



Functional Properties of Microorganisms in Fermented Foods

Jyoti P. Tamang^{1*}, Dong-Hwa Shin^{2,3}, Su-Jin Jung³ and Soo-Wan Chae^{3,4}

¹ Department of Microbiology, School of Life Sciences, Sikkim University, Gangtok, India, ² Shindonghwa Food Research Institute, Jeonju, South Korea, ³ Clinical Trial Center for Functional Foods, Chonbuk National University Hospital, Jeonju, South Korea, ⁴ Division of Pharmacology, Chonbuk National University Medical School, Jeonju, South Korea

Fermented foods have unique functional properties imparting some health benefits to consumers due to presence of functional microorganisms, which possess probiotics properties, antimicrobial, antioxidant, peptide production, etc. Health benefits of some global fermented foods are synthesis of nutrients, prevention of cardiovascular disease, prevention of cancer, gastrointestinal disorders, allergic reactions, diabetes, among others. The present paper is aimed to review the information on some functional properties of the microorganisms associated with fermented foods and beverages, and their health-promoting benefits to consumers.

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*Correspondence:

Jyoti P. Tamang
jyoti_tamang@hotmail.com

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INTRODUCTION

Existing scientific data show many fermented foods have both nutritive and non-nutritive components in foods, which have the potential to modulate specific target functions in the body relevant to well-being and health of the consumers. However, 90% of naturally fermented foods and alcoholic beverages in different countries and regions of the world are still at home production under traditional conditions. Naturally fermented foods and beverages contain both functional and non-functional microorganisms (Tamang et al., 2016). Functional microorganisms transform the chemical constituents of raw materials of plant/animal sources during food fermentation thereby enhancing the bio-availability of nutrients, enriching sensory quality of the food, imparting bio-preservative effects and improvement of food safety, degrading toxic components and anti-nutritive factors, producing antioxidant and antimicrobial compounds, stimulating the probiotic functions, and fortifying with some health-promoting bioactive compounds (Tamang et al., 2009, 2016; Farhad et al., 2010; Bourdichon et al., 2012; Thapa and Tamang, 2015). Among bacteria associated with fermented foods and alcoholic beverages, lactic acid bacteria (LAB) mostly species of *Enterococcus*, *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Pediococcus*, *Weissella*, etc. are widely present in many fermented foods and beverages (Axelsson et al., 2012; Holzapfel and Wood, 2014). Species of *Bacillus* are also present in legume-based fermented foods (Kubo et al., 2011; Tamang, 2015). Species of *Bifidobacterium*, *Brachy bacterium*, *Brevibacterium*, and *Propionibacterium* are isolated from cheese, and species of *Arthrobacter* and *Hafnia* from fermented meat products (Bourdichon et al., 2012). Several genera with hundred of species of yeasts have been isolated from fermented foods, alcoholic beverages and non-food mixed amylolytic starters which mostly include *Candida*, *Debaryomyces*, *Geotrichum*, *Hansenula*, *Kluyveromyces*, *Pichia*, *Rhodotorula*, *Saccharomyces*, *Saccharomycopsis*, *Schizosaccharomyces*, *Torulopsis*, *Wickerhamomyces*, and *Zygosaccharomyces* (Tamang and Fleet, 2009; Lv et al., 2013). Species of *Actinomucor*, *Amylomyces*, *Aspergillus*, *Monascus*, *Mucor*, *Neurospora*, *Penicillium*, *Rhizopus*, and *Ustilago* are reported for many fermented foods, Asian non-food amylolytic starters, and alcoholic beverages (Chen et al., 2014).

Functional properties of microorganisms in fermented foods include probiotics properties (Hill et al., 2014), antimicrobial properties (Meira et al., 2012), antioxidant (Perna et al., 2013), peptide production (De Mejia and Dia, 2010), fibrinolytic activity (Kotb, 2012), poly-glutamic acid (Chettri and Tamang, 2014), degradation of antinutritive compounds (Babalola, 2014), etc. which may be important criteria for selection of starter culture(s) to be used in the manufacture of functional foods (Badis et al., 2004). Some genera and species of microorganisms are used as commercial starters in food fermentation (Table 1), and some of products are commercialized and marketed globally as functional foods, health foods, therapeutic foods and nutraceuticals foods (Bernardeau et al., 2006; Bourdichon et al., 2012; Thapa and Tamang, 2015). The present paper is aimed to review the information on some functional properties of the microorganisms associated with fermented foods and beverages, and their health-promoting benefits to consumers.

Probiotic Microorganisms

Probiotics are defined as live microorganisms that, when administered in adequate amounts, confer a health benefit on the host (Hill et al., 2014). Probiotic organisms used in foods must have the ability to resist gastric juices, exposure to bile, and be able to proliferate and colonize the digestive tract (Saad et al., 2013). The beneficial effects of probiotic foods on human health and nutrition are constantly increasing (de LeBlanc et al., 2007; Monteagudo-Mera et al., 2012), and probiotics are popularly using bio-ingredients in many functional fermented foods (Chávarri et al., 2010). The most commonly used probiotic bacteria belong to the heterogeneous group of LAB (*Lactobacillus*, *Enterococcus*) and to the genus *Bifidobacterium*, however, yeasts and other microbes have also been developed as potential probiotics during recent years (Ouweland et al., 2002). Some popular commercial probiotic cultures which are available in global markets include *Bacillus coagulans* BC30 marketed by Ganeden Biotech, Inc., Cleveland, OH, USA; *Lactobacillus acidophilus* NCFM, *Lactobacillus rhamnosus* HN001 (DR20) and *Bifidobacterium lactis* HN019 (DR10) marketed by Danisco (Madison, WI, USA), *L. casei* strain Shirota and *B. breve* strain Yakult marketed by Yakult (Tokyo, Japan), *L. fermentum* VRI003 (PCC) marketed by Probiomics (Eveleigh, NSW, Australia), *L. rhamnosus* R0011 marketed by Institut Rosell (Montreal, QC, Canada), *Streptococcus oralis* KJ3 marketed by Orogenics, Inc. (Alachua, FL, USA), and *Saccharomyces cerevisiae* (boulardii) marketed by Biocodex (Creswell, OR, USA; US Probiotics Home, 2011).

Products containing probiotic bacteria generally include foods and supplements (Varankovich et al., 2015). Fermented milk products are the most traditional source of probiotic strains of lactobacilli (Bernardeau et al., 2006; Shah, 2015); however, commercial probiotic lactobacilli have also been added to meat products, snacks, fruit juice, etc. (Ranadheera et al., 2010). Probiotic properties of *Lactobacillus plantarum* isolated from *kimchi*, Korean fermented vegetable product, has been reported (Ji et al., 2013), and is also found to prevent the growth of *Helicobacter pylori* (Lim and Im, 2009). Probiotic strain *L. acidophilus* La-5 produces conjugated linoleic acid (CLA),

an anti-carcinogenic agent (Macouzet et al., 2009). *Pediococcus pentosaceus* CIAL-86 isolated from wine shows anti-adhesion activity against *Escherichia coli* CIAL-153, indicating its probiotic potential in wine (García-Ruiz et al., 2014).

Antimicrobial Properties

Many species of LAB isolated from fermented vegetable and milk products have antimicrobial activities due to production of antimicrobial compounds such as bacteriocin and nisin (Tamang et al., 2009; Khan et al., 2010; Gaggia et al., 2011; Jiang et al., 2012; Grosu-Tudor and Zamfir, 2013). Many strains of LAB isolated from *kimchi* produce antimicrobial compounds such as bacteriocin by *L. lactis* BH5 (Hur et al., 2000) and *L. citreum* GJ7 (Chang et al., 2008), and pediocin by *P. pentosaceus* (Shin et al., 2008). Species of LAB isolated from *kimchi* show strong antimicrobial activity against *Listeria monocytogenes*, *Staphylococcus aureus*, *E. coli*, and *Salmonella typhimurium* (Lee et al., 2009). *Weissella cibaria* isolated from fermented cabbage product shows antimicrobial activity against Gram-positive and Gram-negative pathogens (Patel et al., 2014). *Lactococcus lactis* isolated from *dahi*, Indian curd, produces nisin Z that inhibits *L. monocytogenes* and *S. aureus* (Mitra et al., 2010). Several LAB species isolated from Romanian traditional fermented fruits and vegetables have antimicrobial activity against *L. monocytogenes*, *E. coli*, *Salmonella*, and *Bacillus* (Grosu-Tudor and Zamfir, 2013). Microorganisms as protective cultures, e.g., bacteriocin producers, may have several advantages, as they can contribute to the flavor, texture and nutritional value of the product besides the production of bacteriocin (Gaggia et al., 2011).

