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Effects of acidic and neutral biochars on properties and cadmium retention of soils

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| 1           | Effects of acidic and neutral biochars on properties and   |
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| 2           | cadmium retention of soils   |
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| - 11 | ACCEPTED MANUSCRIPT                           |
|------|---|
| 24   | Abbreviations                                 |
| 25   | ANC = Acid neutralization capacity            |
| 26   | Ba = Soil from Mt Bygalore (Ferrosol, acidic) |
| 27   | CEC = Cation exchange capacity                |
| 28   | CL biochar = Chicken litter biochar           |
| 29   | EC = Electrical conductivity                  |
| 30   | Mt = Soil from Mt Shank (Dermosol, alkaline)  |
| 31   | PyC = Pyrogenic carbon                        |
| 32   | TOC = Total organic carbon                    |
| 33   | WS biochar = Wood shaving biochar             |
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#### 44 Abstract

In this study, an acidic biochar and a neutral biochar were applied at 5 wt% into two 45 soils for an 11-month incubation experiment. One Ferrosol soil (Ba) was slightly acidic 46 with low organic matter and the other Dermosol soil (Mt) was slightly alkaline with high 47 organic matter. The acidic (pH=3.25) wood shaving (WS) biochar had no marked impact on 48 nutrient levels, cation exchange capacity (CEC), pH and acid neutralisation capacity (ANC) 49 of either soil. By contrast, the neutral (pH=7.00) chicken litter (CL) biochar significantly 50 increased major soluble nutrients, pH, ANC of soil Ba. In terms of C storage, 87.9% and 51 52 69.5% WS biochar-C can be sequestrated as TOC by soil Ba and Mt, respectively, whereas only 24.0% of CL biochar-C stored in soil Ba and negligible amount in Mt as TOC. 53 Biochars did not have significant effects on soil sorption capacity and sorption reversibility 54 except that CL biochar increased sorption of soil Ba by around 25.4% and decreased 55 desorption by around 50.0%. Overall, the studied acidic C rich WS biochar held little 56 agricultural or remedial values but was favourable for C sequestration. The neutral mineral 57 rich CL biochar may provide short-term agricultural benefit and certain sorption capacities 58 59 of lower sorption capacity soils, but may be unlikely to result in heightened C sequestration 60 in soils. This is the first study comprehensively examining functions of acidic and neutral biochars for their benefits as a soil amendment and suggests the importance of pre-testing 61 biochars for target purposes prior to their large scale production. 62

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Keyword: Acidic biochar, neutral biochar, soil, C sequestration, sorption

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#### 1. Introduction

Biochars are carbon-rich products produced through thermochemical processing of 70 biomass under an oxygen-deficient environment (Cao and Harris, 2010; Venegas et al., 71 2015). It has been a hot topic of research in recent years due to its versatile role in soil 72 biogeochemical processes. The most important environmental functions of biochar include 73 74 acting as a long-term carbon sink for climate change mitigation (Lehmann, 2007b; McBeath and Smernik, 2009; Bird et al., 2017), a soil conditioner that increases soil 75 nutrient retention, cation exchange capacity (CEC), soil fertility and crop yield (Liang et al., 76 77 2006; Downie et al., 2011; Qu et al., 2016), and an in situ immobilizer for soil heavy metal contaminants (Cao et al., 2009; Uchimiya et al., 2011a; Qian et al., 2013; Lu et al., 2017). 78 Biochars perform in these processes by increasing soil organic carbon and reducing 79 greenhouse gas emission (Lehmann, 2007b), increasing soil pH, acid neutralisation capacity 80 (ANC) and CEC (Glaser et al., 2002; Cheng et al., 2006; Yuan and Xu, 2011; Venegas et 81 al., 2015) as well as increasing soil intrinsic binding sites for soil contaminants (Cao and 82 Harris, 2010; Beesley et al., 2011; Uchimiya et al., 2011c). 83

84 Biochars are commonly alkaline (Jiang et al., 2012a) which contribute to their liming effects and enhanced sorption of soils for cationic contaminants. However, biochar 85 pH values can range from acidic to alkaline (Chan and Xu, 2009) and lower pH biochars 86 are normally neglected. Biochar pH increases with increasing pyrolysis temperature as 87 88 more acidic functional groups can be removed at higher temperatures (Ippolito et al., 2016). Biochars produced under low pyrolysis temperatures can be acidic (Novak et al., 2009; 89 90 Hagner et al., 2016; Zhang et al., 2017). For example, birch (Betula spp.) biochar produced at 300 °C and 375 °C were shown to be acidic (pH=5.1 and 5.2, respectively) (Hagner et al., 91 2016). Pecan shell (350 °C) and switchgrass (250 °C) biochar had a pH of 5.9 and 5.4, 92 respectively (Novak et al., 2009). In limited studies about lower pH biochars, low-93

temperature (250 and 350 °C) acidic switchgrass biochars were found to lower pH and
initially increase plant-available nutrients in aridic calcareous soils (Ippolito et al., 2012;
Ippolito et al., 2016). Similarly, neutral biochars may also behave differently from normally
available alkaline biochars in envrionmetal processes after being added into soils. From this
sense, a more comprehensive examination of potential benefits of acidic and neutral biochar
on soil nutrient leaching, C storage and contamination remediation is essential.

