

Interaction of the capsular polysaccharide A from *Bacteroides fragilis* with DC-SIGN on human dendritic cells is necessary for its processing and presentation to T cells

Karien Bloem^{1,2}, Juan J. García-Vallejo¹, Ilona M. Vuist¹, Brian A. Cobb³, Sandra J. van Vliet¹ and Yvette van Kooyk^{1*}

¹ Department of Molecular Cell Biology and Immunology, VU University Medical Center, Amsterdam, Netherlands

² Centre for Specialized Nutrition, Danone Research, Wageningen, Netherlands

³ Department of Pathology, Case Western Reserve University School of Medicine, Cleveland, OH, USA

Edited by:

Paul A. Ramsland, Burnet Institute, Australia

Reviewed by:

Meredith O'Keefe, Burnet Institute for Medical Research, Australia
Mihaela Gadjeva, Harvard Medical School, USA

*Correspondence:

Yvette van Kooyk, Department of Molecular Cell Biology and Immunology, VU University Medical Center, P. O. Box 7057, Van der Boechorststraat 7, 1007 MB Amsterdam, Netherlands.
e-mail: y.vankooyk@vumc.nl

The zwitterionic capsular polysaccharide A (PSA) of *Bacteroides fragilis* is the first carbohydrate antigen described to be presented in major histocompatibility complex (MHC) class II for the induction of CD4⁺ T cell responses. However, the identity of the receptor mediating binding and internalization of PSA in antigen presenting cells remains elusive. C-type lectins are glycan-binding receptors known for their capacity to target ligands for antigen presentation to T cells. Here, we investigated whether C-type lectins were involved in the internalization of PSA and identified dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin (DC-SIGN) as the main receptor for PSA on human dendritic cells (DC). The induction of PSA-specific T cell proliferation appeared to be completely dependent on DC-SIGN. These data reveal a crucial role for DC-SIGN in the endocytosis and routing of PSA in human DC for the efficient stimulation of PSA-specific CD4⁺ T cells.

Keywords: C-type lectin, dendritic cell, polysaccharide, pathogen, T cell proliferation

INTRODUCTION

In the defense against infection, specialized antigen presenting cells (APCs), such as dendritic cells (DCs) are able to endocytose pathogens into acidic compartments rich in endoproteases to allow the processing required for the extraction of antigenic epitopes. These peptide epitopes are then loaded onto major histocompatibility complex (MHC) class II and presented to CD4⁺ T cells. Until recently, peptides originating from the enzymatic degradation of protein antigens were thought to be the only class of molecules capable of being presented in MHC class II. However, these peptides can be posttranslationally modified with glycans, as has been shown for the MHC class II presentation of glycopeptides derived from the tumor antigen MUC1 (Vlad et al., 2002) or type II collagen (Michaëlsson et al., 1994).

Although DCs are able to endocytose different types of glycoconjugates and some of these glycoconjugates facilitate the induction of essential T cell help leading to the efficient production of glycan-specific antibodies (Astronomo and Burton, 2010); it was not until recently that the direct presentation of a polysaccharide on MHC class II was described. Zwitterionic polysaccharides (ZPSs) purified from the capsule of different bacteria were shown to activate CD4⁺ T cell responses without prior conjugation to a peptide (Tzianabos et al., 2000a,b). ZPSs are composed of repeating carbohydrate units in a polysaccharide structure, which contain both negatively and positively charged motifs, causing the formation of a helical structure (Choi et al., 2002), thereby resembling the structure of peptides in MHC class II. Removal of the positive charge in the ZPS of *B. fragilis*, polysaccharide A (PSA),

led to the inability to fit into the MHC class II groove and thus failure to activate T cells. Strikingly, removal of the positive charge does not alter endocytosis, demonstrating that PSA is internalized independently of the charge of the molecule (Cobb and Kasper, 2008).

Protein antigens are normally processed by endoproteases in acidic compartments within the endo-lysosomal pathway. The short peptides generated are then loaded predominantly on MHC class II for presentation to CD4⁺ T cells. PSA also needs to be processed before it can be presented on MHC class II. We have previously shown that cleavage of PSA is a non-enzymatic oxidative reaction facilitated by the production of nitric oxide (NO) by inducible NO synthase (iNOS, EC.1.12.13.39), resulting in glycan structures of approximately 15 repeating units (Cobb et al., 2004). Furthermore, the formation of abscesses was completely absent in iNOS^{-/-} mice challenged with PSA or live *B. fragilis*, indicating that the iNOS-dependent degradation of PSA is a necessary event for presentation of PSA to T cells.

The mechanism of PSA internalization by DCs in order to enter the endo-lysosomal pathway for MHC class II presentation has remained unresolved until now. We suspected that since the internalization of PSA is a receptor-mediated event (Cobb et al., 2004) and DCs are equipped with a wide variety of antigen-uptake receptors including several C-type lectin receptors (CLRs) with specificity for glycans, CLRs would be likely candidates for the endocytosis of PSA. In addition, CLRs can target their ligands to the endo-lysosomal pathway for MHC class II presentation (van Vliet et al., 2008) and could therefore route PSA to MHC class

II loading compartments. We here show that PSA is a ligand for the CLR dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin (DC-SIGN) and that blocking DC-SIGN on DCs resulted in inhibition of PSA-specific T cell proliferation.

