



Love, S., & Miners, J. S. (2016). Cerebral Hypoperfusion and the Energy Deficit in Alzheimer's Disease. Brain Pathology, 26(5), 607-617. DOI: 10.1111/bpa.12401

Peer reviewed version

License (if available): CC BY-NC

Link to published version (if available): 10.1111/bpa.12401

Link to publication record in Explore Bristol Research PDF-document

This is the author accepted manuscript (AAM). The final published version (version of record) is available online via Wiley at http://onlinelibrary.wiley.com/doi/10.1111/bpa.12401/abstract. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: http://www.bristol.ac.uk/pure/about/ebr-terms.html

Cerebral hypoperfusion and the energy deficit in Alzheimer's disease

Seth Love; J. Scott Miners

Dementia Research Group, Institute of Clinical Neurosciences, School of Clinical

Sciences, University of Bristol, Bristol, UK

Short title: Hypoperfusion and energy deficit in Alzheimer's

Keywords

Alzheimer's disease, ischaemia, hypoperfusion, vascular dysfunction, amyloid-β

peptide, endothelin-1, cholinergic innervation, renin-angiotensin system,

endothelial nitric oxide synthase, cerebral amyloid angiopathy, capillary damage

Corresponding author

Seth Love, School of Clinical Sciences, University of Bristol, Learning & Research

level 2, Southmead Hospital, Bristol BS10 5NB, UK

Abstract

There is a perfusion deficit in Alzheimer's disease (AD), commencing in the precuneus and spreading to other parts of the cerebral cortex. The deficit anticipates the development of dementia, contributes to brain damage, and is caused by both functional and structural abnormalities of the cerebral vasculature. Most of the abnormalities are probably secondary to the accumulation of $A\beta$ but the consequent hypoperfusion may, in turn, increase $A\beta$ production. In the early stages of disease, abnormalities that cause vasoconstriction predominate. These include cholinergic vascular denervation, inhibition of endothelial nitric oxide synthase, increased production of endothelin-1 production and possibly also of angiotensin II. Patients with AD also have an increased prevalence of structural disease of cerebral microvessels, particularly CAA and capillary damage, and particularly in the later stages of disease these are likely to make an important contribution to the cerebral hypoperfusion. The metabolic abnormalities that cause early vascular dysfunction offer several targets for therapeutic intervention. However, for intervention to be effective it probably needs to be early. Prolonged cerebral hypoperfusion may induce compensatory circulatory changes that are themselves damaging, including hypertension and small vessel disease. This has implications for the use of antihypertensive drugs once there is accumulation of Aß within the brain.

Introduction

A major contributor to the energy deficit in Alzheimer's disease (AD) is the reduction in cerebral perfusion that results from dysfunction and structural

abnormalities of the cerebral vasculature. As discussed elsewhere in this minisymposium, there are also other contributors to the energy deficit in Alzheimer's disease (AD), including extracranial abnormalities that affect cerebral blood supply and intracellular disturbances of mitochondrial function, but those are not covered in the present review.

Within this review, we consider three questions:

- Is cerebral hypoperfusion in AD caused by reduced demand or reduced supply?
- 2. What are the mechanisms of the hypoperfusion?
- 3. What are the therapeutic implications?

Reduced demand or reduced supply?

The relative contributions of falling metabolic demand and reduction in blood supply vary according to the stage of disease. In preclinical and early AD there is evidence from multiple studies employing a range of methods that the cerebral hypoperfusion is pathological rather than physiological, i.e. the decline in perfusion exceeds the reduction in metabolic demand, and causes tissue damage. Metabolic demand may, in fact, be increased during the earliest stages of amyloid accumulation (11, 27).

Some of the data comes from imaging studies using arterial spin-labeled perfusion magnetic resonance imaging (ASL-MRI) or 2-deoxy-2-(18F)fluoro-D-glucose positron emission tomography (FDG-PET) (there is a near-perfect topographical correspondence between changes in ASL-MRI and FDG-PET in AD – see, for example (24)). In people with mutations that cause autosomal dominant forms of AD, in whom the timing of onset of the AD is highly

predictable, a reduction in glucose uptake is demonstrable by FDG-PET at least 10 years before the onset of clinical disease and before there is detectable atrophy (11). The decline shows a consistent pattern of topographical progression, starting in the precuneus (medial parietal cortex) and extending along the cingulate gyrus, lateral part of the parietal lobe and anterior part of the occipital lobe, then into the rest of the cerebrum.

A similar pattern of progression of hypoperfusion was demonstrated by ASL-MRI in sporadic AD (12). In patients with MCI the hypoperfusion was most pronounced in the precuneus and posterior cingulate cortex but also involved lateral parietal, occipital and frontal cortex. Perfusion declined significantly in the temporal cortex only when patients became demented and did not fall in the hippocampus. Similar findings were reported in earlier studies on people with MCI (5, 30) and in healthy carriers of the *APOE* ϵ 4 allele (65), a strong genetic risk factor for AD. This stereotypical distribution of hypoperfusion does not bear an obvious relationship to the distribution of cerebral atrophy, which correlates with that of neurofibrillary tangle pathology, commencing in the inferomedial part of the temporal lobes before spreading to other parts of the cerebrum (11). The distribution of hypoperfusion/reduced glucose uptake correlates much more closely with that of the preceding accumulation of amyloid (see for example (11, 27)).

We recently used a biochemical approach to assess the adequacy of perfusion and oxygenation of the precuneus in early AD. We compared the levels of two myelin proteins, myelin-associated glycoprotein (MAG), which is highly susceptible to reduced tissue oxygenation, and proteolipid protein-1 (PLP1), which is relatively resistant (8, 9). Both myelin proteins are synthesized in the

oligodendrocyte cell body and require energy-dependent transport to reach their sites of insertion into the myelin sheath (Figure 1). PLP1 is distributed throughout the myelin sheath whereas MAG is inserted only far from the cell body, in the adaxonal loop of myelin, the first part of the sheath to degenerate when blood supply is insufficient to meet the energy demands of the oligodendrocyte. Both myelin proteins are very stable under post-mortem conditions (9), and as they have half-lives of several months (47, 136), a decline in the MAG:PLP1 ratio in post-mortem brain tissue reflects a pathological reduction in ante-mortem perfusion over a relatively long period prior to death. We showed that the MAG:PLP1 ratio, was reduced in the precuneus by ~50% in Braak stage III-IV disease (75), indicating that even in early AD there is a disparity between oligodendrocyte energy demand and supply in the first region of brain to show hypoperfusion.

Tarumi et al (131) used near-infrared spectroscopy to compare the tissue oxygenation index (a measure of the saturation of haemoglobin by oxygen) in frontal cortex of people with amnestic MCI and age-matched controls. In amnestic MCI patients the tissue oxygenation index was significantly reduced, both at rest and after a sit-stand manoeuvre, indicating increased oxygen extraction. Had hypoperfusion been a response to reduced metabolic demand rather than a pathological reduction in blood supply, the tissue oxygenation index would have been higher rather than lower.

Cerebral hypoperfusion predicts the development of dementia in patients with MCI and the rate of cognitive decline in patients with AD (14, 16, 22). The fact that the hypoperfusion actually damages the brain, even in preclinical or early disease, is well demonstrated on imaging of cerebral white matter in

people with mutations that cause autosomal dominant forms of AD. A recent study showed that people with such mutations had a greater volume of white matter hyperintensities (WMH) several years before clinical disease, indicating that the hypoperfusion was severe enough to cause tissue damage (67). The increase in WMH was most pronounced in the parietal and occipital lobes; as noted above, this corresponds approximately to the distribution of early amyloid accumulation and reduced perfusion in the overlying cerebral cortex.

The relationship between cortical amyloid and WMH is likely to be relevant to the pathogenesis of the white matter hypoperfusion (see below). Other evidence for a relationship between amyloid burden and WMH comes from an MRI study of 150 cognitively normal people by Scott et al (122). The authors found that amyloid burden, as assessed by measuring A β 42 level in the CSF, was an independent predictor of total WMH volume. Lee and colleagues (67) found some correlation between WMH and the presence of microbleeds, suggesting a contribution from cerebral amyloid angiopathy. However, the increase in WMH remained significant after controlling for presence of microbleeds, which were calculated to account for 21% of the association between AD mutation status and WMH.

In later AD, it is likely that the decline in perfusion continues to exceed the reduction in metabolic demand but less so than in early disease. Several functional MRI studies have demonstrated an increase in the regional oxygen extraction fraction (rOEF) in the cerebral cortex and white matter in AD (83, 84, 137), indicating a continued pathological reduction in blood supply. Indeed, in the series of Toghi et al (137), rOEF was higher in the cerebral cortex of patients

with AD than in those with vascular dementia (in the white matter the increase was more marked in patients with vascular dementia).

Our own studies found the MAG:PLP1 ratio to be reduced in the cerebral cortex in late, as well as early AD (75, 134). However, within the precuneus, the ratio was not as markedly decreased in brain tissue from patients with Braak stage V-VI disease as in those with earlier (Braak stage III-IV) disease and not significantly so in comparison with Braak stage 0-II disease (Figure 2). We interpreted this lessening of the perfusion deficit as being likely to reflect falling metabolic demand with increasing synaptic and neuronal damage.