Cadmium is one of the most hazardous metals and is readily absorbed from soil to 100 plant with a relatively high transfer coefficient and subsequently to animals and human 101 through fodder and food products (Park et al., 2011; Zhao et al., 2015a). In this study, Cd 102 was selected as a model metal to study contaminant retention cpacity of soils given its wide 103 104 presence in agricultural soils due to Cd-rich phosphate fertilizers applications (Naidu et al., 1994; Ali et al., 2013; Muehe et al., 2013). The objectives of this work were to (i) study 105 effects of acidic and neutral biochars on soil properties especially surface charge properties; 106 (ii) examine the C sequestration capacity of acidic and neutral biochars by evaluating soil 107 total organic carbon (TOC) and stable organic carbon (pyrogenic carbon, PyC) content; (iii) 108 explore sorption behaviours and mechanisms of biochar amended soils for Cd. 109

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## 111 **2. Methods**

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## 2.1 Incubation of biochar-amended soils

Two typical Australian soils (0-10 cm) were sampled for this study. One Ferrosol soil from Mt Bygalore (Ba) of New South Wales (S33°39.88', E146°49.07") of Australia that is lower in organic matter content and slightly acidic (pH=6.14, TOC=16.3 mg/g). The other Dermosol soil was sampled from Mt Shank (Mt) of South Australia (S37°56.78', E140°44.59') that is slightly alkaline higher in organic matter soil (pH=7.87, TOC=78.9 mg/g). One biochar made from chicken litter (CL, 550 °C) and one from wood shavings

119 (WS, 650 °C) were obtained from commercial producers. The biochars were made by 16minute slow pyrolysis (8 minutes in the drier, 8 minutes in the pyrolysis chamber) in a 120 Continuous Flow Pyrolyzer. Biochar products were water mist quenched immediately after 121 pyrolysis, then additional water was hosed onto the bulk product. Before use, both biochars 122 were air dried and sieved to pass through a 2-mm stainless steel sieve to represent their 123 typical use as a soil amendment. Biochars were mixed with soils at the ratios 5 wt% on a 124 dry weight base. Soils with/without biochar amendment were incubated at  $25 \pm 2$  °C for 11 125 months in 2L glass jars with holes on the lids. The jars were maintained at 60% water 126 holding capacity (WHC) and weighed for water replenishment every week within the first 3 127 months and every fortnight thereafter. Incubation of all samples were in duplicates. At the 128 end of the 11 months, the soil samples were dried and sieved (<2 mm or <150 µm) for 129 further chemical analysis and the sorption experiment. These soil samples after 11 months 130 incubation were used for following analysis and sorption experiment. Following the way 131 "soil name + months of incubation + biochar types", the samples were recorded as Ball, 132 Bal1WS, Bal1CL, Mt11, Mt11WS, Mt11CL, respectively. 133

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#### 135 2.2 Soil and biochar characterization before and after treatment

Soil and biochar pH, electrical conductivity (EC) were determined using 1:5 and 1:10 136 sample to water ratio, respectively. Soil TOC were determined by combusting oven dried and 137 ground soil samples (<150 µm) at 1100 °C in a CHN Elemental Analyzer (Euro EA 3000 138 Elemental Analyzer, Eurovector SPA, Milano, Italy) that uses infrared technology to quantify 139 CO<sub>2</sub>. Soil pyrogenic carbon (PyC), was measured by a chemo-thermal oxidation (CTO-375) 140 method. Both TOC and PyC analysis were collaborated with a lab in Switzerland where same 141 analytical procedure as Agarwal and Bucheli, (2011) were applied. Soil and biochar total 142 metals was extracted by microwave digestion in aqua regia following USEPA 3051 40 143

144 method before detecting metals by inductively coupled plasma mass spectrometry (ICP-MS, Agilent 7900, USA). Soil texture was determined by the hydrometer method (Gee et al., 145 1986). Amorphous Al, Mn and Fe (Aloxa, Mnoxa, Feoxa) was extracted by 0.2 M ammonium 146 oxalate/oxalic acid following Rayment and Higginson (1992). The contents (wt%) of carbon 147 (C), hydrogen (H), nitrogen (N), sulphur (S) of biochars were measured by a CHNS analyzer 148 (Vario Micro cube, Elementar, Germany). Ash content (%) was measured by heating samples 149 under 800 °C for 4 h in muffle furnace. The weight percent of oxygen was determined by 150 mass difference (Chen et al., 2008; Cheng et al., 2013; Luo et al., 2016). The acid 151 neutralisation capacity (ANC) of the biochar and soil samples is defined as the quantity of 152 acid or base (cmol  $H^+/kg$ ) required to shift the initial pH of the material to a pH of 4 153 (Venegas et al., 2015). In this study, we used a 1:200 solid to liquid ratio in 0.01 M NaNO<sub>3</sub> 154 for ANC measurement. Specific surface area of char samples (<2mm) were measured with 155 nitrogen adsorption isotherms at liquid nitrogen temperature (-196 °C) by a Surface Area 156 Analyzer (Micromeritics ASAP 2020 M, USA). Biochar samples (=0.2-0.25 g) were 157 outgassed overnight at 60 °C under vacuum at 2 Torr before N<sub>2</sub> adsorption. The molecular 158 surface area of 16.2  $\text{\AA}^2$  for N<sub>2</sub> and the BET (Brunauer-Emmett-Teller) equation (Brunauer et 159 al., 1938) were used to calculate the surface area of the char samples. Fourier Transformed 160 Infrared Spectroscopy (FTIR) (Nicolet IS10, Thermo Fisher, Waltham, MA, USA) spectra 161 were collected to get information on surface functional groups of char samples. This was 162 done by applying dehydrated KBr disc technique, where char samples (150 µm) char sample 163 were mixed with spectroscopic grade KBr at a ratio of 1:50 before scanning to produce 164 sufficient absorbance. Spectra over the 4000-400 cm<sup>-1</sup> range were obtained by the co-addition 165 of 64 scans with a resolution of  $4 \text{ cm}^{-1}$  and a mirror velocity of 0.6329 cm/s. Surface 166 morphology of the CL biochar in soil Ba matrix were examined under an Environmental 167

Scanning Electron Microscope (SEM) (Zeiss Sigma, Germany) equipped with a Bruker
energy dispersive X-ray spectroscopy (EDS) detector