MATERIALS AND METHODS

Fc CHIMERIC PROTEINS AND ANTIBODIES

DC-SIGN-Fc is composed of the extracellular domains of DC-SIGN (amino acid residues 64–404) fused to the Fc domain of human IgG1 (Geijtenbeek et al., 2002). DC-SIGN-Fc was produced in CHO cells and purified with a Hi Trap Protein A HP column (GE Healthcare). The following antibodies were used: α -DC-SIGN [clone AZN-D1 (Geijtenbeek et al., 2000b)], α -dendritic cell immunoreceptor [DCIR, clone 111F8.04 (Dendritics)], and α -mannose receptor [MR, clone 19.2 (BD Bioscience)]. Stainings were performed using 5 μ g/ml of primary antibody and 5 μ g/ml of AlexaFluor 488 F(ab')₂ fragment of goat anti-mouse IgG (Invitrogen) as secondary antibody.

PSA AND Sp1

PSA was purified from *B. fragilis* as previously described (Cobb et al., 2004). Labeling was performed by oxidation of 20% of the galactofuranose side chain sugar. AlexaFluor 488 (AF488) was added via the aldehyde group (Cobb et al., 2004). Sp1 was prepared as described previously (Tzianabos et al., 2000b).

CELLS

Raji and Raji-DC-SIGN cells (Geijtenbeek et al., 2000a) were cultured in RPMI1640 (Invitrogen) supplemented with 10% FBS (BioWhittaker), 1000 U/ml penicillin/streptomycin (Lonza), and 2 mM glutamine (Lonza). Monocytes were isolated from PBMCs from buffy coats of healthy donors (Sanquin) by a lymphoprep gradient (Axis-Shield) and a subsequent percoll gradient centrifugation (Amersham). DCs were generated by culturing purified monocytes in RPMI1640 supplemented with 10% FBS, 1000 U/ml penicillin/streptomycin, and 2 mM glutamine in combination with IL-4 (262.5 U/ml; Biosource) and GM-CSF (112.5 U/ml; Biosource) for 4–7 days. CD4⁺ T cells were isolated from the PBL fraction of buffy coats of healthy donors via negative selection using MACS beads (Milteny Biotec) according to the manufacturer's instructions.

BINDING/INTERNALIZATION ASSAYS

PSA-AF488 was incubated with DCs for 3 h at 37°C at the indicated concentrations in PBS containing calcium. For blocking experiments, 30 μ g/ml PSA-AF488 was incubated with DCs for 45 min at 37°C in the presence of blocking agents and monitored by flow cytometry (FACScan, BD Biosciences). All flow cytometric analysis was performed with Flowjo software (Tree star). DC-SIGN expressing Raji cells were incubated with the indicated concentrations of PSA-AF488 for 3 h at 37°C. Unbound PSA was washed away and the amount of bound/internalized PSA was measured by flow cytometry. CLRs were blocked with a final concentration of 20 μ g/ml of CLR-specific blocking antibodies, 50 mM monosaccharides [α -N-acetyl-D-glucosamine, α -L-Fucose, D-galactose, D-glucose, N-acetyl-D-glucosamine (Sigma Aldrich)], 25 μ g/ml mannan (Sigma Aldrich), 25 μ g/ml Laminarin (Sigma Aldrich),

25 μ g/ml mannose-BSA (Sigma Aldrich), or 10 mM EGTA for 30 min at 37°C prior to the PSA binding assay.

INTERNALIZATION ASSAY

PSA-AF488 was incubated with DCs for 3 h at 37°C at the indicated concentrations in PBS buffer containing calcium. As a positive and negative control, cells were incubated with a targeting antibody against DC-SIGN (AZN-D1) for 3 h at either 37 or 4°C respectively. Cells were washed in ice-cold PBS, fixated in 4% paraformaldehyde (Electron Microscopy Sciences), and measured by imaging flow cytometry (AMNIS Inc.) as previously described (García-Vallejo et al., 2012). Cells that have internalized antigen typically have positive internalization scores while cells that show the antigen still on the membrane have negative scores. Cells with scores around 0 have similar amounts of antigen on the membrane and in intracellular compartments.

DC-SIGN-Fc BINDING ASSAY

Ninety six-well flat-bottomed ELISA plates (Maxisorp, Nunc) were coated with 10 μ g/ml or indicated concentrations of unlabeled PSA or Sp1 in PBS or Lewis^X-polyacrylamide conjugate (PAA, Lectinity) in coating buffer (50 mM Na₂CO₃, pH 9.7) overnight at room temperature. DC-SIGN-Fc binding was measured as previously described (García-Vallejo et al., 2012).

T CELL PROLIFERATION ASSAY

Irradiated (3500 rad) DCs were pre-incubated in the presence or absence of 20 μ g/ml AZN-D1 for 30 min at 37°C. PSA was added at a concentration of 30 μ g/ml and incubated for 3 h at 37°C. Unbound PSA and AZN-D1 were washed away and DCs were incubated with autologous CD4⁺ T cells for 6 days at a ratio of 1:10 in combination with 100 μ M of the NO substrate glyco-SNAP-2 (Calbiochem). PSA binding and uptake and the blocking effect of α -DC-SIGN were monitored by simultaneously incubation of DCs with PSA-AF488 and measured by flow cytometry. Proliferation was measured using [³H]-thymidine incorporation (1 μ Ci/well; Amersham Bioscience) for 16 h. Cells were harvested onto filters and the [³H]-thymidine incorporation was measured with the use of a beta counter (Perkin Elmer).

STATISTICAL ANALYSIS

P values were calculated with a Student's *t*-test. *P* values < 0.05 were considered to be significant.