Mechanisms: metabolic vascular dysfunction

Under normal circumstances, cerebral perfusion is tightly regulated to match the supply of oxygenated blood to metabolic requirements, both of the brain as a whole (through autoregulation – the maintenance of relatively constant blood flow despite changes in perfusion pressure) and of the individual regions within it (through neurovascular coupling) (25, 106). This regulation is effected through multiple neurogenic, myogenic and metabolic pathways. In AD, the activity of several pathways that regulate intracerebral vascular tone and influence neurovascular coupling is abnormally altered. Most of the alterations promote vasoconstriction, acting on smooth muscle cells in the tunica media of arterioles, on pericytes in capillaries or on both types of cell, and reduce tissue oxygenation. Other abnormalities allow inappropriate local vasodilatation, diverting blood away from regions of higher metabolic demand. Both types of alteration have the potential to affect neurological function and, if sustained, to cause permanent damage.

Cerebral cortex

Cholinergic innervation

Arterioles in the cerebral cortex are innervated by cholinergic nerves, originating in the nucleus basalis of Meynert (139, 142). Stimulation of neurons in the nucleus basalis (see (26) for review) or of muscarinic receptors in isolated arterioles (48) causes vasodilatation, partly mediated by stimulation of the production of nitric oxide (NO) (151). Tong and Hamel (139) found a reduction in the cholinergic innervation of cortical blood vessels in AD, mirroring a general loss of cholinergic nerve terminals from the cerebral cortex and in keeping with the loss of neurons from the nucleus basalis from an early stage of disease (see (31, 68) for review). Cholinergic deafferentation reduces blood flow in the cerebral cortex, as was demonstrated after targeted ablation of cholinergic neurons by administration of 192 IgG-saporin (144). Although interpretation of the findings is complicated by possible effects of cholinergic denervation on neuronal activity and metabolic demand, it seems likely that reduced stimulation of muscarinic receptors in the walls of cortical arterioles contributes to the hypoperfusion of the cortex in AD, and possible beneficial effects of cholinesterase inhibitors in AD may relate partly to augmented cerebral perfusion (13, 26). It should be noted that AD is also associated with alterations in a range of other neurotransmitters that have direct or indirect effects on vascular contractility, including glutamate, γ-aminobutyric acid, noradrenaline (norepinephrine), serotonin and dopamine (28, 34, 44, 63, 113).

Perhaps not surprisingly, several of the processes that mediate vascular dysfunction in AD are probably initiated by the accumulation of A β . Topical application of A β 40 or A β 42 to isolated arteries causes vasoconstriction, A β 40 being more potent than A β 42 in this regard (29). The vasoconstriction can be reduced by free radical scavengers and cyclo-oxygenase inhibitors (99, 112, 135, 140). A β 40 enhances the constriction induced by endothelin-1 (EDN1) and reduces the vasodilatation produced by NO (99, 101).

Aβ40 also induces vasoconstriction in vivo, as demonstrated by its application to mouse cortex (89). This can be prevented by administering free radical scavengers or by an M35Nle amino acid substitution in Aβ40 which interferes with its ability to generate reactive oxygen species (89, 90). In a series of studies on mice overexpressing Aβ-precursor protein (APP), Iadecola and colleagues showed that elevated endogenous AB also caused vasoconstriction from an early age (2 months in Tg2576 mice, well before plaque formation), impaired autoregulation and interfered with neurovascular coupling (in this case the functional hyperaemia of the barrel cortex that is normally induced by whisker stimulation) (86-88, 90). Shin et al (125) confirmed that Tg2576 mice had an attenuated hyperaemic response to hypercapnia and whisker stimulation but were unable to demonstrate this until the mice had reached the age of 9 months, i.e. after commencement of vascular deposition of Aβ. The authors suggested that vascular deposition of Aβ was a prerequisite for the vascular dysfunction. It is also possible that the development of vascular dysfunction in these mice simply depends on the concentration of soluble A β , which increases with age, although it is not clear why the two research groups found so marked a

difference in the timing of onset of the dysfunction. The abnormal cerebral vasoconstriction in Tg2576 mice requires the production of free radicals by nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase), as shown by studies in which NADPH oxidase was either inhibited (103), or inactivated by deletion of Nox2 (104). These findings are in keeping with the studies on isolated arteries, described above.

For simplicity, the various processes that contribute to hypoperfusion in AD are considered under separate headings in the present review. However, as noted below, the different pathways overlap and interact substantially – particularly insofar as they involve A β , e.g. in upregulating the production of EDN1 by endothelin-converting enzymes-1 and -2 (ECE1 and ECE2) and that of angiotensin II (Ang II) by angiotensin-converting enzyme (ACE), in reducing NO production by endothelial cells, in binding to and sequestering vascular endothelial growth factor (VEGF) in plaques, and in blocking VEGF receptor 2 (VEGFR-2) signalling in endothelial cells. There is also interaction between the endothelin and renin-angiotensin systems (reviewed in (76)); A β and the cholinergic systems (see (55, 94, 147) for review); and the cholinergic system and VEGF production (53).

Endothelin system

EDN1 is a potent vasoconstrictor and its concentration is increased significantly in cerebral cortex from patients with AD (75, 97, 134). The increase is demonstrable from an early stage of disease, including within the precuneus (75), the first region in which blood flow declines (see above). Paradoxically, the gene that encodes EDN1 is also upregulated by hypoxia (127). The extent to

which EDN1 is increased in AD correlates with the severity of cortical hypoperfusion/tissue hypoxia, as measured by the decline in the MAG:PLP1 ratio.

Both Aβ40 and Aβ42 are capable of increasing EDN1 production *in vitro*: Aβ40 through upregulation of ECE1 in endothelial cells (97, 98), and Aβ42 through upregulation of ECE2 in neurons (96). In cortex from patients with AD the concentration of EDN1 correlates closely with that of Aβ42 but bears no relationship to that of Aβ40 (75), suggesting that the increase in EDN1 and decrease in tissue oxygenation are caused, at least in part, by Aβ42-mediated neuronal upregulation of ECE2. Aβ is a physiological substrate of both ECEs (36-38) and the upregulation of ECE2 and consequent sustained overproduction of EDN1 in AD may simply be an unfortunate side effect of the parenchymal accumulation of substrate in the form of Aβ42 (76). The lack of association between AB40 and EDN1 (or MAG:PLP1) does not discount a role for AB40 in the vascular dysfunction of AD, but such a role is likely to be predominantly episodic: interfering with autoregulation and neurovascular coupling rather than causing sustained hypoperfusion. Palmer et al (98) showed that the enhanced release of EDN1 that follows the addition of Aβ40 to human cerebrovascular endothelial cells in vitro could be prevented by the addition of superoxide dismutase, potentially linking upregulation of ECE1 with the observations of Niwa et al (89, 90) on free-radical mediated vasoconstriction, and suggesting that the episodic cerebral vasoconstriction induced by AB40 results from a free radical-mediated increase in endothelial ECE-1 activity and EDN1 production.

Angiotensin

Increased production of the vasoconstrictor Ang II may contribute to hypoperfusion of the frontal cortex, where ACE activity (74, 77, 78) and Ang II level (unpublished observations) are elevated in AD, perhaps in response to the accumulation of A β 42. Miners et al (78) showed that ACE activity in SH-SY5Y neuroblastoma cells was upregulated by aggregated A β 42 (but not A β 40 or freshly solubilised A β 42). The relationship between A β 42, ACE activity, Ang II production and hypoperfusion is, however, less clear-cut than that between A β 42, ECE1 activity, EDN1 production and hypoperfusion, in that neither ACE activity nor Ang II level was increased in precuneus from patients with AD (75).

Like so many of the dysregulated pathways that are the focus of this review, the renin-angiotensin system has a complex interrelationship with other vasoregulatory processes. ACE cleaves (141) and probably thereby limits the duration of action of the vasodilator bradykinin, the production of which is likely to be elevated in AD, as a result of increased activity of plasma kallikrein (4). Ang II was reported to increase EDN1 production in endothelial (32) and vascular adventitial fibroblasts (1), probably by inducing transcription of the preproendothelin-1 gene (117) and by a mechanism involving NADPH oxidase (2), thereby contributing to EDN1-mediated hypoperfusion. Conversely, hypoxia was shown to upregulate the expression and activation of ACE (62).

Endothelial nitric oxide synthase (eNOS)

NO, a potent vasodilator, is synthesized within the endothelium by eNOS, the activity of which plays an important role in local regulation of the cerebral microcirculation (151). eNOS is activated by a wide range of stimuli, including

acetylcholine (40), bradykinin, oxidative stress, shear stress and hypoxia. Both A β 40 and A β 42 inhibit eNOS activity. A β 40 was reported to do so through a mechanism that depends on protein kinase C (46), and A β 42 through interfering with Akt/GSK-3 β signalling and a mechanism involving interaction of eNOS with heat shock protein 90 (64, 128). Mice partially deficient in eNOS develop cognitive impairment associated with a range of neuropathological abnormalities, including cerebral amyloid angiopathy (CAA) and disruption of the blood-brain barrier (BBB), largely confined to the temporoparietal and retrosplenial granular cortex and hippocampus (129). Jeynes and Provias (52) reported a significant negative correlation between the number of eNOS-immunolabelled capillaries and the density of neurofibrillary tangles and A β plaques in sections of temporal and calcarine cortex in AD.