170 **2.3 Surface charge of soils** 

Effects of biochars on surface charge properties of soil were assessed by CEC and 171 electrophoretic mobility property of soils. CEC is an indicator of abundance of the negative 172 charge on the surface of a material, which can be balanced by exchangeable cations 173 (Mukherjee et al., 2011; Jiang et al., 2012a; Zhao et al., 2015b; Jiang et al., 2016b). In this 174 study, soil CEC was measured by BaCl<sub>2</sub>/NH<sub>4</sub>Cl compulsive exchange method descried by 175 Gillman and Sumpter (1986). Electrophoretic mobility property of soil samples, often 176 expressed as zeta potential is another way to evaluate soil surface charge properties. Zeta 177 178 potential is the electrical potential of a sliding plane which is the interface between the Stern and diffuse layers in the double layer model of colloidal particles (Appel et al., 2003; Jiang et 179 al., 2016b). Its electro-negativity depends on the amount of negative surface charges. Soil and 180 biochar-amended soil samples were ground to <150µm before being dispensed into 0.01M 181 NaNO<sub>3</sub> suspension (0.02%, w/v) to guarantee the good suspension of samples. The 182 suspensions were then equilibrated on a rotatory shaker for 24 h. The pH values of 183 suspension were recorded and zeta potential of these suspensions were measured by Zetasizer 184 Nano ZS instrument (Malvern, ZEN3600, UK). 185

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#### 187 **2.4 Cadmium sorption and desorption**

The Cd sorption experiment was performed using a batch equilibration technique. Our preliminary test showed that the two studied biochars did not show noticeable influence to sorption capacities of both soils under 300  $\mu$ M Cd. Hence, we increased Cd concentration to 1500  $\mu$ M to evaluate the effects of biochars on maximum sorption capacity of the studied soils. Specifically, the sorption was performed by agitating 1g soils in 30 mL of 0.01 M

NaNO<sub>3</sub> solution containing 0, 30, 150, 300, 600, 900, 1200 and 1500  $\mu$ M Cd(NO<sub>3</sub>)<sub>2</sub> on a 193 reciprocating shaker at 200 rpm for 24 h. Solution pH at 0 and 300 µM Cd(NO<sub>3</sub>)<sub>2</sub> was 194 recorded after equilibrium, and the mixtures were centrifuged, filtered through 0.22µm 195 cellulose acetate syringe filter and acidified prior to major element (Cd, K, Ca, Na, Mg, Al, 196 Fe, Mn) analysis by ICP-MS (Agilent 7900, USA). Metrohm Ion chromatography was used 197 for  $PO_4^{3-}$ ,  $Cl^-$ ,  $NO_3^-$ ,  $SO_4^{2-}$  analysis (790 Personal IC, Switzerland). Dissolved organic 198 carbon (DOC) and dissolved inorganic carbon (DIC) were measured with TOC analyzer 199 (Multi N/C 3100, Analytik Jena, Germany). The amount of Cd sorbed was calculated by 200 subtracting the remaining Cd concentration after sorption from the initial Cd concentration. 201 Subsequently, Cd speciation was conducted by PHREEQC Interactive v3.3.7 software and 202 sorption isotherms were fitted in to Langmuir models to evaluate the sorption properties of 203 soil samples. After removing all solutions from sorption experiment, desorption studies were 204 carried out at a low (150 µM) and high (1500 µM) Cd loading to assess the reversibility of 205 sorption. Specifically, 30 mL 0.01M NaNO<sub>3</sub> was added to the decanted centrifuged tubes 206 following sorption experiment and mixed end over end at 200 rpm for another 24 h. 207 Detection of Cd was performed by ICP-MS (Agilent 7900, USA). 208

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#### 210 **2.5 Statistical analyses**

All analysis was conducted in duplicate and expressed as means of the replicates. One-way analysis of variance (ANOVA) were carried out using SPSS version 19.0 to determine the significance of the differences between treatments. The post-hoc least significant difference (LSD) test was employed. When a significant F-value (p<0.05) were obtained, means of treatments were regarded as significantly different.

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#### 217 **3. Results and discussions**

#### 218 **3.1 Basic soil properties**

Pertinent properties of soils, biochars and 11-month incubated soils used in this study are listed in Tables 1, 2 and 3, respectively. The WS biochar contained a higher amount of C (75.6%) and minimal amount of ash (1.65%) while CL biochar contained relatively less carbon (33.7%) but considerable amount of ash (46.2%). High ash contents of biochars can indicate the higher content of inorganic minerals, which could raise pH, EC and CEC of amended soils (Ahmad et al., 2016).

WS biochar was strongly acidic (pH=3.27) and CL biochar was neutral (pH=7.00). 225 Both increase and decrease (Mierzwa-Hersztek et al., 2016) as well as no change (Shafiq, 226 2016) of soil pH can occur upon biochar amendment depending on biochar type and dose. In 227 our study, only CL biochar increased the pH of soil Ba markedly (p < 0.05) by 0.3 units. WS 228 did not noticeably change pH of both soils while CL biochar failed to change pH of soil Mt 229 significantly. Soil Mt held significantly higher intrinsic ANC than soil Ba due may be to its 230 higher CaCO<sub>3</sub> equivalence (23.2%) (Venegas et al., 2015; Jiang et al., 2016b). WS biochar 231 did not change the ANC of both soils whereas CL biochar increased ANC of both soils 232 significantly (p < 0.05). This suggests the mineral rich CL biochar contained high amounts of 233 pH buffering components like CaCO<sub>3</sub>. 234

The EC values of both soils was significantly (p<0.05) increased by CL but (p<0.05) markedly reduced by WS biochar (Table 3). For CL biochar, this was because it contained relatively high amounts of soluble cations and anions (Supporting information Table A1). In biochar application, it was suggested that the EC values of the biochars should be characterized to avoid creating unwanted salt effects (Cantrell et al., 2012). In this study, EC values in CL biochar amended soils were below the saline limits of soils (4 dS/m) (Mohamed et al., 2015) and would not cause salt stress to plant growth.

