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Critical Review

COMPREHENSIVE REVIEW OF SEVERAL SURFACTANTS IN MARINE ENVIRONMENTS: FATE AND ECOTOXICITY

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Abstract: Surfactants are a commercially important group of chemicals widely used on a global scale. Despite high removal efficiencies during wastewater treatment, their high consumption volumes mean that a certain fraction will always enter aquatic ecosystems, with marine environments being the ultimate sites of deposition. Consequently, surfactants have been detected within marine waters and sediments. However, aquatic environmental studies have mostly focused on the freshwater environment, and marine studies are considerably underrepresented by comparison. The present review aims to provide a summary of current marine environmental fate (monitoring, biodegradation, and bioconcentration) and effects data of 5 key surfactant groups: linear alkylbenzene sulfonates, alcohol ethoxysulfates, alkyl sulfates, alcohol ethoxylates, and ditallow dimethyl ammonium chloride. Monitoring data are currently limited, especially for alcohol ethoxysulfates and alkyl sulfates. Biodegradation was shown to be considerably slower under marine conditions, whereas ecotoxicity studies suggest that marine species are approximately equally as sensitive to these surfactants as freshwater species. Marine bioconcentration studies are almost nonexistent. Current gaps within the literature are presented, thereby highlighting research areas where additional marine studies should focus. Environ Toxicol Chem 2016;35:1077-1086. © 2015 SETAC

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INTRODUCTION

Surfactants are a diverse group of economically important chemicals widely used in cleaning detergents, personal care products, and various industrial applications (e.g., oil, textiles, polymers, agriculture, paints) [1]. The global surfactant market is forecast to grow at a compound annual rate of 6.02% from 2015 to 2019 (key regions being Asia-Pacific, Europe, North and South America, and the rest of the world), with rising demand for personal care products being the market driver [2]. Surfactants are comprised of both hydrophobic and hydrophilic moieties and are classified by their ionic properties in water as either anionic (negative charge), nonionic (no charge), cationic (positive charge), or amphoteric (positive/negative charge depending on pH) [3]. Among the most important anionic surfactants (by production volume) are the linear alkylbenzene sulfonates (LAS), alcohol ethoxysulfates (AES), and alkyl sulfates (AS). The most important nonionic surfactants (by volume) are the alcohol ethoxylates (AE). Ditallow dimethyl ammonium chloride (DTDMAC), a quaternary ammonium compound is a historically commonly used cationic surfactant [4]. Given their importance, these 5 surfactant groups (Figure 1) are the focus of the present review.

Commercial LAS contain homologous mixtures from C₁₀ to C_{14} (average chain length, $C_{11,7}$ – $C_{11,8}$) [5] and are currently the most commonly used group of anionic surfactants in Europe,

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with consumption estimates of 497 818 tons in 2013 (European Committee of Organic Surfactants and their Intermediates [CESIO], Brussels, Belgium, personal communication). However, commercial AES are notably becoming increasingly important, with consumption rates increasing to overtake those of LAS in certain parts of the world (e.g., North America) [6]. Alcohol ethoxysulfates contain variable alkyl chain lengths $(C_{12}-C_{18})$ and ethoxylated chain lengths (0-8) [7]. European AES consumption estimates were 456160 tons in 2013 (CESIO, Brussels, Belgium, personal communication). Commercial AS contain variable chain lengths from C_{12} to C_{18} [8]. European AS consumption estimates were 65 885 tons in 2013 (CESIO, Brussels, Belgium, personal communication). Commercial AE contain variable hydrocarbon chain lengths (C_8-C_{18}) and ethoxylated chain lengths (0-22). European AE consumption estimates were 371 609 tons in 2013 (CESIO, Brussels, Belgium, personal communication). The DTDMAC surfactant is no longer used commercially on a large geographic scale and has since been substituted with readily biodegradable alternatives [4].

Following their use, surfactants will typically enter wastewater-treatment plants, where removal has been shown to be highly efficient (95–99% average removal [7–11]). However, given their globally high consumption volumes, there is always a certain fraction that is not removed. Thus, surfactants enter aquatic ecosystems via wastewater discharge [12,13]. The marine environment therefore receives almost continuous surfactant input, either directly (from treated and untreated wastewaters discharges) or indirectly (via contaminated rivers) and is regarded as the final site of deposition for surfactants [14-17]. Consequently, surfactants have been

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Figure 1. General chemical structures of the surfactants (A) linear alkylbenzene sulfonate, (B) alcohol ethoxysulfate, (C) alkyl sulfate, (D) alcohol ethoxylate, and (E) ditallow dimethyl ammonium chloride.

detected within marine waters and sediments [18–21]. Once within the marine environment, surfactants may be removed via volatilization (albeit only to a limited extent), adsorption to particles, abiotic or microbial degradation, or uptake by marine organisms [3]. Degradation and sorption are the main mechanisms involved [1].

As such, marine environments may act as sinks for surfactants as a result of sorption to particles that sink to the sediment bed, potentially leading to their accumulation [3]. Rubio et al. [22] observed <6% LAS recovery from marine sediments and concluded (by fitting experimental data to a Freundlich model) that sorption of surfactants to marine sediments was irreversible. Estuarine and coastal environments are considered the most productive yet sensitive ecosystems on earth; hence, exposures can have significant environmental implications [23,24]. Furthermore, some studies suggest that certain marine organisms are more sensitive than freshwater organisms [25,26] (although it is suggested that such variability in toxicity depends largely on species choice [27]). Marine studies should therefore be strongly considered when addressing the environmental effects of surfactants.

Emphasis on protecting the marine environment has increased over the years, with concerns that hazardous substances may accumulate to significant toxic concentrations to cause unpredictable, potentially irreversible long-term effects. It has also been emphasized that remote ocean areas should remain untouched by hazardous substances and that pristine marine environments should be protected [28]. Similar to other environmental compartments, the most important surfactant-related environmental issues considered in the marine environment are whether there is risk of direct toxicity or whether biodegradation, bioaccumulation, and biomagnification pose reason for concern [3]. However, most reported ecotoxicity studies concerning surfactants have focused on freshwater species, whereas marine studies are comparatively lacking. The same is true of marine biodegradation studies, whereas marine bioaccumulation studies are practically nonexistent [3,29].

An extensive review of currently available marine studies on the 5 previously mentioned surfactants (LAS, AES, AS, AE, and DTDMAC) was therefore conducted in association with the Environmental Risk Assessment of Surfactants Management (although, given the diverse and large-scale application of surfactants, this work would also be of interest to other organizations, such as the Oslo/Paris convention for the protection of the marine environment of the northeast Atlantic and the European Oilfield Specialty Chemicals Association), taking into account monitoring, biodegradation, bioaccumulation, and ecotoxicity studies. Comparison is also made with general freshwater data. The present review ultimately aims to provide a comprehensive summary of such studies and to highlight current gaps within the literature.