White matter

As noted above, most of the studies on mechanisms of vascular dysfunction in AD have used rodent models and have focussed on cerebral cortex. Whilst several of the local metabolic abnormalities that contribute to hypoperfusion of the cerebral cortex may also apply in the white matter, our studies have highlighted differences that are relevant to our understanding of the pathogenesis of ischaemic white matter damage in AD and have implications for treatment.

Whereas we found the concentration of EDN1 to be elevated approximately twofold in the cerebral cortex in AD, presumably in response to A β 42-induced upregulation of ECE2, EDN1 level was significantly reduced in the underlying white matter (8, 75). This reduction occurred in association with a modest decline in MAG and in MAG:PLP1 in the white matter (9) (in keeping with other

evidence of ischaemic white matter damage in AD, e.g. on neuroimaging, as discussed above). The relationship between MAG:PLP1 and EDN1 in the white matter was the converse of that in the cortex: in the cortex MAG:PLP1 and EDN1 correlated negatively and in the white matter they correlated positively (75). White matter hypoperfusion in AD is not therefore caused by increased white matter EDN1, which falls as would be expected physiologically in response to hypoperfusion. However, MAG:PLP1 in the white matter did correlate positively with the concentration of EDN1 *in the overlying cortex*, suggesting that hypoperfusion of the white matter in AD results partly from vasoconstriction of perforating arterioles as they traverse the cortex (Figure 3) (23, 75). This mechanism of white matter hypoperfusion is likely to be relevant to other Aβ-dependent processes that increase vasoconstriction within the cortex, including cholinergic denervation, Ang II production and reduction in activity of eNOS.

Mechanisms: structural abnormalities of the cerebral vasculature

The abnormalities described above affect vascular function but are not associated with long-lasting structural alterations. As recently reviewed (71), patients with AD also have an increased prevalence of structural disease of cerebral microvessels, particularly CAA and capillary damage. They may also have more severe non-amyloid small vessel disease (SVD) than elderly people without AD but most of the cited evidence is indirect, based on the identification of white matter abnormalities on neuroimaging, and as likely to have resulted from CAA or capillary damage as from SVD. An MRI-based study of regional

cerebrovascular resistance (CVRi) found this to be increased in several regions of brain that are not affected by CAA and have a predilection for SVD, including thalamus and caudate nucleus (85), but the pathological substrate of the increased CVRi remains to be demonstrated. For further consideration of the possible association of SVD with AD, see (71).

Cerebral amyloid angiopathy

Most patients with AD have CAA, in some series over 90%, compared to about 30% in elderly controls (21, 42, 69, 72, 73, 143, 146). In many cases the CAA is relatively mild, affecting only occasional arterioles in the leptomeninges, but some patients have widespread involvement of cortical and meningeal arterioles, as well as deposition of A β in the adventitia of meningeal venules. In patients with AD, possession of *APOE* ϵ 4 is a risk factor for more-severe CAA (21, 111, 121) and is strongly associated with capillary CAA (6, 69, 133) (in controls, arteriolar A β amyloid angiopathy is more strongly associated with *APOE* ϵ 2). In the majority of patients CAA is restricted to the cerebral cortex and overlying leptomeninges but it may also involve the cerebellum (particularly the meningeal vessels) and occasionally the brain stem. A β may also accumulate in the walls of capillaries, sometimes extensively so. Capillary CAA predominantly involves the entorhinal and occipital cortex but can be present in other parts of the neocortex and is often, but not always, associated with severe arteriolar CAA.

CAA has several adverse effects on cerebral perfusion. Perhaps the most widely recognised are cerebral micro-haemorrhages and larger lobar haemorrhages, but there is also extensive documentation of ischaemic abnormalities (predominantly cortical microinfarcts) (3, 18, 41, 43, 93, 138),

some of which are caused by local thrombosis, some by the marked narrowing of severely affected blood vessels, and some probably by impaired neurovascular coupling. Evidence of neurovascular decoupling in human patients comes from MRI studies of occipital vascular reactivity in response to visual stimulation in patients with probable CAA (35, 107, 126). Peca et al (107) also found that impaired neurovascular coupling, as evidenced by lower functional MRI responses to visual stimuli, was associated with more microbleeds and a higher volume of white matter lesions, linking impairment of neurovascular coupling with severity of tissue damage.

Capillary damage

The capillary bed constitutes much the largest part of the cerebral vasculature and is also the most important in terms of metabolic homeostasis. Yet the contribution of capillary damage to hypoperfusion in AD has been somewhat neglected, perhaps because of the small size and inconspicuous histological appearance of individual capillaries.

Despite the hypoperfusion, the density of capillaries in the cerebral cortex is unchanged or reduced in AD (8, 15, 17, 45, 60, 134), and more of them show degenerative changes in AD than in age-matched controls (7, 20, 51, 123). Both endothelial cells and pericytes are affected, their degeneration eventually leaving residual 'string' vessels consisting solely of tubes of collagen. These degenerative changes occur despite a significant increase in the concentration of VEGF in AD (54, 75, 130, 134), which would be expected to promote angiogenesis, with the formation of new capillaries (92, 150). Several factors may contribute to this lack of angiogenic response. A β peptides have direct anti-angiogenic activity (99, 100,

102) and also bind to VEGF receptor 2, blocking VEGF signalling. In addition, A β within plaques binds and thereby potentially sequesters VEGF, interfering with its biological availability (105, 148).

At the level of the capillary bed, degeneration of pericytes has emerged as a key contributor to hypoperfusion. Changes in the contractile activity of pericytes modulate capillary calibre and cerebral blood flow and probably play an important role in neurovascular coupling (10, 49, 108). Dore-Duffy et al reported that pericytes in primary cultures express both EDN1 and its two receptors (EDNRA and EDNRB) (33). The authors also provided in vivo evidence that EDN1 (which is elevated in AD – see above) contributes to the regulation of capillary perfusion through binding to EDNRA receptors in pericytes. Experimental traumatic brain injury in mice caused an increase in the number of smooth muscle actin-positive pericytes around capillaries, a rise in capillary EDN1, and reduced capillary diameter. These changes could be prevented by administration of an EDNRA antagonist. Pericytes are also important for maintenance of the blood-brain barrier (10).

A series of studies by Zlokovic and colleagues have shown that loss of pericytes exacerbates multiple pathological processes in AD (145), including hypoperfusion and disruption of the blood-brain barrier (10, 123), accumulation of A β 40 and A β 42, tau pathology and neuronal loss (120). Montagne et al (80) quantified blood-brain barrier permeability in the hippocampus of young and older adult volunteers by dynamic contrast-enhanced MRI, and showed the degree of increase in permeability to correlate with the CSF:plasma albumin ratio (a marker of blood-brain barrier breakdown) and the CSF concentration of soluble platelet-derived growth factor receptor β (a marker of pericyte injury).

Both blood-brain barrier permeability and pericyte injury were more pronounced in participants with MCI than in older individuals who were cognitively normal.

Therapeutic implications

There is therefore overwhelming evidence of a wide range of functional and structural abnormalities of the cerebral microvasculature in AD, that contribute to hypoperfusion and the resulting energy deficit as well as to other aspects of the disease. Implications for therapy are both specific and general. Specific implications concern the potential for targeting of particular pathways or receptors to ameliorate the vascular abnormalities – particularly those that are not the result of structural changes. General implications relate to the timing and broader consequences of intervention.

Several of the pathways implicated in abnormal microvascular function in AD are potentially amenable to treatment. The cholinergic system is, of course, already routinely targeted in AD patients through the administration of cholinesterase inhibitors. These drugs improve cerebral perfusion in mild to moderate disease (19), and several studies found evidence of an association between cognitive response and cerebral blood flow (13, 26, 124). The potential for intervention in the renin-angiotensin system has been extensively reviewed (56-59) and the effects on cognition and cerebral blood flow of losartan, an angiotensin receptor antagonist, are currently being tested in a multicentre UK clinical trial (115).

Another potential target is the endothelin system. Bosentan, a non-selective EDNR antagonist (119), improves pulmonary blood flow and exercise

tolerance in patients with pulmonary hypertension, another disease in which there is elevated production of EDN1 (114, 118). Bosentan also preserves endothelium-dependent aortic and carotid vasodilatation in Tg2576 mice (39). Selective EDNRA receptor antagonists such as zibotentan (50, 81) offer theoretical advantages, in that they target the predominant type of EDN1 receptor responsible for mediating vasoconstriction in both smooth muscle cells of cerebral arterioles and pericytes that surround capillaries (33, 91). For discussion see (95).

Interventions aimed at reversing functional abnormalities of the vasculature in AD have the potential not only to improve symptoms but also to slow the progression of disease. Hypoperfusion probably increases the production of A β 42, thereby accelerating the progression of disease. Simulation of neuronal ischaemia in vitro, or experimental cerebral hypoperfusion in animal models increases A β 42 production through multiple mechanisms (reviewed in (70, 71)), including upregulation of amyloid- β precursor protein and β -secretase and possibly reduced neprilysin-mediated degradation. Indirect evidence of a hypoperfusion-induced increase in A β comes from observations in patients who had survived a recent cardiac arrest (149) or diffuse traumatic brain injury with cerebral oedema (and therefore almost certainly hypoperfusion) (79). Both groups had elevated serum A β 42 over several days. In the patients with diffuse traumatic brain injury, A β 42 was also monitored in the CSF where the level declined, arguing against non-specific leakage of A β from damaged brain tissue as the explanation for the rising level in the serum.