The CL biochar increased soluble anions (Cl<sup>-</sup>, PO<sub>4</sub><sup>3-</sup> and SO<sub>4</sub><sup>2-</sup>) of both soils 242 significantly (Table A1). As for soluble cations (Table A1), CL biochar increased soluble Na 243 of soil Ba by 10 fold and other mineral element Mg, K, Ca, Fe, Al by 1-3 fold. In contrast, 244 CL biochar increased only soluble Na and Ca of soil Mt at a magnitude of 2.2 and 1.7 fold, 245 respectively. Soil Mt had a high initial K, Mg, Fe, Al concentration, hence effects from CL 246 biochar was not significant. These soluble minerals stored in biochar can serve as plant 247 nutrients (fertilizer) during drought or nutrient deficient conditions (Cao et al., 2014; Hagner 248 et al., 2016). Hence, the WS biochar did not provide much fertilizer benefit while CL could 249 250 potentially be a source of plant nutrients (K, Ca, Mg, P, Fe).

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#### 3.2 Surface charges of soils 252

Adsorption of heavy metals in soil is mainly governed by interactions with negatively 253 charged soil components (Naidu et al., 1997b; Bolan et al., 1999; Chintala et al., 2013). This 254 surface charge of soil samples evaluated by cation exchange capacity (CEC) and zeta 255 potential are shown in Fig 1. Results showed that CEC of soil Mt (33.4 cmol+/kg) was 256 around three times that of soil Ba (11.6 cmol+/kg) (Fig. 4). The higher TOC content, CaCO<sub>3</sub> 257 equivalence as well as amorphous Mn, Al, Fe oxides of soil Mt than soil Ba (Table 1) can be 258 responsible for higher negative charge of soil Mt. In addition, higher pH of soil Mt can also 259 contribute to its higher CEC (Naidu et al., 1990; Yu, 1997). 260

Both biochars failed to change soil CEC significantly (p>0.05). This indicated low 261 CEC values of these two studied biochars. Normally biochars are considered to develop more 262 oxygen-containing functional groups (Bruun et al., 2011; Manyà, 2012) and hence increased 263 CEC and negative charge of soils (Lehmann, 2007a; Maia et al., 2011; Mohamed et al., 2015; 264 Zhang et al., 2017). Our study, however, suggested the increase of soil functional groups 265 caused by biochars was not significant enough to change soil surface charge. Previously, 266

Slavich et al., (2013) observed no effect of a green waste biochar (10 t/ha) on exchangeable cations and CEC of a Ferrosol in a 36-month field experiment. Schulz and Glaser (2012) found biochar (5%) failed to change CEC of an infertile sandy soil in a greenhouse experiment. Given that CEC is the indicator of capacity of soils to retain nutrients, our studied biochars did not increase the ability of studied soils to hold nutrients.

Surface charge as indicated by zeta potential, the electrical potential of a sliding plane of soil colloids (Zhao et al., 2015b) were negative at original soil pH (0.01M NaNO<sub>3</sub>) for all soil samples (Fig 1). Hence, all soil samples carried negative surface charge. And the zeta potential values after both biochar amendment were comparable (p>0.05) to that of biocharfree soils. These results were consistent with results from CEC on that both biochar amendment did not noticeably change the surface charge characteristics of the studied soils.

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#### 279 **3.3** C sequestration potential

Soil PyC is the fraction of TOC that was assumed to be more stable and therefore 280 resistant to thermal oxidation (combustion at 375°C) than other soil organic matter. Table 3, 281 Fig 2a and 2b show the TOC and PyC content in studied soil samples. Results showed that 282 after 11 months, TOC content of soil Ba (13.2 mg/g) increased by 253% (p < 0.05) due to 283 addition of WS biochar (to 46.4 mg/g) and CL biochar by 88.9% (to 24.9 mg/g), respectively. 284 The PyC content of soil Ba (0.61 mg/g) was increased significantly (p < 0.05, Table 3) to 3.25 285 mg/g by WS but was not significantly changed by CL biochar. Regarding soil Mt, after 11 286 months, WS biochar increased its TOC (81.7 mg/g) by 11.1% (to 90.8 mg/g) but this increase 287 was not statistically significant (p>0.05). Hence, we conclude that both biochars did not show 288 statistically significant change of TOC and PyC level of soil Mt but increased that of soil Ba 289 noticeably. 290

- 291 Given that the C content of WS and CL biochar were 75.6% and 33.7%, respectively (Table 2) and biochar amendment ratio was 5 wt%, the percentage of biochar-C sequestered 292
- as TOC and PvC by each soil were calculated by the following equations: 293

$$Biochar C sequestered as TOC(\%) = \frac{(TOC \ biochar \ amended \ soils - TOC \ control \ soils)/gsoil}{Biochar \ C/gsoil} * 100$$

$$Biochar C sequestered \ as PyC(\%) = \frac{(PyC \ biochar \ amended \ soils - PyC \ control \ soils)/gsoil}{Picket \ biochar \ amended \ soils - PyC \ control \ soils)/gsoil} * 100$$

Biochar C sequestered as  $PyC(\%) = \frac{\alpha}{2}$ Biochar C/gsoil

The calculated percentage (Table 4) showed that, within a time range of 11 months, 294 87.9% and 69.5% of WS and CL biochar-C was sequestrated by soil Ba as TOC. Comparably, 295 only 24.0% of WS biochar-C was stored as TOC in soil Mt while CL biochar-C totally 296 degraded in soil Mt. Similarly, soil Ba stored 6.96% and 3.54% of WS and CL biochar-C as 297 thermally stable OC (PyC), whereas no contribution of biochar-C to PyC of soil Mt was 298 299 noted.