FATE

Monitoring

The amount of marine sediment monitoring data found, from highest to lowest, was AE > LAS > DTDMAC > AES > AS (Table 1). Detected concentrations ranged from 0.0074 mg/kg to 9.19 mg/kg in AE [30], <0.003 mg/kg to 15.63 mg/kg in LAS [13,21,30,31], 0.0048 mg/kg to >25 mg/kg in DTDMAC [30], 0.061 mg/kg to 14.32 mg/kg in AES [21,32–34], and 0.13 mg/kg in AS [21]. Measured concentrations in sediment were higher when sampling was undertaken in close proximity to direct discharge from sewage-treatment plant effluents, whereas samples taken from offshore locations were not impacted by direct discharge and consequently showed much lower surfactant concentration levels. Based on the upper range values, detected concentration levels are, from highest to lowest, DTDMAC > LAS > AES > AE > AS. High

Table 1. Summary of marine data values reported for the surfactants: Linear alkylbenzene sulfonate, alcohol ethoxysulfate, alkyl sulfate, alcohol ethoxylate, and ditallow dimethyl ammonium chloride^a

	LAS	AES	AS	AE	DTDMAC	Total
Degradation	37	8	7	13	ND	65
Bioconcentration	3	ND	1	ND	ND	4
Monitoring	19	4	1	20	10	54
Acute algae toxicity	26	8	12	ND	2	48
Acute invertebrate toxicity	127	9	240	51	4	431
Acute fish toxicity	23	3	34	1	1	62
Chronic algae/marine plant toxicity	19	10	7	1	ND	37
Chronic invertebrate toxicity	41	ND	50	4	1	96
Chronic fish toxicity	8	ND	8	ND	ND	16
Marine bacteria toxicity	1	ND	ND	ND	ND	1
Total	304	42	360	90	18	

^aNote that classification of chronic studies is somewhat subjective.

LAS = linear alkylbenzene sulfonate; AES = alcohol ethoxysulfate; AS = alkyl sulfate; AE = alcohol ethoxylate; DTDMAC = ditallow dimethyl ammonium chloride; ND = no data found.

DTDMAC concentrations can be attributed to their higher persistence levels (because of their poor biodegradability coupled with their positive cationic charge, causing them to strongly adsorb to negatively charged sediment surfaces). As mentioned, DTDMAC commercial consumption has been considerably reduced on a geographic scale because of their high persistence levels [4]. The concentration trend seen in the other surfactants corresponds to their reported consumption rates. Monitoring data for AS and AES are comparatively scarce in comparison with the other surfactants reviewed in the present study. Lara-Martin et al. [19] argued (in the case of AES) that the main reason for this is that these compounds are not volatile and do not fluoresce; therefore, conventional highperformance liquid chromatography with ultraviolet or fluorescence detectors and gas chromatography with mass spectrometry cannot be used. However, gas chromatography with mass spectrometry and liquid chromatography with mass spectrometry are now being applied more frequently for specific analysis of surfactants [35-41].

Biodegradation

Evidence on surfactant removal by abiotic or biotic degradation is important, since this largely determines their

fate and persistence in the environment. The number of marine degradation studies found during the present review, from highest to lowest, is LAS > AE > AES > AS (no marine DTDMAC studies were found; Table 1). Marine studies are evidently scarce, despite the role of these environments as ultimate recipients of domestic and industrial wastewaters. The most common endpoints from the obtained studies were parent disappearance in LAS and AS and ¹⁴CO₂ evolution in AE and AES. Table 2 provides a summary of the marine degradation data found as well as a comparison with typical freshwater halflife values. Studies that reported half-lives >60 d were considered unreliable based on European Centre for Ecotoxicology and Toxicology of Chemicals recommendations [28] and therefore were excluded. Based on the mean half-lives presented, marine degradability of the 5 surfactants, from highest to lowest, is AS > LAS > AE > AES (although there is evidently large overlap between half-life data across all 4 surfactants, suggesting similarity). This trend is consistent with previous results in that AS exceeds all other anionic surfactants in biodegradation rates [4,42,43]. However, it should be noted that although AS presents the lowest mean half-life of the 4 surfactants examined, it is also the most data-deficient; hence, the calculated mean is not considered as reliable as that of LAS

Table 2. Summary of marine degradation data ranges for the surfactants and half-life comparison with typical freshwater values^a

Compound		Typical freshwater degradation					
	Primary degradation rate (d ⁻¹)	Lag (d)	Mineralization (%)	Half-life (d)	References	Half-life (d)	Reference
LAS	0.02 - 0.19 (0.11; $n = 9$)	0-6.67 (1.45; $n=15$)	10-60.4 (24.52; $n=13$)	0.3 - 45 (8.67; $n = 37$)	[11,15,23,24,29,42– 44,46,48,50,51,55,83–86]	0.025–0.5 (0.16)	[10]
AES	0.1 - 0.39 (0.28; <i>n</i> =4)	0.65 - 26.5 (10.15; $n = 3$)	0-96.7 (71.79; $n=7$)	1 - 49.8 (14.07; $n = 8$)	[11,83–85]	0.042–1.4 (0.72)	[7]
AS	—	7 - 14 (10.5; $n = 2$)	0 (n = 1)	0.26 - 20 (6.78; $n = 7$)	[11,42,43,49,84]	0.3–1.0 (0.75)	[8]
AE	0.02 - 0.34 (0.14; $n = 13$)	0-3 (0.86; $n=7$)	8-87.2 (49.2; $n=11$)	2.3n-28 (11.76; $n = 13$)	[83,87,88]	0.17–1.0 (0.46)	[9]
DTDMAC	—	—	—	—	_	—	—

^aMarine studies were conducted at 20 °C to 25 °C (with the exception of George [49], Leon et al. [50], and Mauffret et al. [51], which included studies performed at -1.8-0.65 °C, 10 °C, and 10-18 °C, respectively). Representative freshwater data were collected at lower temperatures than marine studies (7–27 °C). Data are based on first-order degradation models. Mean values are given in parentheses, followed by the number of data values used. LAS = linear alkylbenzene sulfonate; AES = alcohol ethoxysulfate; AS = alkyl sulfate; AE = alcohol ethoxylate; DTDMAC = ditallow dimethyl ammonium chloride.