The timing of intervention is likely to be critical, as prolonged hypoperfusion causes permanent brain damage (as discussed above), and as the

disease progresses the balance tends to shift from metabolic to structural vascular dysfunction (Figure 4). It seems possible too that prolonged cerebral hypoperfusion may induce compensatory changes in the circulation that are themselves damaging, including hypertension and SVD. Mid-life hypertension is significantly associated with AD (109, 110) Several clinical studies have reported an association between hypertension before the age of 65 years (particularly if there is elevation of diastolic blood pressure) and later development of AD (61, 66, 82). Hypertension in cognitively normal adults with at least 1 *APOE* $\epsilon 4$ allele was associated with increased binding of the Aβ tracer F18–labelled florbetapir (116). The conventional interpretation is that hypertension increases the risk of developing AD by promoting the accumulation of A\u03c3. However, another possible explanation is that hypertension is a physiological response to tonic cerebral vasoconstriction induced by mid-life accumulation of Aβ; a means of maintaining cerebral perfusion. There is experimental evidence that this is the case, in that cerebroventricular infusion of A\beta was shown to cause a progressive, highly significant rise in blood pressure in rats (132). This has obvious implications for autoregulation and for the treatment of hypertension once Aβ has begun to accumulate, and suggests that blood pressure in such patients should be lowered only cautiously, ideally with monitoring of the effects on cerebral perfusion.

Acknowledgements

This work was supported by Alzheimer's Research UK (ART-PG2011-1 and ARUK-PG2015-11). The South West Dementia Brain Bank is part of the Brains for Dementia Research program, jointly funded by Alzheimer's Research UK and

Alzheimer's Society, and is supported by BRACE (Bristol Research into Alzheimer's and Care of the Elderly) and the Medical Research Council.

References

- 1. An SJ, Boyd R, Wang Y, Qiu X, Wang HD (2006) Endothelin-1 expression in vascular adventitial fibroblasts. *Am J Physiol Heart Circ Physiol* **290**:H700-8.
- 2. An SJ, Boyd R, Zhu M, Chapman A, Pimentel DR, Wang HD (2007) NADPH oxidase mediates angiotensin II-induced endothelin-1 expression in vascular adventitial fibroblasts. *Cardiovasc Res* **75**:702-9.
- Arvanitakis Z, Capuano AW, Leurgans SE, Buchman AS, Bennett DA,
 Schneider JA (2016) The relationship of cerebral vessel pathology to brain microinfarcts. *Brain Pathol*.
- 4. Ashby EL, Love S, Kehoe PG (2012) Assessment of activation of the plasma kallikrein-kinin system in frontal and temporal cortex in Alzheimer's disease and vascular dementia. *Neurobiol Aging* **33**:1345-55.
- 5. Asllani I, Habeck C, Scarmeas N, Borogovac A, Brown TR, Stern Y (2008)

 Multivariate and univariate analysis of continuous arterial spin labeling

 perfusion MRI in Alzheimer's disease. *J Cereb Blood Flow Metab* 28:725-36.
- 6. Attems J, Jellinger KA (2004) Only cerebral capillary amyloid angiopathy correlates with Alzheimer pathology--a pilot study. *Acta Neuropathol* **107**:83-90.
- 7. Baloyannis SJ, Baloyannis IS (2012) The vascular factor in Alzheimer's disease: a study in Golgi technique and electron microscopy. *J Neurol Sci* **322**:117-21.

- 8. Barker R, Ashby EL, Wellington D, Barrow VM, Palmer JC, Kehoe PG, Esiri MM, Love S (2014) Pathophysiology of white matter perfusion in Alzheimer's disease and vascular dementia. *Brain* **137**:1524-32.
- 9. Barker R, Wellington D, Esiri MM, Love S (2013) Assessing white matter ischemic damage in dementia patients by measurement of myelin proteins.

 J Cereb Blood Flow Metab 33:1050-7.
- 10. Bell RD, Winkler EA, Sagare AP, Singh I, LaRue B, Deane R, Zlokovic BV (2010) Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* **68**:409-27.
- 11. Benzinger TL, Blazey T, Jack CR, Jr., Koeppe RA, Su Y, Xiong C, Raichle ME, Snyder AZ, Ances BM, Bateman RJ, Cairns NJ, Fagan AM, Goate A, Marcus DS, Aisen PS, Christensen JJ, Ercole L, Hornbeck RC, Farrar AM, Aldea P, Jasielec MS, Owen CJ, Xie X, Mayeux R, Brickman A, McDade E, Klunk W, Mathis CA, Ringman J, Thompson PM, Ghetti B, Saykin AJ, Sperling RA, Johnson KA, Salloway S, Correia S, Schofield PR, Masters CL, Rowe C, Villemagne VL, Martins R, Ourselin S, Rossor MN, Fox NC, Cash DM, Weiner MW, Holtzman DM, Buckles VD, Moulder K, Morris JC (2013) Regional variability of imaging biomarkers in autosomal dominant Alzheimer's disease. *Proc Natl Acad Sci U S A* 110:E4502-9.
- 12. Binnewijzend MA, Kuijer JP, Benedictus MR, van der Flier WM, Wink AM, Wattjes MP, van Berckel BN, Scheltens P, Barkhof F (2013) Cerebral blood flow measured with 3D pseudocontinuous arterial spin-labeling MR imaging in Alzheimer disease and mild cognitive impairment: a marker for disease severity. *Radiology* **267**:221-30.

- 13. Blin J, Ivanoiu A, Coppens A, De Volder A, Labar D, Michel C, Laterre EC (1997) Cholinergic neurotransmission has different effects on cerebral glucose consumption and blood flow in young normals, aged normals, and Alzheimer's disease patients. *Neuroimage* **6**:335-43.
- 14. Borroni B, Perani D, Broli M, Colciaghi F, Garibotto V, Paghera B, Agosti C, Giubbini R, Di Luca M, Padovani A (2005) Pre-clinical diagnosis of Alzheimer disease combining platelet amyloid precursor protein ratio and rCBF spect analysis. *J Neurol* **252**:1359-62.
- 15. Bouras C, Kovari E, Herrmann FR, Rivara CB, Bailey TL, von Gunten A, Hof PR, Giannakopoulos P (2006) Stereologic analysis of microvascular morphology in the elderly: Alzheimer disease pathology and cognitive status. *J Neuropathol Exp Neurol* **65**:235-44.
- 16. Brown DR, Hunter R, Wyper DJ, Patterson J, Kelly RC, Montaldi D, McCullouch J (1996) Longitudinal changes in cognitive function and regional cerebral function in Alzheimer's disease: a SPECT blood flow study. *J Psychiatr Res* **30**:109-26.
- 17. Buee L, Hof PR, Bouras C, Delacourte A, Perl DP, Morrison JH, Fillit HM (1994) Pathological alterations of the cerebral microvasculature in Alzheimer's disease and related dementing disorders. *Acta Neuropathol* 87:469-80.
- 18. Cadavid D, Mena H, Koeller K, Frommelt RA (2000) Cerebral β amyloid angiopathy is a risk factor for cerebral ischemic infarction. A case control study in human brain biopsies. *J Neuropathol Exp Neurol* **59**:768-73.
- 19. Ceravolo R, Volterrani D, Tognoni G, Dell'Agnello G, Manca G, Kiferle L,
 Rossi C, Logi C, Strauss HW, Mariani G, Murri L (2004) Cerebral perfusional

- effects of cholinesterase inhibitors in Alzheimer disease. *Clin*Neuropharmacol 27:166-70.
- 20. Challa VR, Thore CR, Moody DM, Anstrom JA, Brown WR (2004) Increase of white matter string vessels in Alzheimer's disease. *J Alzheimers Dis* **6**:379-83; discussion 443-9.
- 21. Chalmers K, Wilcock GK, Love S (2003) APOE ε4 influences the pathological phenotype of Alzheimer's disease by favouring cerebrovascular over parenchymal accumulation of A beta protein. *Neuropathol Appl Neurobiol* **29**:231-8.
- 22. Chao LL, Buckley ST, Kornak J, Schuff N, Madison C, Yaffe K, Miller BL, Kramer JH, Weiner MW (2010) ASL perfusion MRI predicts cognitive decline and conversion from MCI to dementia. *Alzheimer Dis Assoc Disord* 24:19-27.
- 23. Charidimou A, Pantoni L, Love S (2016) The concept of sporadic cerebral small vessel disease: a road map on key definitions and current concepts.

 Int J Stroke 11:6-18.
- 24. Chen Y, Wolk DA, Reddin JS, Korczykowski M, Martinez PM, Musiek ES, Newberg AB, Julin P, Arnold SE, Greenberg JH, Detre JA (2011) Voxel-level comparison of arterial spin-labeled perfusion MRI and FDG-PET in Alzheimer disease. *Neurology* **77**:1977-85.
- 25. Cipolla MJ (2009) Control of cerebral blood flow. In: The Cerebral Circulation, Chapter 5, pp. 41-52, Morgan & Claypool Life Sciences: San Rafael, CA.