Antioxidant Activity

Antioxidant activities in fermented foods include 1,1-diphenyl-2-picryl hydrazyl (DPPH) radical scavenging activity, 2,2'-azino-bis (3-ethylbenzo-thiazoline-6-sulfonic acid; ABTS) radical scavenging activity, total phenol content (TPC) estimation, and reducing power assay (Liu and Pan, 2010; Abubakr et al., 2012). Many Asian fermented soybean foods have antioxidant properties, e.g., *natto*, *Bacillus*-fermented soybean food of Japan (Ping et al., 2012), *chungkokjang* and *jang*, fermented soybean foods of Korea (Shon et al., 2007; Shin and Jeong, 2015), *douchi*, a fermented soybean food of China (Wang et al., 2007a), *kinema*, *Bacillus*-fermented soybean food of India and Nepal (Moktan et al., 2008; Tamang, 2015), *bekang* and *tungrymbai*, *Bacillus*-fermented soybean foods of India (Chettri and Tamang, 2014), *thua nao*, *Bacillus*-fermented soybean food of Thailand (Dajanta et al., 2013), and *tempe* mold-fermented soybean food of Indonesia (Nurrahman et al., 2013). Antioxidant activities have also been observed in *kimchi* (Park et al., 2011) and yogurt (Sabeena et al., 2010).

Peptide Production

Bioactive peptides are formed during food fermentation by proteolytic microorganisms (De Mejia and Dia, 2010). In fermented foods peptides have some functional properties such as immunomodulatory (Qian et al., 2011), antithrombic (Singh et al., 2014), and antihypertensive properties (Phelan and Kerins, 2011). Species of *Bacillus* are involved in enzymatic hydrolysis of

TABLE 1 | Some functional microorganisms used as commercial starters in food fermentation (amended and compiled from references: Mogensen et al., 2002; Bernardeau et al., 2006; Bourdichon et al., 2012; Thapa and Tamang, 2015).

Group	Genera/species	Product/application(s)
Bacteria	<i>Acetobacter aceti</i> subsp. <i>aceti</i>	Vinegar
	<i>A. pasteurianus</i> subsp. <i>pasteurianus</i>	Vinegar, cocoa
	<i>Bacillus acidopulluliticus</i>	Pullulanases (food additive)
	<i>B. coagulans</i>	Cocoa; glucose isomerase (food additive), fermented soybeans
	<i>B. licheniformis</i>	Protease (food additive)
	<i>B. subtilis</i>	Fermented soybeans, protease, glycolipids, riboflavin-B ₂ (food additive)
	<i>Bifidobacterium animalis</i> subsp. <i>lactis</i> , <i>B. breve</i>	Fermented milks with probiotic properties; common in European fermented milks
	<i>Brachybacterium alimentarium</i>	Gruyère and Beaufort cheese
	<i>Brevibacterium flavum</i>	Malic acid, glutamic acid, lysine, monosodium glutamate (food additives)
	<i>Corynebacterium ammoniagenes</i>	Cheese ripening
	<i>Enterobacter aerogenes</i>	Bread fermentation
	<i>Enterococcus durans</i>	Cheese and sourdough fermentation
	<i>E. faecium</i>	Soybean, dairy, meat, vegetables
	<i>Klebsiella pneumoniae</i> subsp. <i>ozaenae</i>	<i>Tempe</i> ; production of vitamin B ₁₂
	<i>Lactobacillus acetotolerans</i>	Ricotta cheese, vegetables
	<i>L. acidophilus</i>	Fermented milks, probiotics, vegetables
	<i>L. alimentarius</i>	Fermented sausages; ricotta; meat, fish
	<i>L. brevis</i>	Bread fermentation; wine; dairy
	<i>L. buchneri</i>	Malolactic fermentation in wine; sourdough
	<i>L. casei</i> subsp. <i>casei</i>	Dairy starter; cheese ripening; green table olives
	<i>L. delbruecki</i> subsp. <i>bulgaricus</i>	Yogurt and other fermented milks, mozzarella
	<i>L. fermentum</i>	Fermented milks, sourdough, urease (food additive)
	<i>L. ghanensis</i>	Cocoa
	<i>L. helveticus</i>	Starter for cheese; cheese ripening, vegetables
	<i>L. hilgardii</i>	Malolactic fermentation of wine
	<i>L. kefir</i>	Fermented milk (<i>kefir</i>), reduction of bitter taste in citrus juice
	<i>L. kimchii</i>	<i>Kimchi</i>
	<i>L. oeni</i>	Wine
	<i>L. paracasei</i> subsp. <i>paracasei</i>	Cheese fermentation, probiotic cheese, probiotics, wine, meat
	<i>L. pentosus</i>	Meat fermentation and biopreservation of meat; green table olives; dairy, fruits, wine
	<i>L. plantarum</i> subsp. <i>plantarum</i>	Fermentation of vegetables, malolactic fermentation, green table olives; dairy, meat
	<i>L. sakei</i> subsp. <i>sakei</i>	Fermentation of cheese and meat products; beverages
	<i>L. salivarius</i> subsp. <i>salivarius</i>	Cheese fermentation
	<i>L. sanfranciscensis</i>	Sourdough
	<i>L. vermouthensis</i>	Dry sausages
	<i>Lactococcus lactis</i> subsp. <i>lactis</i>	Dairy starter, Nisin (protective culture)
	<i>L. lactis</i> , <i>L. mesenteroides</i> subsp. <i>cremoris</i> , <i>L. mesenteroides</i> subsp. <i>dextranicum</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i>	Dairy starter
	<i>Oenococcus oeni</i>	Malolactic fermentation of wine
	<i>Pediococcus acidilactici</i>	Meat fermentation and biopreservation of meat; cheese starter
	<i>P. pentosaceus</i>	Meat fermentation and biopreservation of meat
<i>Propionibacterium acidipropionici</i>	Meat fermentation and biopreservation of meat	
<i>P. arabinosum</i>	Cheese fermentation; probiotics	

(Continued)

TABLE 1 | Continued

Group	Genera/species	Product/application(s)
Yeasts	<i>P. freudenreichii</i> subsp. <i>freudenreichii</i>	Cheese fermentation (Emmental cheese starter)
	<i>Streptococcus natalensis</i>	Natamycin (food additive)
	<i>Weissella ghanensis</i>	Cocoa
	<i>Zymomonas mobilis</i> subsp. <i>mobilis</i>	Beverages
	<i>Candida famata</i>	Fermentation of blue vein cheese and biopreservation of citrus; meat
	<i>C. guilliermondii</i>	Citric acid (food additive)
	<i>C. krusei</i>	<i>Kefir</i> fermentation; sourdough fermentation
	<i>Debaryomyces hansenii</i>	Ripening of smear cheeses; meat
	<i>Geotrichum candidum</i>	Ripening of soft and semisoft cheeses or fermented milks; meat
	<i>Kluyveromyces marxianus</i>	Cheese ripening; lactase (food additive)
	<i>S. bayanus</i>	<i>Kefir</i> fermentation; juice and wine fermentation
	<i>S. cerevisiae</i>	Beer, bread, invertase (food additive)
	<i>S. cerevisiae</i> subsp. <i>boulardii</i>	Used as probiotic culture
	<i>S. florentinus</i>	<i>Kefir</i> fermentation
<i>S. pastorianus</i>	Beer	
<i>S. sake</i>	<i>Sake</i> fermentation	
<i>S. unisporus</i>	<i>Kefir</i> fermentation	
<i>Schizosaccharomyces pombe</i>	Wine	
<i>Zygosaccharomyces rouxii</i>	Soy sauce	
Filamentous moulds	<i>Aspergillus flavus</i>	α -amylases (food additive)
	<i>A. niger</i>	Beverages; industrial production of citric acid; amyloglucosidases, pectinase, cellulase, glucose oxidase, protease (food additives)
	<i>A. oryzae</i> , <i>A. sojae</i>	Soy sauce, beverages; α -amylases, amyloglucosidase, lipase (food additives)
	<i>Penicillium camemberti</i>	White mold cheeses (camembert type)
	<i>P. notatum</i>	Glucose oxidases (food additive)
	<i>P. roqueforti</i>	Blue mold cheeses
	<i>Rhizopus oligosporus</i>	<i>Tempe</i> fermentation
	<i>R. oryzae</i>	Soy sauce, <i>koji</i>

protein producing peptides and amino acids, which claim to have health benefits (Nagai and Tamang, 2010). Inhibitory properties of Angiotensin converting enzyme (ACE) have been studied in various fermented milk products such as *kefir* (Quiros et al., 2005), *koumiss* (Chen et al., 2010), yogurt (Papadimitriou et al., 2007), fermented camel milk (Moslehishad et al., 2013), cheese (Meyer et al., 2009), and fermented fish products (Ichimura et al., 2003).