(the second most rapidly degradable surfactant examined in the present review). Current studies suggest that mineralization in marine conditions is highest in AES (mean, 71.8%), followed by AE (mean, 49.2%) and LAS (mean, 24.5%; Table 2); AS mineralization data are currently severely limited. There is considerable variability among studies in all 4 surfactants (shown by large data ranges), with many studies revealing marine surfactant biodegradation to be a long process. It is argued that such data variability may be explained by natural variation (between different environments) and microbial variation by different pretreatment methods (e.g., acclimation history of associated microbial communities to surfactants [24,44,45]), humic substance association, cometabolic transformation, or nutrient-level variation [46]. Primary biodegradability of LAS homologs has been shown to increase with alkyl chain length [23,47], although Larson et al. [24] found that LAS mineralization was relatively unaffected by such structural differences. The biodegradability of AE and AES is relatively unaffected by alkyl and ethoxylated chain lengths, although branching does hinder it [4]. Studies that included marine sediments showed increased extent [24,44] and rates [15,48] of biodegradation as a result of additional microbial biomass, organic matter, or nutrients [3].

Although the majority of marine studies obtained in the present review were conducted at 20 °C to 25 °C, some were conducted at lower temperatures (-1.8–0.65 °C [49], 10 °C [50], 10–18°C [51]), demonstrating comparatively longer half-lives. Temperature is widely known to influence biodegradation, with higher temperatures causing enhanced microbial metabolic activity and reduced lag times and lower temperatures inhibiting degradation [3,49,50,52]. Terzić et al. [23] detected a lag time of 2 d for LAS at 14 °C but not at 23 °C (although this difference was detected within freshwater estuary layers as opposed to saline layers). Antarctic coastal half-lives generally have been shown to be far longer than those in temperate waters [49]. Some studies have therefore argued that marine surfactant contamination may worsen in winter months as a result of reduced biodegradation rates [23,48,50,52]. In contrast, however, other studies suggest that seasonal temperature variations have little to no effect on surfactant biodegradation under realistic environmental conditions [49,53].

Comparison with typical freshwater half-life values suggests that (like other chemicals) degradation of surfactants is much slower in marine than freshwater environments, thereby corresponding with previous reports [3,24,29,44]. It was suggested that this comparatively slower degradation might be explained by marine microbial communities being less active than their freshwater counterparts toward xenobiotic chemicals. It was also suggested that complexation with calcium and magnesium ions in seawater reduces bioavailability (particularly at low concentrations), thereby inhibiting biodegradation [44]. In contrast, Quiroga and Sales [54] found no difference in the extent of LAS biodegradation over 21 d under varying salinities (16-65%); however, induction periods were notably shorter under higher salinities (50-65%). It was argued that this may have been an artifact of bacterial culture dilution causing reduced culture numbers (and consequently reduced biodegradation). Many of the representative freshwater data in Table 2 reportedly used temperatures that were comparatively lower than (most) marine studies found in the present review (freshwater temperatures: AE, ~12 °C; AES, 22 ± 3 °C; AS, 10–27 °C; LAS, 7–27 °C). However, despite these lower temperatures, half-lives were still faster in freshwater than marine conditions. Perales et al. [29] concluded that under similar temperatures (and initial concentrations), LAS degraded more slowly in marine than freshwater conditions, demonstrating slower half-life and lag times in marine conditions.

Bioconcentration

Bioconcentration and bioaccumulation measure the net accumulation of a chemical within an organism as a result of uptake via exposure to the material (from either the surrounding environment only [bioconcentration] or the surrounding environment and food [bioaccumulation]). Such accumulations may eventually lead to concentration levels capable of causing toxic effects within the organism or net accumulation of the chemical to predator organisms through the food chain (biomagnification).

Current aquatic bioconcentration/bioaccumulation studies in surfactants have almost exclusively focused on freshwater species, generally concluding that surfactants (LAS, AES, AS, and AE) possess low accumulation potential because of metabolism and subsequent elimination from the organism [4]. Marine studies are practically nonexistent. Guarino et al. [55] reported sodium lauryl sulfate (AS) bioaccumulation in the spiny dogfish (Squalus acanthias) following a 1-mg/kg dose via injection over 2h to 144h. However, the study was nonstandard, given the method of exposure used (injection into body tissue). According to Sáez et al. [56], C₁₁LAS body concentrations between 1 mg/kg and 3 mg/kg wet weight were found in bivalves and fish collected from less contaminated sites $(0.005 \text{ mg/L }C_{11}\text{LAS}$ in water samples) and more contaminated sites (0.05 mg/L C₁₁LAS in water samples), respectively, of the Bay of Cadiz (Spain) [56,57], with bioconcentration factors (BCFs) said to range from 60 L/kg to 200 L/kg [58]. Furthermore, body concentrations of LAS metabolites (i.e., sulfophenylcarboxylic acids) were below detection limits, suggesting that they were eliminated from organisms [56]. Renaud et al. [59] reported a mean BCF of 120 L/kg in the marine shrimp Palaemonetes varians after 7-d C12-6-LAS exposure under realistic environmental concentrations. The estimated BCF at steady state (BCF_{ss}) was 159 L/kg, which was reached after 11.5-d exposure. Major accumulation was found in the cephalothorax circulatory system (gills, heart, hepatopancreas) and ocular peduncle but not flesh (implying limited transfer potential to human consumers). Depuration was rapid, showing < 1% of initial LAS after 8 d [59]. Álvarez-Muñoz et al. [60] found BCF_{ss} values of 17 L/kg for C₁₀-2-LAS and 387 L/kg for C12-2-LAS in Solea senegalensis under realistic environmental concentrations. Biotransformation and elimination also were reported by identifying and quantifying LAS metabolites in organisms and depuration water. From these limited marine studies, it has been concluded that LAS presents no risk of bioaccumulation at environmental concentration levels [59,60].

Renaud et al. [59] noted that marine BCF values obtained in their study were similar to those previously observed for freshwater organisms and concluded that salinity has no significant impact on LAS bioconcentration. However, it has also been argued that extrapolating freshwater data to marine conditions should be done with caution, given the lower relevance of freshwater data compared with marine data, particularly for surfactants that are used offshore [3]. Freshwater results have shown that the accumulation potentials of surfactants in the aqueous phase are generally below the conventional levels for concern (i.e., log octanol–water partition coefficient values of 3-4) [3]. Freshwater studies have also shown high rates of biotransformation in surfactants (in vitro studies in freshwater fish have also shown this to be the case [61]); hence, their accumulation potential is considered to be low [62-64]. Moreover, some freshwater bioconcentration data generated with radiolabeled test compounds should be regarded as overestimates if parent and metabolites have not been quantified separately and the BCF is based on the total combined fractions [3]. Slight increases in LAS and AE alkyl chain lengths (and decreasing ethoxylated chain length in AE)-that is, increasing hydrophobicity-were shown to significantly increase bioconcentration potential in Pimephales promelas [63,65], whereas increasing hydrophilicity reduces bioconcentration potential [3]. Furthermore, the closer the positioning of the *p*-sulfophenyl group to the terminal carbon of the alkyl chain in LAS, the higher the BCF [65]. Biomagnification is also considered unlikely since surfactants lack the properties necessary to remain stable in the environment (i.e., they are rapidly degraded and metabolized and are not highly hydrophobic) [3].

