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# Elucidation of the elusive structure and formula of the active pharmaceutical ingredient bismuth subgallate by continuous rotation electron diffraction†

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**Bismuth subgallate has been used in wound and gastrointestinal therapy for over a century. The combination of continuous rotation electron diffraction and sample cooling finally revealed its structure as a coordination polymer. The structure provides insight regarding its formula, poor solubility, acid resistance and previously unreported gas sorption properties.**

When the active pharmaceutical ingredient (API) in a drug has both an unknown structure and unclear formula, an unsatisfactory situation results for all stakeholders, from patients to regulatory bodies. Structure determination is paramount for understanding physical and chemical properties such as solubility, which the pharmaceutical properties are often critically dependent on. However, structure determination is unfortunately not always trivial. Complications such as crystal size, poor stability during characterization, and complexity of the structures can impede structure investigations by conventional techniques. We now resolve this situation for bismuth(III) gallate (bismuth subgallate) using a combination of continuous rotation electron diffraction and sample cooling. The compound has been used in medicine for over a century to treat wounds, gastrointestinal disorders and syphilis.<sup>1</sup> Today it is the API of numerous over-the-counter drugs including Devrom<sup>®</sup>, Stryphnasal<sup>®</sup> N and Sulbogin<sup>®</sup> (Suile<sup>™</sup>).<sup>2</sup> In addition to efficacy as an antimicrobial agent, drugs containing the title compound are also used as astringents, haemostatic agents, and internal deodorants.<sup>3</sup> Until now, due to lack of

structural evidence, it has been presented as a molecular compound, either with Bi<sup>3+</sup> chelated by two deprotonated phenolate groups, one phenol group intact and a coordinated hydroxyl ion, or with Bi<sup>3+</sup> coordinated by a carboxylate group and two hydroxyl ions (*i.e.* in the US pharmacopeia, Chemspider database *etc.*). However, we have discovered that both structure and formula are actually somewhat different from earlier proposals.

While bismuth compounds have historically been used in cosmetics and in medicine for over three centuries,<sup>4</sup> today much focus is on their applications in treating *Helicobacter pylori* infections due to their implications on society – at least half of the world's population is infected with *H. pylori*.<sup>5</sup> These infections are linked to peptic and duodenal ulcers, gastritis and stomach cancer.<sup>6</sup> Orally administered bismuth compounds have been reported to form crystalline coatings over ulcers hastening healing.<sup>7</sup> Antibiotic resistance is a growing threat to global health, and it has been demonstrated that the co-administration of bismuth compounds with organic antibiotics and proton-pump inhibitors are effective in the treatment of antibiotic resistant strains of *H. pylori*.<sup>8</sup> Structure determination of bismuth compounds used as APIs<sup>7a,9</sup> has been impeded by their tendency to form as nanocrystalline powders rather than large crystals which prevents structure determination by the conventional method of choice: single crystal X-ray diffraction. Additionally, their pronounced tendency of oligomerization results in complex species such as [Bi<sub>38</sub>O<sub>44</sub>(Hsal)<sub>26</sub>(Me<sub>2</sub>CO)<sub>16</sub>(H<sub>2</sub>O)<sub>2</sub>](Me<sub>2</sub>CO)<sub>4</sub>,<sup>9d</sup> which complicates structure determination by X-ray powder diffraction (XPD). Moreover, variations in pH may have a large impact.<sup>10</sup> Consequently, despite their current and historical applications in medicine, details of their structures are just emerging.<sup>7a,9d,e</sup> Obviously, this lack of structural information is linked to the still sketchy knowledge of the mode of action of these drugs. Therefore it is paramount to develop and implement new structure characterization methods in order to overcome issues preventing crystal structure elucidation.