RESULTS

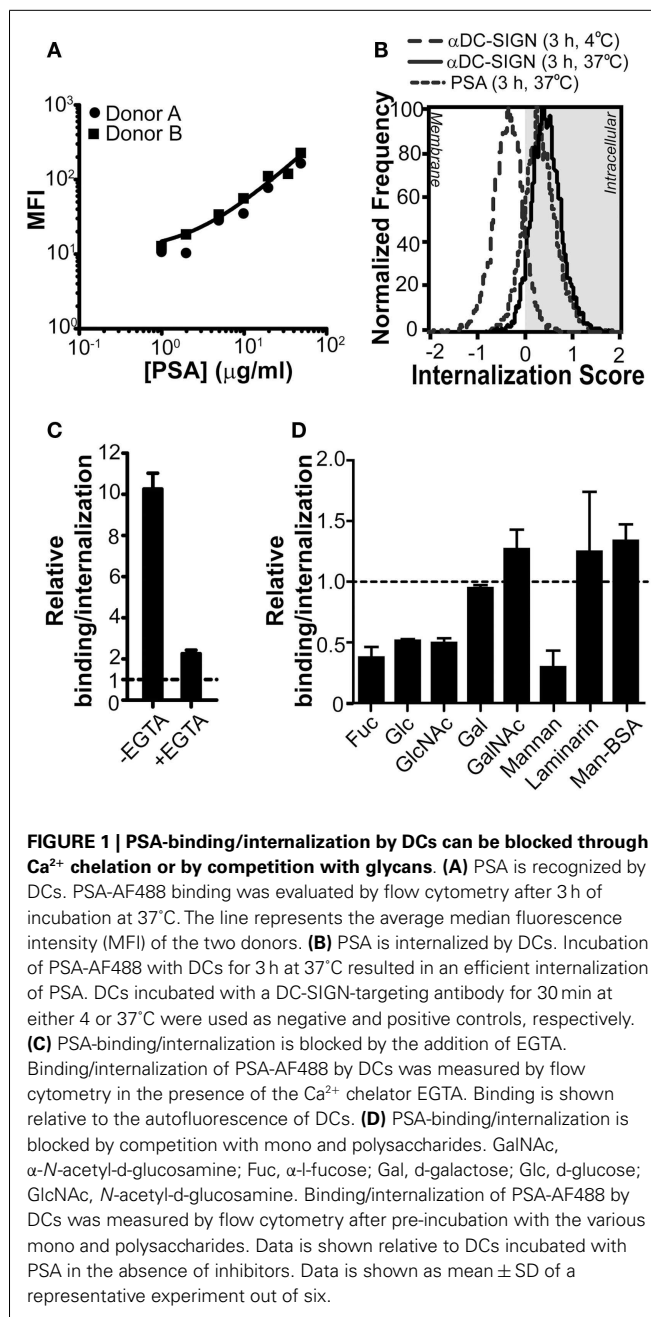
THE INTERACTION OF PSA WITH DCs IS CARBOHYDRATE-DEPENDENT

Although the unique feature of PSA to activate specific CD4⁺ T cell responses in humans has long been demonstrated (Tzianabos et al., 2000b), the molecular mechanisms leading to recognition and internalization, a crucial step in the processing and loading of PSA onto MHC class II, are still unclear. Since PSA is a complex carbohydrate and CLRs on DCs are involved in the recognition of glycans, CLRs appear to be the prime candidates for the recognition of PSA by APCs. In addition, several CLRs are antigen-uptake receptors for presentation on MHC class II or MHC class I (Sancho and Reis e Sousa, 2012). Therefore, we first analyzed whether DCs, known to express a wide variety of

CLRs, were able to bind PSA. Our results show that human DCs efficiently bound PSA in a dose-dependent manner (**Figure 1A**). Although DCs exposed to PSA for a prolonged period of time at 37°C are expected to internalize any bound material, we could not deduce this from classical flow cytometry data. We therefore measured PSA-incubated DCs using imaging flow cytometry. As a positive and negative control we treated DCs with a monoclonal antibody against DC-SIGN, known to induce DC-SIGN internalization (Engering et al., 2002a), at 37 or 4°C, respectively. Triggering of DC-SIGN induced internalization only at 37°C. The signal corresponding to PSA perfectly overlapped with that of the positive control, indicating that PSA was efficiently internalized under these conditions (**Figure 1B**). To determine whether CLRs play a role in the binding/internalization of PSA we analyzed binding/internalization by flow cytometry in the presence of EGTA, a Ca²⁺ chelator. Results indicate that PSA binding/internalization by DCs was indeed Ca²⁺-dependent, a typical characteristic of CLRs (**Figure 1C**). In order to elucidate the identity of the CLR(s) involved in the recognition of PSA, we incubated DCs with PSA in the presence of different mono and polysaccharides as competitive inhibitors for CLRs. Binding/internalization of PSA could be blocked in the presence of fucose and mannan (**Figure 1D**), in concentrations previously shown to block DC-SIGN-dependent ligand binding (Curtis et al., 1992). In addition, high concentrations of glucose and *N*-acetyl-*D*-glucosamine (GlcNAc) could also block binding/internalization of PSA, although to a lesser extent (Curtis et al., 1992; Lee et al., 2011). Altogether these results indicate the involvement of a mannose/fucose-specific CLR, such as DC-SIGN or MR. Other mono and polysaccharides, like laminarin (a dectin-1-specific glycan) and GalNAc (a macrophage galactose-type lectin-specific sugar) did not alter PSA binding/internalization (**Figure 1D**). Also, Mannose-BSA (a MR-specific ligand) did not inhibit PSA binding/internalization (**Figure 1D**), suggesting that MR is not involved in the recognition of PSA by DCs. Therefore, we concluded that the CLR DC-SIGN is a likely PSA receptor on DCs.

