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# Structure revision of the *Penicillium* alkaloids haenamindole and citreoindole

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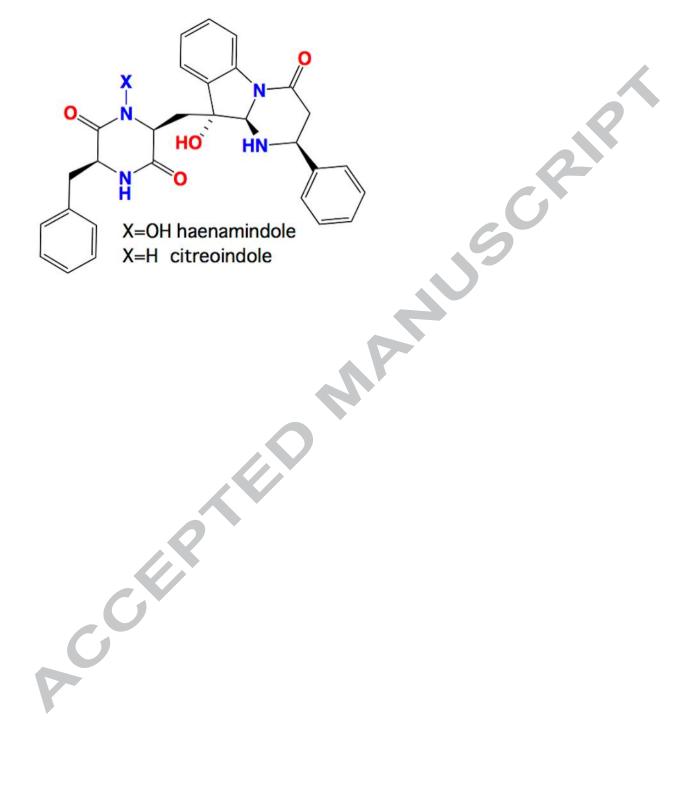
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#### ABSTRACT

Herein, we describe the isolation of rare alkaloids, haenamindole and citreoindole, from a South China Sea deep-sea fungus, *Penicillium citrinum* (MF006), and their structure revision based on detailed spectroscopic and  $C_3$  Marfey's analysis.

KEYWORDS: haenamindole; citreoindole; structure revision, deep-sea fungus

#### **GRAPHICAL ABSTRACT**



The genus *Penicillium* has been a prolific source of structurally diverse metabolites for over 90 years, with the antibiotic  $\beta$ -lactam penicillins, for example, heralding a revolution in the treatment of infectious disease. Notwithstanding past achievements, *Penicillium* isolates remain a valuable resource for microbial biodiscovery.

In our ongoing microbial biodiscovery program we used Dragendorff's reagent to screen a library of 946 marine-derived fungi for the presence of alkaloids, with an extract obtained from a South China Sea (-1500 m) sediment-derived *Penicillium citrinum* (MF006) exhibiting a positive response. As *P. citrinum* isolates have previously been reported to produce cytotoxic<sup>1-3</sup> and antibacterial<sup>4-6</sup> alkaloids, we were keen to investigate the alkaloids produced by MF006. A potato dextrose agar slant of MF006 was used to inoculate seed broth cultivations of (4 × 250 mL flasks, containing 40 mL of a liquid medium consisting of potato infusion (20%), glucose (2.0%), artificial sea salt (3.5%) and distilled water), which were incubated at 28 °C and 160 rpm for 3 d. Seed culture aliquots (5 mL) were used to inoculate larger cultivations (12 × 1 L flasks, each containing rice (100 g) and artificial seawater (3.5%; 30 mL)), which were incubated under static conditions at 25 °C for 21 d. The resulting rice-based cultivations were subjected to solvent extraction and reversed–phase HPLC fractionation to yield the known but otherwise rare alkaloids haenamindole (1)<sup>7</sup> and citreoindole (2).<sup>8</sup> Our re-investigation of 1 and 2 confirmed the need for structure revisions.