Hence, overall, both biochars had the capacity to increase total OC of soil Ba but 300 301 acidic WS biochar showed a higher capacity within a period of 11 months. This may be due to the higher C content of WS biochar than CL biochar (Table 2). In addition, WS biochar 302 had a smaller O/C ratio (0.25) than CL biochar (0.41) (Table 2), suggesting C in WS biochar 303 may have more stable conjugated aromatic structures (Slavich et al., 2013) that resist 304 degradation more. Meanwhile, soil Ba displayed a better C sequestration effect than soil Mt. 305 One of the reasons may be lower pH of soil Ba than soil Mt as lower pH was suggested to 306 favour C stabilization (Skjemstad et al., 1996; Clough and Skjemstad, 2000). According to 307 PyC content obtained by CTO-375 method, soil Ba can store only <7% while soil Mt stored 308 negligible percentage of biochar-C as stable OC in the 11-month period. Our previous study 309 showed that combustion of biochars (including biochars used in this study) at 375 °C for 24 h 310 almost oxidized all the C in biochars (data not shown). This was also noted by other studies 311 (Quénéa et al., 2006; Roth et al., 2012; Kerré et al., 2016). Comparing to other commonly 312 used methods, the CTO-375 method can remove all of the char PyC and tends to obtain the 313

314 smallest PyC value that represents highly condensed soot PyC (Hammes et al., 2007; Poot et al., 2009; Qi et al., 2017). Hence, on one hand, the CTO-375 method may have 315 underestimated the PyC introduced from biochar into soils. Better methods to quantify PyC 316 dynamics in biochar-amended soils are needed in the future. On the other hand, the better 317 survival of biochars through CTO-375 combustion in soil Ba matrix than their bare biochar 318 counterparts suggests possible incorporation of biochar-C into soil aggregates which provides 319 physical protection of biochar-C from thermal degradation. This is also one of the most 320 important mechanisms for TOC and PyC to be stabilized into soils. 321

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#### 323 **3.4 Sorption Properties**

#### 324 **3.4.1 Sorption isotherms**

The studied soils and biochars contain very low amounts of heavy metal(loid)s and only negligible amount of Cd (supporting information Table A2). The sorption isotherms of all soils were L type (Limousin et al., 2007) where the ratio of equilibrium solute concentration in solution to that on soils decreased with increasing solute concentration and suggested the soil surfaces were progressively get saturated by Cd. The sorption isotherms of all soil samples were fitted into Langmuir models, with linear form of this model as follows (Yan et al., 2015):

$$\frac{C_e}{q_e} = \frac{Ce}{q_{max}} + \frac{1}{bq_{max}}$$

where  $C_e$  (mg/L) and  $q_e$  (mg/g) are the equilibrium concentrations of Cd in solution and on soil samples, respectively. Langmuir constants  $q_{max}$  (mg/g) and b (L/g) represent maximum sorption capacity and sorption strength, respectively.

The Cd sorption isotherms (Fig 3a and 3b) of biochar free and biochar amended soils all followed the Langmuir model ( $R^2 = 0.82-0.99$ ) (Table 5). The amount of Cd sorbed on all

337 soil samples increased with increasing initial Cd concentrations. An assessment of the sorption maxima (q<sub>max</sub>) calculated from Langmuir model showed that soil Mt had 338 significantly higher  $q_{max}$  (10.2 mg/g) than soil Ba (3.71 mg/g). This may be due to difference 339 in properties of the two soils. Higher pH and higher capacity to maintain pH (higher ANC) of 340 soil Mt should favour the higher sorption capacity of soil Mt. This is because higher pH 341 affords more negative surface charge (Naidu et al., 1994; Zhou and Wong, 2001; Forján et al., 342 2016; Jiang et al., 2016a) and more hydrolysis  $Cd^{2+}$  that is with higher affinity than free  $Cd^{2+}$ 343 ions to sorbent surface (Davis and Leckie, 1978; Naidu et al., 1994; Yu, 1997; Jiang et al., 344 2016a). Higher TOC and CaCO<sub>3</sub> content (Uchimiya et al., 2011b; Forján et al., 2016; Usman 345 et al., 2016) and Fe, Al and Mn oxides (Yu, 1997; Jiang et al., 2012b) of soil Mt could also 346 drive its high sorption capacity for Cd. WS biochar amendment did not noticeably change 347 sorption capacity of both soils. CL biochar application increased q<sub>max</sub> of soil Ba by 25.6% 348 and but showed no effect on sorption capacities of soil Mt. This indicated application of both 349 biochars to alkaline organic matter rich soils may not show its remedial function and acidic 350 351 biochar would not show remedial effects for both soils.

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353 **3.4.2 Proportions of specific sorption** 

Sorption of metal cations are relevant to negative surface charge (Naidu et al., 1994; 354 Naidu et al., 1997a; Chintala et al., 2013). Mechanisms involved in metal ions attraction from 355 soil solutions to soil surfaces include non-specific sorption (outer-sphere complexes) through 356 cation exchange reactions where charge of soils are balanced by sorbed metal ions, specific 357 sorption (inner-sphere complexes) where chemical bonds form between metals and soil 358 surface (Evans, 1989; Naidu et al., 1994; Echeverria et al., 1998; Strawn and Sparks, 2000; 359 Appel and Ma, 2002; Bradl, 2004; Bolan et al., 2014) and co-precipitation. On a relative basis, 360 specific sorption and co-precipitation can retain Cd nearly irreversibly in many cases while 361

nonspecific sorption renders the metals most labile and easily reversible (Appel and Ma,2002).