Production of Enzymes by Microorganisms

Another important reason to ferment foods is to coax microorganisms into producing enzymes that also provide very useful services. During food fermentation microorganisms produce enzymes to break down complex compounds to simple

bio-molecules for several biological activities such as proteinase, amylase, mannase, cellulase, and catalase in many Asian fermented soybean foods by *Bacillus* spp. (Tamang and Nikkuni, 1996; Chettri and Tamang, 2014). Common genera of mycelial fungi in fermented foods and beverages such as *Actinomucor*, *Amylomyces*, *Aspergillus*, *Monascus*, *Mucor*, *Neurospora*, and *Rhizopus* produce various carbohydrases such as α -amylase, amyloglucosidase, maltase, invertase, pectinase, β -galactosidase, cellulase, hemi-cellulase; acid and alkaline proteases; and lipases (Nout and Aidoo, 2002). Taka-amylase A (TAA), an enzyme produced by *Aspergillus oryzae* in *koji* has many uses in industry (Suganuma et al., 2007). Dry, solid, cake-like mixed amyolytic starters used for alcohol production in the Himalayas have yeasts *Saccharomycopsis fibuligera*, *S. capsularis* and *Pichia burtonii* with high amylase activities (Tsuyoshi et al., 2005; Tamang et al., 2007).

Bacillus subtilis subsp. *natto* in *natto* produces nattokinase showing fibrinolytic activity (Mine et al., 2005; Kotb, 2012). Among bacteria isolated from fermented foods, *B. subtilis* and *B. amyloliquefaciens* (Chang et al., 2012; Zeng et al., 2013; Singh et al., 2014), *Vagococcus carniphilus*, *V. lutrae*, *Enterococcus faecalis*, *E. faecium*, *E. gallinarum*, and *P. acidilactici* (Singh et al., 2014), and *Virgibacillus halodenitrificans* SK1-3-7 isolated from fish sauce fermentation (Montriwong et al., 2012) produce fibrinolytic enzymes.

Increase in Isoflavones and Saponin and Production of PGA

Isoflavones are daidzein, genistein and glycitein, each of which exists in four chemical forms viz., aglycones, β -glucoside, acetylglucoside, and malonylglucoside in soybeans (Kudou et al., 1991). Isoflavone glucosides are hydrolyzed into their corresponding aglycones during fermentation of some Asian fermented soybean foods such as *sufu* and *douchi* of China (Wang et al., 2007b; Yin et al., 2007), *miso* and *natto* of Japan (Chiou and Cheng, 2001), *chungkokjang* and *doenjang* of Korea (Lee et al., 2007), *tempe* of Indonesia (Lu et al., 2009), and *thua nao* of Thailand (Dajanta et al., 2009). During *tempe* fermentation, isoflavone particularly Factor-II and aglycone contents are found to increase (Nakajima et al., 2005). Isoflavones in *doenjang* increase the activation of an LDL-C receptor, which is beneficial to prevent vascular diseases (Kwak et al., 2012).

Soybean saponins, which are oleanane triterpenoid glycosides, are again of two types viz., Group A and DDMP (2,3-dihydro-2,5-dihydroxy-6-methyl-4H-pyran-4-one; Paucar-Menacho et al., 2010). DDMP and their derivatives, Groups B and E saponins show health promoting benefits such as prevention of hypercholesterolemia (Murata et al., 2006), suppression of colon cancer cell proliferation (Ellington et al., 2006), and anti-peroxidation of lipids (Ishii and Tanizawa, 2006). Saponin contents are increased in *natto*, which are generated by *Bacillus natto* (Yanagisawa and Sumi, 2005). *Kinema* has high content of Group B saponin, which may indicate its health-promoting benefits to consumers (Omizu et al., 2011).

Poly-glutamic acid (PGA) is not synthesized by ribosomal proteins (Oppermann-Sanio and Steinbüchel, 2002), but is produced by some strains of *Bacillus* spp. in fermented soybean foods of Asia (Urushibata et al., 2002; Meerak et al., 2007; Nishito et al., 2010; Chettri and Tamang, 2014). *B. subtilis* and *B. licheniformis* are widely used industrial producers of γ -PGA (Stanley and Lazazzera, 2005). It is safe eating the viscous materials of Asian fermented soybean foods since PGA is completely biodegradable and water-soluble and non-toxic to human (Yoon et al., 2000).

Degradation of Anti-nutritive Compounds

Some microorganisms present in fermented foods may degrade anti-nutritive substances and thereby convert the substrates into consumable products (Nout, 1994; Tamang, 2015). Various steps employed during the processing of *gari* and *fufu*, fermented cassava products of Africa, such as peeling, washing, grating,

fermentation, dewatering and roasting minimizes the residual cyanide contents of the product (Babalola, 2014). Bitter varieties of cassava tubers contain the cyanogenic glycoside linamarin and lotaustralin, which are detoxified by species of *Leuconostoc*, *Lactobacillus*, and *Streptococcus* during traditional method to *gari* and *fufu* productions to yield hydrocyanic acid (HCN) which has low boiling point and escapes from the dewatered pulp during toasting rendering the product safe for human consumption (Lambri et al., 2013; Babalola, 2014; Bamidele et al., 2015). In *tempe*, *Rhizopus oligosporus* eliminates the flatulence causing indigestible oligosaccharides such as stachyose and verbascose into the absorbable monosaccharides and disaccharides (Hesseltine, 1983; Sanchez, 2008). Degradation of anti-nutritive compounds by *B. subtilis* has been reported in *kinema* (Sarkar et al., 1997). Phytic acid is reduced during fermentation of *idli* (Reddy and Salunkhe, 1980) and *rabadi*, a fermented cereal food of India (Gupta et al., 1992).

HEALTH BENEFITS OF FERMENTED FOODS

Ethnic foods have in-built systems both as foods and medicine to meet up hungry and also curative (Shin and Jeong, 2015; Thapa and Tamang, 2015). The highest longevity observed among the people of Okinawa prefecture in Japan is mostly due to their traditional and cultural foods such as *natto*, *miso*, *tofu*, *shoyu*, fermented vegetables, cholesterol-free, low-fat, and high bioactive-compounded foods in addition to active physical activity, sound environment, happiness and other several factors (Willcox et al., 2004). Korean *kimchi* has been claimed to possess health-promoting benefits (Cheigh, 1999; Lee et al., 2011; Park et al., 2014; Han et al., 2015). *Kimchi* has also anti-aging effect (Kim et al., 2002). *Natto* has several health benefits such as high contents of nattokinase, isoflavones, saponins, vitamin K, unsaturated fatty acids, probiotics and immunomodulating activities mostly produced by *B. subtilis* (*natto*; Tsubura, 2012; Nagai, 2015). *Kinema* has also some health promoting benefits (Omizu et al., 2011; Tamang, 2015). Indian popular fermented milk *dahi* has anti-carcinogenic property (Arvind et al., 2010). Lactic acid produced in *kimchi* may prevent fat accumulation and to improve obesity-induced heart diseases (Park et al., 2008). Anti-obesity effects have been reported in *kimchi* (Kim et al., 2011; Park et al., 2012) and in *doenjang* (Kwak et al., 2012) based on clinical trials (Cha et al., 2012; Jung et al., 2014). Red wine has anti-aging property due to presence of melatonin that regulates the body clock (Corder et al., 2006; Walker, 2014).