The HRESI(+)MS data for 1 revealed a sodium adduct ion consistent with a molecular formula  $(C_{29}H_{28}N_4O_5, \Delta mmu + 0.6)$  requiring 18 double bond equivalents (DBE). The 1D NMR (DMSOd<sub>6</sub>) data for 1 (ESI) revealed resonances for two mono-substituted and an *ortho*-disubstituted benzene ring, as well as three ester/amide carbonyls ( $\delta_c$  166.1, 159.8 and 168.8), accounting for

15 DBE and requiring that **1** be hexacyclic. The NMR data for **1** (ESI, Table S1) proved to be identical with that for haenamindole (**1a**) ( $[\alpha]_{D}$  –86.4, *c* 0.3, MeOH), first reported in 2015 by Kim and co-workers from a marine sediment-derived *Penicillium* sp. KCB12F005 ( $[\alpha]_{D}$  –89.4, *c* 0.05, MeOH).<sup>7</sup> A C<sub>3</sub> Marfey's analysis<sup>9</sup> on **1** confirmed the presence of L-Phe and L- $\beta$ -Phe residues, with a diagnostic ROESY correlation from H-10 to H-12 supporting the 10*R*, 12*S* and 24*S* configuration as previously noted by Kim and co-workers for **1a**.

We were however skeptical about the configurational assignment made by Kim and coworkers, where the absence of ROESY correlations between H-21 and H-24, and H-10 and 9-OH, was taken as evidence of *trans* relationships, and hence a 9*S*, 21*R* configuration. Interpretations based on the absence of ROESY/NOESY correlations should be approached with great caution. In our hands we observed ROESY correlations between H-21 and H-24, and H-10 and 9-OH, consistent with *cis* relationships and a 9*R*, 21*S* configuration. Based on these observations we propose the revised structure for haenamindole (1) as indicated (Figure 1).

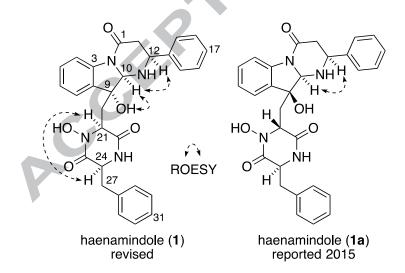


Figure 1. Revised (1) and reported (1a) structures for haenamindole

The HRESI(+)MS data for **2** revealed a sodium adduct ion consistent with a molecular formula  $(C_{29}H_{28}N_4O_4, \Delta mmu +0.0)$  for a deoxy analogue of **1**. On reviewing the literature, the NMR (DMSO-*d*<sub>6</sub>) data for **2** (ESI, Table S2) proved identical with that for citreoindole (**2a**) ( $[\alpha]_D$  – 38.1, *c* 0.06, MeOH), first reported in 1991 by Matsunaga and co-workers from a hybrid strain derived from *Penicillium citreo-viride* B. IFO 6200 and 4692 (no reported  $[\alpha]_D$ ).<sup>8</sup> Matsunaga and co-workers used chiral HPLC on a hydrolysate of **2a** to detect only L-Phe. By contrast, we employed C<sub>3</sub> Marfey's methodology<sup>9</sup> to identify both L-Phe and L- $\beta$ -Phe residues in **2**, consistent with the biosynthetically related co-metabolite **1**.

While we accepted the H-21 to H-24 NOESY correlation observed by Matsunaga and coworkers, and the associated 21*S* assignment, by misidentifying the L- $\beta$ -Phe residue we had concerns regarding other conformational assignments. Based on the above, as well as spectroscopic and biogenetic considerations, we propose that **1** and **2** possess the same absolute configuration, and attribute the revised structure for citreoindole (**2**) as indicated (Figure 2).

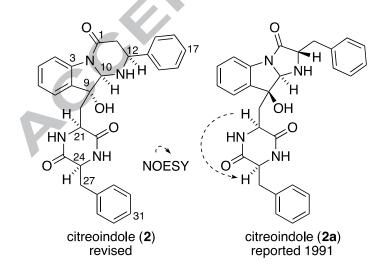


Figure 2. Revised (2) and reported (2a) structures for citreoindole

Acceleration

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#### SUPPORTING INFORMATION

General experimental conditions, including fungal taxonomy and cultivations, and the extraction, isolation and characterization of **1** and **2**, as well as tabulated 1D and 2D NMR data, and selected spectra. This material is available free of charge via the Internet at http://pubs.acs.org

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#### Highlights

Deep-sea

Accepter