The nonspecifically sorbed metals were estimated by the amount released from 364 desorption process (Jiang et al., 2012b; Dai et al., 2015). Data from Fig 4 shows that at lower 365 Cd loading rate (150µM), non-specific sorption accounted for around 0.19% of the total 366 sorption of soil Mt, and this ratio increased to around 0.35% at a higher intial Cd 367 concentration (1500µM) but was still minimal. The ratio of non-specific sorption for soil Ba 368 was relatively higher which was around 2.21% and 8.17%, at 150 and 1500 µM Cd loading, 369 respectively. Hence, both soils, especially soil Mt, were dorminated by specific sorption. This 370 specific sorption can be through Cd complexation by multidentate ligands on the surface of 371 particulate organic matter and nonbridging -OH sites of various hydro(oxide)s (Uchimiya et 372 al., 2011c). On the one hand, the large number of functional groups of soil organic matter can 373 retain heavy metal mostly by surface complexation and surface precipitation (Bradl, 2004). 374 On the other, various Fe, Al and Mn oxides could have a good contribution to the specific 375 sorption capacity of metal ions (Yu, 1997) with correlation of specific sorption with free Fe 376 oxides having been reported (Jiang et al., 2012b). 377

Biochar amendments did not noticeably influence the ratio of non-specific sorption of the two soils except a 50.0% reduction caused by CL biochar on that of soil Ba. The Pand Si-related minerals may be responsible for the CL manure biochar to increase the nonelectrostatic Cd sorption of soil Ba (Dai et al., 2015). The combination of Cd with Si, Al, P may be supported by the EDS spectrum of CL biochar in soil Ba matrix (Fig 5).

383

#### 384 3.4.3 Sorption mechanisms

385 Heavy metal can be removed by biochar through direct electrostatic interactions 386 and cation exchange (nonspecific) as well as surface complexation with functional groups

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- (specific) (Trakal et al., 2014; Inyang et al., 2015; Lu et al., 2017) and co-precipitation.
  Hence, sorptive mechanisms of biochar-amended soils for Cd will be discussed from nonspecific and specific aspects in this section. To make the discussion easy, co-precipitation
  was included in the specific irreversible sorption.
- 391

#### 3.4.3.1 Non-specific sorption

The surface of all soil samples were negatively charged as indicated by negative 392 zeta potential (Table 3 and Fig 1). Hence, the sorption systems were favourable and direct 393 electrostatic interaction can contribute to the sorption of Cd. In addition, given measurable 394 CEC of both soils (Table 3 and Fig 1), Cd can be sorbed on soil constituents by replacing 395 readily exchangeable cations such as  $Ca^{2+}$  and  $Mg^{2+}$  (Uchimiya et al., 2011b). In this study, 396 Ca and Mg was released after Cd loading (mmol/g, after subtracting the amount released 397 from control sample, 0mM) for both soils, suggesting cation exchange as one of the 398 mechanisms of Cd sorption (Cui et al., 2016; Usman et al., 2016) (Table A3). Both Ca and 399 Mg release were higher at higher initial Cd loading in all soil samples. Soil Ba had more 400 release of Mg while soil Mt had higher release of Ca. Assuming 1 mg/kg Mg or Ca will be 401 released to sorb 1 mg/kg Cd, we calculated the total contribution of Ca + Mg release to Cd 402 retention (Rexchange) (Table A3). We can see that Rexchange of soil Mt (5.27%) at lower Cd 403 concentration (150µM) was slightly higher than soil Ba (3.40%), and this may be due to 404 higher CEC of soil Mt. However at higher concentration (1500µM), Rexchange of soil Mt 405 reduced to around 3.30% and that of soil Ba increased to 4.82%. Higher Rexchange of soil Ba 406 at higher Cd loading (1500µM) may be because not all cations were released from soil Ba 407 at 150µM, while the decrease of R<sub>exchange</sub> of soil Mt at higher concentration was likely due 408 to a higher increase of Cd sorption than cation release. Overall, WS biochar did not 409 infuence the amount of cation release markedly while CL biochar increased the release of 410

Mg in soil Ba and Mt at both Cd laodings. This indicated CL biochar increase the amountof Cd sorbed through increasing Mg exchange in both soils.

413

#### 414 **3.4.3.2 Specific sorption**

As pH can be the governing factor for metal sorption in aqueous systems, we cannot 415 ignore the changes of pH during sorption. The comparison of equilibrium pH of Cd-soil 416 reaction systems at an initial Cd loading of 0 and 300 µM showed that pH drop of soil Mt 417 (0.01-0.14 units) was not as significant as that of soil Ba (0.2-0.35) (Table A4). This drop 418 of pH after metal immobilization in sorption systems suggests either ion exchange of Cd 419 with dissociable proton (proton exchange) (Uchimiya et al., 2010; Inyang et al., 2012) or 420 421 forming of Cd(OH)<sub>2</sub> precipitants (Dong et al., 2014). For soil Mt, a pH decrease in the soil Mt system may be higher than the obtained value as its high ANC could counter-balanced 422 part of the pH drop (Inyang et al., 2012). Given that Cd(OH)<sub>2</sub> precipitation can form when 423 solution pH ranges from 8 to 9 (Sun et al., 2014; Zhao et al., 2016), both precipitation of 424 Cd(OH)<sub>2</sub> and ion exchange may have occurred in soil Mt system. For soil Ba, Cd(OH)<sub>2</sub> 425 may not be likely to form given the equilibrium pH range from 5.6-6.5, hence proton 426 exchange should have caused the pH drop. The precipitation of CdCO<sub>3</sub> in soil Mt was 427 indicated by saturation index calculated by PHREEQC Interactive v3.3.7. Sun et al. (2014) 428 429 also reported surface precipitation of Cd as carbonate in crop straws and wood shaving biochar sorption systems. 430