Ethnic people have customary belief in medicinal values of some of their ethnic foods including fermented foods and beverages, however, clinical trials and validation of the health benefits claims of almost all naturally fermented foods and beverages of the world need to be studied. Some health benefits of fermented foods are listed in **Table 2**.

Synthesis of Nutrient

Enrichment of substrates with vitamins, essential amino acids, and bioactive compounds occur during food fermentation

(Holzapfel et al., 1995; Steinkraus, 1996; Thapa and Tamang, 2015). In *tempe*, mold-fermented soybean food of Indonesia, contents of folic acid, niacin, riboflavin, nicotinamide and pyridoxine are found to be increased by *Rhizopus oligosporus* (Astuti, 2015), whereas vitamin B₁₂ is synthesized by non-pathogenic strains of *Klebsiella pneumoniae* and *Citrobacter freundii* (Liem et al., 1977; Okada, 1989; Keuth and Bisping, 1994). Contents of thiamine, riboflavin and methionine in *idli*, a rice-legume based fermented food of India and Sri Lanka enhance during fermentation (Ghosh and Chattopadhyay, 2011). Similarly, vitamins B complex and C, lysine and tryptophane, and iron contents have been found to increase during fermentation of *pulque*, an alcoholic drink of Mexico made from cactus plant (Ramirez et al., 2004). Riboflavin and niacin contents are increased in many *Bacillus*-fermented Asian fermented foods (Sarkar et al., 1998; Kim and Hahm, 2002; Nagai, 2015). Riboflavin and folic acid were found to be synthesized in *kimchi* by *L. mesenteroides* and *L. sakei* (Jung et al., 2013). Yeasts

Saccharomyces cerevisiae, *Candida tropicalis*, *Aureobasidium* sp., and *Pichia manschuria* isolated from *idli* and *jalebi*, fermented cereal foods of India and Pakistan produce vitamin B₁₂ (Syal and Vohra, 2013). Free amino acids are increased in fermented soybean foods (Nikkuni et al., 1995; Sarkar and Tamang, 1995; Tamang and Nikkuni, 1998; Kiers et al., 2000; Dajanta et al., 2011).

Prevention of Hypertension and Heart Disease

Antihypertensive properties of many fermented milk products have been validated using animal models and clinical trials (Seppo et al., 2002; Sipola et al., 2002). Consumption of fermented milks or probiotic bacteria (Agerholm-Larsen et al., 2000) and fermented soybean foods (Liu and Pan, 2010) lowers the risk of heart diseases. Fermented whole grain foods can lower the serum LDL-cholesterol values, hypertriacylglycerolaemia,

TABLE 2 | Some bioactive compounds in fermented foods and their health benefits.

Bioactive compounds	Synthesized in fermented foods	Health benefits	Reference
Genistein	<i>Doenjang</i>	Facilitates the β-oxidation of fatty acid, reducing body weight	Kwak et al., 2012
Lipoteichoic acid from <i>L. rhamnosus</i> GG	Fermented milk	Oral photoprotective agent against UV-induced carcinogenesis	Weill et al., 2013
Isocyanate and sulphide indole-3-carbinol	<i>Kimchi</i>	Prevention of cancer, detoxification of heavy metals in liver, kidney, and small intestine	Kwak et al., 2014
Ornithine		Anti-obesity efficacy	Park et al., 2012
Vitamin A, Vitamin C, fibers		Suppression of cancer cells	Han et al., 2015
Capsaicin, Allicin		Prevention of cancer, suppression of <i>Helicobacter pylori</i>	Lim and Im, 2009
Chlorophyll		Helps in prevention of absorbing carcinogen	Ferruzzi and Blakeslee, 2007
S-adenosyl-L-methionine (SAM)		Treatment of depression	Lee and Lee, 2009
HDMPPA (an antioxidant)		Therapeutic application in human atherosclerosis	Kim et al., 2007
Nattokinase, antibiotics, Vitamin K	<i>Natto</i>	Antitumor, immunomodulating	Nagai, 2015
Vitamin C	Sauerkraut	Scurvy	Peñas et al., 2013
Glucosinolates		Activation of natural antioxidant enzymes	Martinez-Villaluenga et al., 2012
Antioxidant genestein, daidzein, tocopherol, superoxide dismutase	<i>Tempe</i>	Prevents oxidative stress causing non-communicable disease such as hyperlipidemia, diabetes, cancer (breast and colon), prevents the damage of pancreatic beta cell	Astuti, 2015
Phenolics- resveratrol	<i>Wine (red)</i>	Anti inflammatory	Jeong et al., 2010
Phenolics, succinic acid		Digestive aid	Jackson, 2008
Phenolics, resveratrol, flavonoids – quercetin, Vitamins C and E, mineral selenium		Prevent cardiovascular diseases, reduce incidence of heart attacks and mortality rate	Walker, 2014
Melatonin, resveratrol		Antioxidant and anti-aging property	Fernández-Mar et al., 2012
Resveratrol		Anti-diabetic	Ramadori et al., 2009

hypertension, coronary heart disease, insulin resistance, and hyperhomocysteinaemia (Anderson, 2003). Consumption of some fermented foods reduces the cholesterol level in *tempe* (Hermosilla et al., 1993), fermented soybean foods (Lee, 2004), and *kefir* (Otes and Cagindi, 2003). *Calpis*, the Japanese fermented sour milk containing two peptides VPP and IPP has shown hypotensive effect (Nakamura et al., 1996). *L. helveticus* in fermented milk reduces elevated blood pressure (Aihara et al., 2005; Shah, 2015). *Monascus purpureus* in fermented red-rice of China locally called *angkak*, prohibits creation of cholesterol by blocking a key enzyme, HMG-CoA reductase due to presence of mevinolin citrinin (Pattanagul et al., 2008).

Drinking of fermented tea of China prevents heart disease (Mo et al., 2008). Some Asian fermented soybean foods have antihypertensive properties as observed in *natto* (Nagai, 2015) and *tempe* (Astuti, 2015). Isoflavone in *doenjang*, mold-fermented soybean food of Korea, plays an important role in preventing cardiovascular diseases (Kwak et al., 2012; Shin et al., 2015). Fermented whole-grain intake appears to protect from development of heart disease and diabetes (Anderson, 2003). Moderate consumption of wine is healthier (Walker, 2014). Polyphenols in red wine probably are synergists of the tocopherol (Vitamin E) and ascorbic acid (Vitamin C), thus they inhibit lipid peroxidation (Feher et al., 2007). Regular consumption of the Korean fermented soybean foods by hypertensive and Type 2 diabetic patients results in favorable changes in cardiovascular risk factors (Jung et al., 2014) and reduction of hypocholesterolemic effect (Lim et al., 2014). ACEs inhibitory peptides derived from food proteins are used for treating hypertension (Jakubczyk et al., 2013). Fermented foods, which are rich in fibrinolytic enzymes, are useful for thrombolytic therapy to prevent rapidly emerging heart diseases (Mine et al., 2005; Singh et al., 2014).

Prevention from Cancer

Some LAB-fermented foods have antimutagenic and anticarcinogenic activities (Lee et al., 2004). *Kefir* is used for the treatment of cancer (Otes and Cagindi, 2003; Yanping et al., 2009). Sauerkraut, fermented vegetable of Germany, contains *s*-methylmethionine, which reduces tumorigenesis risk in the stomach (Kris-Etherton et al., 2002). Consumption of fermented milk products containing live cells of *L. acidophilus* decreases β -glucuronidase, azoreductase, and nitroreductase (catalyze conversion of procarcinogens to carcinogens), probably removes procarcinogens, and activate the immune system of consumers (Goldin and Gorbach, 1984; Macouzet et al., 2009). Similarly, Indian *dahi* has anti-carcinogenic property (Mohania et al., 2013). Cancer preventive potential of *W. cibaria*, and *L. plantarum* has been reported in *kimchi* (Kwak et al., 2014). Consumption of yogurt can reduce bladder, colon and cervical cancer has been observed (Chandan and Kilara, 2013).

