani origin and did not have a family history of PAH or pulmonary veno-occlusive disease (PVOD). At presentation, he was 22 years old and had a reduced transfer coefficient for carbon monoxide (K_{CO} ; 31% predicted) despite preserved spirometry. High-resolution computed tomography of his chest showed subtle but extensive (50%–75% involvement) ground glass opacification. No interlobular septal thickening or mediastinal lymphadenopathy was observed. No suspicion of PVOD/pulmonary capillary hemangiomatosis (PCH) was raised from the radiological appearances. Histopathology was reviewed by 2 independent pathologists, each confirming the predominant histological pattern to be one of pulmonary arterial vasculopathy. The pulmonary arteries showed eccentric and concentric intimal fibrosis and medial hypertrophy (A and B), as well as some lesions with features of recanalized thrombus (C). Several concentrically muscularized arterioles were also observed (D). No complex plexiform lesions were present. There was patchy thickening of the alveolar septa with capillary congestion and pigmented intra-alveolar macrophages similar to PCH (E and F). Venous remodeling was difficult to trace and infrequent but present. Fibrous thickening of the intima in septal veins (G and I) and a microvessel (H).

clinical diagnosis of PVOD/PCH and a large kindred and a single family with a possible diagnosis of PAH.^{20,31,32} As expected, we identified a high frequency of biallelic *EIF2AK4* mutations in patients with a clear clinical presentation of PVOD/PCH. However, we also found biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH.

The discovery of biallelic *EIF2AK4* mutations in PVOD/PCH raised the possibility of rapid molecular diagnosis in the majority of patients with familial and up to 25% of patients with sporadic PVOD/PCH.^{14,15} In the present study, the presence of biallelic *EIF2AK4* mutations was associated with a poor prognosis, even in patients who have a clinical diagnosis of PAH and who did not develop pulmonary edema in response to pulmonary

artery vasodilator therapies. Therefore, early identification of these patients through genetic testing may prompt early referral for lung transplantation similar to patients with clinically diagnosed PVOD/PCH.¹⁸

The presence of biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH raises the question whether *EIF2AK4* mutations can cause classic idiopathic PAH or whether there are cases of PVOD/PCH caused by *EIF2AK4* mutations that are wrongly classified even by expert centers. We further show that phenotypic, radiological, and histological assessments can be difficult to interpret. The presence of subtle or infrequent features may lead to an incorrect diagnosis of PAH in patients with biallelic *EIF2AK4* mutations. This study suggests that patients with pathogenic biallelic

EIF2AK4 mutations may present with a spectrum of phenotypic, radiological, and histological features that can overlap with PAH.

Patients with PAH with biallelic *EIF2AK4* mutations demonstrated a reduced Kco despite normal spirometry, which is characteristic of patients with PVOD/PCH. The reduced Kco likely reflects widespread reduction in alveolar gas exchange caused by endothelial proliferation and patchy thickening of the blood-gas barrier by the process of capillary hemangiomas. Ultrastructural thickening of the capillary basement membrane may also play a role.³³ In keeping with previous reports in PVOD/PCH, we also show that patients with PAH with biallelic mutations in *EIF2AK4* are younger at diagnosis than patients with either *BMP2* mutations or no known mutation.^{14,20} However, the presence of these characteristic features has a low positive predictive value for the identification of patients with biallelic *EIF2AK4* mutations.

In contrast to previous descriptions of patients with PVOD, none of the patients with clinically diagnosed PAH and biallelic *EIF2AK4* mutations developed pulmonary edema in response to pulmonary artery vasodilator therapies. For example, intravenous prostanoids were used in 50% of these patients. In patients with classic PVOD, pulmonary edema with intravenous prostanoids has been reported in up to 44% of patients after a median treatment duration of just 9 days.⁴ Presumably, the extent and severity of the pulmonary venous involvement in these patients might underlie the differing responses to prostanoids.

It is generally considered that high-resolution CT imaging is a useful noninvasive test to assist in the diagnosis of suspected PVOD/PCH.¹¹ Although there was an increased prevalence of mediastinal lymphadenopathy and interlobular septal thickening in patients with PAH with biallelic *EIF2AK4* mutations, we found that radiological features at the time of diagnosis could not accurately determine the underlying genotype.⁶ The differing radiological features of all patients with biallelic *EIF2AK4* mutations compared with patients with PVOD without mutations is of interest. This may reflect differences between the younger-onset genetic cases of PVOD compared with the predominantly older group of patients without *EIF2AK4* mutations in whom other nongenetic factors such as exposure to inorganic solvents may play an important role.³⁴

Histological examination (usually postmortem or from explanted lungs) is often considered essential for diagnostic confirmation of PVOD/PCH but may be confounded by the heterogeneous nature of vascular pathology.³⁵ Surgical biopsy of the lung in patients with severe PAH is contraindicated, and a limitation of this study is that lung tissue from only 1 patient with biallelic *EIF2AK4* mutations was available for analysis. This patient had a rare and predicted deleterious homozy-

gous missense mutation in *EIF2AK4*. The predominant feature on assessment of the explanted lung tissue was pulmonary arteriopathy, as usually seen in PAH. Although only infrequent, fibrosis of the septal venules and the possible presence of siderophages in the alveolar space were observed. These features are found in patients with PVOD/PCH. This case supports the hypothesis that patients with biallelic *EIF2AK4* mutations may present with a spectrum of venous and arterial involvement.

There are increasing reports of phenotypic, radiological, and histological similarities between PAH and PVOD/PCH.^{6,12,13} Tenorio et al³¹ reported a homozygous missense mutation in *EIF2AK4* in a large kindred of Iberian Romani with apparent heritable PAH. This kindred is likely to have PVOD/PCH because these diagnoses were not confirmed histologically and PVOD was suspected in half the patients. More recently, Best et al³² also report 2 sisters with apparent heritable PAH-carrying biallelic *EIF2AK4* mutations. These patients also had a reduced Kco but had not had high-resolution CT assessment of their lung parenchyma, which may have altered their clinical diagnosis. Taken together, these previous reports are compatible with the findings in this larger cohort that patients with a clinical presentation of idiopathic or heritable PAH may in fact have underlying PVOD/PCH as determined by genetic analysis.

A strength of this study is the centralized reporting of radiographic features. However, the data collection was retrospective and incomplete in some cases. Assessing rare diseases such as PAH and PVOD/PCH with a prospective study recruiting incident cases would take a prohibitively long time. This is especially true for the assessment of survival and response to therapy. In this study including prevalent and retrospectively recruited patients, we demonstrated a worse prognosis in patients with a clinical diagnosis of PAH and biallelic *EIF2AK4*. However, the inclusion of prevalent and retrospectively recruited patients can introduce bias such as immortal time bias, when there are long periods between diagnosis and enrollment in the study. The effect of immortal time bias and other confounders such as the inclusion of prevalent and incident cases can be difficult to predict. All groups are likely to include patients who died before study enrollment and thus would not feature in any analysis. When we attempted to eliminate these sources of bias in a sensitivity analysis restricted to prospectively recruited patients from the United Kingdom, the study did not have sufficient power to show a difference in survival between different genotypes. Further studies of survival and response to therapy are needed to definitively show whether misclassified patients with PAH with biallelic *EIF2AK4* mutations and patients with classic PVOD with these mutations have a similarly poor prognosis.

The genetic architecture of idiopathic and heritable PAH remains to be fully elucidated. Ongoing analysis of whole-genome sequence data in our cohort is likely to reveal novel rare variation underlying this condition. Mutations in *BMPR2* account for ≈17% of cases of idiopathic PAH, and other known PAH genes account for ≈1% to 2% of all cases.^{21,36} In the present study, *BMPR2* mutations were found in 11% of patients without a family history of PAH. It is worth noting that patients with the sporadic form of the disease with no reported family history represent a higher burden of *BMPR2* mutations (n=89) compared with those with a family history (n=49). This has important implications for clinical genetic testing in patients with sporadic and familial disease.

In previous studies, mutations in both *EIF2AK4* alleles are required to cause PVOD and PCH.^{14,15} In autosomal recessive disorders, it is unusual for the heterozygous state to manifest the disease phenotype, and heterozygous *EIF2AK4* variants thus would not be expected to be pathogenic. In this study, we found a significant overrepresentation of heterozygous rare and predicted deleterious *EIF2AK4* variants in patients with PAH compared with control subjects and report 2 patients with rare variants in both *BMPR2* and *EIF2AK4*. Recently, the possibility that heterozygous *EIF2AK4* variants influence the penetrance of *BMPR2* mutations has been raised in a single family with PAH.³⁷ Further studies are required to determine whether heterozygous *EIF2AK4* variants contribute to pathogenesis in PAH.

CONCLUSIONS

We demonstrate that biallelic *EIF2AK4* mutations are found in patients diagnosed clinically with idiopathic and familial PAH. These patients may have subtle features suggestive of PVOD/PCH on close inspection and are likely to have underlying PVOD/PCH. The spectrum of phenotypic, radiological, and histological features found in patients with biallelic *EIF2AK4* mutations made by current clinical assessments is wider and less clear-cut than previously recognized. This may lead to misclassification of patients as having PAH rather than PVOD and hinders accurate risk stratification. Ascertaining the *EIF2AK4* mutation status of patients through clinical genetic testing provides additional information to aid risk stratification and to guide management. In a young patient presenting with apparent PAH, the presence of a low Kco with normal spirometry strongly suggests the presence of underlying biallelic *EIF2AK4* mutations. Patients with an apparent clinical diagnosis of PAH and biallelic *EIF2AK4* mutations have a worse prognosis compared with patients with *BMPR2* mutations and those without these mutations. Clinical genetic testing should aid identification of this high-risk

group and facilitate early referral for lung transplantation and appropriate management.

AUTHORS

Charaka Hadinnapola, MA, MB, BChir; Marta Bleda, PhD; Matthias Haimel, BSc; Nicholas Screatton, BM, BCh, FRCR, FRCP; Andrew Swift, FRCP, PhD; Peter Dorfmueller, MD, PhD; Stephen D. Preston, FRCPATH; Mark Southwood, PhD; Jules Hernandez-Sanchez, PhD; Jennifer Martin, BSc; Carmen Treacy, BSc; Katherine Yates, BSc; Harm Bogaard, MD, PhD; Colin Church, FRCP, PhD; Gerry Coghlan, MD, FRCP; Robin Condliffe, MD; Paul A. Corris, MBBS, FRCP; Simon Gibbs, MD, FRCP; Barbara Girerd, PhD; Simon Holden, FRCP, PhD; Marc Humbert, MD, PhD; David G. Kiely, MD; Allan Lawrie, PhD; Rajiv Machado, PhD; Robert MacKenzie Ross, MB, BChir; Shahin Moledina, MBChB; David Montani, MD, PhD; Michael Newnham, MBBS; Andrew Peacock, MD; Joanna Pepke-Zaba, PhD, FRCP; Paula Rayner-Matthews, BSc; Olga Shamardina, PhD; Florent Soubrier, MD, PhD; Laura Southgate, PhD; Jay Suntharalingam, MD, FRCP; Mark Toshner, MD; Richard Trembath, FRCP, FMedSci; Anton Vonk Noordegraaf, MD; Martin R. Wilkins, MD, FRCP, FMedSci; Stephen J. Wort, PhD, FRCP; John Wharton, PhD; NIHR BioResource–Rare Diseases Consortium; UK National Cohort Study of Idiopathic and Heritable PAH; Stefan Graf, PhD*; Nicholas W. Morrell, MD, FRCP, FMedSci*

ACKNOWLEDGMENTS

The authors acknowledge the help of all the pulmonary hypertension centers, research nurses, and clinical staff involved in the recruitment of patients. They thank the patients and their families who were recruited to this study and the Pulmonary Hypertension Association (United Kingdom).