EFFECTS

The number of reported marine ecotoxicity data on the 5 surfactants, from highest to lowest, is AS > LAS > AE > AES > DTDMAC (Table 1). The taxa most commonly used, from highest to lowest, are invertebrates > algae > fish (1 marine bacteria study was found for LAS [66]); however, the exact marine species used vary considerably between studies. In the present review, data are considered acute if they were described as such in the study, if the endpoint was presented as a 50% effective or lethal concentration (EC50 or LC50), or if exposure duration was equal to or less than that recommended in standardized test guidelines (e.g., 96-h fish exposure, 48-h invertebrate exposure). Data are considered chronic if they were described as such in the study; if the end point was presented as a lowest-observed-effect concentration (LOEC), a no-observed-effect concentration (NOEC), a 10% lethal or effective concentration (LC10 or EC10), or an effect that would influence the long-term well-being of the test species (e.g., reduced byssal activity in *Mytilus edulis*); or if exposure duration was considerably longer than would be expected from a standardized acute study. Acute studies (particularly lethality) are most abundant (especially in invertebrates), whereas chronic studies are much less common (particularly in fish). Marine study data are currently lacking for acute algae and chronic fish for AE, chronic invertebrate and fish for AES, and chronic algae and fish for DTDMAC. It is also apparent (from Supplemental Data, Tables S1 and S2) that marine sediment studies are scarce, relative to seawater studies, and mostly use bulk sediment (mg/kg) as an exposure metric. These marine sediment studies all used LAS as a test compound, with acute toxicity ranging from 4.18 mg/kg to 4.77 mg/kg (mean, 4.48 mg/kg) [67,68], 2 mg/kg to 295 mg/kg (mean, 102.06 mg/kg) [14,25,69–73], and 876 mg/kg to 2180 mg/kg (mean, 1528 mg/kg) [73,74] in algae, invertebrates, and fish, respectively. Chronic toxicity ranged from 0.35 mg/kg to 561 mg/kg (mean, 105.4 mg/kg) [25,70,73,75] and 223 mg/kg to 755 mg/kg (mean, 601 mg/kg) [73,74] in invertebrates and fish, respectively (chronic algal sediment data were lacking). Adsorption of surfactants to marine sediments is believed to be one of their main removal mechanisms [1]; thus, marine sediment toxicity data should be considered important. However, given that exposure of

benthic organisms occurs via interstitial water, it is suggested that an alternative exposure metric (rather than bulk sediment) should be adopted in such studies (i.e., porewater). Rico-Rico et al. [69] showed that C_{12} -2-LAS effect concentrations in porewater were similar to those obtained via water-only exposures in the marine benthic amphipod *Corophium volutator*, suggesting that this would be a suitable alternative metric.

There is evidently considerable variability among the marine data ranges reviewed in the present study (Figures 2 and 3) and, consequently, reason for caution when drawing conclusions. It is suggested that variable test conditions (e.g., temperature, salinity [76,77]), species, and chemical structure among studies may partially explain this [4]. It is also apparent that the vast majority of current marine data are based on nominal rather than measured concentrations (see Supplemental Data, Tables S1 and S2), thereby potentially limiting their reliability. This is because of the difficulty in the separation and quantification of individual components of surfactant products at the required low concentration levels in such toxicity tests. Furthermore, real exposures (i.e., truly dissolved fractions) are often unknown. In general, surfactant toxicity varies among homologs depending on their respective alkyl chain lengths and ethoxylated chain lengths, with longer alkyl chains corresponding to higher toxicity (e.g., C14LAS is more toxic than C₁₀LAS), whereas increasing ethoxylated chain lengths reduce toxicity [4,10]. For instance, the growth inhibition rate was considerably lower in 3 marine microalgae (Tetraselmis suecica, Isochrysis galbana, and Rhodomonas salina) when exposed to C₁₁LAS (72-h EC50 values for reduced algal biomass: 13.37 mg/L, 7.7 mg/L, and 4.43 mg/L, respectively) relative to C13LAS (72-h values for reduced algal biomass values: 1.23 mg/L, 0.54 mg/L, and 0.36 mg/L, respectively) [78]. Likewise, 48-h LC50 values in Crassostrea gigas larvae were lower in C10-12LAS (0.56 mg/L) compared with C₁₂₋₁₄LAS (0.1 mg/L) exposures [79]. Currently, typical LAS homologs within the marine environment have not been studied in detail. Rather, C_{11.6} LAS has been emphasized as a representative homolog for aquatic environmental testing in general and has largely been adopted in marine testing [10]. However, it appears that representative homologs for environmental testing for the other surfactants examined in the present study have not been established (for AE and AES surfactants, this is normally because of the added complexity resulting from variable ethoxylated chain lengths). However, it has been noted that AES toxicity peaks at C₁₆ alkyl chains [4] and that most AS studies have focused on C_{12} homologs. Toxicity also varies depending on whether a surfactant is linear or branched, with branching leading to increased toxicity (however, it has been shown that this is not always the case [9]).

Comparison of freshwater and marine surfactant toxicity has been largely inconsistent within the literature. Ezemonye et al. [71] reported significantly lower 10-d LC50 values in freshwater *Desmoscaris trispinosa* (139 mg/kg) compared with brackish water *Palaemonetes africanus* (259 mg/kg) under LAS exposures. Van de Plassche and de Bruijn [80] considered it unlikely that marine and freshwater organisms differed in sensitivity to AE, although they did conclude that marine species were more sensitive to LAS than freshwater species. In contrast, Swedmark et al. [81] concluded that marine and freshwater fish species show similar sensitivity to LAS. Similarly, Temara et al. [58] argued that there was no significant difference between mean acute LC50 values of





Figure 2. Comparison of acute and chronic aquatic ecotoxicity data ranges of the surfactants (**A**) linear alkylbenzene sulfonate, (**B**) alcohol ethoxysulfate, (**C**) alkyl sulfate, (**D**) alcohol ethoxylate, and (**E**) ditallow dimethyl ammonium chloride from marine studies reviewed in the present study (see Supplemental Data, Tables S1 and S2) and typical freshwater data [4,8–11,89–92]. Blue bars show marine data; red bars show freshwater data. Mean values are presented within individual bars. Acute endpoints include 50% lethal (effective) concentrations and others (Supplemental Data, Table S1). Chronic end points include lowest-observed-effect concentration, no-observed-effect concentration, 10% lethal concentration, and others (Supplemental Data, Table S2). Numbers of marine data (*n*) used to calculate mean values for each type of study are given below bars. Data have been logarithmically scaled.

freshwater and marine organisms to LAS (4.1 mg/L and 4.3 mg/L, respectively); however, comparison of mean chronic NOEC values revealed significantly higher sensitivity in marine organisms than freshwater organisms (0.3 mg/L and 2.3 mg/L, respectively). Figure 2 reveals that marine data generally seem to fall within freshwater data ranges, leading to the conclusion that marine species are approximately equally sensitive to surfactants as freshwater species, thereby agreeing with previous conclusions made by the European Centre for Ecotoxicology and Toxicology of Chemicals [82]. The comparison of marine and freshwater mean toxicity values provided within the present review also largely suggests this; however, there are occasional discrepancies because of limited and highly variable data (e.g., mean AES

acute invertebrate toxicities were 156.4 mg/L and 15.75 mg/L for marine and freshwater species, respectively; however, this marine value is derived from just 9 data values, ranging from 9 mg/L to 1000 mg/L).