- 26. Claassen JA, Jansen RW (2006) Cholinergically mediated augmentation of cerebral perfusion in Alzheimer's disease and related cognitive disorders: the cholinergic-vascular hypothesis. *J Gerontol A Biol Sci Med Sci* **61**:267-71.
- 27. Cohen AD, Price JC, Weissfeld LA, James J, Rosario BL, Bi W, Nebes RD, Saxton JA, Snitz BE, Aizenstein HA, Wolk DA, Dekosky ST, Mathis CA, Klunk WE (2009) Basal cerebral metabolism may modulate the cognitive effects of Abeta in mild cognitive impairment: an example of brain reserve. *J Neurosci* **29**:14770-8.
- 28. Cohen Z, Bonvento G, Lacombe P, Hamel E (1996) Serotonin in the regulation of brain microcirculation. *Prog Neurobiol* **50**:335-62.
- 29. Crawford F, Suo Z, Fang C, Mullan M (1998) Characteristics of the in vitro vasoactivity of β-amyloid peptides. *Exp Neurol* **150**:159-68.
- 30. Dai W, Lopez OL, Carmichael OT, Becker JT, Kuller LH, Gach HM (2009)

 Mild cognitive impairment and alzheimer disease: patterns of altered

 cerebral blood flow at MR imaging. *Radiology* **250**:856-66.
- 31. Davies P, Maloney AJ (1976) Selective loss of central cholinergic neurons in Alzheimer's disease. *Lancet* **2**:1403.
- 32. Dohi Y, Hahn AW, Boulanger CM, Buhler FR, Luscher TF (1992) Endothelin stimulated by angiotensin II augments contractility of spontaneously hypertensive rat resistance arteries. *Hypertension* **19**:131-7.
- Dore-Duffy P, Wang S, Mehedi A, Katyshev V, Cleary K, Tapper A, Reynolds
 C, Ding Y, Zhan P, Rafols J, Kreipke CW (2011) Pericyte-mediated
 vasoconstriction underlies TBI-induced hypoperfusion. *Neurol Res* 33:176-86.

- 34. Drake CT, Iadecola C (2007) The role of neuronal signaling in controlling cerebral blood flow. *Brain Lang* **102**:141-52.
- 35. Dumas A, Dierksen GA, Gurol ME, Halpin A, Martinez-Ramirez S, Schwab K, Rosand J, Viswanathan A, Salat DH, Polimeni JR, Greenberg SM (2012)

 Functional magnetic resonance imaging detection of vascular reactivity in cerebral amyloid angiopathy. *Ann Neurol* **72**:76-81.
- 36. Eckman EA, Adams SK, Troendle FJ, Stodola BA, Kahn MA, Fauq AH, Xiao HD, Bernstein KE, Eckman CB (2006) Regulation of steady-state β-amyloid levels in the brain by neprilysin and endothelin-converting enzyme but not angiotensin-converting enzyme. *J Biol Chem* **281**:30471-8.
- 37. Eckman EA, Reed DK, Eckman CB (2001) Degradation of the Alzheimer's amyloid β peptide by endothelin-converting enzyme. *J Biol Chem* **276**:24540-8.
- 38. Eckman EA, Watson M, Marlow L, Sambamurti K, Eckman CB (2003)

 Alzheimer's disease β-amyloid peptide is increased in mice deficient in endothelin-converting enzyme. *J Biol Chem* **278**:2081-4.
- 39. Elesber AA, Bonetti PO, Woodrum JE, Zhu XY, Lerman LO, Younkin SG, Lerman A (2006) Bosentan preserves endothelial function in mice overexpressing APP. *Neurobiol Aging* **27**:446-50.
- 40. Elhusseiny A, Hamel E (2000) Muscarinic--but not nicotinic--acetylcholine receptors mediate a nitric oxide-dependent dilation in brain cortical arterioles: a possible role for the M5 receptor subtype. *J Cereb Blood Flow Metab* **20**:298-305.

- 41. Ellis RJ, Olichney JM, Thal LJ, Mirra SS, Morris JC, Beekly D, Heyman A

 (1996) Cerebral amyloid angiopathy in the brains of patients with

 Alzheimer's disease: the CERAD experience, Part XV. *Neurology* **46**:1592-6.
- 42. Esiri MM, Wilcock GK (1986) Cerebral amyloid angiopathy in dementia and old age. *J Neurol Neurosurg Psychiatry* **49**:1221-6.
- 43. Esiri MM, Wilcock GK, Morris JH (1997) Neuropathological assessment of the lesions of significance in vascular dementia. *J Neurol Neurosurg**Psychiatry 63:749-53.
- 44. Fergus A, Lee KS (1997) GABAergic regulation of cerebral microvascular tone in the rat. *J Cereb Blood Flow Metab* **17**:992-1003.
- 45. Fischer VW, Siddiqi A, Yusufaly Y (1990) Altered angioarchitecture in selected areas of brains with Alzheimer's disease. *Acta Neuropathol* **79**:672-9.
- 46. Gentile MT, Vecchione C, Maffei A, Aretini A, Marino G, Poulet R, Capobianco L, Selvetella G, Lembo G (2004) Mechanisms of soluble β-amyloid impairment of endothelial function. *J Biol Chem* **279**:48135-42.
- 47. Greer JM, Lees MB (2002) Myelin proteolipid protein--the first 50 years. *Int J Biochem Cell Biol* **34**:211-5.
- 48. Hamel E (2004) Cholinergic modulation of the cortical microvascular bed.

 *Prog Brain Res 145:171-8.**
- 49. Hamilton NB, Attwell D, Hall CN (2010) Pericyte-mediated regulation of capillary diameter: a component of neurovascular coupling in health and disease. *Front Neuroenergetics* **2**.
- 50. Haque SU, Dashwood MR, Heetun M, Shiwen X, Farooqui N, Ramesh B, Welch H, Savage FJ, Ogunbiyi O, Abraham DJ, Loizidou M (2013) Efficacy of

- the specific endothelin a receptor antagonist zibotentan (ZD4054) in colorectal cancer: a preclinical study. *Mol Cancer Ther* **12**:1556-67.
- 51. Hunter JM, Kwan J, Malek-Ahmadi M, Maarouf CL, Kokjohn TA, Belden C, Sabbagh MN, Beach TG, Roher AE (2012) Morphological and pathological evolution of the brain microcirculation in aging and Alzheimer's disease. *PLoS One* **7**:e36893.
- 52. Jeynes B, Provias J (2009) Significant negative correlations between capillary expressed eNOS and Alzheimer lesion burden. *Neurosci Lett* **463**:244-8.
- 53. Kakinuma Y, Furihata M, Akiyama T, Arikawa M, Handa T, Katare RG, Sato T (2010) Donepezil, an acetylcholinesterase inhibitor against Alzheimer's dementia, promotes angiogenesis in an ischemic hindlimb model. *J Mol Cell Cardiol* **48**:680-93.
- 54. Kalaria RN, Cohen DL, Premkumar DR, Nag S, LaManna JC, Lust WD (1998)

 Vascular endothelial growth factor in Alzheimer's disease and

 experimental cerebral ischemia. *Brain Res Mol Brain Res* **62**:101-5.
- 55. Kar S, Quirion R (2004) Amyloid β peptides and central cholinergic neurons: functional interrelationship and relevance to Alzheimer's disease pathology. *Prog Brain Res* **145**:261-74.
- 56. Kehoe PG (2003) The renin-angiotensin-aldosterone system and Alzheimer s disease? *J Renin Angiotensin Aldosterone Syst* **4**:80-93.
- 57. Kehoe PG, Miners S, Love S (2009) Angiotensins in Alzheimer's disease friend or foe? *Trends Neurosci* **32**:619-28.

- 58. Kehoe PG, Passmore PA (2012) The renin-angiotensin system and antihypertensive drugs in Alzheimer's disease: current standing of the angiotensin hypothesis? *J Alzheimers Dis* **30 Suppl 2**:S251-68.
- 59. Kehoe PG, Wilcock GK (2007) Is inhibition of the renin-angiotensin system a new treatment option for Alzheimer's disease? *Lancet Neurol* **6**:373-8.
- 60. Kitaguchi H, Ihara M, Saiki H, Takahashi R, Tomimoto H (2007) Capillary beds are decreased in Alzheimer's disease, but not in Binswanger's disease.