They acknowledge the support of the NIHR Rare Diseases Translational Research Collaboration, Imperial NIHR Clinical Research Facility, Cambridge NIHR Biomedical Research Center, Netherlands CardioVascular Research Initiative, Dutch Heart Foundation, Dutch Federation of University Medical Centres, Netherlands Organisation for Health Research and Development, and Royal Netherlands Academy of Sciences.

SOURCES OF FUNDING

The NIHR BioResource for Rare Diseases provided funding for sequencing and analysis. The study was supported by a British Heart Foundation Special Project Grant and a Medical Research Council (UK) Experimental Challenge Award.

DISCLOSURES

None.

AFFILIATIONS

Department of Medicine, University of Cambridge, UK (C.H., M.B., M.H., J.M., C.T., K.Y., M.N., M.T., S. Graf, N.W.M.). NIHR BioResource–Rare Diseases (M.H., J.M., K.Y., P.R.-M.,

O.S., S. Gräf, N.W.M.). Papworth Hospital, Cambridge, UK (N.S., S.D.P., M.S., J.H.-S., J.P.-Z., M.T.). Sheffield University, UK (A.S., A.L.). Université Paris-Sud, France (P.D., B.G., M.H., D.M.). VU University Medical Centre, Amsterdam, the Netherlands (H.B., A.V.N.). Golden Jubilee Hospital, Glasgow, UK (C.C., A.P.). Royal Free Hospital, London, UK (G.C.). Royal Hallamshire Hospital, Sheffield, UK (R.C., D.G.K.). Newcastle University, UK (P.A.C.). Imperial College London, UK (S. Gibbs, M.R.W., J.W.). Addenbrooke's Hospital, Cambridge, UK (S.H.). University of Lincoln, UK (R.M.). Royal United Hospitals Bath NHS Foundation Trust, UK (R.M.R., J.S.). Great Ormond Street Hospital, London, UK (S.M.). Hôpital Pitié Salpêtrière, Paris, France (F.S.). King's College London, UK (L.S., R.T.). St George's, University of London, UK (L.S.). Royal Brompton Hospital, London, UK (S.J.W.). Department of Haematology, University of Cambridge, UK (S. Gräf).

FOOTNOTES

Received March 14, 2017; accepted August 25, 2017.

The online-only Data Supplement is available with this article at <http://circ.ahajournals.org/lookup/suppl/doi:10.1161/CIRCULATIONAHA.117.028351/-DC1>.

Circulation is available at <http://circ.ahajournals.org>.

REFERENCES

- Simonneau G, Gatzoulis MA, Adatia I, Celermajer D, Denton C, Ghofrani A, Gomez Sanchez MA, Krishna Kumar R, Landzberg M, Machado RF, Olschewski H, Robbins IM, Souza R. Updated clinical classification of pulmonary hypertension. *J Am Coll Cardiol*. 2013;62(suppl):D34–D41. doi: 10.1016/j.jacc.2013.10.029.
- Galie N, Humbert M, Vachiery JL, Gibbs S, Lang I, Torbicki A, Simonneau G, Peacock A, Vonk Noordegraaf A, Beghetti M, Ghofrani A, Gomez Sanchez MA, Hansmann G, Klepetko W, Lancellotti P, Matucci M, McDonagh T, Pierard LA, Trindade PT, Zompatori M, Hoeper M. 2015 ESC/ERS guidelines for the diagnosis and treatment of pulmonary hypertension: the Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS); endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Respir J*. 2015;46:903–975. doi: 10.1183/13993003.01032-2015.
- Resten A, Maitre S, Humbert M, Rabiller A, Sitbon O, Capron F, Simonneau G, Musset D. Pulmonary hypertension: CT of the chest in pulmonary venoocclusive disease. *AJR Am J Roentgenol*. 2004;183:65–70. doi: 10.2214/ajr.183.1.1830065.
- Montani D, Achouh L, Dorfmueller P, Le Pavec J, Sztrymf B, Tcherakian C, Rabiller A, Haque R, Sitbon O, Jais X, Dartevielle P, Maître S, Capron F, Musset D, Simonneau G, Humbert M. Pulmonary veno-occlusive disease: clinical, functional, radiologic, and hemodynamic characteristics and outcome of 24 cases confirmed by histology. *Medicine (Baltimore)*. 2008;87:220–233. doi: 10.1097/MD.0b013e31818193bb.
- Trip P, Girerd B, Bogaard HJ, de Man FS, Boonstra A, Garcia G, Humbert M, Montani D, Vonk-Noordegraaf A. Diffusion capacity and BMPR2 mutations in pulmonary arterial hypertension. *Eur Respir J*. 2014;43:1195–1198. doi: 10.1183/09031936.00136413.
- Rajaram S, Swift AJ, Condliffe R, Johns C, Elliot CA, Hill C, Davies C, Hurdman J, Sabroe I, Wild JM, Kiely DG. CT features of pulmonary arterial hypertension and its major subtypes: a systematic CT evaluation of 292 patients from the ASPIRE Registry. *Thorax*. 2015;70:382–387. doi: 10.1136/thoraxjnl-2014-206088.
- Trip P, Nossent EJ, de Man FS, van den Berk IA, Boonstra A, Groepenhoff H, Leter EM, Westerhof N, Grünberg K, Bogaard HJ, Vonk-Noordegraaf A. Severely reduced diffusion capacity in idiopathic pulmonary arterial hypertension: patient characteristics and treatment responses. *Eur Respir J*. 2013;42:1575–1585. doi: 10.1183/09031936.00184412.
- Mandel J, Mark EJ, Hales CA. Pulmonary veno-occlusive disease. *Am J Respir Crit Care Med*. 2000;162:1964–1973. doi: 10.1164/ajrcm.162.5.9912045.
- Pietra GG, Edwards WD, Kay JM, Rich S, Kernis J, Schloo B, Ayres SM, Bergofsky EH, Brundage BH, Detre KM. Histopathology of primary pulmonary hypertension. A qualitative and quantitative study of pulmonary blood vessels from 58 patients in the National Heart, Lung, and Blood Institute, Primary Pulmonary Hypertension Registry. *Circulation*. 1989;80:1198–1206.
- Lantuéjoul S, Sheppard MN, Corrin B, Burke MM, Nicholson AG. Pulmonary veno-occlusive disease and pulmonary capillary hemangiomatosis: a clinicopathologic study of 35 cases. *Am J Surg Pathol*. 2006;30:850–857. doi: 10.1097/01.pas.0000209834.69972.e5.
- Montani D, Lau EM, Dorfmueller P, Girerd B, Jais X, Savale L, Perros F, Nossent E, Garcia G, Parent F, Fadel E, Soubrier F, Sitbon O, Simonneau G, Humbert M. Pulmonary veno-occlusive disease. *Eur Respir J*. 2016;47:1518–1534. doi: 10.1183/13993003.00026-2016.
- Dorfmueller P, Humbert M, Perros F, Sanchez O, Simonneau G, Müller KM, Capron F. Fibrous remodeling of the pulmonary venous system in pulmonary arterial hypertension associated with connective tissue diseases. *Hum Pathol*. 2007;38:893–902. doi: 10.1016/j.humpath.2006.11.022.
- Ghigna MR, Guignabert C, Montani D, Girerd B, Jais X, Savale L, Hervé P, Thomas de Montpréville V, Mercier O, Sitbon O, Soubrier F, Fadel E, Simonneau G, Humbert M, Dorfmueller P. BMPR2 mutation status influences bronchial vascular changes in pulmonary arterial hypertension. *Eur Respir J*. 2016;48:1668–1681. doi: 10.1183/13993003.00464-2016.
- Eyries M, Montani D, Girerd B, Perret C, Leroy A, Lonjou C, Chelghoum N, Coulet F, Bonnet D, Dorfmueller P, Fadel E, Sitbon O, Simonneau G, Tregouët DA, Humbert M, Soubrier F. EIF2AK4 mutations cause pulmonary veno-occlusive disease, a recessive form of pulmonary hypertension. *Nat Genet*. 2014;46:65–69. doi: 10.1038/ng.2844.
- Best DH, Sumner KL, Austin ED, Chung WK, Brown LM, Borczuk AC, Rosenzweig EB, Bayrak-Toydemir P, Mao R, Cahill BC, Tazelaar HD, Leslie KO, Hemnes AR, Robbins IM, Elliott CG. EIF2AK4 mutations in pulmonary capillary hemangiomatosis. *Chest*. 2014;145:231–236.
- Dever TE, Feng L, Wek RC, Cigan AM, Donahue TF, Hinnebusch AG. Phosphorylation of initiation factor 2 alpha by protein kinase GCN2 mediates gene-specific translational control of GCN4 in yeast. *Cell*. 1992;68:585–596.
- Harding HP, Zhang Y, Zeng H, Novoa I, Lu PD, Calton M, Sadri N, Yun C, Popko B, Paules R, Stojdl DF, Bell JC, Hettmann T, Leiden JM, Ron D. An integrated stress response regulates amino acid metabolism and resistance to oxidative stress. *Mol Cell*. 2003;11:619–633.
- Wille KM, Sharma NS, Kulkarni T, Lammi MR, Barney JB, Bellot SC, Cantor RS, Naftel DC, Diaz-Guzman E, McGiffin DC. Characteristics of patients with pulmonary venoocclusive disease awaiting transplantation. *Ann Am Thorac Soc*. 2014;11:1411–1418. doi: 10.1513/AnnalsATS.201408-354OC.
- Palmer SM, Robinson LJ, Wang A, Gossage JR, Bashore T, Tapson VF. Massive pulmonary edema and death after prostacyclin infusion in a patient with pulmonary veno-occlusive disease. *Chest*. 1998;113:237–240.
- Montani D, Girerd B, Jais X, Levy M, Amar D, Savale L, Dorfmueller P, Seferian A, Lau EM, Eyries M, Le Pavec J, Parent F, Bonnet D, Soubrier F, Fadel E, Sitbon O, Simonneau G, Humbert M. Clinical phenotypes and outcomes of heritable and sporadic pulmonary veno-occlusive disease: a population-based study. *Lancet Respir Med*. 2017;5:125–134. doi: 10.1016/S2213-2600(16)30438-6.
- Evans JD, Girerd B, Montani D, Wang XJ, Galie N, Austin ED, Elliott G, Asano K, Grunig E, Yan Y, Jing ZC, Manes A, Palazzini M, Wheeler LA, Nakayama I, Satoh T, Eichstaedt C, Hinderhofer K, Wolf M, Rosenzweig EB, Chung WK, Soubrier F, Simonneau G, Sitbon O, Graf S, Kaptoge S, Di Angelantonio E, Humbert M, Morrell NW. BMPR2 mutations and survival in pulmonary arterial hypertension: an individual participant data meta-analysis. *Lancet Respir Med*. 2016;4:129–137.
- Runo JR, Vnencak-Jones CL, Prince M, Loyd JE, Wheeler L, Robbins IM, Lane KB, Newman JH, Johnson J, Nichols WC, Phillips JA 3rd. Pulmonary veno-occlusive disease caused by an inherited mutation in bone morphogenetic protein receptor II. *Am J Respir Crit Care Med*. 2003;167:889–894. doi: 10.1164/rccm.200208-861OC.
- Aldred MA, Vijaykrishnan J, James V, Soubrier F, Gomez-Sanchez MA, Martensson G, Galie N, Manes A, Corris P, Simonneau G, Humbert M, Morrell NW, Trembath RC. BMPR2 gene rearrangements account for a significant proportion of mutations in familial and idiopathic pulmonary arterial hypertension. *Hum Mutat*. 2006;27:212–213. doi: 10.1002/humu.9398.