Based on the data ranges shown in Figure 3 and the mean toxicities shown in Figure 2, it is concluded that the toxicity of the 5 surfactants, from highest to lowest, is LAS > AS > AE > DTDMAC > AES. Consistently, LAS produced the lowest effect concentration range in all study types (as low as 0.025 mg/L in acute algae studies), whereas AES consistently showed the highest (as high as 1000 mg/L in acute invertebrate studies; Figure 3). The second lowest mean toxicity range was produced by AS, whereas AE produced a lower toxic concentration mean and range compared with DTDMAC



Figure 3. Comparison of acute and chronic marine ecotoxicity data ranges of the surfactants alcohol ethoxylate (AE), alcohol ethoxysulfate (AES), alkyl sulfate (AS), ditallow dimethyl ammonium chloride (DTDMAC), and linear alkylbenzene sulfonate (LAS) from marine studies reviewed in the present study (Supplemental Data, Tables S1 and S2). Black bars represent AE; grey bars represent AES; yellow bars represent AS; blue bars represent DTDMAC; red bars represent LAS. Acute end points include 50% lethal (effective) concentrations and others (see Supplemental Data, Table S1). Chronic end points include lowest-observed-effect concentration, 10% lethal concentration, and others (see Supplemental Data, Table S2). Data have been logarithmically scaled.

(Figures 2 and 3). Acute toxicity for the 5 surfactants examined (for algae, invertebrates, and fish) was generally equisensitive across the board.

CONCLUSIONS

Having reviewed the available marine fate and effects data on the surfactants LAS, AES, AS, and DTDMAC, we can draw several conclusions.

Although these surfactants have been detected in marine environments, monitoring data are limited, especially for AES (4 studies) and AS (2 studies). Marine biodegradation data are scarce, highly variable, and predominantly based on LAS (33 of the 59 reported data were LAS data). Based on mean half-lives given in Table 2, marine degradation is generally a rapid process (although some studies suggest otherwise) but is comparatively slower than freshwater degradation (approximately 54, 20, 9, and 26 times slower for LAS, AES, AS, and AE, respectively).

Marine concentration data are limited to only a few reliable studies for LAS. Based on the available marine data for LAS and other freshwater data for other surfactants, bioconcentration and biomagnification of these surfactants are not expected. Areas where marine ecotoxicology studies are lacking have been identified for the 5 surfactants, particularly highlighting a shortage of chronic and sediment studies (see Table 1) and a current lack of attention toward AES (despite its growing commercial importance).

Available studies show wide variation in marine toxicity; in general, however, marine data fall within typical freshwater data ranges, suggesting approximately equal sensitivity to freshwater species. Although there is similarity in sensitivity for marine and freshwater species for the 3 common trophic levels (i.e., fish, invertebrates, and algae), there is still uncertainty because there are other key marine taxa that cannot be tested (e. g., echinoderms, mollusks, cephalopods, ctenophora [82]); hence, marine species are underrepresented.

The present review has revealed that the vast majority of available marine ecotoxicity studies are based on nominal rather than measured concentrations (likely the result of technical difficulties associated with analytical measurements of components of these surfactants). Consequently, it could be argued that the reliability of numerous current marine ecotoxicity studies is limited.

It is clear that there is a real limitation in the required fate and toxicity data for these surfactants, which results in uncertainties in their risk assessment in the marine environment. More information needs to be generated in laboratory and field studies on the occurrence, distribution, and degradation of these surfactants in the marine environment, including a better understanding of their mobility and interstitial partitioning as well as the development of estuarine and marine exposure models.

Recognition of the economic and ecological importance of the marine environment and its sensitivity toward anthropogenic impacts is growing. Consequently, increased emphasis is being placed on their protection. Commercial surfactants are consumed on a massive global scale, with growing demands showing no signs of slowing. Given that marine environments are considered the ultimate disposal sites of surfactants, additional marine studies—in particular bioconcentration studies, chronic AES studies, and AES and AS monitoring studies—within the present review are encouraged.

Supplemental Data—The Supplemental Data are available on the Wiley Online Library at DOI: 10.1002/etc.3297.

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Data availability—Data references are provided.