Cadmium can be sorbed by complexing with functional groups present on sorbent
surface (Sun et al., 2014; Cui et al., 2016). Soil organic matter and various soil hydr(oxide)s
contain large number of functional groups (Bradl, 2004; Uchimiya et al., 2011c). FTIR
spectrum (Fig 6) confirmed our WS and CL biochar contained phenolic hydroxyl (-OH)
groups at around 3450-3475 cm<sup>-1</sup> (Tsai et al., 2012; Wang et al., 2012), aromatic C=C

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functional groups at 1590 - 1641 cm<sup>-1</sup> (Wang et al., 2011; Zaafouri et al., 2016), aliphatic 436 ether C-O stretching at 1085/1110 cm<sup>-1</sup> (Tsai et al., 2012; Yang and Jiang, 2014) as well as 437 aromatic carbonyl/carboxylic groups (-COO/C=O) at 1427-1463 cm<sup>-1</sup> with a weaker signal 438 (Yuan et al., 2011; Choppala et al., 2012; Guo and Chen, 2014). Hence, surface 439 complexation should be an important mechanism for Cd sorption to soil Mt and Ba given 440 that around 99.7% and 91.8% of the sorption was specific (Fig 4). 441

442

#### 443

## 4. Conclusions and implications

Chicken litter (CL) biochar had a statistically significant liming effect for the 444 slightly acidic soil Ba as indicated by the significantly increased pH and acid neutralization 445 capacity. Also, CL biochar can serve as short-term nutrient source (increased Mg, Ca, K, P 446 levels) for lower-nutrient soil Ba but cannot improve the capacity of both soils to retain 447 nutrients (no increase in CEC). Acidic wood shaving (WS) biochar held little value in soil 448 liming and soil nutrient supply but it had a higher C sequestration potential (87.9% WS-C 449 in soil Ba and 69.5% WS-C in soil Mt) than CL biochar. The slightly acidic soil Ba could 450 sequester more C from biochars as TOC (87.9% WS-C and 69.5% CL-C) than alkaline soil 451 Mt (24.0% WS-C and -0.27% CL-C). The WS biochar did not change the sorption capacity 452 and sorption reversibility of both soils for Cd while CL biochar improved sorption of the 453 acidic soil Ba by 25.4% and reduced its desorption by around 50.0%. Hence based on the 454 11-month experiment. WS biochar did not offer much value for nutrient supply or remedial 455 aspects of studied soils but can be quite favourable for C sequestration, especially in the 456 acidic soil. In contrast, CL biochar can increase soil nutrient levels and sorption capacity for 457 Cd in soils with lower sorption capacities (like Ba) while offering no C sequestration 458 benefits based on the 11-month experiment. Future longer term experiment will be needed 459

to evaluate these biochars. This study indicates biochars should be properly evaluated
before large scale production and application according to the target use.

462

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Table 1. Characteristics of original soil samples

 Table 2. Characteristics of biochar samples

Table 3. Characteristics of biochar amended soil

Table 4. C sequestered from biochars as TOC and PyC in soils

 Table 5. Langmuir isotherm model parameters

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| Table 1.  | Characteristics  | of original   | soil | samples |
|-----------|------------------|---------------|------|---------|
| I UNIC II | Character istics | or or ignitur | DOM  | Samples |

| Soils | <mark>Soil type</mark> | Sand (%)  | Silt (%)  | Clay (%)  | <mark>Texture</mark><br>class | TOC<br>(mg/g) | CaCO <sub>3</sub><br>equivalence<br>(%) | pH <sup>a</sup> | EC <sup>a</sup> CEC<br>(dS/m) (cmol+/kg) | Fe <sub>oxa</sub> <sup>b</sup><br>(mg/kg) | Al <sub>oxa</sub> <sup>b</sup><br>(mg/kg) | Mn <sub>oxa</sub> <sup>b</sup><br>(mg/kg) |
|-------|------------------------|-----------|-----------|-----------|-------------------------------|---------------|---|-----------------|--|---|---|---|
| Ba    | <b>Ferrosol</b>        | 54.9±0.88 | 30.9±2.17 | 14.2±1.31 | <mark>Loam</mark>             | 16.3±0.85     | 2.00±0.50                               | $6.14 \pm 0.05$ | 50.0±0.14 11.6±0.51                      | 4370±37.2                                 | 1310±11.8                                 | 190±4.93                                  |
| Mt    | <b>Dermosol</b>        | 50.9±1.17 | 38.9±2.50 | 10.1±1.34 | <mark>Loam</mark>             | 78.9±5.64     | 23.2±0.67                               | $7.87 \pm 0.04$ | 260±50.2 37.8±1.71                       | 14900±858                                 | 2290±983                                  | 410±19.1                                  |

Note: <sup>a</sup> pH and EC were in 1:5 solid : water ratio. <sup>b</sup>Fe<sub>oxa</sub>, Al<sub>oxa</sub>, and Mn<sub>oxa</sub> - ammonium oxalate/oxalic acid extractable Fe, Al, Mn. Chip Manus

 Table 2. Characteristics of biochar samples

| Chars | C (%)     | H (%)     | N (%)           | S (%)         | O (%)     | O/C <sup>a</sup> | H/C <sup>a</sup> | Ash (%)   | SSA <sup>c</sup><br>(m <sup>2</sup> /g) | pH <sup>b</sup> | EC <sup>b</sup><br>(dS/m) | CaCO <sub>3</sub><br>equivalence<br>(%) | ANC <sup>d</sup><br>(cmol H <sup>+</sup> /kg) |
|-------|-----------|-----------|-----------------|---------------|-----------|------------------|------------------|-----------|---|-----------------|---------------------------|---|---|
| WS    | 75.6±0.23 | 3.23±0.01 | $0.25 \pm 0.00$ | 0.32±0.09     | 18.9±0.03 | 0.25             | 0.04             | 1.66±0.10 | 0.81±0.13                               | 3.25±0.07       | 421±15.0                  | 1.67±0.18                               | 2.00±0.07                                     |
| CL    | 33.7±2.33 | 2.41±0.12 | 3.81±0.30       | $0.40\pm0.04$ | 13.5±2.80 | 0.40             | 0.07             | 46.2±0.00 | 6.05±0.93                               | 7.00±0.16       | 9190±439                  | 10.1±0.23                               | 222±12.4                                      |

Note: <sup>a</sup>Molecular ratio of O/C and H/C are ash free base. <sup>b</sup>pH and EC were in 1:10 solid : water ratio. <sup>c</sup>SSA - (BET) specific surface area. <sup>d</sup>ANC - acid neutralization capacity.