Protection against Gastrointestinal Disorders

Lactic acid bacteria present in fermented foods may decrease number of incidence, duration and severity of

some gastrointestinal disorders (Verna and Lucak, 2010). Administration of some strains of *Lactobacillus* improves the inflammatory bowel disease, paucities and ulcerative colitis (Orel and Trop, 2014). *L. rhamnosus* GG is effective in the treatment of acute diarrhea (Szajewska et al., 2007) and administration of *L. helveticus*-fermented milk in healthy older adults produced improvements in cognition function (Chung et al., 2014). Consumption of fermented milk products containing live bacteria has immunomodulation capacity (Granier et al., 2013), and cures diarrhea (Balamurugan et al., 2014). Korean *kimchi* is suitable for control of inflammatory bowel diseases (Lim et al., 2011).

Anti-allergic Reactions

Lactobacillus kefiranofaciens M1 isolated from *kefir* grains has an anti-allergic effect (Hong et al., 2010). Digestion of caseins during maturation of fermented milk products has shown to facilitate loss of allergenic reactivity thus increases tolerance (Alessandri et al., 2012). *Chongkokjang* has anti-allergic effect such as dermis thickness, decreased ear thickness, auricular lymph node and infiltrating mast cells (Lee et al., 2014). *Lactobacillus* species isolated from *kimchi* are found to modulate Th1/Th2 balance by producing a large amount of IL-12 and IFN- γ with ability to alleviate atopic dermatitis and food allergy (Won et al., 2011). Fermented fish oil, which is rich with Omega-3 polyunsaturated fatty acids, can reduce sensitization of allergy (Han et al., 2012).

Protection from Diabetes and Osteoporosis

Intake of high fiber foods may decrease the insulin requirements in diabetic persons (Meyer et al., 2000), and may increase the sensitivity to insulin for non-diabetic persons (Fukagawa et al., 1990; Anderson, 2003). Probiotic *dahi*-supplemented diet significantly delays the glucose intolerance, hyperglycemia, hyperinsulinemia, oxidative stress and dyslipidemia indicating a lower risk of diabetes (Yadav et al., 2007). Daily consumption of *chungkokjang* may increase the insulin resistivity thus controls diabetics (Shin et al., 2011; Tolhurst et al., 2012).

Vitamin K2 present in *natto* stimulates the formation of bone, which may help to prevent osteoporosis in older women in Japan (Yanagisawa and Sumi, 2005). Mineral such magnesium, calcium, phosphorus, potassium, and also protein present in yogurt may function together to promote formation of healthy bones (Chandan and Kilara, 2013).

Alleviation of Lactose Malabsorption

Some people suffer from lactose malabsorption, a condition in which lactose, the principal carbohydrate of milk, is not completely digested into glucose and galactose due to lack of β -D-galactosidase (Shah, 2015). *L. delbrueckii* subsp. *bulgaricus* and *S. thermophilus* used in production of yogurt contain substantial quantities of β -D-galactosidase which improve the symptoms of lactose malabsorption in lactose intolerant people (Shah et al., 2013). Consumption of fresh yogurt (with live yogurt cultures) has been demonstrated better lactose digestion and absorption than with the consumption of a pasteurized product

(Pedone et al., 2000). *Kefir* can minimize the symptoms of lactose intolerance by providing extra source of β -galactosidase (Hertzler and Clancy, 2003).

HEALTH RISK OF FERMENTED FOODS

One of the important health risks in fermented foods is presence of biogenic amines. Biogenic amines are low molecular weight organic compounds by microbial decarboxylation of their precursor amino acids or by transamination of aldehydes and ketones by amino acid transaminases (Zhai et al., 2012), which are present in some fermented foods such as *sauerkraut*, fish products, cheese, wine, beer, dry sausages, etc. (Halász et al., 1994; Suzzi and Gardini, 2003; Spano et al., 2010; Visciano et al., 2014). Enterobacteriaceae and enterococci are major biogenic amine producers in foods (Nout, 1994). Foods with high levels of biogenic amines could be considered as unhealthy (Latorre-Moratalla et al., 2010). High levels (>100 mg/kg) of histamine and tyramine can cause adverse effects to human health (Rauscher-Gabernig et al., 2009). Fermentation of cabbage with certain lactic starters such as *L. casei* subsp. *casei*, *L. plantarum* and *L. curvatus* could reduce the biogenic amine content of *sauerkraut* (Rabie et al., 2011). The ingestion of food containing small amounts of histamine has little effect in healthy individuals, but it can result in histamine intolerance in persons characterized by impairment of diamine oxidase activity, either due to genetic predisposition, gastrointestinal diseases, or medication with monoamine oxidase inhibitors (Maintz and

Novak, 2007). A maximum limit of 100 mg/kg of histamine in food indicates a safe level for consumption (Halász et al., 1994).

CONCLUSION

Some fermented foods and beverages have health benefits due to presence of functional microorganisms. Although, some fermented foods and beverages are marketed globally as health foods, functional foods, therapeutic foods, nutraceutical foods, bio-foods, however, due to urbanization, changes in life-style, and the shifting from traditional food habits to commercial fast foods, the production and consumption of traditional fermented foods is in decline mostly in Asia and Africa. Reliance on fewer providers of fermented foods is also leading to a decline in the biodiversity of microorganisms. We recommend that validation of health claims by clinical trials and animal models of some common fermented foods of the world may be studied in details, and also introduction of new fermented food products containing well-validated functional microorganism(s) may emerge in global food market.

AUTHOR CONTRIBUTIONS

JPT (70% – data collection, analysis, writing), D-HS (10% – data collection), S-JJ (10% – data collection) and S-WC (10% – data collection).