REFERENCES

- 1. Ying GG. 2006. Fate, behavior and effects of surfactants and their degradation products in the environment. *Environ Int* 32:417–431.
- TechNavio. 2015. Global surfactants market 2015–2019. [cited 2015 October 30]. Available from: http://www.technavio.com/report/globalsurfactants-market-2015-2019
- McWilliams P, Payne G. 2001. Bioaccumulation potential of surfactants: A review. European Oilfield Specialty Chemicals Association, Aberdeen, UK.
- 4. Madsen T, Boyd HB, Nylén D, Pedersen AR, Petersen GI, Simonsen F. 2001. Environmental and Health Assessment of Substances in Household Detergents and Cosmetic Detergent Products. Danish Environmental Protection Agency, Copenhagen, Denmark.
- 5. Organisation for Economic Co-operation and Development. 2005. Linear alkylbenzene sulfonate (LAS): SIDS initial assessment report. United Nations Environment Programme, Nairobi, Kenya.
- Cowan-Ellsberry C, Belanger S, Dorn P, Dyer S, McAvoy D, Sanderson H, Versteeg D, Ferrer D, Stanton K. 2014. Environmental safety of the use of major surfactant classes in North America. *Crit Rev Environ Sci Technol* 44:1893–1993.
- Human and Environmental Risk Assessment on Ingredients of Household Cleaning Products. 2004. Alcohol ethoxysulphates (AES): Environmental risk assessment. [cited 2015 October 30]. Available from: http://www.heraproject.com/files/1-E-04-HERA% 20AES%20ENV%20%20web%20wd.pdf
- Human and Environmental Risk Assessment on Ingredients of Household Cleaning Products. 2002. Alkyl sulphates: Environmental risk assessment. [cited 2015 October 30]. Available from: http://www.heraproject. com/files/3-E-417F36A9-DB35-F780-97A4CF8B60763 C35. pdf
- Human and Environmental Risk Assessment on Ingredients of Household Cleaning Products. 2009. Alcohol ethoxylates, Ver 2.0. [cited 2015 October 30]. Available from: http://www.heraproject.com/ files/34-F-09%20HERA%20AE%20Report%20Version%202%20-% 203%20Sept%2009.pdf
- Human and Environmental Risk Assessment on Ingredients of Household Cleaning Products. 2013. LAS: Linear alkylbenzene sulphonate (CAS No. 68411-30-3). [cited 2015 October 30]. Available from: http://www. heraproject.com/files/HERA-LAS%20revised%20April%202013% 20Final1.pdf
- Pérez-Carrera E, León VM, Lara-Martín PA, González-Mazo E. 2010. Influence of the hydrophilic moiety of anionic and nonionic surfactants on their aerobic biodegradation in seawater. *Sci Total Environ* 408:922–930.
- Lara-Martín PA, Gómez-Parra A, Sanz JL, González-Mazo E. 2010. Anaerobic degradation pathway of linear alkylbenzene sulfonates (LAS) in sulfate-reducing marine sediments. *Environ Sci Technol* 44:1670–1676.
- Hampel M, Mauffret A, Pazdro K, Blasco J. 2012. Anionic surfactant linear alkylbenzene sulfonates (LAS) in sediments from the Gulf of Gdańsk (southern Baltic Sea, Poland) and its environmental implications. *Environ Monit Assess* 184:6013–6023.
- Hampel M, Moreno-Garrido I, Gonzalez-Mazo E, Blasco J. 2009. Suitability of marine prosobranch snail *Hydrobia ulvae* for sediment toxicity assessment: A case study with the anionic surfactant linear alkylbenzene sulphonate (LAS). *Ecotoxicol Environ Saf* 72:1303–1308.
- Quiroga JM, Sales D, Gómez-Parra A. 1992. Influence of sediment on the biodegradation kinetics of linear alkylbenzene sulphonates in the marine environment. *Toxicol Environ Chem* 37:85–93.
- León VM, Gómez-Parra A, Rubio A, González-Mazo E. 2011. Sorption of linear alkylbenzene sulfonates and sulfophenylcarboxylic acids on marine sediments. Repositorio Instituto Español de Oceanografía, Madrid, Spain.
- 17. Kusk KO, Peterson S. 1997. Acute and chronic toxicity of tributyltin and linear alkylbenzene sulfonate to the marine copepod *Acartia tonsa*. *Environ Toxicol Chem* 16:1629–1633.

- Petrovic M, Fernández-Alba AR, Borrull A, Marce RM, González-Mazo E, Bareló D. 2002. Occurrence and distribution of nonionic surfactants, their degradation products, and linear alkylbenzene sulfonates in coastal waters and sediments in Spain. *Environ Toxicol Chem* 21:37–46.
- Lara-Martin PA, Gómez-Parra A, González-Mazo E. 2008. Sources, transport and reactivity of anionic and non-ionic surfactants in several aquatic ecosystems in SW Spain: A comparative study. *Environ Pollut* 156:36–45.
- González-Mazo E, Quiroga JM, Sales D, Gómez-Parra A. 1997. Levels of linear alkylbenzenesulfonate (LAS) in waters and sediments of the coastal ecosystems of the Gulf of Cadiz. *Toxicol Environ Chem* 59:77–87.
- Oliver-Rodríguez B, Zafra-Gómez A, Camino-Sánchez FJ, Conde-González JE, Pérez-Trujillo JP, Vílchez JL. 2013. Multi-residue method for the analysis of commonly used commercial surfactants, homologues and ethoxymers, in marine sediments by liquid chromatography-electrospray mass spectrometry. *Microchem J* 110: 158–168.
- Rubio JA, González-Mazo E, Gómez-Parra A. 1996. Sorption of linear alkylbenzenesulfonates (LAS) on marine sediment. *Mar Chem* 54:171–177.
- Terzić S, Hršak D, Ahel M. 1992. Primary biodegradation kinetics of linear alkylbenzene sulphonates in estuarine waters. *Water Res* 26: 585–591.
- Larson RJ, Rothgeb TM, Shimp RJ, Ward TE, Ventullo RM. 1993. Kinetics and practical significance of biodegradation of linear alkylbenzene sulfonate in the environment. J Am Oil Chem Soc 70:645.
- Bressan M, Brunetti R, Casellato S, Fava GC, Giro P, Marin M, Negrisolo P, Tallandini L, Thomann S, Tosoni L, Turchetta M. 1989. Effects of linear alkylbenzene sulfonate (LAS) on benthic organisms. *Tenside Surf Det* 26:148–158.
- Lewis MA. 1991. Chronic and sublethal toxicities of surfactants to aquatic animals: A review and risk assessment. Water Res 25:101–113.
- Wheeler JR, Leung KMY, Morritt D, Sorokin N, Rogers H, Toy R, Holt M, Whitehouse P, Crane M. 2002. Freshwater to saltwater toxicity extrapolation using species sensitivity distributions. *Environ Toxicol Chem* 21:2459–2467.
- European Centre for Ecotoxicology and Toxicology of Chemicals. 2009. Collation of existing marine biodegradation data and its use in environmental risk assessment. Technical Report 108. Brussels, Belgium.
- Perales JA, Manzano MA, Garrido MC, Sales D, Quiroga JM. 2007. Biodegradation kinetics of linear alkylbenzene sulphonates in sea water. *Biodegradation* 18:63–70.
- Leonards P, Eadsforth C, Schowanek D. 2013. MonitoringBase surfactants—A database specifically for storage of environmental data on surfactants in Europe. *Tenside Surf Det* 50:325–331.
- Cantarero S, Camino-Sánchez FJ, Zafra-Gómez A, Ballesteros O, Navalón A, Vílchez JL, Verge C, Reis MS, Saraiva PM. 2012. Evaluation of the presence of major anionic surfactants in marine sediments. *Mar Pollut Bull* 64:587–594.
- 32. Fernández-Ramos C, Ballesteros O, Zafra-Gómez A, Camino-Sánchez FJ, Blanc R, Navalón A, Vílchez JL. 2014. Environmental monitoring of alcohol sulfates and alcohol ethoxysulfates in marine sediments. *Environ Sci Pollut Res Int* 21:4286–4296.
- 33. Fernández-Ramos C, Ballesteros O, Zafra-Gómez A, Camino-Sánchez FJ, Blanc R, Navalón A, Pérez-Trujillo JP, Vílchez JL. 2014. Evaluation of the levels of alcohol sulfates and ethoxysulfates in marine sediments near wastewater discharge points along the coast of Tenerife Island. *Mar Pollut Bull* 79:107–113.
- 34. Lara-Martín PA, Gómez-Parra A, González-Mazo E. 2005. Determination and distribution of alkyl ethoxysulfates and linear alkylbenzene sulfonates in coastal marine sediments from the Bay of Cadiz (southwest of Spain). *Environ Toxicol Chem* 24:2196–2202.
- 35. Neubecker TA. 1985. Determination of alkylethoxylated sulfates in wastewaters and surface waters. *Environ Sci Technol* 19:1232–1236.
- Popenoe DD, Morris SJ, III, Horn PS, Norwood KT. 1994. Determination of alkyl sulfates and alkyl ethoxysulfates in wastewater treatment plant influents and effluents and in river water using liquid chromatography/ion spray mass spectrometry. *Anal Chem* 66: 1620–1629.
- Fendinger NJ, Begley WM, McAvoy DC, Eckhoff WS. 1995. Measurement of alkyl ethoxylated surfactants in natural waters. *Environ Sci Technol* 29:856–863.
- Dunphy JC, Pessler DG, Morrall SW, Evans KA, Robaugh DA, Fujimoto G, Negahban A. 2001. Derivatization LC/MS for the simultaneous determination of fatty alcohol and alcohol ethoxylated