PSA INTERACTS WITH DC-SIGN

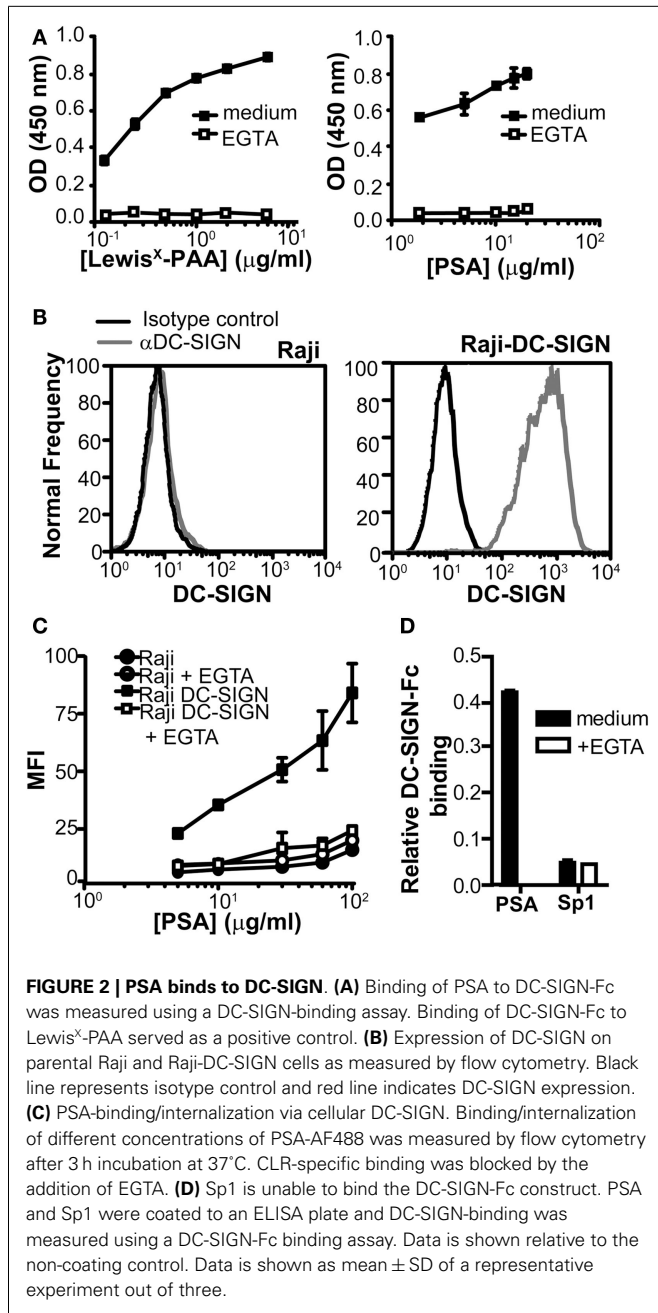
To examine whether DC-SIGN directly binds to PSA, we used a DC-SIGN-Fc chimeric protein consisting of the extracellular domains, including the carbohydrate recognition domain of DC-SIGN fused to the human IgG1 Fc tail. We compared DC-SIGN-binding to PSA to the well-known DC-SIGN ligand Lewis^X. Concentration-dependent binding of DC-SIGN-Fc to both PSA and Lewis^X was observed and the interaction appeared to be Ca²⁺-dependent, since it could be blocked by the addition of EGTA (**Figure 2A**). To confirm the binding of PSA at the cellular level, we used a Raji cell line transduced with DC-SIGN (Geijtenbeek et al., 2000a). Raji-DC-SIGN cells expressed high levels of DC-SIGN, which was absent on the parental Raji cells (**Figure 2B**). PSA bound the Raji-DC-SIGN cells in a concentration-dependent manner, which could be blocked by EGTA to the level of the parental Raji cells (**Figure 2C**). Several ZPSs have similar functional characteristics as PSA, including the induction of CD4⁺ T cell proliferation and the development of peritoneal abscesses (Velez et al., 2009). One of these ZPSs is Sp1, the capsular polysaccharide from type