The structure determination of bismuth subgallate required recently developed crystallographic techniques, as all attempts by conventional methods failed. Single crystal X-ray diffraction

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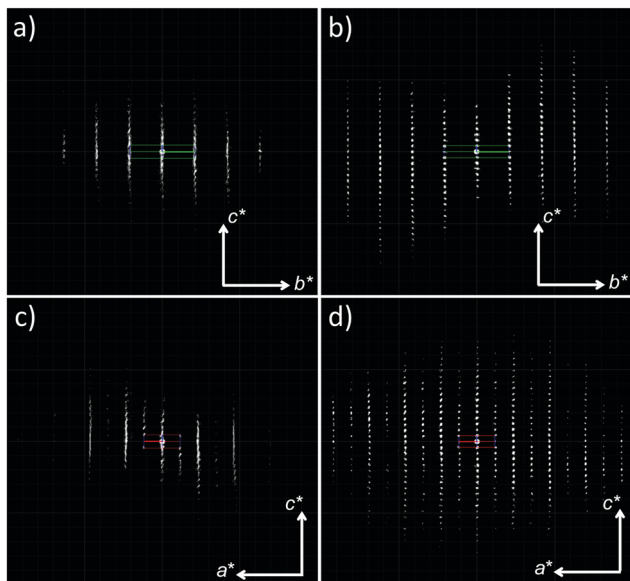


Fig. 1 Electron diffraction data collected on bismuth subgallate by (a and c) conventional RED and (b and d) continuous rotation electron diffraction with sample cooling.

was not plausible due to small crystal size (ca.  $6 \times 4 \times 0.2 \mu\text{m}$ , Fig. S1, ESI<sup>†</sup>). From XPD data (Fig. S2, ESI<sup>†</sup>) unit cell parameters, several possible space groups, and  $\text{Bi}^{3+}$  positions were determined, however efforts to locate any other atom in the structure were unsuccessful. We then turned to electron diffraction (ED),<sup>11</sup> where 3D data can be collected from nano-sized single crystals in a transmission electron microscope (TEM).<sup>12</sup> However, a major limitation with crystalline organic or metal-organic compounds is their tendency to be quickly destroyed under the high-energy electron beam. Rotation electron diffraction (RED)<sup>11b,c</sup> data were initially collected under ambient temperature. Beam damage, poor resolution and significant diffuse scattering were observed in all data sets (Fig. 1a and c), and not even the unit cell parameters could be confirmed with this data. Data collection was reattempted, but with two significant changes in an effort to acquire better quality diffraction data. Firstly, continuous rotation data collection was performed by acquiring electron diffraction frames on a high-speed hybrid detection camera (Timepix Quad) in video mode following the method described by Gemmi *et al.* In this method the sample is continuously rotated throughout the entire data acquisition process.<sup>13</sup> This significantly reduced the data collection time from one hour to three minutes. Secondly, the sample was cooled to 173 K on a cryo-transfer sample holder before insertion into the microscope, and further cooled down to 98 K before data collection. As a result of these two actions, beam damage was minimized, the resolution of the data was greatly improved, and no obvious diffuse scattering was observed (Fig. 1b and d and Fig. S3, ESI<sup>†</sup>) in agreement with the XPD data (Fig. S2 and S4, ESI<sup>†</sup>). This suggests that disorder, as indicated by the presence of diffuse scattering in earlier data sets, was not present in the original sample, but was introduced in the TEM without sample cooling and fast data collection. Metal-organic frameworks<sup>14</sup> and coordination polymers<sup>15</sup> can

exhibit diffuse scattering as a result of the degradation of long-range order upon removal of solvent molecules.

The new ED data indicated similar unit cell parameters as determined by XPD and the extinction symbol,  $P-na$ , was unambiguously determined from the reflection conditions. Structure determination was performed in the space group  $Pmna$  using the ED data and to our surprise the positions of all non-hydrogen atoms in the crystal structure, including those of water molecules in the pores (Fig. S5, ESI<sup>†</sup>), were determined in the initial structure solution. Due to high  $R$ -values as a consequence of dynamical scattering common to electron diffraction data<sup>12a</sup> and low data completeness, Rietveld refinement was performed using XPD data (Fig. S4, ESI<sup>†</sup>).