24. Machado RD, Aldred MA, James V, Harrison RE, Patel B, Schwalbe EC, Gruenig E, Janssen B, Koehler R, Seeger W, Eickelberg O, Olschewski H, Elliott CG, Glissmeyer E, Carlquist J, Kim M, Torbicki A, Fijalkowska A, Szweczyk G, Parma J, Abramowicz MJ, Galie N, Morisaki H, Kyotani S, Nakanishi N, Morisaki T, Humbert M, Simonneau G, Sitbon O, Soubrier F, Coulet F, Morrell NW, Trembath RC. Mutations of the TGF-beta type II receptor BMPR2 in pulmonary arterial hypertension. *Hum Mutat*. 2006;27:121–132. doi: 10.1002/humu.20285.
25. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P, Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol*. 2016;17:122. doi: 10.1186/s13059-016-0974-4.
26. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB, Tukiainen T, Birnbaum DP, Kosmicki JA, Duncan LE, Estrada K, Zhao F, Zou J, Pierce-Hoffman E, Berghout J, Cooper DN, Deflaux N, DePristo M, Do R, Flannick J, Fromer M, Gauthier L, Goldstein J, Gupta N, Howrigan D, Kiezun A, Kurki MI, Moonshine AL, Natarajan P, Orozco L, Peloso GM, Poplin R, Rivas MA, Ruano-Rubio V, Rose SA, Ruderfer DM, Shakir K, Stenson PD, Stevens C, Thomas BP, Tiao G, Tusie-Luna MT, Weisburd B, Won HH, Yu D, Altshuler DM, Ardissino D, Boehnke M, Danesh J, Donnelly S, Elosua R, Florez JC, Gabriel SB, Getz G, Glatt SJ, Hultman CM, Kathiresan S, Laakso M, McCarrroll S, McCarthy MI, McGovern D, McPherson R, Neale BM, Palotie A, Purcell SM, Saleheen D, Scharf JM, Sklar P, Sullivan PF, Tuomilehto J, Tsuang MT, Watkins HC, Wilson JG, Daly MJ, MacArthur DG; Exome Aggregation Consortium. Analysis of protein-coding genetic variation in 60,706 humans. *Nature*. 2016;536:285–291. doi: 10.1038/nature19057.
27. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS, Sunyaev SR. A method and server for predicting damaging missense mutations. *Nat Methods*. 2010;7:248–249. doi: 10.1038/nmeth0410-248.
28. Ng PC, Henikoff S. Predicting deleterious amino acid substitutions. *Genome Res*. 2001;11:863–874. doi: 10.1101/gr.176601.
29. Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM, Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet*. 2014;46:310–315. doi: 10.1038/ng.2892.
30. Staples J, Qiao D, Cho MH, Silverman EK, Nickerson DA, Below JE; University of Washington Center for Mendelian Genomics. PRIMUS: rapid reconstruction of pedigrees from genome-wide estimates of identity by descent. *Am J Hum Genet*. 2014;95:553–564. doi: 10.1016/j.ajhg.2014.10.005.
31. Tenorio J, Navas P, Barrios E, Fernández L, Nevado J, Quezada CA, López-Meseguer M, Arias P, Mena R, Lobo JL, Alvarez C, Heath K, Escibano-Subias P, Lapunzina P. A founder EIF2AK4 mutation causes an aggressive form of pulmonary arterial hypertension in Iberian Gypsies. *Clin Genet*. 2015;88:579–583. doi: 10.1111/cge.12549.
32. Best DH, Sumner KL, Smith BP, Damjanovich-Colmenares K, Nakayama I, Brown LM, Ha Y, Paul E, Morris A, Jama MA, Dodson MW, Bayrak-Toydemir P, Elliott CG. EIF2AK4 mutations in patients diagnosed with pulmonary arterial hypertension. *Chest*. 2017;151:821–828. doi: 10.1016/j.chest.2016.11.014.
33. Villaschi S, Pietra GG. Alveolo-capillary membrane in primary pulmonary hypertension. *Appl Pathol*. 1986;4:132–137.
34. Montani D, Lau EM, Descatha A, Jaïs X, Savale L, Andujar P, Bensefa-Colas L, Girerd B, Zeng L, Le Pavec J, Seferian A, Perros F, Dorfmüller P, Fadel E, Soubrier F, Sitbon O, Simonneau G, Humbert M. Occupational exposure to organic solvents: a risk factor for pulmonary veno-occlusive disease. *Eur Respir J*. 2015;46:1721–1731. doi: 10.1183/13993003.00814-2015.
35. Pietra GG, Capron F, Stewart S, Leone O, Humbert M, Robbins IM, Reid LM, Tudor RM. Pathologic assessment of vasculopathies in pulmonary hypertension. *J Am Coll Cardiol*. 2004;43(suppl S):25S–32S. doi: 10.1016/j.jacc.2004.02.033.
36. Machado RD, Southgate L, Eichstaedt CA, Aldred MA, Austin ED, Best DH, Chung WK, Benjamin N, Elliott CG, Eyries M, Fischer C, Graf S, Hinderhofer K, Humbert M, Keiles SB, Loyd JE, Morrell NW, Newman JH, Soubrier F, Trembath RC, Viales RR, Grunig E. Pulmonary arterial hypertension: a current perspective on established and emerging molecular genetic defects. *Hum Mutat*. 2015;36:1113–1127.
37. Eichstaedt CA, Song J, Benjamin N, Harutyunova S, Fischer C, Grünig E, Hinderhofer K. EIF2AK4 mutation as “second hit” in hereditary pulmonary arterial hypertension. *Respir Res*. 2016;17:141. doi: 10.1186/s12931-016-0457-x.

Phenotypic Characterization of *EIF2AK4* Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically With Pulmonary Arterial Hypertension

Charaka Hadinnapola, Marta Bleda, Matthias Haimel, Nicholas Sreaton, Andrew Swift, Peter Dorfmueller, Stephen D. Preston, Mark Southwood, Jules Hernandez-Sanchez, Jennifer Martin, Carmen Treacy, Katherine Yates, Harm Bogaard, Colin Church, Gerry Coghlan, Robin Condliffe, Paul A. Corris, Simon Gibbs, Barbara Girerd, Simon Holden, Marc Humbert, David G. Kiely, Allan Lawrie, Rajiv Machado, Robert MacKenzie Ross, Shahin Moledina, David Montani, Michael Newnham, Andrew Peacock, Joanna Pepke-Zaba, Paula Rayner-Matthews, Olga Shamardina, Florent Soubrier, Laura Southgate, Jay Suntharalingam, Mark Toshner, Richard Trembath, Anton Vonk Noordegraaf, Martin R. Wilkins, Stephen J. Wort, John Wharton, NIHR BioResource-Rare Diseases Consortium; UK National Cohort Study of Idiopathic and Heritable PAH, Stefan Graf and Nicholas W. Morrell

Circulation. 2017;136:2022-2033; originally published online September 28, 2017;
doi: 10.1161/CIRCULATIONAHA.117.028351

Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2017 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://circ.ahajournals.org/content/136/21/2022>

Free via Open Access

Data Supplement (unedited) at:

<http://circ.ahajournals.org/content/suppl/2017/09/25/CIRCULATIONAHA.117.028351.DC1>

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

Reprints: Information about reprints can be found online at:
<http://www.lww.com/reprints>

Subscriptions: Information about subscribing to *Circulation* is online at:
<http://circ.ahajournals.org/subscriptions/>

APPENDIX

Name	Institution	Country
Principal Investigators BRIDGE Consortium Projects		
Timothy Aitman	Imperial College/University of Edinburgh University of Oxford/Oxford University	UK
David Bennett	Hospitals	UK
Mark Caulfield	Queen Mary University of London University of Cambridge/Cambridge University	UK
Patrick Chinnery	Hospitals	UK
Daniel Gale	University College London	UK
Ania Koziell	Guy's and St Thomas' NHS Foundation Trust	UK
Taco W Kuijpers	Emma Children's Hospital AMC, Amsterdam	Netherlands
Michael A Laffan	Imperial College Healthcare NHS Trust/Imperial College London University of Cambridge/Cambridge University	UK
Eamonn Maher	Hospitals University of Cambridge/Cambridge University	UK
Hugh S Markus	Hospitals University of Cambridge/Cambridge University	UK
Nicholas Morrell	Hospitals University of Cambridge/ NHS Blood and Transplant/ Wellcome Trust Sanger Institute	UK
Willem H Ouwehand	Cambridge University Hospitals	UK
David Perry	University of Cambridge/Cambridge University Hospitals	UK
F Lucy Raymond	University of Oxford/Oxford University	UK
Irene Roberts	Hospitals NHS FT University of Cambridge/Cambridge University	UK
Kenneth Smith	Hospitals	UK

Adrian Thrasher	Great Ormond Street Hospital	UK
Hugh Watkins	University of Oxford/Oxford University Hospitals NHS FT	UK
Catherine Williamson	King's College London	UK
Geoffrey Woods	University of Cambridge/Cambridge University Hospitals	UK
NIIHR BioResource - Rare Diseases - Management Team		
Sofie Ashford	University of Cambridge	UK
John R Bradley	Cambridge University Hospitals	UK
Debra Fletcher	University of Cambridge	UK
Tracey Hammerton	University of Cambridge	UK
Roger James	University of Cambridge	UK
Nathalie Kingston	University of Cambridge	UK
Willem H Ouwehand	University of Cambridge	UK
Christopher J Penkett	University of Cambridge	UK
F Lucy Raymond	University of Cambridge/Cambridge University Hospitals	UK
Kathleen Stirrups	University of Cambridge	UK
Marijke Veltman	University of Cambridge	UK
Tim Young	University of Cambridge	UK
Enrolment and Ethics		
Sofie Ashford	University of Cambridge	UK
Matthew Brown	University of Cambridge	UK
Naomi Clements-Brod	University of Cambridge	UK
John Davis	University of Cambridge	UK
Eleanor Dewhurst	University of Cambridge	UK
Marie Erwood	University of Cambridge	UK
Amy Frary	University of Cambridge	UK
Rachel Linger	University of Cambridge	UK
Jennifer Martin	University of Cambridge	UK