 Neurosci Lett 417:128-31.
- 61. Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, Iivonen S, Mannermaa A, Tuomilehto J, Nissinen A, Soininen H (2002) Apolipoprotein E ε4 allele, elevated midlife total cholesterol level, and high midlife systolic blood pressure are independent risk factors for late-life Alzheimer disease. *Ann Intern Med* 137:149-55.
- 62. Krick S, Hanze J, Eul B, Savai R, Seay U, Grimminger F, Lohmeyer J, Klepetko W, Seeger W, Rose F (2005) Hypoxia-driven proliferation of human pulmonary artery fibroblasts: cross-talk between HIF-1α and an autocrine angiotensin system. *FASEB J* **19**:857-9.
- 63. Krimer LS, Muly EC, 3rd, Williams GV, Goldman-Rakic PS (1998)Dopaminergic regulation of cerebral cortical microcirculation. *Nat Neurosci* 1:286-9.
- 64. Lamoke F, Mazzone V, Persichini T, Maraschi A, Harris MB, Venema RC, Colasanti M, Gliozzi M, Muscoli C, Bartoli M, Mollace V (2015) Amyloid β peptide-induced inhibition of endothelial nitric oxide production involves oxidative stress-mediated constitutive eNOS/HSP90 interaction and disruption of agonist-mediated Akt activation. *J Neuroinflammation* **12**:84.

- 65. Langbaum JB, Chen K, Caselli RJ, Lee W, Reschke C, Bandy D, Alexander GE, Burns CM, Kaszniak AW, Reeder SA, Corneveaux JJ, Allen AN, Pruzin J, Huentelman MJ, Fleisher AS, Reiman EM (2010) Hypometabolism in Alzheimer-affected brain regions in cognitively healthy Latino individuals carrying the apolipoprotein E €4 allele. *Arch Neurol* 67:462-8.
- 66. Launer LJ, Ross GW, Petrovitch H, Masaki K, Foley D, White LR, Havlik RJ (2000) Midlife blood pressure and dementia: the Honolulu-Asia aging study. *Neurobiol Aging* **21**:49-55.
- 67. Lee S, Viqar F, Zimmerman ME, Narkhede A, Tosto G, Benzinger TL, Marcus DS, Fagan AM, Goate A, Fox NC, Cairns NJ, Holtzman DM, Buckles V, Ghetti B, McDade E, Martins RN, Saykin AJ, Masters CL, Ringman JM, Ryan NS, Frster S, Laske C, Schofield PR, Sperling RA, Salloway S, Correia S, Jack C, Weiner M, Bateman RJ, Morris JC, Mayeux R, Brickman AM, Dominantly Inherited Alzheimer N (2016) White matter hyperintensities are a core feature of Alzheimer's disease: Evidence from the Dominantly Inherited Alzheimer Network. *Ann Neurol*.
- 68. Liu AK, Chang RC, Pearce RKB, Gentleman SM (2015) Nucleus basalis of Meynert revisited: anatomy, history and differential involvement in Alzheimer's and Parkinson's disease. *Acta Neuropathol* **129**:527-40.
- 69. Love S, Chalmers K, Ince P, Esiri M, Attems J, Jellinger K, Yamada M, McCarron M, Minett T, Matthews F, Greenberg S, Mann D, Kehoe PG (2014) Development, appraisal, validation and implementation of a consensus protocol for the assessment of cerebral amyloid angiopathy in postmortem brain tissue. *Am J Neurodegener Dis* **3**:19-32.

- 70. Love S, Miners JS (2015) White matter hypoperfusion and damage in dementia: post-mortem assessment. *Brain Pathol* **25**:99-107.
- 71. Love S, Miners JS (2016) Cerebrovascular disease in ageing and Alzheimer's disease. *Acta Neuropathol* **131**:645-58.
- 72. Love S, Nicoll JA, Hughes A, Wilcock GK (2003) APOE and cerebral amyloid angiopathy in the elderly. *Neuroreport* **14**:1535-6.
- 73. Masuda J, Tanaka K, Ueda K, Omae T (1988) Autopsy study of incidence and distribution of cerebral amyloid angiopathy in Hisayama, Japan. *Stroke* **19**:205-10.
- 74. Miners JS, Ashby E, Van Helmond Z, Chalmers KA, Palmer LE, Love S, Kehoe PG (2008) Angiotensin-converting enzyme (ACE) levels and activity in Alzheimer's disease, and relationship of perivascular ACE-1 to cerebral amyloid angiopathy. *Neuropathol Appl Neurobiol* **34**:181-93.
- 75. Miners JS, Palmer J, Love S (in press) Pathophysiology of hypoperfusion of the precuneus in early Alzheimer's disease. *Brain Pathol*.
- 76. Miners JS, Palmer JC, Tayler H, Palmer LE, Ashby E, Kehoe PG, Love S (2014) Aβ degradation or cerebral perfusion? Divergent effects of multifunctional enzymes. *Front Aging Neurosci* 6:238.
- 77. Miners JS, van Helmond Z, Raiker M, Love S, Kehoe PG (2010) ACE variants and association with brain A β levels in Alzheimer's disease. *Am J Transl Res* **3**:73-80.
- 78. Miners S, Ashby E, Baig S, Harrison R, Tayler H, Speedy E, Prince JA, Love S, Kehoe PG (2009) Angiotensin-converting enzyme levels and activity in Alzheimer's disease: differences in brain and CSF ACE and association with *ACE1* genotypes. *Am J Transl Res* **1**:163-77.

- 79. Mondello S, Buki A, Barzo P, Randall J, Provuncher G, Hanlon D, Wilson D, Kobeissy F, Jeromin A (2014) CSF and plasma amyloid-β temporal profiles and relationships with neurological status and mortality after severe traumatic brain injury. *Sci Rep* **4**:6446.
- 80. Montagne A, Barnes SR, Sweeney MD, Halliday MR, Sagare AP, Zhao Z, Toga AW, Jacobs RE, Liu CY, Amezcua L, Harrington MG, Chui HC, Law M, Zlokovic BV (2015) Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **85**:296-302.
- 81. Morris CD, Rose A, Curwen J, Hughes AM, Wilson DJ, Webb DJ (2005)

 Specific inhibition of the endothelin A receptor with ZD4054: clinical and pre-clinical evidence. *Br J Cancer* **92**:2148-52.
- 82. Morris MC, Scherr PA, Hebert LE, Glynn RJ, Bennett DA, Evans DA (2001)

 Association of incident Alzheimer disease and blood pressure measured from 13 years before to 2 years after diagnosis in a large community study.

 Arch Neurol 58:1640-6.
- 83. Nagata K, Kondoh Y, Atchison R, Sato M, Satoh Y, Watahiki Y, Hirata Y, Yokoyama E (2000) Vascular and metabolic reserve in Alzheimer's disease.

 Neurobiol Aging 21:301-7.
- 84. Nagata K, Sato M, Satoh Y, Watahiki Y, Kondoh Y, Sugawara M, Box G, Wright D, Leung S, Yuya H, Shimosegawa E (2002) Hemodynamic aspects of Alzheimer's disease. *Ann N Y Acad Sci* **977**:391-402.
- 85. Nation DA, Wierenga CE, Clark LR, Dev SI, Stricker NH, Jak AJ, Salmon DP, Delano-Wood L, Bangen KJ, Rissman RA, Liu TT, Bondi MW (2013) Cortical and subcortical cerebrovascular resistance index in mild cognitive impairment and Alzheimer's disease. *J Alzheimers Dis* 36:689-98.

- 86. Niwa K, Carlson GA, Iadecola C (2000) Exogenous Aβ1-40 reproduces cerebrovascular alterations resulting from amyloid precursor protein overexpression in mice. *J Cereb Blood Flow Metab* **20**:1659-68.
- 87. Niwa K, Kazama K, Younkin L, Younkin SG, Carlson GA, Iadecola C (2002)

 Cerebrovascular autoregulation is profoundly impaired in mice

 overexpressing amyloid precursor protein. *Am J Physiol Heart Circ Physiol*283:H315-23.
- 88. Niwa K, Kazama K, Younkin SG, Carlson GA, Iadecola C (2002) Alterations in cerebral blood flow and glucose utilization in mice overexpressing the amyloid precursor protein. *Neurobiol Dis* **9**:61-8.
- 89. Niwa K, Porter VA, Kazama K, Cornfield D, Carlson GA, Iadecola C (2001)

 Aβ-peptides enhance vasoconstriction in cerebral circulation. *Am J Physiol Heart Circ Physiol* **281**:H2417-24.
- 90. Niwa K, Younkin L, Ebeling C, Turner SK, Westaway D, Younkin S, Ashe KH, Carlson GA, Iadecola C (2000) A β 1-40-related reduction in functional hyperemia in mouse neocortex during somatosensory activation. *Proc Natl Acad Sci U S A* **97**:9735-40.
- 91. Noll G, Wenzel RR, Luscher TF (1996) Endothelin and endothelin antagonists: potential role in cardiovascular and renal disease. *Mol Cell Biochem* **157**:259-67.
- 92. Nor JE, Christensen J, Mooney DJ, Polverini PJ (1999) Vascular endothelial growth factor (VEGF)-mediated angiogenesis is associated with enhanced endothelial cell survival and induction of Bcl-2 expression. *Am J Pathol* **154**:375-84.