Crystal structure determination revealed that this API is in fact a coordination polymer.<sup>15</sup> All three phenolate oxygen atoms of the gallate ligand coordinate to  $\text{Bi}^{3+}$ , while the carboxylic acid group does not (Fig. 2). The phenolate oxygens in the 3- and 5-positions coordinate to separate  $\text{Bi}^{3+}$ . The phenolate oxygen in the 4-position bridges these two  $\text{Bi}^{3+}$ . Each  $\text{Bi}^{3+}$  is coordinated by two gallate ligands resulting in  $[-\text{Bi}-\text{O}-]_{\infty}$  rods parallel to the  $a$ -axis (Fig. 1b). The rods appear V-shaped when viewed down their lengths due to the positioning of the protruding ligands (Fig. 1d). Additionally, one water molecule also bridges every pair of adjacent  $\text{Bi}^{3+}$ . The coordination geometry around  $\text{Bi}^{3+}$  is that of a severely distorted octahedron.

Although hydrogen atoms were not located in the difference Fourier maps it is evident from the classical hydrogen bond pattern that the carboxylic acid remains protonated. The carboxylic acid groups protrude out perpendicular to the lengths of the rods and form hydrogen bonds with carboxylic acid groups on neighbouring rods with  $\text{O}(\text{H}) \cdots \text{O}$  distances of  $2.82(3) \text{ \AA}$  and  $\text{CO}(\text{H}) \cdots \text{O}$  angles of  $119(1)^{\circ}$  (Fig. 2c and Fig. S6, ESI<sup>†</sup>).

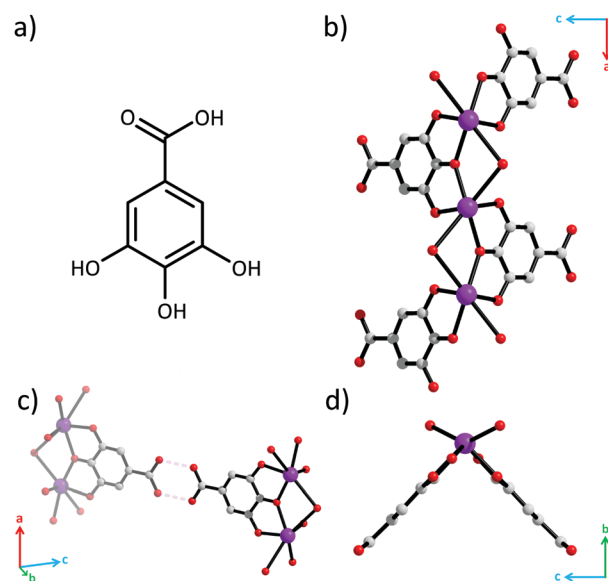


Fig. 2 (a) Gallic acid. (b)  $[-\text{Bi}-\text{O}-]_{\infty}$  rods in bismuth subgallate. Bismuth, carbon and oxygen atoms are represented as purple, grey and red spheres. Hydrogen atoms are omitted for clarity. (c) Hydrogen bonds between carboxylic acids. (d) Rods viewed down the  $a$ -axis.



This classical hydrogen bond motif holds the rods together into zigzag-shaped layers (Fig. 3). The layers stack resulting in channels parallel with the *b*-axis.

Two water molecules per asymmetric unit were located in the pores forming hydrogen bonds with one another as well as with the carboxylic acid and the bridging water molecule in the rods (Fig. S5, ESI†). The pore volume was determined by the SQUEEZE routine in PLATON<sup>16</sup> as 45 Å<sup>3</sup> per Bi<sup>3+</sup> which suggests that fewer than three water molecules can reside in the pores, assuming non-hydrogen atoms occupy an estimated volume of 18 Å<sup>3</sup>.