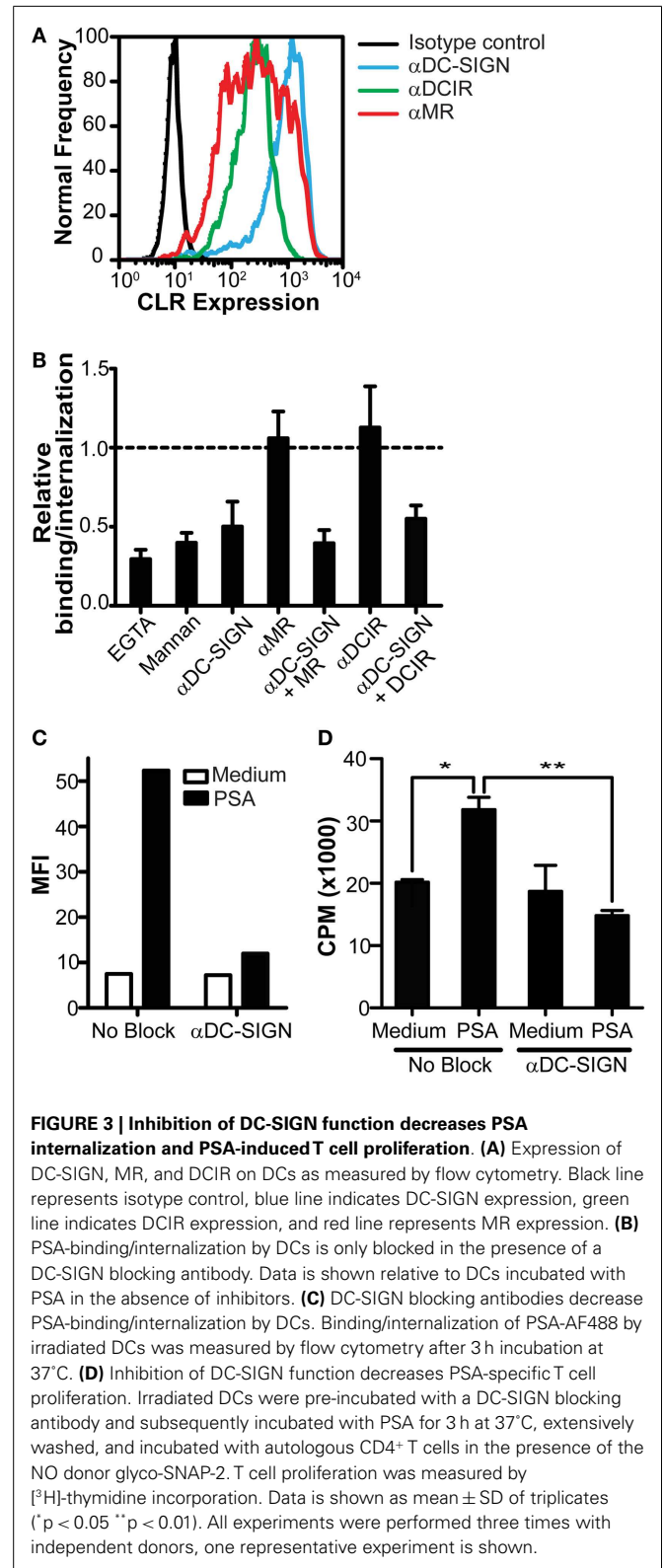
1 *Streptococcus pneumoniae*. Although the biological properties of Sp1 mimic those of PSA, Sp1 was not recognized by DC-SIGN (**Figure 2D**), suggesting that DC-SIGN is a PSA-specific receptor and not a general binding partner for all ZPSs.

BLOCKING DC-SIGN FUNCTION INHIBITS PSA-INDUCED T CELL PROLIFERATION

Since we identified DC-SIGN as a specific PSA receptor, we continued our research to determine the contribution of DC-SIGN in the presentation of PSA by DCs to CD4⁺ T cells. DCs express different CLRs, such as DCIR and MR, that share overlapping glycan specificity with DC-SIGN (**Figure 3A**). To rule out a possible



involvement of these CLRs in the recognition of PSA, we tested the ability of different CLR blocking antibodies to inhibit PSA binding. Based on titration of the DC-SIGN antibody in previously published experiments (Geijtenbeek et al., 2000b), a concentration of 20 μg/ml blocking antibody was used. We observed that only the DC-SIGN blocking antibody decreased binding/internalization significantly, which was not further enhanced in combination with anti-MR or DCIR blocking antibodies (Figure 3B). These data confirmed DC-SIGN as the main receptor on DCs for the recognition of PSA. DC-SIGN functions as internalization receptor that mediates antigen routing to MHC class II loading compartments (Engering et al., 2002a). To assess the contribution of DC-SIGN in the MHC class II presentation of PSA to T cells, DCs



were pulsed with PSA in the presence or absence of DC-SIGN blocking antibodies for 3 h, after which unbound PSA was washed away. PSA binding/uptake by DCs was clearly decreased in the

presence of α -DC-SIGN (Figure 3C). The observed increase in CD4⁺ T cell proliferation by PSA-incubated DCs confirms the presence of PSA-specific T cells in blood (Figure 3D). Furthermore, PSA-specific T cell proliferation was completely inhibited in the presence of blocking antibodies to DC-SIGN (Figure 3D); demonstrating a DC-SIGN-dependent role in the uptake and processing of PSA for MHC class II presentation and subsequent T cell activation.

In conclusion, we have shown that DC-SIGN plays a crucial role in the binding and internalization of the T cell activating ZPSs PSA by human DCs. In addition, inhibiting DC-SIGN function decreases MHC class II presentation and subsequent T cell activation.

DISCUSSION

PSA is one of the ZPSs that can be presented in MHC class II for recognition by CD4⁺ T cells (Tzianabos et al., 2000b). Although internalization and NO-dependent processing of PSA is required for optimal presentation in MHC class II (Cobb et al., 2004; Duan et al., 2008), the identity of the receptor mediating binding and internalization of PSA has remained elusive. DC express several CLRs, specialized in glycan recognition and routing of internalized ligands to MHC class II or MHC class I loading compartments (Sancho and Reis e Sousa, 2012). We therefore hypothesized that a CLR might be involved and in this study we identified DC-SIGN as crucial receptor for PSA internalization and MHC-II presentation by human DCs.