Sofia Papadia	University of Cambridge	UK
Karola Rehnstrom	University of Cambridge	UK
Hannah Stark	University of Cambridge	UK
BRIDGE-BPD Consortium		
David Allsup	Department of Haematology, Castle Hill Hospital, Hull and East Yorkshire NHS Foundation Trust	UK
Steve Austin	Department of Haematology, Guys and St Thomas' NHS Foundation Trust	UK
Tamam Bakchoul	Institut für Immunologie und Transfusionsmedizin, Ernst-Moritz-Arndt-University of Greifswald, Greifswald	Germany
Tadbir K Bariana	The Katharine Dormandy Haemophilia Centre and Thrombosis Unit, Royal Free London NHS Foundation Trust/University College London	UK
Paula Bolton-Maggs	NHS Blood and Transplant, Manchester	UK
Elizabeth Chalmers	Royal Hospital for Children, NHS Greater Glasgow and Clyde	UK
Peter Collins	Arthur Bloom Haemophilia Centre, University Hospital of Wales Heath Park, Cardiff, Wales	UK
Wendy N Erber	Pathology and Laboratory Medicine, University of Western Australia, Crawley, Western Australia	Australia
Tamara Everington	Salisbury Hospital, Salisbury NHS Foundation Trust	UK
Remi Favier	Haematological Laboratory, Trousseau Children's Hospital and INSERM U1009, Paris	France
Kathleen Freson	Department of Cardiovascular Sciences, Center for Molecular and Vascular Biology, University of Leuven	Belgium
Bruce Furie	Beth Israel Deaconess Medical Centre, Harvard Medical School, Boston	USA

Michael Gattens	Cambridge University Hospitals NHS Foundation Trust	UK
Keith Gomez	The Katharine Dormandy Haemophilia Centre and Thrombosis Unit, Royal Free London NHS Foundation Trust/University College London Department of Haematology, University of Cambridge/MRC-BSU	UK
Daniel Greene	Institute for Immunology and Transfusion Medicine, Ernst-Moritz-Arndt-University of Greifswald, Greifswald	UK
Andreas Greinacher	The Royal London Hospital, Barts Health NHS Foundation Trust	Germany
Daniel Hart	Maastricht University, Maastricht	UK
Johan WM Heemskerk	Maastricht University Medical Centre, Maastricht	Netherlands
Yvonne Henskens	Southampton General Hospital, University Hospital Southampton NHS FT	Netherlands
Rashid Kazmi	Oxford Haemophilia and Thrombosis Centre, Oxford University Hospitals NHS Trust, The Churchill Hospital, Oxford	UK
David Keeling	Cambridge University Hospitals NHS Foundation Trust	UK
Anne M Kelly	Imperial College Healthcare NHS Trust/Imperial College London	UK
Michael A Laffan	Division of Hematology, Children's Hospital of Philadelphia/ Department of Pediatrics, Perelman School of Medicine at the University of Pennsylvania, Philadelphia	USA
Michele P Lambert	Imperial College Healthcare NHS Trust/Imperial College London	UK
Claire Lentaigne	Department of Haematology, Great Ormond Street Hospital for Children NHS Trust, London	UK
Ri Liesner		UK

Sarah Mangles	Haemophilia, Haemostasis and Thrombosis Centre, Hampshire Hospitals NHS Foundation Trust, Aldermaston Road, Basingstoke	UK
Mary Mathias	Department of Haematology, Great Ormond Street Hospital for Children NHS Trust, London	UK
Carolyn M Millar	Imperial College Healthcare NHS Trust/Imperial College London	UK
Andrew Mumford	University of Bristol/University Hospitals Bristol NHS Foundation Trust	UK
Paquita Nurden	Institut Hospitalo-Universitaire LIRYC, PTIB, Hôpital Xavier Arnoz, Pessac	France
Willem H Ouwehand	University of Cambridge/ NHS Blood and Transplant/ Wellcome Trust Sanger Institute	UK
Sofia Papadia	Department of Haematology, University of Cambridge	UK
Jeanette Payne	Department of Haematology, Sheffield Children's Hospital NHS Foundation Trust	UK
John Pasi	Barts and The London School of Medicine and Dentistry, Haemophilia Centre, The Royal London Hospital, London	UK
David J Perry	Cambridge University Hospitals NHS Foundation Trust	UK
Kathelijne Peerlinck	Department of Cardiovascular Sciences, Center for Molecular and Vascular Biology, University of Leuven	Belgium
Michael Richards	Leeds Teaching Hospitals NHS Foundation Trust, Leeds	UK
Matthew Rondina	Madsen Health Center, Salt Lake City	USA
Catherine Roughley	Haemophilia Centre, Kent & Canterbury Hospital, East Kent Hospitals University Foundation Trust	UK
Sol Schulman	Beth Israel Deaconess Medical Centre, Harvard Medical School, Boston	USA

Harald Schulze	Lehrstuhl für Experimentelle Biomedizin, Universitätsklinikum Würzburg, Würzburg	Germany
Marie Scully	University College London Hospital	UK
Suthesh Sivapalaratnam	The Royal London Hospital, Barts Health NHS Foundation Trust	UK
R Campbell Tait	Glasgow Royal Infirmary, NHS Greater Glasgow and Clyde	UK
Kate Talks	Haematology Department, Royal Victoria Infirmary, Newcastle upon Tyne	UK
Jecko Thachil	Haematology Department, Manchester Royal Infirmary, Oxford Road, Manchest	UK
Ernest Turro	Department of Haematology, University of Cambridge/MRC-BSU	UK
Cheng-Hock Toh	The Roald Dahl Haemophilia Centre, Royal Liverpool Hospital, Liverpool	UK
Chris Van Geet	Department of Cardiovascular Sciences, Center for Molecular and Vascular Biology, University of Leuven	Belgium
Minka De Vries	Maastricht University Medical Centre, Maastricht	Netherlands
Timothy Q Warner	Barts Health NHS Foundation Trust	UK
Sarah Westbury	University of Bristol/University Hospitals Bristol NHS Foundation Trust	UK
Cambridge Translational GenOmics Laboratory		
Abigail Furnell	University of Cambridge	UK
Rutendo Mapeta	University of Cambridge	UK
Ilenia Simeoni	University of Cambridge	UK
Simon Staines	University of Cambridge	UK
Jonathan Stephens	University of Cambridge	UK
Kathleen Stirrups	University of Cambridge	UK
Deborah Whitehorn	University of Cambridge	UK
Paula Rayner-Matthews	University of Cambridge	UK

Christopher Watt	University of Cambridge	UK
Clinical Bioinformatics		
Antony Attwood	University of Cambridge	UK
Louise Daugherty	University of Cambridge	UK
Sri VV Deevi	University of Cambridge	UK
Csaba Halmagyi	University of Cambridge	UK
Fengyuan Hu	University of Cambridge	UK
Roger James	University of Cambridge	UK
Vera Matser	University of Cambridge	UK
Stuart Meacham	University of Cambridge	UK
Karyn Megy	University of Cambridge	UK
Christopher J Penkett	University of Cambridge	UK
Olga Shamardina	University of Cambridge	UK
Kathleen Stirrups	University of Cambridge	UK
Catherine Titterton	University of Cambridge	UK
Salih Tuna	University of Cambridge	UK
Ping Yu	University of Cambridge	UK
Julie von Ziegenweldt	University of Cambridge	UK
Genetic Epidemiology		
William Astle	University of Cambridge	UK
Marta Bleda	University of Cambridge	UK
Keren Carss	University of Cambridge	UK
Stefan Graf	University of Cambridge	UK
Daniel Greene	University of Cambridge	UK
Matthias Haimel	University of Cambridge	UK
Hana Lango-Allen	University of Cambridge	UK
Ernest Turro	University of Cambridge	UK
MRC Biostatistics Unit		
William Astle	University of Cambridge	UK
Daniel Greene	University of Cambridge	UK

Sylvia Richardson	University of Cambridge	UK
Ernest Turro	University of Cambridge	UK

High Performance Computing Service

Paul Calleja	University of Cambridge	UK
Stuart Rankin	University of Cambridge	UK
Wojciech Turek	University of Cambridge	UK

Administrative Support

Christine Bryson	University of Cambridge	UK
Julie Anderson	University of Cambridge	UK
Debra Fletcher	University of Cambridge	UK
Coleen McJannet	University of Cambridge	UK
Sophie Stock	University of Cambridge	UK
Tim Young	University of Cambridge	UK

SPEED

Evangeline Wassmer	Birmingham Children's Hospital NHS Foundation Trust	UK
Aman Sohal	Birmingham Children's Hospital NHS Foundation Trust	UK
Saikat Santra	Birmingham Children's Hospital NHS Foundation Trust	UK
Julie Vogt	Birmingham Children's Hospital NHS Foundation Trust	UK
Manali Chitre	Cambridge University Hospitals NHS Foundation Trust	UK
Deepa Krishnakumar	Cambridge University Hospitals NHS Foundation Trust	UK
Gautum Ambegaonkar	Cambridge University Hospitals NHS Foundation Trust	UK
Anna Maw	Cambridge University Hospitals NHS Foundation Trust	UK

Ruth Armstrong	Cambridge University Hospitals NHS Foundation Trust	UK
Simon Holden	Cambridge University Hospitals NHS Foundation Trust	UK
Soo-Mi Park	Cambridge University Hospitals NHS Foundation Trust	UK
Sarju Mehta	Cambridge University Hospitals NHS Foundation Trust	UK
Joan Paterson	Cambridge University Hospitals NHS Foundation Trust	UK
Jenny Carmichael	Cambridge University Hospitals NHS Foundation Trust	UK
Louise Allen	Cambridge University Hospitals NHS Foundation Trust	UK
Anke Hensiek	Cambridge University Hospitals NHS Foundation Trust	UK
Helen Firth	Cambridge University Hospitals NHS Foundation Trust	UK
Penelope Stein	Cambridge University Hospitals NHS Foundation Trust	UK
Patrick Deegan	Cambridge University Hospitals NHS Foundation Trust	UK
Rainer Doffinger	Cambridge University Hospitals NHS Foundation Trust	UK
Alasdair Parker	Cambridge University Hospitals NHS Foundation Trust	UK
Maria Bitner-Glindzicz	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Richard Scott	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Jane Hurst	Great Ormond Street Hospital for Children NHS Foundation Trust	UK

Elisabeth Rosser	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Melissa Lees	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Emma Clement	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Robert Henderson	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Dorothy Thompson	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Alice Gardham	Great Ormond Street Hospital for Children NHS Foundation Trust	UK
Paul Gissen	Great Ormond Street Hospital for Children NHS Foundation Trust/University College London	UK
Dragana Josifova	Guy's and St Thomas' NHS Foundation Trust	UK
Ellen Thomas	Guy's and St Thomas' NHS Foundation Trust	UK
Chris Patch	Guy's and St Thomas' NHS Foundation Trust	UK
Charu Deshpande	Guy's and St Thomas' NHS Foundation Trust	UK
Frances Flinter	Guy's and St Thomas' NHS Foundation Trust	UK
Muriel Holder	Guy's and St Thomas' NHS Foundation Trust	UK
Natalie Canham	London North West Healthcare NHS Trust	UK
Emma Wakeling	London North West Healthcare NHS Trust	UK
Susan Holder	London North West Healthcare NHS Trust	UK
Neeti Ghali	London North West Healthcare NHS Trust	UK
Angie Brady	London North West Healthcare NHS Trust	UK
Virginia Clowes	London North West Healthcare NHS Trust	UK
Robert MacLaren	Moorfields Eye Hospital NHS Foundation Trust	UK
Andrew Webster	Moorfields Eye Hospital NHS Foundation Trust/University College London	UK