surfactants in water and wastewater samples. *Environ Sci Technol* 35:1223–1230.

- Pojana G, Cassani G, Marcomini A. 2004. Determination, aerobic biodegradation and environmental occurrence of aliphatic alcohol polyethoxylate sulfates (AES). *Int J Environ Anal Chem* 84:729–738.
- Eadsforth CV, Sherren AJ, Selby MA, Toy R, Eckhoff WS, McAvoy DC, Matthijs E. 2006. Monitoring of environmental fingerprints of alcohol ethoxylates in Europe and Canada. *Ecotoxicol Environ* Saf 64:14–29.
- Lara-Martín PA, González-Mazo E, Brownawell BJ. 2012. Environmental analysis of alcohol ethoxylates and nonylphenol ethoxylate metabolites by ultra-performance liquid chromatography-tandem mass spectrometry. *Anal Bioanal Chem* 402:2359–2368.
- Cook TM, Goldman CK. 1974. Degradation of anionic detergents in Chesapeake Bay. Chesapeake Science 15:52–55.
- 43. Vives-Rego J, Vaqué MD, Sanchez Leal J, Parra J. 1987. Surfactant biodegradation in sea water: Primary biodegradation and bacterial population response in sea water. *Tenside Surf Det* 24:20–22.
- Shimp RJ. 1989. LAS biodegradation in estuaries. *Tenside Surf Det* 26:390–393.
- Goodhead AK, Head IM, Snape JR, Davenport RJ. 2014. Standard inocula preparations reduce the bacterial diversity and reliability of regulatory biodegradation tests. *Environ Sci Pollut Res Int* 21:9511–9521.
- Vives-Rego J, López-Amorós R, Guindulain T, García MT, Comas J, Sánchez-Leal J. 2000. Microbial aspects of linear alkylbenzene sulfonate degradation in coastal water. J Surfactants Deterg 3:303–307.
- García MT, Campos E, Ribosa I, Latorre A, Sánchez-Leal J. 2005. Anaerobic digestion of linear alkyl benzene sulfonates: Biodegradation kinetics and metabolite analysis. *Chemosphere* 60:1636–1643.
- Sales D, Perales JA, Manzano MA, Quiroga M. 1999. Anionic surfactant biodegradation in seawater. *Boletín Instituto Español de Oceanografia* 15:517–522.
- George AL. 2002. Seasonal factors affecting surfactant biodegradation in Antarctic coastal waters: Comparison of a polluted and pristine site. *Mar Environ Res* 53:403–415.
- Leon VM, Gómez-Parra A, González-Mazo E. 2004. Biodegradation of linear alkylbenzene sulfonates and their degradation intermediates in seawater. *Environ Sci Technol* 38:2359–2367.
- Mauffret A, Rottiers A, Federie T, Gillan DC, Hampel M, Blasco J, Temara A. 2009. Colonized beads as inoculum for marine biodegradability assessment: Application to linear alkylbenzene sulfonate. *Environ Int* 35:885–892.
- Quiroga JM, Sales D. 1989. Effect of temperature on kinetic of anionic surfactant degradation in sea-water. J Dispers Sci Technol 10:773–784.
- Lee D, Guckert JB, Belanger SE, Feijtel TCJ. 1997. Seasonal temperature declines do not decrease periphytic surfactant biodegradation or increase algal species sensitivity. *Chemosphere* 35:1143–1160.
- Quiroga JM, Sales D. 1990. Experimental variables in biodegradation of surfactant in marine environment. *Bull Environ Contam Toxicol* 44:851–858.
- 55. Guarino AM, Rieck G, Arnold S, Fenstermacher P, Bend J, Knutson MJ, Anderson JB. 1976. Distribution and toxicity of selected water pollutants in the spiny dogfish, *Squalus acanthias. Bull Mt Desert Isl Biol Lab Salisb Cove Maine* 16:50–53.
- 56. Sáez M, León VM, Gómez-Parra A, González-Mazo E. 2000. Extraction and isolation of linear alkylbenzene sulphonates and their intermediate metabolites from various marine organisms. *J Chromatogr A* 889:99–104.
- 57. León VM, González-Mazo E, Gómez-Parra A. 2000. Handling of marine and estuarine samples for the determination of linear alkylbenzene sulphonates and sulfophenyl carboxylic acids. *J Chromatogr A* 889:211–219.
- Temara A, Carr G, Webb S, Vergsteeg D, Feijtel T. 2001. Marine risk assessment: Linear alkylbenzene sulphonates (LAS) in the North Sea. *Mar Pollut Bull* 42:635–642.
- 59. Renaud F, Warnau M, Oberhänsli F, Teyssie J-L, Temara A, Rouleau C, Metian M. 2014. Bioconcentration of the anionic surfactant linear alkylbenzene sulfonate (LAS) in the marine shrimp *Palaemonetes varians*: A radiotracer study. *Mar Pollut Bull* 85:244–247.
- Álvarez-Muñoz D, Sáez M, Gómez-Parra A, González-Mazo E. 2007. Experimental determination of bioconcentration, biotransformation, and elimination of linear alkylbenzene sulfonates in *Solea senegal*ensis. Environ Toxicol Chem 26:2579–2586.
- 61. Dyer SD, Bernhard M-J, Versteeg DJ. 2003. Identification of an in vitro method for estimating the bioconcentration of surfactants in fish.

Procter & Gamble, Cincinnati, OH, USA. [cited 2015 October 30]. Available from: http://www.erasm.org/study.htm.