Zwitterionic polysaccharides are generally composed of repeating oligosaccharide units. These oligosaccharide units are different for PSA and Sp1, explaining why PSA interacted with the purified DC-SIGN-Fc molecule, while the binding of Sp1 to DC-SIGN-Fc could not be detected. The oligosaccharide structure in PSA is [\rightarrow 3)- α -D-AAT Galp-(1 \rightarrow 4)-[β -D-Galf-(1 \rightarrow 3)] α -D-GalpNAc-(1 \rightarrow 3)-[4,6-pyruvate]- β -D-Galp-(1 \rightarrow) (Tzianabos et al., 1992), while Sp1 is composed of the following oligosaccharide structure: [\rightarrow 3)- α -2,4-dideoxy-4-amino-D-FucNAc-(1 \rightarrow 4)- α -D-GalAp-(1 \rightarrow 3)- α -D-GalAp-(1 \rightarrow) (Velez et al., 2009). DC-SIGN has specificity for mannose- and fucose-containing glycan structures, and can therefore interact with a wide variety of pathogens (Appelmelk et al., 2003). However, both fucose and mannose glycans are not present in the PSA oligosaccharide. Since the β -D-Galf glycan forms a side group in PSA, it is the most easily accessible carbohydrate for the binding to DC-SIGN. Sp1 from *S. pneumoniae*, lacks this side group, potentially explaining the inability of DC-SIGN to bind to Sp1. It goes beyond the scope of this study to determine the exact glycan moiety in PSA that interacts with DC-SIGN. Together, our results indicate that DC-SIGN is a PSA-specific receptor and not a general binding partner for all ZPSs.

Next to the DC-SIGN-mediated PSA binding at the molecular level, we show an enhanced binding of PSA to DC-SIGN expressing Raji cells over the parental cell line. Raji cells are frequently used as model for PSA internalization and presentation in MHC class II (Kalka-Moll et al., 2002; Cobb et al., 2004; Cobb and Kasper, 2008; Duan et al., 2008; Ryan et al., 2011). In our assays PSA internalization was also detectable in the parental cell line,

however this could only be observed when cells were incubated with high concentrations of PSA. DC-SIGN expression in Raji cells clearly increased PSA uptake and blocking DC-SIGN function on DCs significantly reduced PSA internalization and MHC-II presentation, while inhibition of other CLRs with overlapping glycan specificity, such as MR and DCIR, had no effect. These data indicate a crucial role of DC-SIGN in the binding and internalization of PSA by DCs.

DC-SIGN-binding ligands are quickly internalized and routed to endosomes and lysosomes (Engering et al., 2002a). Although the presence of DC-SIGN-binding glycans on protein antigens is necessary for the DC-SIGN-mediated routing to MHC class II loading compartments, it is a protein-derived peptide that is presented in MHC class II (Aarnoudse et al., 2008). Therefore, PSA is the first glycan structure internalized in a DC-SIGN-dependent manner that is presented solely onto MHC class II in the absence of a protein.

The presentation of PSA in MHC class II has previously been shown by the activation of human CD4⁺ T cell responses *in vitro* using PBMCs as APCs (Tzianabos et al., 2000b; Velez et al., 2009). Capture of PSA in this setting could be mediated by the small DC-SIGN⁺ subpopulation present in PBMCs (Engering et al., 2002b) or by the non-specific uptake typically observed in long term *in vitro* cell cultures using high PSA doses. DCs, on the other hand, are more likely candidates for the encounter of PSA under physiological conditions. We here demonstrate the ability of DCs to present DC-SIGN-internalized PSA in MHC class II, resulting in the activation of autologous CD4⁺ T cell responses. The induction of PSA-specific T cell proliferation could already be induced after short incubation times using relatively low concentrations of PSA, more closely mimicking the physiological setting. Activation of PSA-specific CD4⁺ T cells requires not only internalization of PSA, but also NO-dependent PSA processing and loading onto MHC class II (Cobb et al., 2004). Therefore, PSA loaded DCs were co-cultured with autologous CD4⁺ T cells in the presence of a NO donor (glyco-SNAP-2), to ensure the availability of sufficient NO to cleave the PSA. The use of glyco-SNAP-2 was necessary, since compared to DCs present *in vivo*, *in vitro* cultured monocyte-derived DCs hardly produce NO (Nishioka et al., 2003).

Besides being an internalization receptor, DC-SIGN has also been described to modify TLR signaling. Geijtenbeek et al. (2003) showed that binding of the mannosylated ligand ManLAM to DC-SIGN results in an increased IL-10 secretion by LPS-stimulated DCs. Surprisingly, the effect of DC-SIGN stimulation on TLR-induced cytokine responses is dependent on the interacting glycan. Whereas both mannose- and fucose-expressing ligands increase the IL-10 secretion by LPS-stimulated DCs, the effect on IL-12 and IL-6 secretion is different, whereby fucose-expressing ligands decrease the LPS-induced IL-12 and IL-6 production, while mannose-expressing ligands increase the LPS-induced IL-12 and IL-6 secretion (Gringhuis et al., 2009). The effects of PSA-mediated DC-SIGN stimulation still needs to be further investigated.

PSA-induced T cell proliferation has been reported in mice as well (Tzianabos and Kasper, 2002). In the murine genome

eight different homologs of DC-SIGN are present (SIGNR1-8). The internalization capacity of these lectins has been investigated only for SIGNR1, SIGNR3, and SIGNR5, whereby SIGNR1 and SIGNR3 were reported to function as antigen-uptake receptors, whereas this was not the case for SIGNR5 (Takahara et al., 2004; Powlesland et al., 2006). SIGNR1 has been identified on lamina propria DCs in the murine intestine, where it mediates the induction of oral tolerance through the generation of Tr1 cells (Zhou et al., 2010). PSA derived from *B. fragilis*, a commensal bacterium in the gut, has also been described to induce regulatory T cells (Round and Mazmanian, 2010). Preliminary results from our laboratory indicate SIGNR1 as PSA-binding receptor (data not shown); however we cannot exclude the involvement of other receptors in the binding and internalization of PSA in mice *in vivo*.