Anthony Moore	Moorfields Eye Hospital NHS Foundation Trust/University College London	UK
Gavin Arno	Moorfields Eye Hospital NHS Foundation Trust/University College London	UK
Michel Michaelides	Moorfields Eye Hospital NHS Foundation Trust/University College London	UK
Julia Rankin	Royal Devon & Exeter NHS Foundation Trust	UK
Manju Kurian	UCL Great Ormond Street Institute of Child Health	UK
Elaine Murphy	University College London Hospitals NHS Foundation Trust	UK
Keren Carss	University of Cambridge	UK
Alba Sanchis-Juan	University of Cambridge	UK
Marie Erwood	University of Cambridge	UK
Eleanor Dewhurst	University of Cambridge	UK
Detelina Grozeva	University of Cambridge (CIMR Medical Genetics)	UK
F Lucy Raymond	University of Cambridge/Cambridge University Hospitals	UK
Evan Reid	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
Geoff Woods	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
Marc Tischkowitz	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
Richard Sandford	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
PAH		
Nicholas Morrell	University of Cambridge/Cambridge University Hospitals	UK
Stefan Gräf	University of Cambridge	UK

Marta Bleda	Department of Medicine, University of Cambridge	UK
Charaka Hadinnapola	Department of Medicine, University of Cambridge	UK
Matthias Haimel	Department of Medicine, University of Cambridge	UK
Simon Holden	Cambridge University Hospitals NHS Foundation Trust	UK
Jennifer Martin	Department of Medicine, University of Cambridge	UK
Sonia Ali	Imperial and Hammersmith	UK
Harm Boggard	VU University Medical Center, Amsterdam	Netherlands
Colin Church	Golden Jubilee National Hospital	UK
Paul Corris	Newcastle Freeman	UK
Gerry Coghlan	Royal Free	UK
Amanda Creaser-Myers	Sheffield CRF, Royal Hallamshire	UK
Victoria Cookson	GOSH	UK
Rosa DaCosta	Royal Brompton	UK
Natalie Dormand	Royal Brompton	UK
Pavandeep K Ghataorhe	Imperial and Hammersmith	UK
Simon Gibbs	Imperial and Hammersmith	UK
Alan Greenhalgh	Newcastle Freeman	UK
Marc Humbert	University of South Paris	France
Anna Huis in't Veld	VU University Medical Center, Amsterdam	Netherlands
Fiona Kennedy	Golden Jubilee National Hospital	UK
David Kiely	Sheffield CRF, Royal Hallamshire	UK
Allan Lawrie	Sheffield CRF, Royal Hallamshire	UK
Rob Mackenzie Ross	Bath	UK
Rajiv Machado	University of Lincoln	UK
Larahmie Masati	Imperial and Hammersmith	UK
Sharon Meehan	Imperial and Hammersmith	UK

Shahin Moledina	GOSH	UK
Shokri Othman	Imperial and Hammersmith	UK
Andrew Peacock	Golden Jubilee National Hospital	UK
Joanna Pepke-Zaba	Papworth Hospital	UK
Val Pollock	Golden Jubilee National Hospital	UK
Gary Polwarth	Papworth Hospital	UK
Christopher J Rhodes	Imperial and Hammersmith	UK
Kevin Rue-Albrecht	Imperial and Hammersmith	UK
Gwen Schotte	VU University Medical Center, Amsterdam	Netherlands
Debbie Shipley	Newcastle Freeman	UK
Laura Southgate	Kings College, London	UK
Respiratory Nurse Specialists	Bath	UK
Jay Suntharalingam	Bath	UK
Yvonne Tan	Royal Free	UK
Mark Toshner	Papworth Hospital	UK
	Department of Medicine, University of Cambridge	UK
Carmen Treacy	Kings College, London	UK
Richard Trembath	VU University Medical Center, Amsterdam	Netherlands
Anton Vonk Noordegraaf	Imperial and Hammersmith	UK
Ivy Wanjiku	Imperial and Hammersmith	UK
John Wharton	Imperial and Hammersmith	UK
Martin Wilkins	Royal Brompton	UK
John Wort	Imperial and Hammersmith	UK
John Wharton		
PID		
Kenneth Smith	University of Cambridge	UK
Taco Kuijpers	Emma Children's Hospital, Amsterdam	Netherlands
	UCL Great Ormond Street Institute of Child Health	UK
Adrian Thrasher		UK
James Thaventhiran	University of Cambridge	UK

Matthew Brown	University of Cambridge	UK
Hana Lango Allen	University of Cambridge	UK
Ilenia Simeoni	University of Cambridge	UK
Emily Staples	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
Crina Samarghitean	University of Cambridge	UK
Hana Alachkar	Salford Royal NHS Foundation	UK
Richard Antrobus	University Hospitals Birmingham	UK
Gururaj Arumugakani	Leeds Teaching Hopsital	UK
Chiara Bacchelli	UCL Great Ormond Street Institute of Child Health	UK
Helen Baxendale	Papworth Hospital	UK
Claire Bethune	Plymouth Hopsital	UK
Shahnaz Bibi	UCL Great Ormond Street Institute of Child Health	UK
Claire Booth	UCL Great Ormond Street Institute of Child Health	UK
Michael Browning	Leicester Royal Infirmary	UK
Siobhan Burns	Royal Free Hospital	UK
Anita Chandra	Cambridge University Hospitals NHS Foundation Trust	UK
Nichola Cooper	Imperial College Healthcare NHS Trust Cambridge University Hospitals NHS Foundation Trust	UK
Sophie Davies	Royal Hospitals Belfast	UK
Lisa Devlin	University of Cambridge	UK
Rainer Doffinger	Nottingham University Hospitals NHS Trust	UK
Elizabeth Drewe	Royal Hospitals Belfast	UK
David Edgar	Sheffield Teaching Hospitals	UK
William Egner	Barts Health NHS Trust	UK
Rohit Ghurye		

Kimberley Gilmour	UCL Great Ormond Street Institute of Child Health	UK
Sarah Goddard	University Hospitals of North Midlands	UK
Pavel Gordins	Hull & East Yorkshire Hospitals NHS Trust	UK
Sofia Grigoriadou	Barts Health NHS Trust	UK
Scott Hackett	Birmingham Heartlands	UK
Rosie Hague	Royal Hospital for Children, NHS Greater Glasgow and Clyde	UK
Grant Hayman	Epsom & St Helier University Hospitals NHS Trust	UK
Archana Herwadkar	Salford Royal NHS Foundation	UK
Aarnoud Huissoon	Birmingham Heartlands	UK
Stephen Jolles	University Hospital Wales	UK
Peter Kelleher	Imperial College Healthcare NHS Trust	UK
Dinakantha Kumaratne	Cambridge University Hospitals NHS Foundation Trust	UK
Sara Lear	Norfolk & Norwich University Hospital	UK
Hilary Longhurst	Barts Health NHS Trust	UK
Lorena Lorenzo	Barts Health NHS Trust	UK
Jesmee Maimaris	UCL Great Ormond Street Institute of Child Health	UK
Ania Manson	Cambridge University Hospitals NHS Foundation Trust	UK
Elizabeth McDermott	Nottingham University Hospitals NHS Trust	UK
Sai Murng	Gartnavel General Hospital, NHS Greater Glasgow and Clyde	UK
Sergey Nejentsev	University of Cambridge	UK
Sadia Noorani	Sandwell and West Birmingham Hospitals	UK
Eric Oksenhendler	Hopital St Louis, Paris	France
Mark Ponsford	University Hospital Wales	UK

Waseem Qasim	UCL Great Ormond Street Institute of Child Health	UK
Isabella Quinti	Sapienza Universita di Roma	Italy
Alex Richter	University Hospitals Birmingham	UK
Ravishankar Sargur	Sheffield Teaching Hospitals	UK
Sinisa Savic	Leeds Teaching Hopsital	UK
Suranjith Seneviratne	Royal Free Hospital	UK
Carrock Sewell	Scunthorpe General Hospital	UK
Hans Stauss	Royal Free Hospital	UK
Moira Thomas	Gartnavel General Hospital, NHS Greater Glasgow and Clyde	UK
Steve Welch	Birmingham Heartlands	UK
Lisa Willcocks	Cambridge University Hospitals NHS Foundation Trust	UK
Nigel Yeatman	Barts Health NHS Trust	UK
Patrick Yong	Frimley Park Hospital	UK

SUPPLEMENTAL MATERIAL:

Phenotypic characterisation of *EIF2AK4* mutation carriers in a large cohort of patients diagnosed clinically with pulmonary arterial hypertension

Hadinnapola et al.

Supplemental Methods:

Whole genome sequencing

Genomic DNA was extracted from whole blood samples prior to assessment of concentration by Qubit, and quality by gel electrophoresis. After fragmentation of DNA into 200bp fragments (Covaris E220, Covaris Inc, Woburn, USA) DNA libraries were created using Tru SeqDNA LT Prep kit (Illumina Inc, San Diego, USA). The libraries underwent next generation sequencing using 100-150 base pair paired-end sequencing using Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, USA).

Variant calling

Reads were aligned against the Genome Reference Consortium human genome (build 37) (GRCh37) and variants were called using the Isaac Aligner and Variant Caller respectively (version 2, Illumina Inc.). Genebuilds for *BMPR2* and *EIF2AK4* genes were based on Ensembl v75. Variants from these genes were extracted and annotated using Ensembl's Variant Effect Predictor (VEP) v84¹. VEP was also used to annotate data from the Exome Aggregation Consortium's (ExAC) database².

Deletions (resulting in the loss of more than 50bp) were identified by applying Isaac copy number variant caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina).

To be called by both Canvas and Manta deletions required a reciprocal overlap of $\geq 20\%$. Overlapping deletions represented in the Zarrei dataset with a reciprocal overlap of $\geq 50\%$ and deletions with a non-PAH BRIDGE control frequency of more than 1 in 1,000 were excluded³.

Analysis of computed tomographic images of the chest

CT images of the chest, where available, were reviewed independently by 2 cardiothoracic radiologists (AS and NS), with specialist imaging experience in pulmonary hypertension, blinded to the underlying diagnoses using a customised proforma (Supplemental Table 4). In addition to CT scans of patients with *EIF2AK4* mutations or with a clinical diagnosis of PVOD in the cohort, CT scans of patients from Papworth Hospital and the Royal Hallamshire Hospital with normal spirometry ($FEV_1 > 80\%$ predicted and $FVC > 80\%$ predicted) and either *BMPR2* mutations (n=21) or no variants in the known PAH genes (n=21) were analysed (Supplemental Table 5). A consensus read was undertaken for individual CT features and a mutually agreed overall radiological diagnosis was recorded.

Histology

The explanted lung tissue of one patient with a clinical diagnosis of idiopathic PAH and biallelic *EIF2AK4* mutations was available for further analysis. Four micrometre (μm) tissue sections were cut from formalin-fixed paraffin wax embedded blocks from the explanted lung tissue. Representative sections from each lobe of both lungs were stained with Elastic-Van Gieson and Haematoxylin and Eosin stains. Two expert histopathologists examined the sections independently by light microscopy.

Statistical analysis

Statistical analysis was performed in R (www.r-project.org).

Differences between groups of categorical variables were assessed using the Fisher Exact test. Where one of the variables was an ordinal the Cochran-Armitage test was applied using the `chisq_test` function from the “coin” package ⁴. Differences in continuous variables were assessed using the Mann–Whitney U test (2 comparator groups) and the Kruskal-Wallis test (3 or more comparator groups). Post-hoc pairwise comparisons were performed using Dunn’s Test for multiple testing.

Semi-parametric Cox-proportional hazards models were used to assess survival between groups using the “survival” package in R ⁵. Survival time from diagnosis to death and diagnosis to death or transplantation was assessed. Patients were censored at the date of transplantation for the primary survival analysis. Age at diagnosis and gender were used as covariates in the models.