REFERENCES

- Abubakr, M. A. S., Hassan, Z., Imdakim, M. M. A., and Sharifah, N. R. S. (2012). Antioxidant activity of lactic acid bacteria (LAB) fermented skim milk as determined by 1,1-diphenyl-2-picrylhydrazyl (DPPH) and ferrous chelating activity (FCA). *Afr. J. Microbiol. Res.* 6, 6358–6364.
- Agerholm-Larsen, L., Bell, M. L., Grunwald, G. K., and Astrup, A. (2000). The effect of probiotic milk product on plasma cholesterol: a meta-analysis of short-term intervention studies. *Eur. J. Clin. Nutr.* 54, 856–860.
- Aihara, K., Kajimoto, O., Hirata, H., Takahashi, R., and Nakamura, Y. (2005). Effect of powdered fermented milk with *Lactobacillus helveticus* on subjects with high-normal blood pressure or mild hypertension. *J. Am. Col. Nutri.* 24, 257–265.
- Alessandri, C., Sforza, S., Palazzo, F., Lambertini, S., Paoletta, D., Zennaro, C., et al. (2012). Tolerability of a fully matured cheese in cow's milk allergic children: biochemical, immunochemical, and clinical aspects. *PLoS ONE* 7:e40945.
- Anderson, J. W. (2003). Whole grains protect against atherosclerotic cardiovascular disease. *Proc. Nutri. Soc.* 62, 135–142. doi: 10.1079/PNS2002222
- Arvind, K., Nikhlesh, K. S., and Pushpalata, R. S. (2010). Inhibition of 1,2-dimethylhydrazine induced colon genotoxicity in rats by the administration of probiotic curd. *Mol. Biol. Rep.* 37, 1373–1376. doi: 10.1007/s11033-009-9519-1
- Astuti, M. (2015). "Health benefits of tempe", in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York, NY: CRC Press), 371–394.
- Axelsson, L., Rud, I., Naterstad, K., Blom, H., Renckens, B., Boekhorst, J., et al. (2012). Genome sequence of the naturally plasmid-free *Lactobacillus plantarum* strain NC8 (CCUG 61730). *J. Bacteriol.* 194, 2391–2392. doi: 10.1128/JB.00141-12
- Babalola, O. O. (2014). Cyanide content of commercial gari from different areas of Ekiti State, Nigeria. *World J. Nutri. Health* 2, 58–60.
- Badis, A., Guetarni, D., Moussa-Boudjemaa, B., Henni, D. E., Tornadijo, M. E., and Kihal, M. (2004). Identification of cultivable lactic acid bacteria isolated from Algerian raw goat's milk and evaluation of their technological properties. *Food Microbiol.* 21, 343–349. doi: 10.1016/j.fm.2003.11.006
- Balamurugan, R., Chandragunasekaran, A. S., Chellappan, G., Rajaram, K., Ramamoorthi, G., and Ramakrishna, B. S. (2014). Probiotic potential of lactic acid bacteria present in home made curd in Southern India. *Indian J. Med. Res.* 140, 345–355.
- Bamidele, O. P., Fasogbon, M. B., Oladiran, D. A., and Akande, E. O. (2015). Nutritional composition of fufu analog flour produced from Cassava root (*Manihot esculenta*) and Cocoyam (*Colocasia esculenta*) tuber. *Food Sci. Nutr.* 3, 597–603. doi: 10.1002/fsn3.250
- Bernardeau, M., Guguen, M., and Vernoux, J. P. (2006). Beneficial lactobacilli in food and feed: long-term use, biodiversity and proposals for specific and realistic safety assessments. *FEMS Microbiol. Rev.* 30, 487–513. doi: 10.1111/j.1574-6976.2006.00020.x
- Bourdichon, F., Casaregola, S., Farrokh, C., Frisvad, J. C., Gerds, M. L., Hammes, W. P., et al. (2012). Food fermentations: microorganisms with technological beneficial use. *Int. J. Food Microbiol.* 154, 87–97. doi: 10.1016/j.ijfoodmicro.2011.12.030
- Cha, Y. S., Yang, J. A., Back, H. I., Kim, S. R., Kim, M. G., Jung, S. J., et al. (2012). Visceral fat and body weight are reduced in overweight adults by the supplementation of Doenjang, a fermented soybean paste. *Nutri. Res. Pract.* 6, 520–526.
- Chandan, R. C., and Kilara, A. (2013). *Manufacturing Yogurt and Fermented Milks*, 2nd Edn. (Chichester: John Wiley & Sons), 477.
- Chang, C. T., Wang, P. M., Hung, Y. F., and Chung, Y. C. (2012). Purification and biochemical properties of a fibrinolytic enzyme from *Bacillus subtilis* – fermented red bean. *Food Chem.* 133, 1611–1617.
- Chang, J. Y., Lee, H. J., and Chang, H. C. (2008). Identification of the agent from *Lactobacillus plantarum* KFRI464 that enhances bacteriocin production by *Leuconostoc citreum* GJ7. *J. Appl. Microbiol.* 103, 2504–2515. doi: 10.1111/j.1365-2672.2007.03543.x