In conclusion, we have shown that DC-SIGN plays a crucial role in the binding and internalization of the T cell activating

ZPS PSA by human DCs. In addition, inhibiting DC-SIGN function decreases MHC class II presentation and subsequent T cell activation. These data indicate that DC-SIGN behaves as an antigen-uptake receptor for PSA, resulting in loading onto MHC class II molecules and the activation of PSA-specific CD4⁺ T cell responses.

ACKNOWLEDGMENTS

We would like to thank W. W. Unger and S. C. Bruijns for helpful suggestions in setting up the T cell proliferation assays. Karien Bloem was supported by the Dutch Top Institute Pharma, project T1-214, Juan J. García-Vallejo was supported by the Dutch Asthma Foundation (3.210.040), Ilona M. Vuist was supported by MS Research and Sandra J. van Vliet was supported by a VENI grant (863.10.017) from the Netherlands Organization for Scientific Research (NWO).

REFERENCES

- Aarnoudse, C. A., Bax, M., Sánchez-Hernández, M., García-Vallejo, J. J., and van Kooyk, Y. (2008). Glycan modification of the tumor antigen gp100 targets DC-SIGN to enhance dendritic cell induced antigen presentation to T cells. *Int. J. Cancer* 122, 839–846.
- Appelmelk, B. J., van Die, I., van Vliet, S. J., Vandenbroucke-Grauls, C. M., Geijtenbeek, T. B. H., and van Kooyk, Y. (2003). Cutting edge: carbohydrate profiling identifies new pathogens that interact with dendritic cell-specific ICAM-3-grabbing nonintegrin on dendritic cells. *J. Immunol.* 170, 1635–1639.
- Astronomo, R. D., and Burton, D. R. (2010). Carbohydrate vaccines: developing sweet solutions to sticky situations? *Nat. Rev. Drug Discov.* 9, 308–324.
- Choi, Y., Roehrl, M. H., Kasper, D. L., and Wang, J. Y. (2002). A unique structural pattern shared by T-cell-activating and abscess-regulating zwitterionic polysaccharides. *Biochemistry* 41, 15144–15151.
- Cobb, B. A., and Kasper, D. L. (2008). Characteristics of carbohydrate antigen binding to the presentation protein HLA-DR. *Glycobiology* 18, 707–718.
- Cobb, B. A., Wang, Q., Tzianabos, A. O., and Kasper, D. L. (2004). Polysaccharide processing and presentation by the MHCII pathway. *Cell* 117, 677–687.
- Curtis, B. M., Scharnowske, S., and Watson, A. J. (1992). Sequence and expression of a membrane-associated C-type lectin that exhibits CD4-independent binding of human immunodeficiency virus envelope glycoprotein gp120. *Proc. Natl. Acad. Sci. U.S.A.* 89, 8356–8360.
- Duan, J., Avci, F. Y., and Kasper, D. L. (2008). Microbial carbohydrate depolymerization by antigen-presenting cells: deamination prior to presentation by the MHCII pathway. *Proc. Natl. Acad. Sci. U.S.A.* 105, 5183–5188.
- Engering, A., Geijtenbeek, T. B. H., van Vliet, S. J., Wijers, M., van Liempt, E., Demaurex, N., et al. (2002a). The dendritic cell-specific adhesion receptor DC-SIGN internalizes antigen for presentation to T cells. *J. Immunol.* 168, 2118–2126.
- Engering, A., van Vliet, S. J., Geijtenbeek, T. B. H., and van Kooyk, Y. (2002b). Subset of DC-SIGN(+) dendritic cells in human blood transmits HIV-1 to T lymphocytes. *Blood* 100, 1780–1786.
- García-Vallejo, J. J., Ambrosini, M., Overbeek, A., van Riel, H., Bloem, K., Unger, W. W. J., et al. (2012). Multivalent glycopeptide dendrimers for the targeted delivery of antigens to dendritic cells. *Mol. Immunol.* 53, 387–397.
- Geijtenbeek, T. B., Kwon, D. S., Torensma, R., van Vliet, S. J., van Duijnhoven, G. C., Middel, J., et al. (2000a). DC-SIGN, a dendritic cell-specific HIV-1-binding protein that enhances trans-infection of T cells. *Cell* 100, 587–597.
- Geijtenbeek, T. B., Torensma, R., van Vliet, S. J., van Duijnhoven, G. C., Adema, G. J., van Kooyk, Y., et al. (2000b). Identification of DC-SIGN, a novel dendritic cell-specific ICAM-3 receptor that supports primary immune responses. *Cell* 100, 575–585.
- Geijtenbeek, T. B. H., van Duijnhoven, G. C. F., van Vliet, S. J., Krieger, E., Vriend, G., Figdor, C. G., et al. (2002). Identification of different binding sites in the dendritic cell-specific receptor DC-SIGN for intercellular adhesion molecule 3 and HIV-1. *J. Biol. Chem.* 277, 11314–11320.
- Geijtenbeek, T. B. H., van Vliet, S. J., Koppel, E. A., Sanchez-Hernandez, M., Vandenbroucke-Grauls, C. M. J. E., Appelmelk, B., et al. (2003). Mycobacteria target DC-SIGN to suppress dendritic cell function. *J. Exp. Med.* 197, 7–17.
- Gringhuis, S. I., den Dunnen, J., Litjens, M., van der Vlist, M., and Geijtenbeek, T. B. H. (2009). Carbohydrate-specific signaling through the DC-SIGN signalosome tailors immunity to *Mycobacterium tuberculosis*, HIV-1 and *Helicobacter pylori*. *Nat. Immunol.* 10, 1081–1088.
- Kalka-Moll, W. M., Tzianabos, A. O., Bryant, P. W., Niemeyer, M., Ploegh, H. L., and Kasper, D. L. (2002). Zwitterionic polysaccharides stimulate T cells by MHC class II-dependent interactions. *J. Immunol.* 169, 6149–6153.
- Lee, R. T., Hsu, T.-L., Huang, S. K., Hsieh, S.-L., Wong, C.-H., and Lee, Y. C. (2011). Survey of immune-related, mannose/fucose-binding C-type lectin receptors reveals widely divergent sugar-binding specificities. *Glycobiology* 21, 512–520.
- Michaëlsson, E., Malmström, V., Reis, S., Engström, A., Burkhardt, I. H., and Holmdahl, R. (1994). T cell recognition of carbohydrates on type II collagen. *J. Exp. Med.* 180, 745–749.
- Nishioka, Y., Wen, H., Mitani, K., Robbins, P. D., and Lotze, M. T. (2003). Differential effects of IL-12 on the generation of alloreactive CTL mediated by murine and human dendritic cells: a critical role for nitric oxide. *J. Leukoc. Biol.* 73, 621–629.
- Powlesland, A. S., Ward, E. M., Sadhu, S. K., Guo, Y., Taylor, M. E., and Drickamer, K. (2006). Widely divergent biochemical properties of the complete set of mouse DC-SIGN-related proteins. *J. Biol. Chem.* 281, 20440–20449.
- Round, J. L., and Mazmanian, S. K. (2010). Inducible Foxp3 + regulatory T-cell development by a commensal bacterium of the intestinal microbiota. *Proc. Natl. Acad. Sci. U.S.A.* 107, 12204–12209.
- Ryan, S. O., Bonomo, J. A., Zhao, F., and Cobb, B. A. (2011). MHCII glycosylation modulates *Bacteroides fragilis* carbohydrate antigen presentation. *J. Exp. Med.* 208, 1041–1053.
- Sancho, D., and Reis e Sousa, C. (2012). Signaling by myeloid C-type lectin receptors in immunity and homeostasis. *Annu. Rev. Immunol.* 30, 491–529.
- Takahara, K., Yashima, Y., Omatsu, Y., Yoshida, H., Kimura, Y., Kang, Y.-S., et al. (2004). Functional comparison of the mouse DC-SIGN, SIGNR1, SIGNR3 and Langerin, C-type lectins. *Int. Immunol.* 16, 819–829.
- Tzianabos, A. O., Chandraker, A., Kalka-Moll, W., Stingle, E., Dong, V. M., Finberg, R. W., et al. (2000a). Bacterial pathogens induce abscess formation by CD4(+) T-cell activation via the CD28-B7-2 costimulatory pathway. *Infect. Immun.* 68, 6650–6655.
- Tzianabos, A. O., Finberg, R. W., Wang, Y., Chan, M., Onderdonk, A. B., Jennings, H. J., et al. (2000b). T cells activated by zwitterionic molecules prevent abscesses induced by