The proportional hazards assumptions were tested by assessing Schoenfeld residuals over log time ⁶. The goodness of fit of the model was assessed by plotting the log of cumulative hazard of Cox-Snell residuals against the log of time and confirming the simple regression has 0 intercept and slope of 1 ⁷.

The inclusion of retrospectively recruited and prevalent patients in a survival analysis assessing time from diagnosis to death/transplantation can cause immortal time bias. The immortal time is the period between diagnosis and enrolment in the study and so patients

had to have survived till this point. Patients with worse prognosis diagnosed at a similar time may not have survived long enough to enrol in the study. To further explore this potential bias, a sensitivity analysis was performed including only on UK patients recruited prospectively to the study. In this multivariate Cox-proportional hazards model, the survival period was defined as the time period from date of diagnosis to date of death and patients only entered the risk set after enrolment into the study (consent date).

Supplemental Tables

Supplemental Table 1. NIHR BioResource – Rare Diseases Collaboration. See spreadsheet.

Centre	Principle Investigator	Clinicians and research staff
Freeman Hospital, Newcastle, UK	Paul A Corris	Alan Greenhalgh, Debbie Shipley, Margaret Day
Golden Jubilee National Hospital, Glasgow, UK	Andrew Peacock	Colin Church, Val Irvine, Fiona Kennedy
Great Ormond Street Hospital, London, UK	Shahin Moledina	Victoria Cookson
Hammersmith Hospital and Imperial College, London, UK	Martin R Wilkins	Simon Gibbs, John Wharton, Sonia Ali, Larahmie Masati, Sharon Meehan, Ivy Wanjiku, Shokri Othman
Papworth Hospital, Cambridge, UK	Joanna Pepke-Zaba	Mark Toshner, Gary Polwarth
Royal Brompton Hospital, London, UK	Stephen J Wort	Rosa DaCosta, Natalie Dormand, Alice Parker
Royal Free Hospital, London, UK	Gerry Coghlan	Yvonne Tan, Dipa Ghedia
Royal Hallamshire Hospital, Sheffield, UK	David G Kiely	Robin Condliffe, Amanda Creaser-Myers, Stephen Roney, Sara Walker
Royal United Hospitals Bath NHS Foundation Trust, Bath, UK	Jay Suntharalingam	Robert MacKenzie Ross, Mark Grover, Ali Grove, Jill Peel, Ann Coy
University of South Paris	Marc Humbert	David Montani, Florent Soubrier, Barbara Girerd, Mélanie Eyries
VU University Medical Center, Amsterdam, Netherlands	Anton Vonk Noordegraaf	Harm Bogaard, Anna Huis in't Veld, Gwen Schotte, Ale Struiksmma
Supplemental Table 2. Specialist pulmonary hypertension centres participating in the study		

Recruiting cohorts	n
Genomics England	1965
Specialist Pathology: Evaluating Exomes in Diagnostics	1356
Primary Immune Disorders	1299
Bleeding and Platelet Disorders	1004
Pulmonary Arterial Hypertension	932
Multiple Primary Malignant Tumours	376
Hypertrophic Cardiomyopathy	187
Cerebral Small Vessel Diseases	183
Steroid Resistant Nephrotic Syndrome	161
Intrahepatic Cholestasis of Pregnancy	140
Stem Cell & Myeloid Disorders	132
Primary Membranoproliferative Glomerulonephritis	128
Neuropathic Pain Disorder	114
Leber Hereditary Optic Neuropathy	59
Control	15
Ehlers-Danlos Syndromes	15
Supplemental Table 3. NIHR BioResource - Rare Diseases Study recruiting cohorts and GEL	

Parameter	Response
ID	
Date of birth	
Unenhanced CT	(Y/N)
CTPA	(Y/N)
HRCT	(Y/N)
Expiratory CT	(Y/N)
Pulmonary artery diameter (cm)	
Aorta diameter (cm)	
Ground glass opacification centrilobular pattern DENSITY	(None / Subtle / Present)
Ground glass centrilobular pattern EXTENT	(0, <5%, 5-25, 25-50, >50)
Ground glass DISTRIBUTION	(central (C)/peripheral (P)/zonal (Z) or diffuse (D))
Non-specific mosaic pattern / GGO	
Neovascularity vessels	(Y/N)
Arterio-venous malformations	(Y/N)
Bronchial arteries	(Y/N)
Largest bronchial artery size	
Interlobular septal thickening	(None, Subtle, Present)
Mediastinal lymphadenopathy	(Y/N)
Emphysema	(Y/N) and % of parenchyma involved
Fibrosis	(Y/N) and % of parenchyma involved
Pleural effusion	(Y/N)
Air trapping	(Y/N)
Comments	
Likely diagnosis	Any suspicion of PVOD or PCH / PAH
Supplemental Table 4. Proforma used in analysis of CT scans	

Group	n
PAH patients with <i>BMP2</i> variants	21
PAH patients with biallelic <i>EIF2AK4</i> variants	7
PVOD patients	14
PAH patients with heterozygous <i>EIF2AK4</i> variants	4
PAH patients with no variants in the previously reported PAH genes	21

Supplemental Table 5. CT scans of patients with PVOD and patients with PAH carrying biallelic *EIF2AK4* mutations were reassessed by radiologists blinded to the diagnosis. For comparison CT scans of PAH patients with normal spirometry ($FEV_1 > 80\%$ predicted and $FVC > 80\%$ predicted) who either had no mutations in the previously reported PAH genes or carried *BMP2* mutations were assessed.

Supplemental Table 6. Page 1/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
BRIDGE control	c.292C>G	missense variant	p.L98V	0	1	0.00001656	probably damaging (0.999)	deleterious (0)	25.7	Heterozygous variant
BRIDGE control	c.354_355delTG	frameshift variant	p.C118Wfs*7	0	2	Not found in ExAC			35	Heterozygous variant
BRIDGE control	c.745C>T	stop gained & splice region variant	p.R249*	0	1	0.00007451			39	Heterozygous variant
BRIDGE control	c.746G>A	missense variant & splice region variant	p.R249Q	0	1	2.48E-05	probably damaging (0.999)	deleterious (0.02)	34	Heterozygous variant
BRIDGE control	c.767G>T	missense variant	p.C256F	0	1	1.66E-05	possibly damaging (0.904)	deleterious (0.02)	28.4	Heterozygous variant
BRIDGE control	c.985G>A	missense variant	p.E329K	0	1	Not found in ExAC	probably damaging (0.981)	deleterious (0.01)	34	Heterozygous variant
BRIDGE control	c.1153dupG	frameshift variant	p.V385Gfs*30	0	1	0.00003308			32	Heterozygous variant
BRIDGE control	c.1190T>A	missense variant	p.I397N	0	1	Not found in ExAC	possibly damaging (0.67)	deleterious (0)	32	Heterozygous variant

Supplemental Table 6. Summary of rare (MAF < 0.0001) and predicted deleterious (CADD score > 15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 2/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
BRIDGE control	c.1215C>G	stop gained	p.Y405*	0	2	Not found in ExAC			29.4	Heterozygous variant
BRIDGE control	c.1331A>G	missense variant	p.Y444C	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28.7	Heterozygous variant
BRIDGE control	c.1345C>T	missense variant	p.R449C	0	1	0.00001654	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.2249T>A	missense variant & splice region variant	p.L750Q	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28	Heterozygous variant
BRIDGE control	c.2298delG	frameshift variant	p.N767Tfs*24	0	1	Not found in ExAC			28.3	Heterozygous variant
BRIDGE control	c.2720A>T	missense variant	p.Y907F	0	4	1.66E-05	probably damaging (1)	deleterious (0)	31	Heterozygous variant
BRIDGE control	c.2828C>T	missense variant	p.T943M	0	1	0.00003311	probably damaging (1)	deleterious (0)	34	Heterozygous variant
BRIDGE control	c.3104_3106delTCT	inframe deletion	p.F1035del	0	1	Not found in ExAC			22	Heterozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 3/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
BRIDGE control	c.3217C>T	missense variant	p.R1073C	0	1	0.0000166	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3223T>G	missense variant	p.F1075V	0	1	0.0000083	probably damaging (0.997)	deleterious (0)	32	Heterozygous variant
BRIDGE control	c.3344C>T	missense variant	p.P1115L	0	1	8.26E-06	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3358-3C>T	splice region variant & intron variant	p.NA	0	1	Not found in ExAC			17.15	Heterozygous variant
BRIDGE control	c.3406C>T	stop gained & splice region variant	p.R1136*	0	1	Not found in ExAC			40	Heterozygous variant
BRIDGE control	c.3430A>T	missense variant	p.R1144W	0	1	0.0000248	probably damaging (1)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.3986T>C	missense variant	p.F1329S	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Heterozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 4/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
BRIDGE control	c.3992T>C	missense variant	p.F1331S	0	1	8.28E-06	possibly damaging (0.872)	deleterious (0.01)	28.4	Heterozygous variant
BRIDGE control	c.4039G>A	missense variant	p.A1347T	0	1	8.28E-05	probably damaging (1)	deleterious (0)	34	Heterozygous variant
BRIDGE control	c.4388_4389+12 delAGGTAAAGAC GTCA	splice donor variant & coding sequence variant & intron variant	p.NA	0	1	Not found in ExAC			36	Heterozygous variant
BRIDGE control	c.4397C>A	missense variant	p.S1466Y	0	2	Not found in ExAC	probably damaging (0.988)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.4729G>A	missense variant & splice region variant	p.V1577M	0	1	Not found in ExAC	probably damaging (0.999)	deleterious (0)	29.6	Heterozygous variant
BRIDGE control	c.4751dupT	frameshift variant	p.L1585ifs*11	0	1	Not found in ExAC			34	Heterozygous variant
BRIDGE control	c.4920_4931delT AGAGATGACTA	inframe deletion	p.R1641_Y1644 del	0	1	Not found in ExAC			23	Heterozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 5/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
PAH	c.44C>T	missense variant	p.P15L	1	0	8.32E-06	unknown (0)	deleterious low confidence (0.03)	23.5	Heterozygous variant
PAH	c.220G>A	missense variant	p.D74N	1	0	1.66E-05	possibly damaging (0.954)	deleterious (0)	32	Heterozygous variant
PAH	c.1072_1073dup GT	frameshift variant	p.V359*	1	0	Not found in ExAC			32	Heterozygous variant
PAH	c.1660G>T	missense variant & splice region variant	p.D554Y	1	0	Not found in ExAC	probably damaging (0.966)	deleterious (0)	28	Heterozygous variant
PAH	c.2446C>T	stop gained	p.Q816*	1	0	Not found in ExAC			41	Heterozygous variant
PAH	c.2516T>C	missense variant	p.I839T	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	28.9	Heterozygous variant
PAH	c.3218G>T	missense variant	p.R1073L	1	0	Not found in ExAC	probably damaging (0.995)	deleterious (0.01)	35	Heterozygous variant
PAH	c.3604C>T	missense variant	p.H1202Y	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	29.7	Heterozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 6/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
PAH	c.3711_3713del GAG	inframe deletion	p.R1238del	1	0	0.0000083			21.6	Heterozygous variant
PAH	c.3722A>G	missense variant	p.E1241G	1	0	Not found in ExAC	probably damaging (0.971)	deleterious (0)	27.2	Heterozygous variant
PAH	c.4646G>A	missense variant	p.R1549H	1	0	0.0000910	probably damaging (0.998)	deleterious (0.01)	35	Heterozygous variant
PAH	c.145-2A>G	splice acceptor variant	p.NA	1	0	Not found in ExAC			23.9	Additional second (likely trans) variant identified
PAH	c.257+4A>C	splice region variant & intron variant	p.NA	1	0	8.28E-06			15.5	Additional second (likely trans) variant identified
PAH	c.1392delT	frameshift variant	p.R465Vfs*38	1	0	2.48E-05			35	Additional second (likely trans) variant identified
PAH	c.1739dupA	frameshift variant	p.R581Efs*9	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 7/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
PAH	c.1820T>G	missense variant & splice region variant	p.V607G	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	27.3	Additional second (likely trans) variant identified
PAH	c.2727C>G	missense variant	p.S909R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Additional second (likely trans) variant identified
PAH	c.2827A>G	missense variant	p.T943A	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	26.4	Additional second (likely trans) variant identified
PAH	c.2841delG	frameshift variant	p.I948Sfs*35	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified
PAH	c.3055_3064delC TGACCAACG	frameshift variant	p.L1019Wfs*9	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Additional second (likely trans) variant identified