- 93. Okazaki H, Reagan TJ, Campbell RJ (1979) Clinicopathologic studies of primary cerebral amyloid angiopathy. *Mayo Clin Proc* **54**:22-31.
- 94. Pákáski M, Kálmán J (2008) Interactions between the amyloid and cholinergic mechanisms in Alzheimer's disease. *Neurochem Int* **53**:103-11.
- 95. Palmer J, Love S (2011) Endothelin receptor antagonists: potential in Alzheimer's disease. *Pharmacol Res* **63**:525-31.
- 96. Palmer JC, Baig S, Kehoe PG, Love S (2009) Endothelin-converting enzyme-2 is increased in Alzheimer's disease and up-regulated by Aβ. *Am J Pathol* **175**:262-70.
- 97. Palmer JC, Barker R, Kehoe PG, Love S (2012) Endothelin-1 is elevated in Alzheimer's disease and upregulated by amyloid-β. *J Alzheimers Dis* **29**:853-61.
- 98. Palmer JC, Tayler HM, Love S (2013) Endothelin-converting enzyme-1 activity, endothelin-1 production, and free radical-dependent vasoconstriction in Alzheimer's disease. *J Alzheimers Dis* **36**:577-87.
- 99. Paris D, Humphrey J, Quadros A, Patel N, Crescentini R, Crawford F, Mullan M (2003) Vasoactive effects of A β in isolated human cerebrovessels and in a transgenic mouse model of Alzheimer's disease: role of inflammation. Neurol Res 25:642-51.
- 100. Paris D, Patel N, DelleDonne A, Quadros A, Smeed R, Mullan M (2004)

 Impaired angiogenesis in a transgenic mouse model of cerebral

 amyloidosis. *Neurosci Lett* **366**:80-5.
- 101. Paris D, Town T, Parker TA, Humphrey J, Mullan M (1998) Isoform-specific vasoconstriction induced by apolipoprotein E and modulation of this effect by Alzheimer's β-amyloid peptide. *Neurosci Lett* **256**:73-6.

- 102. Paris D, Townsend K, Quadros A, Humphrey J, Sun J, Brem S, Wotoczek-Obadia M, DelleDonne A, Patel N, Obregon DF, Crescentini R, Abdullah L, Coppola D, Rojiani AM, Crawford F, Sebti SM, Mullan M (2004) Inhibition of angiogenesis by Aβ peptides. *Angiogenesis* **7**:75-85.
- 103. Park L, Anrather J, Zhou P, Frys K, Pitstick R, Younkin S, Carlson GA, Iadecola C (2005) NADPH-oxidase-derived reactive oxygen species mediate the cerebrovascular dysfunction induced by the amyloid β peptide. *J*Neurosci 25:1769-77.
- 104. Park L, Zhou P, Pitstick R, Capone C, Anrather J, Norris EH, Younkin L, Younkin S, Carlson G, McEwen BS, Iadecola C (2008) Nox2-derived radicals contribute to neurovascular and behavioral dysfunction in mice overexpressing the amyloid precursor protein. *Proc Natl Acad Sci U S A* **105**:1347-52.
- 105. Patel NS, Mathura VS, Bachmeier C, Beaulieu-Abdelahad D, Laporte V, Weeks O, Mullan M, Paris D (2010) Alzheimer's β-amyloid peptide blocks vascular endothelial growth factor mediated signaling via direct interaction with VEGFR-2. *J Neurochem* **112**:66-76.
- 106. Paulson OB, Strandgaard S, Edvinsson L (1990) Cerebral autoregulation.

 *Cerebrovasc Brain Metab Rev 2:161-92.
- 107. Peca S, McCreary CR, Donaldson E, Kumarpillai G, Shobha N, Sanchez K, Charlton A, Steinback CD, Beaudin AE, Fluck D, Pillay N, Fick GH, Poulin MJ, Frayne R, Goodyear BG, Smith EE (2013) Neurovascular decoupling is associated with severity of cerebral amyloid angiopathy. *Neurology* 81:1659-65.

- 108. Peppiatt CM, Howarth C, Mobbs P, Attwell D (2006) Bidirectional control of CNS capillary diameter by pericytes. *Nature* **443**:700-4.
- 109. Power MC, Weuve J, Gagne JJ, McQueen MB, Viswanathan A, Blacker D
 Blood Pressure. The AlzRisk Database. Alzheimer Research Forum.
- 110. Power MC, Weuve J, Gagne JJ, McQueen MB, Viswanathan A, Blacker D

 (2011) The association between blood pressure and incident Alzheimer
 disease: a systematic review and meta-analysis. *Epidemiology* **22**:646-59.
- 111. Premkumar DR, Cohen DL, Hedera P, Friedland RP, Kalaria RN (1996)
 Apolipoprotein E-€4 alleles in cerebral amyloid angiopathy and
 cerebrovascular pathology associated with Alzheimer's disease. Am J Pathol
 148:2083-95.
- 112. Price JM, Sutton ET, Hellermann A, Thomas T (1997) β -amyloid induces cerebrovascular endothelial dysfunction in the rat brain. *Neurol Res* **19**:534-8.
- 113. Raichle ME, Hartman BK, Eichling JO, Sharpe LG (1975) Central noradrenergic regulation of cerebral blood flow and vascular permeability. *Proc Natl Acad Sci U S A* **72**:3726-30.
- 114. Raja SG, Dreyfus GD (2008) Current status of bosentan for treatment of pulmonary hypertension. *Ann Card Anaesth* **11**:6-14.
- 115. Reducing pathology in Alzheimer's disease through angiotensin targeting.
- 116. Rodrigue KM, Rieck JR, Kennedy KM, Devous MD, Sr., Diaz-Arrastia R, Park DC (2013) Risk factors for β-amyloid deposition in healthy aging: vascular and genetic effects. *JAMA Neurol* **70**:600-6.

- 117. Rossi GP, Sacchetto A, Cesari M, Pessina AC (1999) Interactions between endothelin-1 and the renin-angiotensin-aldosterone system. *Cardiovasc Res* **43**:300-7.
- 118. Rubin LJ, Badesch DB, Barst RJ, Galie N, Black CM, Keogh A, Pulido T, Frost A, Roux S, Leconte I, Landzberg M, Simonneau G (2002) Bosentan therapy for pulmonary arterial hypertension. *N Engl J Med* **346**:896-903.
- 119. Rubin LJ, Roux S (2002) Bosentan: a dual endothelin receptor antagonist.

 Expert Opin Investig Drugs 11:991-1002.
- 120. Sagare AP, Bell RD, Zhao Z, Ma Q, Winkler EA, Ramanathan A, Zlokovic BV (2013) Pericyte loss influences Alzheimer-like neurodegeneration in mice.

 Nat Commun 4:2932.
- 121. Schmechel DE, Saunders AM, Strittmatter WJ, Crain BJ, Hulette CM, Joo SH, Pericak-Vance MA, Goldgaber D, Roses AD (1993) Increased amyloid β-peptide deposition in cerebral cortex as a consequence of apolipoprotein E genotype in late-onset Alzheimer disease. *Proc Natl Acad Sci U S A* **90**:9649-53.
- 122. Scott JA, Braskie MN, Tosun D, Thompson PM, Weiner M, DeCarli C,
 Carmichael OT, Alzheimer's Disease Neuroimaging I (2015) Cerebral
 amyloid and hypertension are independently associated with white matter
 lesions in elderly. *Front Aging Neurosci* **7**:221.
- 123. Sengillo JD, Winkler EA, Walker CT, Sullivan JS, Johnson M, Zlokovic BV (2013) Deficiency in mural vascular cells coincides with blood-brain barrier disruption in Alzheimer's disease. *Brain Pathol* **23**:303-10.

- 124. Shimizu S, Hanyu H, Iwamoto T, Koizumi K, Abe K (2006) SPECT follow-up study of cerebral blood flow changes during Donepezil therapy in patients with Alzheimer's disease. *J Neuroimaging* **16**:16-23.
- 125. Shin HK, Jones PB, Garcia-Alloza M, Borrelli L, Greenberg SM, Bacskai BJ, Frosch MP, Hyman BT, Moskowitz MA, Ayata C (2007) Age-dependent cerebrovascular dysfunction in a transgenic mouse model of cerebral amyloid angiopathy. *Brain* **130**:2310-9.
- 126. Smith EE, Vijayappa M, Lima F, Delgado P, Wendell L, Rosand J, Greenberg SM (2008) Impaired visual evoked flow velocity response in cerebral amyloid angiopathy. *Neurology* **71**:1424-30.
- 127. Stow LR, Jacobs ME, Wingo CS, Cain BD (2011) Endothelin-1 gene regulation. *FASEB J* **25**:16-28.
- 128. Suhara T, Magrane J, Rosen K, Christensen R, Kim HS, Zheng B, McPhie DL, Walsh K, Querfurth H (2003) Aβ42 generation is toxic to endothelial cells and inhibits eNOS function through an Akt/GSK-3beta signaling-dependent mechanism. *Neurobiol Aging* **24**:437-51.
- 129. Tan XL, Xue YQ, Ma T, Wang X, Li JJ, Lan L, Malik KU, McDonald MP, Dopico AM, Liao FF (2015) Partial eNOS deficiency causes spontaneous thrombotic cerebral infarction, amyloid angiopathy and cognitive impairment. *Mol Neurodegener* **10**:24.
- 130. Tarkowski E, Issa R, Sjogren M, Wallin A, Blennow K, Tarkowski A, Kumar P (2002) Increased intrathecal levels of the angiogenic factors VEGF and TGF-β in Alzheimer's disease and vascular dementia. *Neurobiol Aging* 23:237-43.