We propose that charge balance of Bi<sup>3+</sup> is achieved through deprotonation of the three phenolate groups. Bi–O distances of 2.03(2), 2.227(8), and 2.03(2) Å are observed (Fig. S6, ESI†) which are more consistent with Bi<sup>3+</sup>–O<sup>−</sup> distances in structures where two Bi<sup>3+</sup> are bridged by a deprotonated phenolate group (*ca.* 2.2 Å),<sup>17</sup> rather than the longer protonated Bi<sup>3+</sup>–OH phenol bridge (*ca.* 2.6 Å).<sup>18</sup> The rather long Bi<sup>3+</sup>–OH<sub>2</sub> distance of 2.83(2) Å observed in the title compound is more consistent with typical bond lengths of water bridging two Bi<sup>3+</sup> (*ca.* 2.8 Å)<sup>17b,19</sup> rather than hydroxyl groups (*ca.* 2.2 Å).<sup>20</sup>

Thus we propose that the chemical composition of bismuth subgallate should be formulated as [Bi(C<sub>6</sub>H<sub>2</sub>(O)<sub>3</sub>COOH)(H<sub>2</sub>O)]<sub>*n*</sub>·2*n*H<sub>2</sub>O.

Thermogravimetric analysis performed in air (Fig. S7, ESI†) indicated a weight loss of 11% (*calc.* 12%) between 30–150 °C, attributed to the loss of the three water molecules per Bi<sup>3+</sup> in

agreement with previous reports.<sup>21</sup> However we attribute the loss as two water molecules from the pores and the third as the bridging water molecule in the rods. Subsequent loss of the organic ligand occurred between 260–300 °C.

The X-ray thermodiffraction data (Fig. S8, ESI†) indicate a partial degradation in crystallinity starting at 150 °C, coinciding with the removal of the water molecules. The material becomes amorphous at 300 °C. Heating a pristine sample in air to 175 °C followed by cooling back down to 50 °C and leaving it in air for one hour allowed full crystallinity to be recovered (Fig. S9 and S10, ESI†). Alternatively, adding a drop of water immediately restores full crystallinity. The crystal structure of bismuth subgallate was found to remain intact after stirring samples in aqueous HCl solutions at 37 °C (Fig. S11, ESI†) indicating stability under acidic conditions similar to that of gastric acid (pH 1.5–3.5). The poor solubility and good stability of the material in acidic media are attributed to the extended structure and the strong Bi<sup>3+</sup>–O<sub>(phenolate)</sub><sup>−</sup> bonds.

Bismuth compounds tend to crystallize as dense phases and we are only aware of a few porous bismuth coordination polymers.<sup>22</sup> The N<sub>2</sub> adsorption isotherm of bismuth subgallate recorded at 78 K showed an IUPAC Type II isotherm indicative of a non-porous material (Fig. S12, ESI†).<sup>23</sup> However, at 273–293 K noticeable uptake of both CO<sub>2</sub> and N<sub>2</sub> was detected, with high selectivity towards CO<sub>2</sub> (Fig. 4 and Fig. S13–S15, ESI†). The uptake of both gases unexpectedly increased when the temperature was raised from 273 to 283 K (CO<sub>2</sub>: from 0.94 mmol g<sup>−1</sup> to 1.04 mmol g<sup>−1</sup>, N<sub>2</sub>: 0.10 mmol g<sup>−1</sup> to 0.18 mmol g<sup>−1</sup>, at 101 kPa). As adsorption is an exothermic process, such increase in gas uptake with increasing temperature was unexpected. Both the uptake of CO<sub>2</sub> and N<sub>2</sub> reduced when the temperature was further increased from 283 to 293 K (CO<sub>2</sub>: 1.01 mmol g<sup>−1</sup>, N<sub>2</sub>: 0.16 mmol g<sup>−1</sup>, at 101 kPa). The unexpected increase in gas uptake at 283 K suggests that the gas accessible voids of bismuth subgallate are temperature dependent, which implies that the structure of bismuth subgallate is slightly expanding or rearranging with varying temperatures.