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 8/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
PAH	c.3325G>A	missense variant	p.G1109R	1	0	0.0000082	probably damaging (1)	deleterious (0.02)	35	Additional second (likely trans) variant identified
PAH	c.3884T>G	missense variant	p.L1295R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Additional second (likely trans) variant identified
PAH	c.4400dupT	frameshift variant	p.E1468Rfs*14	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.4418_4421delC AGA	frameshift variant	p.T1473Rfs*17	1	0	0.0000083			36	Additional second (likely trans) variant identified
PAH	c.4769delT	frameshift variant	p.L1590*	1	0	0.0000083			33	Additional second (likely trans) variant identified
PAH	c.281dupA	frameshift variant	p.N94Lfs*8	2	0	Not found in ExAC			35	Homozygous variant
PAH	c.1159_1160delC T	frameshift variant	p.L387Cfs*27	2	0	Not found in ExAC			29.6	Homozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 6. Page 9/9

Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	<i>EIF2AK4</i> genotype
PAH	c.1795G>C	missense variant	p.G599R	4	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Homozygous variant
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Homozygous variant
PAH	c.3605A>T	missense variant	p.H1202L	2	0	Not found in ExAC	probably damaging (1)	deleterious (0)	31	Homozygous variant
PAH	c.4392dupT	frameshift variant & splice region variant	p.K1465*	2	0	Not found in ExAC			35	Homozygous variant

Supplemental Table 6. Summary of rare (ExAC MAF <0.0001) and predicted deleterious (CADD score >15 and not benign by both PolyPhen-2 and SIFT) *EIF2AK4* variants in NIHR BRIDGE Study. Transcript: ENST00000263791.5. *EIF2AK4* variants are not shared between PAH patients and controls. Biallelic *EIF2AK4* variants are seen only in PAH cases.

Bold - variants identified in more than one patient in the PAH Cohort. MAF - minor allele frequency

Supplemental Table 7. Page 1/4

Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVS	Consequence type	<i>EIF2AK4</i> genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
23	M	British	c.3884T>G	missense variant	C Het			52	3.3	3	97	119	33	Yes	Possible PVOD / PCH		PDE5i + ERA + IV Prostanoid	No	
			c.3055_3064delCTGACCAACG	frameshift variant															
48	M	Other	c.4400dup T	frameshift variant	C Het			46	6.4	3	116	120	45	No	CT not available for analysis		ERA + PDE5i + inhaled Prostanoid	No	
			c.1739dup A	frameshift variant															
38	F	Other Asian	c.2827A>G	missense variant	C Het			40	4.5	2				No	CT not available for analysis		ERA + PDE5i	No	
			c.4418_4421delCAGA	frameshift variant															
			c.145-2A>G	splice acceptor variant															

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

Supplemental Table 7. Page 2/4

Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVS	Consequence type	<i>EIF2AK4</i> genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
70	F	British	c.1392del T	frameshift variant	C Het			76	6.6	3	101	127	33	Unk	Possible PVOD / PCH		PDE5i + ERA + inhaled Prostanoid	No	
			c.257+4A >C	splice region variant & intron variant															
36	F	Indian	c.3605A>T	missense variant	Hom			44	2.7	3	73	83	40	Yes	Possible PVOD / PCH		ERA + PDE5i + inhaled Prostanoid	No	
22	M	Pakistani	c.1795G>C	missense variant	Hom			65	3.0	3	92	93	31	Yes	PAH		ERA + PDE5i + IV Prostanoid	No	Yes
29	M	Pakistani	c.3097C>T	stop gained	Hom			50	4.9	3	99	107	27	Unk	PAH	Sister died from PAH	PDE5i	No	
18	M	Not stated	c.1159_160delCT	frameshift variant	Hom			92		3	86	82	28	No	Possible PVOD / PCH		ERA + IV Prostanoid	No	
25	F	Pakistani	c.1795G>C	missense variant	Hom			57	5.6	3	82	87	33	No	PAH		PDE5i + ERA	No	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

Supplemental Table 7. Page 3/4

Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVS	Consequence type	<i>EIF2AK4</i> genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
24	F	Not stated	c.2446C>T	stop gained	Het (both on same allele) *			60	5.2	3	96	97	81	Unk	CT not available for analysis	Father and sister died of PAH	Unk	Unk	
			c.3218G>T	missense variant															
39	F	British	c.1072_1073dupG T	frameshift variant	Het			54	3.0	2	87	98	72	No	CT not available for analysis		ERA	No	
40	F	British	c.44C>T	missense variant	Het		c.4303-50delT	43	5.6	2	99	96	109	Unk	Possible PVOD / PCH		ERA	No	
44	M	British	c.2516T>C	missense variant	Het	c.853-2A>G (splice acceptor variant)	c.361-180A>G	53	3.8	3	102	98	54	Unk	PAH		PDE5i + ERA	No	
25	F	British	c.3722A>G	missense variant	Het					3	53	49	41	No	CT not available for analysis		PDE5i + ERA + IV Prostanoid	No	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown, *maternally inherited

Supplemental Table 7. Page 4/4

Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVS	Consequence type	<i>EIF2AK4</i> genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV ₁ (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
66	F	Not stated	c.4646G>A	missense variant	Het			44	2.1	3	79	100		Unk	PAH		PDE5i + ERA	No	
72	M	British	c.1660G>T	missense variant & splice region variant	Het			30	2.8	3				No	PAH		IV Prostanoid	No	
59	F	Other	c.3711_3713delGAG	inframe deletion	Het			41	3.4	3	68	68	95	Unk	PAH		ERA + PDE5i	No	
48	F	British	c.3604C>T	missense variant	Het	c.2695C>T (stop gained)		57	4.4	4	90	100	61	Unk	PAH		PDE5i + ERA	No	
70	F	Other White	c.220G>A	missense variant	Het			42	5.4	2				Unk	CT not available for analysis		ERA	Unk	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV₁ – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

Supplemental Table 8. Page 1/2

	PAH patients with <i>BMPR2</i> mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with <i>EIF2AK4</i> heterozygous variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	p
n	64	255	3	7	5	
Age (years)	42 [31 - 52]	53 [39 - 67]	39 [32 - 40]	25 [23 - 38]	63 [27 - 76]	<0.001
Gender (n female [%])	45 [70.3%]	179 [70.2%]	3 [100%]	2 [28.6%]	4 [80%]	0.161
Ethnicity (n white Caucasian [%])	50 [78.1%]	226 [88.6%]	2 [66.7%]	2 [28.6%]	4 [80%]	<0.001
Digital clubbing (n [%])	5 [13.2%]	3 [2.2%]	0 [0%]	2 [40%]	0 [0%]	0.004
BMI	28 [25 - 33]	27 [24 - 31]	24 [24 - 25]	24 [21 - 27]	27 [24 - 32]	0.202
<p>Supplemental Table 8. Phenotype summary of patients with preserved spirometry ($FEV_1 > 80\%$ predicted and $FVC > 80\%$ predicted). PAH patients with biallelic <i>EIF2AK4</i> mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups. mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV_1 – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. * Also includes the 2 patients with heterozygous <i>EIF2AK4</i> variants and a <i>BMPR2</i> mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.</p>						

Supplemental Table 8. Page 2/2

	PAH patients with <i>BMPR2</i> mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with <i>EIF2AK4</i> heterozygous variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	p
mPAP (mmHg)	56 (15)	51 (18)	54 (8)	57 (20)	57 (7)	0.008
CO (L/min)	3 [3 - 4]	4 [3 - 5]	5 [4 - 5]	5 [4 - 6]	3 [3 - 3]	<0.001
PVR (WU)	14 [10 - 18]	10 [7 - 14]	8 [7 - 9]	9 [8 - 15]	14 [11 - 19]	<0.001
Vasoresponders (n [%])	0 [0%]	18 [21.7%]	0 [0%]	0 [0%]		0.016
FEV ₁ (%pred)	97 [88 - 102]	93 [87 - 101]	96 [92 - 97]	97 [89 - 100]	98 [94 - 106]	0.525
FVC (%pred)	102 [96 - 113]	103 [96 - 112]	97 [96 - 98]	107 [90 - 120]	109 [101 - 113]	0.704
KCO (%pred)	80 [71 - 93]	68 [46 - 84]	81 [76 - 95]	33 [30 - 33]	33 [28 - 37]	<0.001
Resting S _A O ₂ (%)	96 [94 - 98]	96 [93 - 98]	98 [98 - 99]	91 [90 - 92]	95 [91 - 95]	0.021
S _A O ₂ post walk test (%)	95 [90 - 98]	91 [85 - 96]	94 [87 - 96]	80 [75 - 84]	85 [85 - 88]	<0.001

Supplemental Table 8. Phenotype summary of patients with preserved spirometry (FEV₁ > 80 % predicted and FVC > 80 % predicted). PAH patients with biallelic *EIF2AK4* mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups. mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV₁ – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. * Also includes the 2 patients with heterozygous *EIF2AK4* variants and a *BMPR2* mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

Supplemental Table 9. Page 1/2				
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no <i>EIF2AK4</i> mutation	p
n		11	10	
Age (years)		26.8 [22.5 - 34.3]	68.3 [63.9 - 72.1]	0.001
Gender (n female [%])		6 [54.5%]	5 [50.0%]	1.000
Ethnicity (n white Caucasian [%])		5 [45.5%]	9 [90.0%]	0.063
mPAP (mmHg)		52 [47 - 63]	48 [42 - 57]	0.342
PCWP (mmHg)		11 [7.5 - 12]	11.5 [9.0 - 12.2]	0.560
FEV ₁ (% pred)		93.1 [82.8 - 98.5]	79.0 [72.3 - 91.0]	0.236
FVC (% pred)		95.5 [84.6 - 108.5]	96.0 [73.0 - 101.0]	0.720
KCO (% pred)		32.0 [28.7 - 33.0]	41.4 [36.8 - 54.0]	0.013
Centrilobular ground glass opacification density	None	2 [18.2%]	6 [60.0%]	0.012
	Subtle	2 [18.2%]	3 [30.0%]	
	Present	7 [63.6%]	1 [10.0%]	
<p>Supplemental Table 9. Phenotypic and radiological characteristics of biallelic <i>EIF2AK4</i> mutation carriers compared to patients with a clinical diagnosis of PVOD and no <i>EIF2AK4</i> mutation.</p> <p>mPAP – mean pulmonary artery pressure, PCWP – pulmonary capillary wedge pressure, FEV₁ – forced expiratory volume 1 s, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.</p>				