- Chávarri, M., Marañón, I., Ares, R., Ibáñez, F. C., Marzo, F., and Villarán, M. C. (2010). Microencapsulation of a probiotic and prebiotic in alginate-chitosan capsules improves survival in simulated gastro-intestinal conditions. *Int. J. Food Microbiol.* 142, 185–189. doi: 10.1016/j.ijfoodmicro.2010.06.022
- Cheigh, H. (1999). Production, characteristics and health functions of kimchi. *Acta Horticult.* 483, 405–420. doi: 10.17660/ActaHortic.1999.483.47
- Chen, B., Wu, Q., and Xu, Y. (2014). Filamentous fungal diversity and community structure associated with the solid state fermentation of Chinese Maotai-flavor liquor. *Int. J. Food Microbiol.* 179, 80–84. doi: 10.1016/j.ijfoodmicro.2014.03.011
- Chen, Y., Wang, Z., Chen, X., Liu, Y., Zhang, H., and Sun, T. (2010). Identification of angiotensin I-converting enzyme inhibitory peptides from koumiss, a traditional fermented mare's milk. *J. Dairy Sci.* 93, 884–892.
- Chettri, R., and Tamang, J. P. (2014). Functional properties of tungrymbai and bekgang, naturally fermented soybean foods of North East India. *Int. J. Fer. Foods* 3, 87–103. doi: 10.5958/2321-712X.2014.01311.8
- Chiu, R. Y. Y., and Cheng, S. L. (2001). Isoflavone transformation during soybean koji preparation and subsequent miso fermentation supplemented with ethanol and NaCl. *J. Agric. Food Chem.* 49, 3656–3660. doi: 10.1021/jf001524l
- Chung, Y. C., Jin, H. M., Cui, Y., Kim, D. S., Jung, J. M., Park, J. I., et al. (2014). Fermented milk of *Lactobacillus helveticus* IDCC3801 improves cognitive functioning during cognitive fatigue tests in healthy older adults. *J. Funct. Foods* 10, 465–474. doi: 10.1016/j.jff.2014.07.007
- Corder, R., Mullen, W., Khan, N. Q., Marks, S. C., Wood, E. G., Carrier, M. J., et al. (2006). Oenology: red wine procyanidins and vascular health. *Nature* 444:566.
- Dajanta, K., Apichartsrangkoon, A., Chukeatirote, E., Richard, A., and Frazier, R. A. (2011). Free-amino acid profiles of thua nao, a Thai fermented soybean. *Food Chem.* 125, 342–347. doi: 10.1016/j.foodchem.2010.09.002
- Dajanta, K., Chukeatirote, E., Apichartsrangkoon, A., and Frazier, R. A. (2009). Enhanced aglycone production of fermented soybean products by *Bacillus* species. *Acta Biol. Szeged.* 53, 93–98.
- Dajanta, K., Janpum, P., and Leksing, W. (2013). Antioxidant capacities, total phenolics and flavonoids in black and yellow soybeans fermented by *Bacillus subtilis*: a comparative study of Thai fermented soybeans (thua nao). *Int. Food Res. J.* 20, 3125–3132.
- de LeBlanc, A. M., Matar, C., and Perdígón, G. (2007). The application of probiotics in cancer. *Br. J. Nutri.* 98, S105–S110. doi: 10.1017/S0007114507839602
- De Mejia, E. G., and Dia, V. P. (2010). The role of nutraceutical proteins and peptides in apoptosis, angiogenesis, and metastasis of cancer cells. *Cancer Metast.* 29, 511–528. doi: 10.1007/s10555-010-9241-4
- Ellington, A. A., Berhow, M. A., and Singletary, K. W. (2006). Inhibition of Akt signaling and enhanced ERK1/2 activity are involved in induction of macroautophagy by triterpenoid B-group soyasaponins in colon cancer cells. *Carcinogenesis* 27, 298–306. doi: 10.1093/carcin/bgi214
- Farhad, M., Kailasapathy, K., and Tamang, J. P. (2010). "Health aspects of fermented foods," in *Fermented Foods and Beverages of the World*, eds J. P. Tamang and K. Kailasapathy (New York, NY: CRC Press), 391–414.
- Feher, J., Lengyel, G., and Lugasi, A. (2007). The cultural history of wine - theoretical background to wine therapy. *Central Eur. J. Med.* 2, 379–391.
- Fernández-Mar, M. I., Mateos, R., García-Parrilla, M. C., Puertas, B., and Cantos-Villar, E. (2012). Bioactive compounds in wine: Resveratrol, hydroxytyrosol and melatonin: a review. *Food Chem.* 130, 797–813.
- Ferruzzi, M. G., and Blakeslee, J. (2007). Digestion, absorption, and cancer preventative activity of dietary chlorophyll derivatives. *Nutr. Res.* 27, 1–12. doi: 10.1016/j.nutres.2006.12.003
- Fukagawa, N. K., Anderson, J., Young, V. R., and Minaker, K. L. (1990). High-carbohydrate, high-fiber diets increase peripheral insulin sensitivity in healthy young and old adults. *Am. J. Clin. Nutri.* 52, 524–528.
- Gaggia, F., Di Gioia, D., Baffoni, L., and Biavati, B. (2011). The role of protective and probiotic cultures in food and feed and their impact in food safety. *Trends Food Sci. Technol.* 22, 58–66. doi: 10.1016/j.tifs.2011.03.003
- García-Ruiz, A., Esteban-Fernández, D. G. A., Requena, T., Bartolomé, B., and Moreno-Arribas, M. V. (2014). Assessment of probiotics properties in lactic acid bacteria isolated from wine. *Food Microbiol.* 44, 220–225. doi: 10.1016/j.fm.2014.06.015
- Ghosh, D., and Chattopadhyay, P. (2011). Preparation of idli batter, its properties and nutritional improvement during fermentation. *J. Food Sci. Technol.* 48, 610–615. doi: 10.1007/s13197-010-0148-4
- Goldin, B. R., and Gorbach, S. L. (1984). The effect of milk and lactobacillus feeding on human intestinal bacterial enzyme activity. *American. J. Clin. Nutri.* 39, 756–761.
- Granier, A., Goulet, O., and Hoarau, C. (2013). Fermentation products: immunological effects on human and animal models. *Pediatr. Res.* 74, 238–244. doi: 10.1038/pr.2013.76
- Grosu-Tudor, S. S., and Zamfir, M. (2013). Functional properties of LAB isolated from Romanian fermented vegetables. *Food Biotechnol.* 27, 235–248. doi: 10.1080/08905436.2013.811082
- Gupta, M., Khetarpaul, N., and Chauhan, B. M. (1992). Rabadi fermentation of wheat: changes in phytic acid content and in vitro digestibility. *Plant Foods Hum. Nutri.* 42, 109–116. doi: 10.1007/BF02196463
- Halász, A., Baráth, A., Simon-Sarkadi, L., and Holzapfel, W. H. (1994). Biogenic amines and their production by microorganisms in food. *Trends Food Sci. Technol.* 5, 42–49. doi: 10.1016/0924-2244(94)90070-1
- Han, E. S., Kim, H. J., and Choi, H. K. (2015). "Health benefits of Kimchi," in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York: CRC Press), 343–370.
- Han, S., Kang, G., Ko, Y., Kang, H., Moon, S., Ann, Y., et al. (2012). Fermented fish oil suppresses T helper 1/2 cell response in a mouse model of atopic dermatitis via generation of CD4+CD25+Foxp3+ T cells. *BMC Immunol* 13:44.
- Hermosilla, J. A. G., Jha, H. C., Egge, H., and Mahmud, M. (1993). Isolation and characterization of hydroxymethylglutaryl coenzyme A reductase inhibitors from fermented soybean extracts. *J. Clin. Biochem. Nutri.* 15, 163–174. doi: 10.3164/jcbn.15.163
- Hertzler, S. R., and Clancy, S. M. (2003). Kefir improves lactose digestion and tolerance in adults with lactose maldigestion. *J. Am. Diet Assoc.* 103, 582–587. doi: 10.1053/jada.2003.50111
- Hesseltine, C. W. (1983). Microbiology of oriental fermented foods. *Ann. Rev. Microbiol.* 37, 575–601. doi: 10.1146/annurev.mi.37.100183.003043
- Hill, C., Guarner, F., Reid, G., Gibson, G. R., Merenstein, D. J., Pot, B., et al. (2014). Expert consensus document: the international scientific association for probiotics and prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* 11, 506–514. doi: 10.1038/nrgastro.2014.66
- Holzapfel, W. H., Giesen, R., and Schillinger, U. (1995). Biological preservation of foods with reference to protective cultures, bacteriocins and food-grade enzymes. *Int. J. Food Microbiol.* 24, 343–362. doi: 10.1016/0168-1605(94)00036-6
- Holzapfel, W. H., and Wood, B. J. B. (2014). *Lactic Acid Bacteria: Biodiversity and Taxonomy*. (New York, NY: Wiley-Blackwell), 632.
- Hong, W., Chen, Y., and Chen, M. (2010). The antiallergic effect of kefir Lactobacilli. *J. Food Sci.* 75, H244–H253.
- Hur, J. W., Hyun, H. H., Pyun, Y. R., Kim, T. S., Yeo, I. H., and Park, H. D. (2000). Identification and partial characterization of lactacin bh5, a bacteriocin produced by *Lactococcus lactis* BH5 isolated from Kimchi. *J. Food Protect.* 63, 1707–1712.
- Ichimura, T., Hu, J., Aita, D. O., and Maruyama, S. (2003). Angiotensin I-Converting enzyme inhibitory activity and insulin secretion stimulative activity of fermented fish sauce. *J. Biosci. Bioengineer.* 96, 496–499. doi: 10.1016/S1389-1723(03)70138-8
- Ishii, Y., and Tanizawa, H. (2006). Effects of soyasaponins on lipid peroxidation through the secretion of thyroid hormones. *Biol. Pharm. Bull.* 29, 1759–1763. doi: 10.1248/bpb.29.1759
- Jackson, R. S. (2008). *Wine Science: Principles and Applications*, 3rd Edn. London: Academic Press, 686–706.
- Jakubczyk, A., Karaś, M., Baraniak, B., and Pietrzak, M. (2013). The impact of fermentation and in vitro digestion on formation angiotensin converting enzyme (ACE) inhibitory peptides from pea proteins. *Food Chem.* 141, 3774–3780. doi: 10.1016/j.foodchem.2013.06.095
- Jeong, J., Junga, H., Leea, S., Leea, H., Hwanga, K. T., and Kimb, T. (2010). Antioxidant, anti-proliferative and anti-inflammatory activities of the extracts from black raspberry fruits and wine. *Food Chem.* 123, 338–344.
- Ji, Y., Kim, H., Park, H., Lee, J., Lee, H., Shin, H., et al. (2013). Functionality and safety of lactic bacterial strains from Korean kimchi. *Food Control* 31, 467–473. doi: 10.1016/j.foodcont.2012.10.034
- Jiang, J., Shi, B., Zhu, D., Cai, Q., Chen, Y., Li, J., et al. (2012). Characterization of a novel bacteriocin produced by *Lactobacillus sakei* LSJ618 isolated from traditional Chinese fermented radish. *Food Control* 23, 338–344.