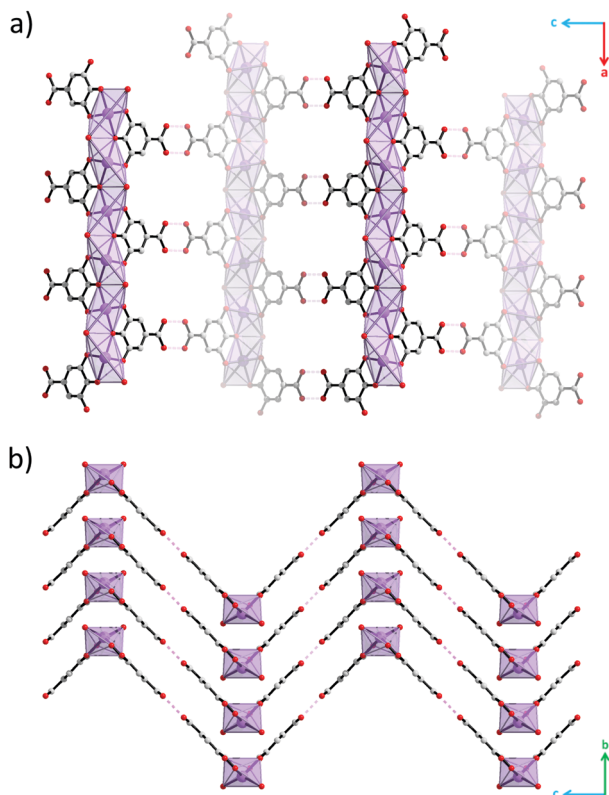


Fig. 3 Crystal structure of bismuth subgallate viewed along (a) [010] and (b) [100]. Bismuth, carbon and oxygen atoms are coloured purple, grey and red respectively. Hydrogen atoms and water molecules in the pores have been omitted for clarity.

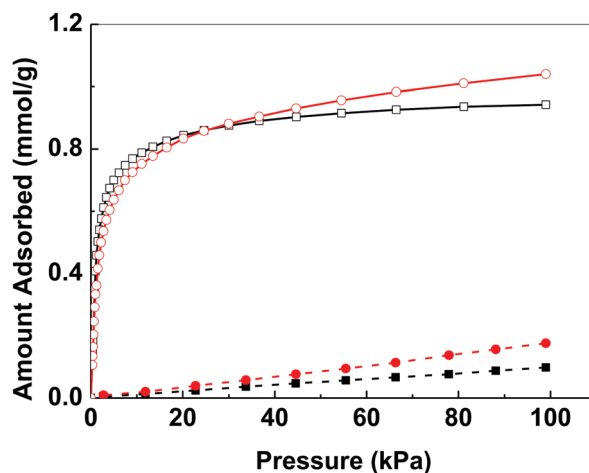


Fig. 4 Gas adsorption isotherms of bismuth subgallate recorded at 273 K (CO<sub>2</sub>: □, N<sub>2</sub>: ■) and 283 K (CO<sub>2</sub>: ○, N<sub>2</sub>: ●).



In conclusion, bismuth subgallate was revealed as a coordination polymer in contrast to the molecular formulas suggested in the past. The structure of bismuth subgallate significantly differs from those of other bismuth-based APIs such as bismuth subsalicylate,<sup>9d,e</sup> which forms isolated clusters, and bismuth subcitrate which can exist as an open framework.<sup>7a</sup> The material also displayed interesting gas sorption properties that warrant further investigations which may open up new application areas for bismuth compounds. Continuous rotation electron diffraction performed under low temperature was paramount for collecting data of sufficient quality. These results lead the way for the structure determination of other beam sensitive crystals, which is important for the development pharmaceutical compounds and understanding their functions.

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## Notes and references

‡ Crystallographic data: C<sub>7</sub>H<sub>9</sub>BiO<sub>8</sub>,  $M_r = 430.1$ , orthorhombic, *Pmna*,  $a = 8.5422(2)$ ,  $b = 4.66793(9)$ ,  $c = 23.7131(4)$  Å,  $V = 945.54(3)$  Å<sup>3</sup>,  $Z = 4$ ,  $\lambda = 0.15406$  Å (Cu<sub>Kα1</sub>),  $R_p = 11.8$ ,  $R_{wp} = 16.43$ ,  $R_{exp} = 9.41$ ,  $GoF = 1.75$ . Structure determination with electron diffraction data and Rietveld refinement with X-ray powder diffraction data were conducted using SHELX<sup>24</sup> and TOPAS<sup>25</sup> respectively. CCDC 1526756 contains the supplementary crystallographic data for this paper.†