Supplemental Table 9. Page 2/2				
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no <i>EIF2AK4</i> mutation	p
Centrilobular ground glass opacification extent	None	2 [18.2%]	7 [70.0%]	0.007
	<5%	1 [9.1%]	1 [10.0%]	
	5-25%	2 [18.2%]	1 [10.0%]	
	25-50%	1 [9.1%]	1 [10.0%]	
	50-75%	2 [18.2%]	0 [0.0%]	
	75-100%	3 [27.3%]	0 [0.0%]	
Interlobular septal thickening	None	7 [63.6%]	2 [20.0%]	0.068
	Subtle	0 [0.0%]	1 [10.0%]	
	Present	4 [36.4%]	7 [70.0%]	
Mediastinal lymphadenopathy	None	4 [36.4%]	2 [20.0%]	0.635
	Present	7 [63.6%]	8 [80.0%]	
Pleural effusion	None	11 [100.0%]	6 [60.0%]	0.035
	Small	0 [0.0%]	4 [40.0%]	
Neovascularity	None	10 [90.9%]	9 [90.0%]	1.000
	Present	1 [9.1%]	1 [10.0%]	
CT diagnosis	PAH	4 [36.4%]	3 [30.0%]	
	Possible PVOD/PCH	7 [63.6%]	7 [70.0%]	
<p>Supplemental Table 9. Phenotypic and radiological characteristics of biallelic <i>EIF2AK4</i> mutation carriers compared to patients with a clinical diagnosis of PVOD and no <i>EIF2AK4</i> mutation. mPAP - mean pulmonary artery pressure, PCWP - pulmonary capillary wedge pressure, FEV₁ - forced expiratory volume 1 s, FVC - forced vital capacity, KCO - transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.</p>				

Group	Time to assessment 1 (days)	n	Change in 6mwd (m)	Change in FC	Time to assessment 2 (days)	n	Change in 6mwd (m)	Change in FC	Number on prostanoid therapy before the 2 nd assessment [%]
PAH <i>BMPR2</i>	357 [314 - 386]	21	+69 [20 - 100]	-1 [-1 - -1]	1120 [1055 - 1174]	18	+45 [31 - 115]	-1 [-1 - -0.5]	5 [23%]
PAH biallelic <i>EIF2AK4</i>	358 [335 - 388]	9	+28 [-13 - 77]	0 [-1 - 0]	1102 [1090 - 1112]	5	+62 [-8 - 132]	0 [0 - 0]	1 [10%]
PAH no mutation	387 [340 - 414]	16	+81 [61 - 151]	-1 [-1 - 0]	1118 [1105 - 1159]	9	+104 [20 - 144]	-1 [-1 - 0]	4 [17%]
p	0.295		0.343	0.039	0.730		0.748	0.044	0.816

Supplemental Table 10. Response to pulmonary artery vasodilator therapies at 1 and 3 years after diagnosis compared to baseline. 6mwd - six-minute walk test distance, FC - functional class. Drop in number of patients between assessment 1 and 2 due to death, transplantation or lack of sufficient follow up time. Data presented as median [IQR] unless stated.

Variable	Hazard Ratio [95% confidence interval]	p
PAH <i>BMPR2</i> mutation*	0.148 [0.055 - 0.396]	<0.001
PAH no mutation*	0.179 [0.073 - 0.440]	<0.001
PVOD*	0.393 [0.075 - 2.065]	0.27
Age at diagnosis	1.043 [1.033 - 1.053]	<0.001
Male gender	1.631 [1.222 - 2.179]	<0.001

Supplemental Table 11. Cox proportional hazards model assessing time to death. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations had an increased risk of death compared to other PAH patients. Number of patients = 858. Events = 194.
* compared to the PAH biallelic *EIF2AK4* mutation carriers

Variable	Hazard Ratio [95% confidence interval]	p
PAH <i>BMPR2</i> mutation*	0.175 [0.066 - 0.462]	<0.001
PAH no mutation*	0.203 [0.083 - 0.501]	<0.001
PVOD*	0.840 [0.222 - 3.193]	0.798
Age at diagnosis	1.036 [1.027 - 1.046]	<0.001
Male gender	1.542 [1.165 - 2.042]	0.002

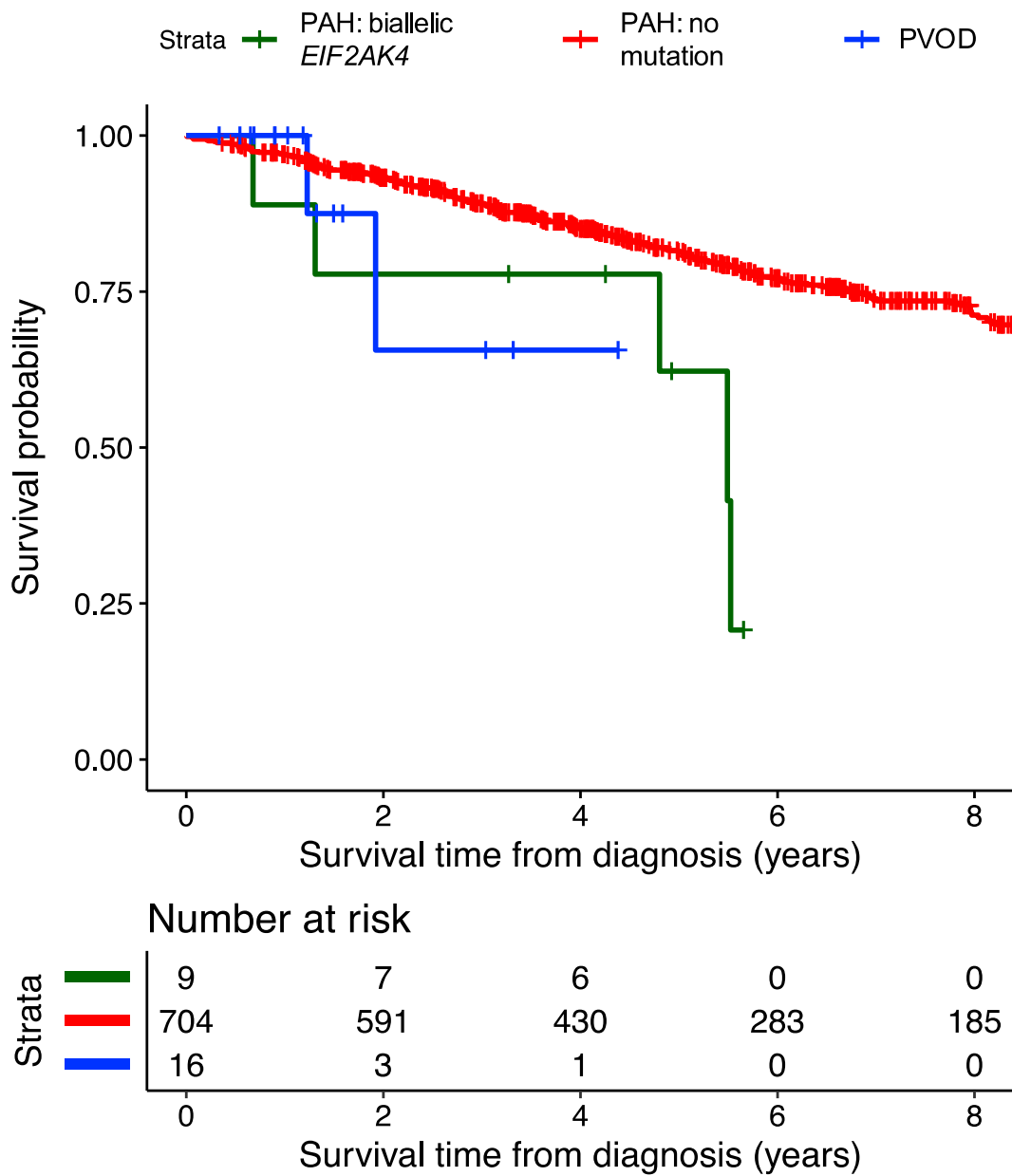
Supplemental Table 12. Cox proportional hazards model assessing time to death or transplantation. Number of patients = 858. Events = 208.
* compared to the PAH biallelic *EIF2AK4* mutation carriers

Variable	Hazard Ratio [95% confidence interval]	p
PAH <i>BMPR2</i> mutation*	0.376 [0.080 - 1.763]	0.215
PAH no mutation*	0.456 [0.109 - 1.905]	0.282
PVOD*	1.029 [0.133 - 7.953]	0.978
Age at diagnosis	1.034 [1.020 - 1.046]	<0.001
Male gender	1.515 [1.000 - 2.296]	0.051

Supplemental Table 13. Sensitivity analysis including only prospectively recruited UK patients. Cox proportional hazards model assessing time to death. Number of patients = 608. Events = 95.
* compared to the PAH biallelic *EIF2AK4* mutation carriers

Supplemental Figures

Figure S1



Supplemental Figure Legends:

Figure S1: Kaplan – Meier survival curves showing survival time (time to death) for patients with a clinical diagnosis of PAH or PVOD.

Supplemental References

1. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P and Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol.* 2016;17:122.
2. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB, Tukiainen T, Birnbaum DP, Kosmicki JA, Duncan LE, Estrada K, Zhao F, Zou J, Pierce-Hoffman E, Berghout J, Cooper DN, Deflaux N, DePristo M, Do R, Flannick J, Fromer M, Gauthier L, Goldstein J, Gupta N, Howrigan D, Kiezun A, Kurki MI, Moonshine AL, Natarajan P, Orozco L, Peloso GM, Poplin R, Rivas MA, Ruano-Rubio V, Rose SA, Ruderfer DM, Shakir K, Stenson PD, Stevens C, Thomas BP, Tiao G, Tusie-Luna MT, Weisburd B, Won HH, Yu D, Altshuler DM, Ardissino D, Boehnke M, Danesh J, Donnelly S, Elosua R, Florez JC, Gabriel SB, Getz G, Glatt SJ, Hultman CM, Kathiresan S, Laakso M, McCarroll S, McCarthy MI, McGovern D, McPherson R, Neale BM, Palotie A, Purcell SM, Saleheen D, Scharf JM, Sklar P, Sullivan PF, Tuomilehto J, Tsuang MT, Watkins HC, Wilson JG, Daly MJ and MacArthur DG. Analysis of protein-coding genetic variation in 60,706 humans. *Nature.* 2016;536:285-91.
3. Zarrei M, MacDonald JR, Merico D and Scherer SW. A copy number variation map of the human genome. *Nat Rev Genet.* 2015;16:172-83.
4. Hothorn T, Hornik K, Wiel MAvd and Zeileis A. A Lego System for Conditional Inference. *The American Statistician.* 2012;60:257-263.
5. Therneau T and Grambsch P. *Modeling Survival Data: Extending the Cox Model.* 1 ed. New York: Springer-Verlag 2000.
6. Grambsch P and Therneau H. Proportional hazards tests and diagnostics based on weighted residuals. *Biometrika.* 1994;81:515-526.
7. Collett D. *Modelling Survival Data in Medical Research.* 3rd ed. London: Chapman & Hall/CRC; 2014.