- 62. Tolls J, Sijm DTHM. 1999. Bioconcentration and biotransformation of the nonionic surfactant octaethylene glycol monotridecyl ether 14 C-C₁₃ EO₈. *Environ Toxicol Chem* 18:2689–2695.
- Tolls J, Haller M, Labee E, Verweij M, Sijm DTHM. 2000. Experimental determination of bioconcentration of the nonionic surfactant alcohol ethoxylate. *Environ Toxicol Chem* 19: 646–653.
- 64. Droge ST, Postama JF, Hermens JLM. 2008. Sediment toxicity of a rapidly biodegrading nonionic surfactant: Comparing the equilibrium partitioning approach with measurements in pore water. *Environ Sci Technol* 42:4215–4221.
- Tolls J, Haller M, de Graaf I, Thijssen MATC, Sijm DTHM. 1997. Bioconcentration of LAS: Experimental determination and extrapolation to environmental mixtures. *Environ Sci Technol* 31: 3426–3431.
- Jurado E, Fernández-Serrano M, Lechuga M, Ríos F. 2012. Environmental impact of ether carboxylic derivative surfactants. *J Surfactants Deterg* 15:1–7.
- 67. Moreno-Garrido I, Hampel M, Lubián LM, Blasco J. 2003. Marine benthic microalgae *Cylindrotheca closterium* (Ehremberg) Lewin and Reimann (Bacillariophyceae) as a tool for measuring toxicity of linear alkylbenzene sulfonate in sediments. *Bull Environ Contam Toxicol* 70:242–247.
- Moreno-Garrido I, Lubián LM, Blasco J. 2007. Sediment toxicity tests involving immobilized microalgae (*Phaeodactylum tricornutum* Bohlin). *Environ Int* 33:481–485.
- Rico-Rico A, Temara A, Hermens JLM. 2009. Equilibrium partitioning theory to predict the sediment toxicity of the anionic surfactant C₁₂-2-LAS to *Corophium volutator*. *Environ Pollut* 157: 575–581.
- Mauffret A, Rico-Rico A, Blasco J. 2010. Exposure of the marine deposit feeder *Hydrobia ulvae* to sediment associated LAS. *Environ Pollut* 158:529–535.
- Ezemonye LIN, Ogeleka DF, Okieimen FE. 2009. Lethal toxicity of industrial detergent on bottom dwelling sentinels. *Int J Sediment Res* 24:479–483.
- Mauffret A, Temara A, Blasco J. 2010. Exposure of the marine deposit feeder *Hydrobia ulvae* to sediment spiked with LAS congeners. *Water Res* 44:2831–2840.
- Hampel M, González-Mazo E, Vale C, Blasco J. 2007. Derivation of predicted no effect concentrations (PNEC) for marine environmental risk assessment: Application of different approaches to the model contaminant linear alkylbenzene sulphonates (LAS) in a site-specific environment. *Environ Int* 33:486–491.
- Hampel M, Ortiz-Delgado JB, Sarasquete C, Blasco J. 2008. Effects of sediment sorbed linear alkylbenzene sulphonate on juveniles of the Senegal sole, *Solea senegalensis*: Toxicity and histological indicators. *Histol Histopathol* 23:87–100.
- Marin MG, Pivotti L, Campesan G, Turchetto M, Tallandini L. 1994. Effects and fate of sediment-sorbed linear alkylbenzene sulphonate (LAS) on the bivalve mollusc *Mytilus galloprovincialis* Lmk. *Water Res* 28:85–90.
- Gaonkar SN, Karande AA, Rege MS. 1986. SDS-Artemia bioassay for evaluating oil-dispersant toxicity. *Pollut Res* 5:33–37.
- Persoone G, Van VA, Steertegem MV, Nayer BD. 1989. Predictive value of laboratory tests with aquatic invertebrates: Influence of experimental conditions. *Aquat Toxicol* 14:149–166.
- Hampel M, Moreno-Garrido I, Sobrino C, Lubián M, Blasco J. 2001. Acute toxicity of LAS homologues in marine microalgae: Esterase activity and inhibition growth as endpoints of toxicity. *Ecotoxicol Environ Saf* 48:287–292.
- 79. Cardwell RD. 1976. Relative acute toxicity of a pesticide, heavy metal, and anionic surfactants to marine organisms. *Proceedings of the National Shellfisheries Association* 66:107.
- Van de Plassche E, de Bruijn J. 1997. Risk assessment of four major surfactant groups in The Netherlands. *Tenside Surf Det* 34:242–249.
- Swedmark M, Braaten B, Emanuelsson E, Granmo A. 1971. Biological effects of surface active agents on marine animals. *Mar Biol* 9:183–201.
- European Centre for Ecotoxicology and Toxicology of Chemicals. 2003. Aquatic hazard assessment II. Technical Report 91. Brussels, Belgium.
- Vashon RD, Schwab BS. 1982. Mineralization of linear alcohol ethoxylates and linear alcohol ethoxy sulfates at trace concentrations in estuarine water. *Environ Sci Technol* 16:433–436.

- Sekiguchi H, Miura K, Oba K. 1975. Biodegradation of some anionic surfactants in river die-away test. *Journal of the Japan Oil Chemists' Society* 24:451–455.
- Sibila MA, Garrido MC, Perales JA, Quiroga JM. 2008. Ecotoxicity and biodegradability of an alkyl ethoxysulphate surfactant in coastal waters. *Sci Total Environ* 394:265–274.
- Hon-Nami H, Hanya T. 1980. Linear alkylbenzene sulfonates in river, estuary and bay water. Water Res 14:1251–1256.
- Larson RJ, Vashon RD, Games LM. 1983. Biodegradation of trace concentrations of detergent chemicals in freshwater and estuarine systems. *Biodeterioration* 5:235–245.
- Henkel KGaA. 1998. Vergleichende Bewertung des Abbauverhaltens von Stoffen im Süßwasser und marinen Abbautestsystemen. Report No. R9701182. Henkel, Düsseldorf, Germany.
- Arthur D. Little. 1991. Environmental and Human Safety of Major Surfactants, Vol 1: Anionic Surfactants. Part 2: Alcohol Ethyxy Sulfates. Final report to the Soap and Detergent Association. Cambridge, MA, USA.
- Roghair CJ, Buijze A, Schoon HNP. 1992. Ecotoxicological risk evaluation of the cationic fabric softener DTDMAC. I. Ecotoxicological effects. *Chemosphere* 24:599–609.
- Leeuwen K, Roghair C, Nijs T, Greef J. 1992. Ecotoxicological risk evaluation of the cationic fabric softener DTDMAC. III. Risk assessment. *Chemosphere* 24:629–639.
- Versteeg DJ, Feijtel TCJ, Cowan CE, Ward TE, Rapaport RA. 1992. An environmental risk assessment for DTDMAC in The Netherlands. *Chemosphere* 24:641–662.