- 1 C. Beck, *J. Am. Med. Assoc.*, 1908, **50**, 1233–1236.
- 2 L.-M. Mai, C.-Y. Lin, C.-Y. Chen and Y.-C. Tsai, *Biomaterials*, 2003, **24**, 3005–3012.
- 3 (a) S. L. Gorbach, *Gastroenterol.*, 1990, **99**, 863–875; (b) M. Sparberg, *Gastroenterol.*, 1974, **66**, 476–480; (c) C.-Y. Lin, Y.-H. Shen, S.-H. Wu, C.-H. Lin, S.-M. Hwang and Y.-C. Tsai, *Biochem. Biophys. Res. Commun.*, 2004, **315**, 830–835.
- 4 (a) D. M. Keogan and D. M. Griffith, *Molecules*, 2014, **19**, 15258–15297; (b) J. A. Salvador, S. A. Figueiredo, R. M. Pinto and S. M. Silvestre, *Future Med. Chem.*, 2012, **4**, 1495–1523.
- 5 (a) R. M. Peek and M. J. Blaser, *Nat. Rev. Cancer*, 2002, **2**, 28–37; (b) A. H. Mendis and B. J. Marshall, *Biological Chemistry of Arsenic, Antimony and Bismuth*, Wiley, John Wiley & Sons Ltd, Singapore, 2010, ch. 10.
- 6 J. M. Liou, M. S. Wu and J. T. Lin, *J. Gastroenterol. Hepatol.*, 2016, **31**, 1918–1926.
- 7 (a) W. Li, L. Jin, N. Zhu, X. Hou, F. Deng and H. Sun, *J. Am. Chem. Soc.*, 2003, **125**, 12408–12409; (b) G. Tytgat, *Digestion*, 1987, **37**, 31–41.
- 8 (a) N. Chiba, *Can. J. Gastroenterol. Hepatol.*, 2000, **14**, 885–889; (b) D. Y. Graham and S.-Y. Lee, *Gastroenterol. Clin. North Am.*, 2015, **44**, 537–563.
- 9 (a) Y. Yang, R. Ouyang, L. Xu, N. Guo, W. Li, K. Feng, L. Ouyang, Z. Yang, S. Zhou and Y. Miao, *J. Coord. Chem.*, 2015, **68**, 379–397; (b) H. Sun, *Biological chemistry of arsenic, antimony and bismuth*, John Wiley & Sons Ltd, Chichester, UK, 2010; (c) H. Li and H. Sun, *Curr. Opin. Chem. Biol.*, 2012, **16**, 74–83; (d) P. C. Andrews, G. B. Deacon, C. M. Forsyth, P. C. Junk, I. Kumar and M. Maguire, *Angew. Chem., Int. Ed.*, 2006, **45**, 5638–5642; (e) V. André, A. Hardeman, I. Halasz, R. S. Stein, G. J. Jackson, D. G. Reid, M. J. Duer, C. Curfs, M. T. Duarte and T. Frišćić, *Angew. Chem., Int. Ed.*, 2011, **50**, 7858–7861.
- 10 N. Yang, Y. An, J. Cai, L. Hu, Y. Zeng, Z. Mao, G. Chen and H. Sun, *Sci. China: Chem.*, 2010, **53**, 2152–2158.
- 11 (a) U. Kolb, T. Gorelik, C. Kübel, M. Otten and D. Hubert, *Ultra-microscopy*, 2007, **107**, 507–513; (b) D. Zhang, P. Oleynikov, S. Hovmöller and X. Zou, *Z. Kristallogr.*, 2010, **225**, 94–102; (c) W. Wan, J. Sun, J. Su, S. Hovmöller and X. Zou, *J. Appl. Crystallogr.*, 2013, **46**, 1863–1873.