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| Soils  | рН <sup>ь</sup>     | $EC^{b} (dS/m)$        | CEC (cmol+/kg) | ANC <sup>c</sup> (cmol H+/kg) | zeta potential (mV) | PyC <sup>d</sup> (mg/g) | TOC (mg/g)             |
|--------|---------------------|------------------------|----------------|-------------------------------|---------------------|-------------------------|------------------------|
| Ba11   | 6.28±0.04           | 348±3.53               | 11.5±1.13      | 6.51±0.72                     | -30.2±0.14          | 0.61±0.31               | 13.2±2.42              |
| Ba11WS | $6.20\pm0.02$       | 3180±2.12 <sup>a</sup> | 11.9±0.88      | 5.50±0.71                     | -30.1±0.71          | $3.25{\pm}0.98^{a}$     | $46.4 \pm 9.46^{a}$    |
| Bal1CL | $6.58 \pm 0.06^{a}$ | 7780±4.95 <sup>a</sup> | 12.6±0.85      | 12.0±1.41 <sup>a</sup>        | -32.1±0.99          | 1.21±0.37               | 24.9±5.89 <sup>a</sup> |
| Mt11   | $7.60\pm0.00$       | 797±4.24               | 36.5±0.94      | 27.0±0.71                     | -27.4±0.71          | 4.33±0.47               | 81.7±26.8              |
| Mt11WS | 7.63±0.01           | 590±14.8 <sup>a</sup>  | 33.4±1.80      | 25.0±0.71                     | -29.6±1.56          | 4.75±2.12               | 90.8±8.33              |
| Mt11CL | 7.57±0.00           | 1170±41.7 <sup>a</sup> | 33.1±2.05      | 48.0±1.41 <sup>a</sup>        | -30.6±0.57          | 4.28±1.65               | 76.5±12.2              |

#### Table 3. Characteristics of biochar amended soil

Notes: <sup>a</sup>Signifies difference from control is significant (*p*<0.05 by one-way AVONA anaylsis). <sup>b</sup>pH and EC were in 1:5 solid : water ratio. <sup>c</sup>ANC - acid neutralization capacity. <sup>d</sup>PyC - pyrogenic carbon.

| C<br>sources      | <mark>Sequestered</mark> as TOC<br>in <mark>acidic</mark> soil Ba (%) | <mark>Sequestered</mark> as TOC in<br><mark>alkaline</mark> soil Mt (%) | <mark>Sequestered</mark> as PyC<br>in <mark>acidic</mark> soil Ba (%) | <mark>Sequestered</mark> as PyC in<br>alkaline soil Mt (%) |
|-------------------|---|---|---|--|
| WS-C <sup>b</sup> | 87.9  | 24.0  | 6.96  | 1.12   |
| CL-C <sup>b</sup> | 69.5  | -   | 3.54  | -  |

## Table 4. C sequestered from biochars as TOC<sup>a</sup> and PyC<sup>a</sup> in soils

Note: <sup>a</sup>TOC and PyC were total organic carbon and pyrogenic carbon, respectively.

<sup>b</sup>WS-C and CL-C were C of wood shaving and chicken litter biochar, respectively. - indicate decrease of soil TOC and PyC by biochar were negligible.

od s.. C and PyC by .

| Samples | $q_{max}(mg/g)$ | b (L/mg) | $\mathbf{R}^2$ |
|---------|-----------------|----------|----------------|
| B11     | 3.50            | 0.19     | 0.99           |
| B11WS   | 3.53            | 0.21     | 0.99           |
| B11CL   | 4.39            | 0.31     | 0.99           |
| M11     | 9.55            | 0.69     | 0.89           |
| M11WS   | 8.69            | 0.78     | 0.90           |
| M11CL   | 9.87            | 0.69     | 0.88           |
|         |                 |          |                |
|         | F               |          |                |
|         | E.              |          |                |
|         |                 |          |                |
|         | TEN -           |          |                |
|         |                 |          |                |

Table 5. Langmuir isotherm model parameters

- Fig 1. CEC and zeta potential of soil samples
- Fig 2. TOC and PyC content of soil samples
- Fig 3. Sorption isotherm of soil samples
- Fig 4. Desorption ratio of Cd of soil Ba and Mt
- Fig 5. SEM image and EDS spectrum of CL biochar in soil Ba matrix

Fig 6. FTIR spectrum of WS and CL biochars



Fig 1. CEC and zeta potential of soil samples





Fig 2a. TOC content of soil samples





Fig 3a. Sorption isotherm of soil Ba

Fig 3b. Sorption isotherm of soil Mt







Fig 5. SEM image and EDS spectrum of CL biochar in soil Ba matrix



Fig 6. FTIR spectrum of WS and CL biochars

# Highlights

- The C rich acidic biochar did not work for nutrients supply or Cd retention but helped in storing C.
- > The neutral mineral rich biochar enhanced nutrient and sorption for some soils.
- > Neutral mineral rich biochar hardly increased soil C after 11 months.