- pathogenic bacteria. *J. Biol. Chem.* 275, 6733–6740.
- Tzianabos, A. O., and Kasper, D. L. (2002). Role of T cells in abscess formation. *Curr. Opin. Microbiol.* 5, 92–96.
- Tzianabos, A. O., Pantostit, A., Baumann, H., Brisson, J.-R., Jennings, H. J., and Kasper, D. L. (1992). The capsular polysaccharide of *Bacteroides fragilis* comprises two ionically linked polysaccharides. *J. Biol. Chem.* 267, 18230–18235.
- van Vliet, S. J., García-Vallejo, J. J., and van Kooyk, Y. (2008). Dendritic cells and C-type lectin receptors: coupling innate to adaptive immune responses. *Immunol. Cell Biol.* 86, 580–587.
- Velez, C. D., Lewis, C. J., Kasper, D. L., and Cobb, B. A. (2009). Type I *Streptococcus pneumoniae* carbohydrate utilizes a nitric oxide and MHC II-dependent pathway for antigen presentation. *Immunology* 127, 73–82.
- Vlad, A. M., Muller, S., Cudic, M., Paulsen, H., Otvos, L., Hanisch, F.-G., et al. (2002). Complex carbohydrates are not removed during processing of glycoproteins by dendritic cells: processing of tumor antigen MUC1 glycopeptides for presentation to major histocompatibility complex class II-restricted T cells. *J. Exp. Med.* 196, 1435–1446.
- Zhou, Y., Kawasaki, H., Hsu, S.-C., Lee, R. T., Yao, X., Plunkett, B., et al. (2010). Oral tolerance to food-induced systemic anaphylaxis mediated by the C-type lectin SIGNR1. *Nat. Med.* 16, 1128–1133.
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Received: 04 March 2013; paper pending published: 02 April 2013; accepted: 21 April 2013; published online: 06 May 2013.
- Citation: Bloem K, García-Vallejo JJ, Vuist IM, Cobb BA, van Vliet SJ and van Kooyk Y (2013) Interaction of the capsular polysaccharide A from *Bacteroides fragilis* with DC-SIGN on human dendritic cells is necessary for its processing and presentation to T cells. *Front. Immunol.* 4:103. doi: 10.3389/fimmu.2013.00103
- This article was submitted to *Frontiers in Molecular Innate Immunity*, a specialty of *Frontiers in Immunology*. Copyright © 2013 Bloem, García-Vallejo, Vuist, Cobb, van Vliet and van Kooyk. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.