- 131. Tarumi T, Dunsky DI, Khan MA, Liu J, Hill C, Armstrong K, Martin-Cook K, Cullum CM, Zhang R (2014) Dynamic cerebral autoregulation and tissue oxygenation in amnestic mild cognitive impairment. *J Alzheimers Dis* **41**:765-78.
- 132. Tayler HM, Palmer JC, Thomas TL, Kehoe PG, Paton JFR, Love S (2014)
 Investigating the relationship between cerebral Aβ and systemic
 hypertension [Abstract]. *Neuropathol Appl Neurobiol* **40 (Suppl 1)**:41.
- 133. Thal DR, Ghebremedhin E, Rub U, Yamaguchi H, Del Tredici K, Braak H

 (2002) Two types of sporadic cerebral amyloid angiopathy. *J Neuropathol Exp Neurol* **61**:282-93.
- 134. Thomas T, Miners S, Love S (2015) Post-mortem assessment of hypoperfusion of cerebral cortex in Alzheimer's disease and vascular dementia. *Brain* **138**:1059-69.
- Thomas T, Thomas G, McLendon C, Sutton T, Mullan M (1996) β-Amyloid-mediated vasoactivity and vascular endothelial damage. *Nature* 380:168-71.
- 136. Toews AD, White FV, Morell P (1988) Metabolism of functional groups modifying the CNS myelin-associated glycoprotein. *J Neurochem* **51**:1646-50.
- 137. Tohgi H, Yonezawa H, Takahashi S, Sato N, Kato E, Kudo M, Hatano K, Sasaki T (1998) Cerebral blood flow and oxygen metabolism in senile dementia of Alzheimer's type and vascular dementia with deep white matter changes.

 Neuroradiology 40:131-7.
- 138. Tomonaga M (1981) Cerebral amyloid angiopathy in the elderly. *J Am Geriatr Soc* **29**:151-7.

- 139. Tong XK, Hamel E (1999) Regional cholinergic denervation of cortical microvessels and nitric oxide synthase-containing neurons in Alzheimer's disease. *Neuroscience* **92**:163-75.
- 140. Townsend KP, Obregon D, Quadros A, Patel N, Volmar C, Paris D, Mullan M (2002) Proinflammatory and vasoactive effects of Aβ in the cerebrovasculature. *Ann N Y Acad Sci* **977**:65-76.
- 141. Tschope C, Schultheiss HP, Walther T (2002) Multiple interactions between the renin-angiotensin and the kallikrein-kinin systems: role of ACE inhibition and AT1 receptor blockade. *J Cardiovasc Pharmacol* **39**:478-87.
- 142. Vaucher E, Hamel E (1995) Cholinergic basal forebrain neurons project to cortical microvessels in the rat: electron microscopic study with anterogradely transported Phaseolus vulgaris leucoagglutinin and choline acetyltransferase immunocytochemistry. *J Neurosci* **15**:7427-41.
- 143. Vinters HV, Gilbert JJ (1983) Cerebral amyloid angiopathy: incidence and complications in the aging brain. II. The distribution of amyloid vascular changes. *Stroke* **14**:924-8.
- 144. Waite JJ, Holschneider DP, Scremin OU (1999) Selective immunotoxin-induced cholinergic deafferentation alters blood flow distribution in the cerebral cortex. *Brain Res* **818**:1-11.
- 145. Winkler EA, Sagare AP, Zlokovic BV (2014) The pericyte: a forgotten cell type with important implications for Alzheimer's disease? *Brain Pathol* **24**:371-86.
- 146. Yamada M, Tsukagoshi H, Otomo E, Hayakawa M (1987) Cerebral amyloid angiopathy in the aged. *J Neurol* **234**:371-6.

- 147. Yan Z, Feng J (2004) Alzheimer's disease: interactions between cholinergic functions and β -amyloid. *Curr Alzheimer Res* **1**:241-8.
- 148. Yang SP, Bae DG, Kang HJ, Gwag BJ, Gho YS, Chae CB (2004) Coaccumulation of vascular endothelial growth factor with β -amyloid in the brain of patients with Alzheimer's disease. *Neurobiol Aging* **25**:283-90.
- 149. Zetterberg H, Mortberg E, Song L, Chang L, Provuncher GK, Patel PP, Ferrell E, Fournier DR, Kan CW, Campbell TG, Meyer R, Rivnak AJ, Pink BA, Minnehan KA, Piech T, Rissin DM, Duffy DC, Rubertsson S, Wilson DH, Blennow K (2011) Hypoxia due to cardiac arrest induces a time-dependent increase in serum amyloid β levels in humans. *PLoS One* **6**:e28263.
- 150. Zhang ZG, Zhang L, Jiang Q, Zhang R, Davies K, Powers C, Bruggen N, Chopp M (2000) VEGF enhances angiogenesis and promotes blood-brain barrier leakage in the ischemic brain. *J Clin Invest* **106**:829-38.
- 151. Zhu J, Song W, Li L, Fan X (2016) Endothelial nitric oxide synthase: a potential therapeutic target for cerebrovascular diseases. *Mol Brain* **9**:30.

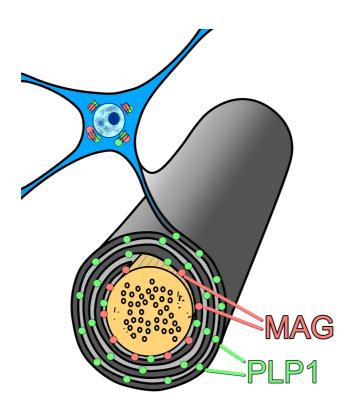


Figure 1. Schematic illustration of the distribution of MAG (pink dots) and PLP1 (green dots) in the myelin sheath. PLP1 is distributed throughout the myelin sheath whereas MAG is inserted only far from the cell body, in the adaxonal loop of myelin, the first part of the sheath to degenerate when blood supply is insufficient to meet the energy demands of the oligodendrocyte. As MAG and PLP1 are stable post mortem and have half-lives of several months, a decline in MAG:PLP1 in post-mortem brain tissue reflects a hypoperfusion-related energy deficit over a relatively long period prior to death. Image adapted from (71).

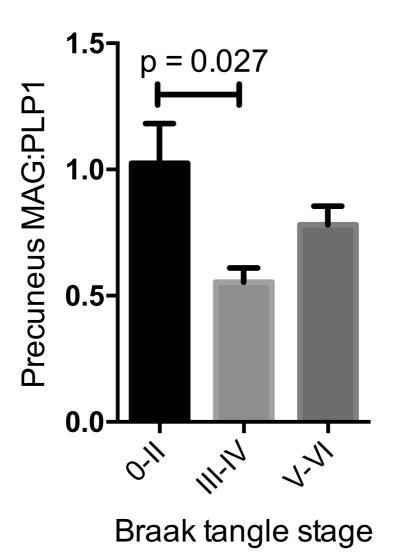


Figure 2. Bar chart showing decline of MAG:PLP1 in the precuneus in AD. The decline is most marked in early disease (Braak tangles stage III-IV). The ratio may rise in late disease as a consequence of falling metabolic demand.

Reproduced from (75).

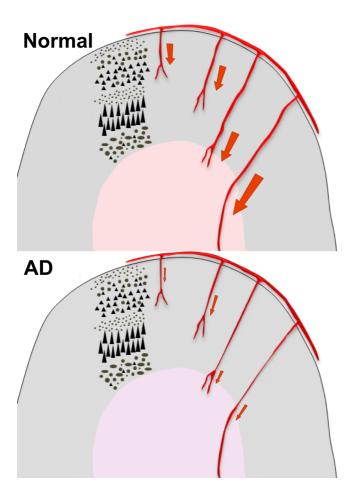


Figure 3. Perforating arterioles that arise from meningeal branches of the major cerebral arteries supply both the cerebral cortex and the underlying white matter. Excessive vasoconstriction within the cerebral cortex in AD affects not only cortical arterioles but also perforating arterioles that traverse the cortex.

Thus vasoconstriction within the cerebral cortex contributes to hypoperfusion of the white matter even if arterioles in the white matter are not themselves constricted. Modified from (23).

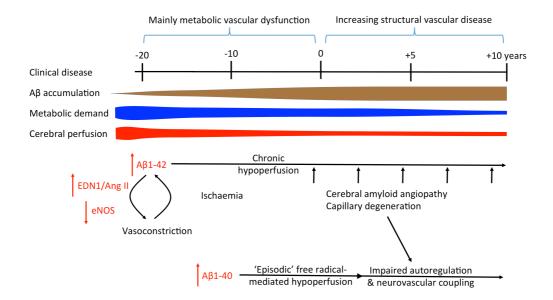


Figure 4. Shift from purely metabolic to structural vascular dysfunction over the course of AD (in relation to the onset of clinical disease, at 0 years). After a brief period of increased metabolic demand and cerebral blood flow, the progressive accumulation of A β in early (preclinical) stages of AD drives several metabolic pathways that lead to excessive vasoconstriction and reduced cerebral perfusion. Cerebral perfusion declines faster than metabolic demand. A β 42-induced metabolic processes may be more important in driving chronic hypoperfusion, and A β 40-induced processes in impairing vascular responsiveness. As the disease progresses, capillary damage and, in many patients, CAA, become increasingly important contributors to both chronic hypoperfusion and abnormalities of autoregulation and neurovascular coupling.