- 12 (a) Y. Yun, X. Zou, S. Hovmöller and W. Wan, *IUCrJ*, 2015, **2**, 267–282; (b) E. Van Genderen, M. Clabbers, P. Das, A. Stewart, I. Nederlof, K. Barentsen, Q. Portillo, N. Pannu, S. Nicolopoulos and T. Gruene, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2016, **72**; (c) M. D. Eddleston, K. E. Hejczyk, E. G. Bithell, G. M. Day and W. Jones, *Chem. – Eur. J.*, 2013, **19**, 7874–7882.
- 13 M. Gemmi, M. G. La Placa, A. S. Galanis, E. F. Rauch and S. Nicolopoulos, *J. Appl. Crystallogr.*, 2015, **48**, 718–727.
- 14 H. Furukawa, K. E. Cordova, M. O’Keeffe and O. M. Yaghi, *Science*, 2013, **341**, 1230444.
- 15 S. Kitagawa, R. Kitaura and S.-i. Noro, *Angew. Chem., Int. Ed.*, 2004, **43**, 2334–2375.
- 16 A. Spek, *J. Appl. Crystallogr.*, 2003, **36**, 7–13.
- 17 (a) L. Liu, L. N. Zakharov, J. A. Golen, A. L. Rheingold and T. A. Hanna, *Inorg. Chem.*, 2008, **47**, 11143–11153; (b) D. Mendoza-Espinosa, A. L. Rheingold and T. A. Hanna, *Dalton Trans.*, 2009, 5226–5238.
- 18 D. Mansfeld, L. Miersch, T. Ruffer, D. Schaarschmidt, H. Lang, T. Böhle, R. W. Troff, C. A. Schalley, J. Müller and M. Mehring, *Chem. – Eur. J.*, 2011, **17**, 14805–14810.
- 19 (a) A. Ilyukhin and A. Poznyak, *Crystallogr. Rep.*, 2000, **45**, 56–63; (b) H. Wullens, B. Tinant, J.-P. Declercq and M. Devillers, *Inorg. Chim. Acta*, 2003, **343**, 335–342.
- 20 (a) H. J. Breunig, N. Haddad, E. Lork, M. Mehring, C. Mügge, C. Nolde, C. I. Rat and M. Schürmann, *Organometallics*, 2009, **28**, 1202–1211; (b) P. C. Junk and L. M. Louis, *Z. Anorg. Allg. Chem.*, 2000, **626**, 556–559; (c) L. J. Barbour, S. J. Belfield, P. C. Junk and M. K. Smith, *Aust. J. Chem.*, 1998, **51**, 337–342; (d) A. Fridrichová, T. s. Svoboda, R. Jambor, Z. k. Padělková, A. Růžička, M. Erben, R. Jirásko and L. Dostál, *Organometallics*, 2009, **28**, 5522–5528.
- 21 Y. M. Yukhin, O. Logutenko, I. Vorsina and V. Evseenko, *Theor. Found. Chem. Eng.*, 2010, **44**, 749–754.
- 22 (a) M. Feyand, E. Mugnaioli, F. Vermoortele, B. Bueken, J. M. Dieterich, T. Reimer, U. Kolb, D. De Vos and N. Stock, *Angew. Chem., Int. Ed.*, 2012, **51**, 10373–10376; (b) M. Savage, S. Yang, M. Suyetin, E. Bichoutskaia, W. Lewis, A. J. Blake, S. A. Barnett and M. Schröder, *Chem. – Eur. J.*, 2014, **20**, 8024–8029; (c) A. Thirumurugan and A. K. Cheetham, *Eur. J. Inorg. Chem.*, 2010, 3823–3828; (d) A. K. Inge, M. Köppen, J. Su, M. Feyand, H. Xu, X. Zou, M. O’Keeffe and N. Stock, *J. Am. Chem. Soc.*, 2016, **138**, 1970–1976.
- 23 K. S. Sing, *Pure Appl. Chem.*, 1985, **57**, 603–619.
- 24 G. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2007, **64**, 112–122.
- 25 A. Coelho, Bruker AXS, Karlsruhe, Germany, 2007.