- Jung, J. Y., Lee, S. H., Jin, H. M., Hahn, Y., Madsen, E. L., and Jeon, C. O. (2013). Metatranscriptomic analysis of lactic acid bacterial gene expression during kimchi fermentation. *Int. J. Food Microbiol.* 163, 171–179. doi: 10.1016/j.ijfoodmicro.2013.02.022
- Jung, S. J., Park, S. H., Choi, E. K., Cha, Y. S., Cho, B. H., Kim, Y. G., et al. (2014). Beneficial effects of Korean traditional diets in hypertensive and Type 2 diabetic patients. *J. Med. Food* 17, 161–171. doi: 10.1089/jmf.2013.3042
- Keuth, S., and Bisping, B. (1994). Vitamin B12 production by *Citrobacter freundii* or *Klebsiella pneumoniae* during tempeh fermentation a proof of enterotoxin absence by PCR. *Appl. Environ. Microbiol.* 60, 1495–1499.
- Khan, H., Flint, S., and Yu, P. L. (2010). Enterocins in food preservation. *Int. J. Food Microbiol.* 141, 1–10. doi: 10.1016/j.ijfoodmicro.2010.03.005
- Kiers, J. L., Van laeken, A. E. A., Rombouts, F. M., and Nout, M. J. R. (2000). In vitro digestibility of *Bacillus* fermented soya bean. *Int. J. Food Microbiol.* 60, 163–169.
- Kim, E. K., An, S. Y., Lee, M. S., Kim, T. H., Lee, H. K., Hwang, W. S., et al. (2011). Fermented kimchi reduces body weight and improves metabolic parameters in overweight and obese patients. *Nutri. Res.* 31, 436–443. doi: 10.1016/j.nutres.2011.05.011
- Kim, H. J., Lee, J. S., Chung, H. Y., Song, S. H., Suh, H., Noh, J. S., et al. (2007). 3-(4'-Hydroxyl-3', 5'-dimethoxyphenyl) propionic acid, an active principle of kimchi, inhibits development of atherosclerosis in rabbits. *J. Agric. Food Chem.* 55, 10486–10492. doi: 10.1021/jf072454m
- Kim, J. H., Ryu, J. D., and Song, Y. O. (2002). The effect of kimchi intake on free radical production and the inhibition of oxidation in young adults and the elderly people. *Korean J. Commun. Nutri.* 7, 257–265.
- Kim, K. Y., and Hahn, Y. T. (2002). Recent studies about physiological functions of Chungkkokjang and Functional enhancement with genetic engineering. *Inst. Mol. Biol. Genet.* 16, 1–18.
- Kotb, E. (ed.). (2012). "Springer briefs microbiol," in *Fibrinolytic Bacterial Enzymes with Thrombolytic Activity* (Berlin: Springer).
- Kris-Etherton, P. M., Hecker, K. D., Bonanome, A., Coval, S. M., Binkoski, A. E., Hilpert, K. F., et al. (2002). Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *Am. J. Med.* 113, 71S–88S. doi: 10.1016/S0002-9343(01)00995-0
- Kubo, Y., Rooney, A. P., Tsukakoshi, Y., Nakagawa, R., Hasegawa, H., and Kimura, K. (2011). Phylogenetic analysis of *Bacillus subtilis* strains applicable to natto (fermented soybean) production. *Appl. Environ. Microbiol.* 77, 6463–6469. doi: 10.1128/AEM.00448-11
- Kudou, S., Fleury, Y., Welti, D., Magnolato, D., Uchida, T., Kitamura, K., et al. (1991). Malonyl isoflavone glycosides in soybean seeds (*Glycine max* Merrill). *Agric. Biol. Chem.* 55, 2227–2233. doi: 10.1271/abb1961.55.2227
- Kwak, C. S., Park, S., and Song, K. Y. (2012). Doenjang, a fermented soybean paste, decreased visceral fat accumulation and adipocyte size in rats fed with high fat diet more effectively than nonfermented soybeans. *J. Med. Food* 15, 1–9. doi: 10.1089/jmf.2010.1224
- Kwak, S. H., Cho, Y. M., Noh, G. M., and Om, A. S. (2014). Cancer preventive potential of Kimchi lactic acid bacteria (*Weissella cibaria*, *Lactobacillus plantarum*). *J. Cancer Prevent.* 19, 253–258. doi: 10.15430/JCP.2014.19.4.253
- Lambri, M., Fumi, M. D., Roda, A., and de Faveri, D. (2013). Improved processing methods to reduce the total cyanide content of cassava roots from Burundi. *Afr. J. Biotechnol.* 12, 2685–2691.
- Latorre-Moratalla, M. L., Bover-Cid, S., Talon, R., Garriga, M., Aymerich, T., Zanardi, E., et al. (2010). Strategies to reduce biogenic amine accumulation in traditional sausage manufacturing. *Food Sci. Technol.* 43, 20–25.
- Lee, C. H. (2004). Creative fermentation technology for the future. *J. Food Sci.* 69, 33–34.
- Lee, H., Yoon, H., Ji, Y., Kim, H., Park, H., Lee, J., et al. (2011). Functional properties of *Lactobacillus* strains isolated from kimchi. *Int. J. Food Microbiol.* 145, 155–161. doi: 10.1016/j.ijfoodmicro.2010.12.003
- Lee, H. R., and Lee, J. M. (2009). Anti-stress effects of kimchi. *Food Sci. Biotechnol.* 18, 25–30.
- Lee, J. K., Jung, D. W., Kim, Y. J., Cha, S. K., Lee, M. K., Ahn, B. H., et al. (2009). Growth inhibitory effect of fermented kimchi on food-borne pathogens. *Food Sci. Biotechnol.* 18, 12–17.
- Lee, J. W., Shin, J. G., Kim, E. H., Kang, H. E., Yim, I. B., Kim, J. Y., et al. (2004). Immunomodulatory and antitumor effects in vivo by the cytoplasmic fraction of *Lactobacillus casei* and *Bifidobacterium longum*. *J. Vet. Sci.* 5, 41–48.
- Lee, Y. J., Kim, J. E., Kwak, M. H., Go, J., Kim, D. S., Son, H. J., et al. (2014). Quantitative evaluation of the therapeutic effect of fermented soybean products containing high concentration of GABA on phthalic anhydride-induced atopic dermatitis in IL4/Luc/CNS-1 Tg mice. *Int. J. Mol. Med.* 33, 1185–1194.
- Lee, Y. W., Kim, J. D., Zheng, J. Z., and Row, K. H. (2007). Comparisons of isoflavones from Korean and Chinese soybean and processed products. *Biochem. Eng. J.* 36, 49–53. doi: 10.1016/j.bej.2006.06.009
- Liem, I. T. H., Steinkraus, K. H., and Cronk, T. C. (1977). Production of vitamin B12 in tempeh, a fermented soybean food. *Appl. Environ. Microbiol.* 34, 773–776.
- Lim, J., Seo, B. J., Kim, J. E., Chae, C. S., Im, S. H., Hahn, Y. S., et al. (2011). Characteristics of immunomodulation by a *Lactobacillus sakei* proBio65 isolated from Kimchi. *Korean J. Microbiol. Biotechnol.* 39, 313–316.
- Lim, J. H., Jung, E. S., Choi, E. K., Jeong, D. Y., Seung-Wha, J. O., Jin, J. H., et al. (2014). Supplementation with *Aspergillus oryzae*-fermented kochujang lowers serum cholesterol in subjects with hyperlipidemia. *Clin. Nutri.* 34, 383–387. doi: 10.1016/j.clnu.2014.05.013
- Lim, S.-M., and Im, D. S. (2009). Screening and characterization of probiotic lactic acid bacteria isolated from Korean fermented foods. *J. Microbiol. Biotechnol.* 19, 178–186. doi: 10.4014/jmb.0804.269
- Liu, C. F., and Pan, T. M. (2010). In vitro effects of lactic acid bacteria on cancer cell viability and antioxidant activity. *J. Food Drug Anal.* 18, 77–86.
- Lu, Y., Wang, W., Shan, Y., Zhiqiang, E., and Wang, L. (2009). Study on the inhibition of fermented soybean to cancer cells. *J. Northeast Agric. Univ.* 16, 25–28.
- Lv, X. C., Huang, X. L., Zhang, W., Rao, P. F., and Ni, L. (2013). Yeast diversity of traditional alcohol fermentation starters for Hong Qu glutinous rice wine brewing, revealed by culture-dependent and culture-independent methods. *Food Control* 34, 183–190. doi: 10.1016/j.foodcont.2013.04.020
- Macouzet, M., Lee, B. H., and Robert, N. (2009). Production of conjugated linoleic acid by probiotic *Lactobacillus acidophilus* La-5. *J. Appl. Microbiol.* 106, 1886–1891. doi: 10.1111/j.1365-2672.2009.04164.x
- Maintz, L., and Novak, N. (2007). Histamine and histamine intolerance. *Am. J. Clin. Nutr.* 85, 1185–1196.
- Martinez-Villaluenga, C., Peñas, E., Sidro, B., Ullate, M., Frias, J., and Vidal-Valverde, C. (2012). White cabbage fermentation improves ascorbin content, antioxidant and nitric oxide production inhibitory activity in LPS-induced macrophages. *LWT-Food Sci. Technol.* 46, 77–83. doi: 10.1016/j.lwt.2011.10.023
- Meerak, J., Lida, H., Watanabe, Y., Miyashita, M., Sato, H., Nakagawa, Y., et al. (2007). Phylogeny of poly- γ -glutamic acid-producing *Bacillus* strains isolated from fermented soybean foods manufactured in Asian countries. *J. Gen. Appl. Microbiol.* 53, 315–323. doi: 10.2323/jgam.53.315
- Meira, S. M. M., Daroit, D. J., and Helfer, V. E. (2012). Bioactive peptides in water soluble extract of ovine cheese from southern Brazil and Uruguay. *Food Res. Int.* 48, 322–329. doi: 10.1016/j.foodres.2012.05.009
- Meyer, J., Butikofer, U., Walther, B., Wechsler, D., and Sieber, R. (2009). Hot topic: changes in angiotensin-converting enzyme inhibition and concentration of the teripeptides Val-Pro-Pro and Ile-Pro-Pro during ripening of different Swiss cheese varieties. *J. Dairy Sci.* 92, 826–836. doi: 10.3168/jds.2008-1531
- Meyer, K., Kushi, L., Jacobs, D., Slavin, J., Sellers, T., and Folsom, A. (2000). Carbohydrates, dietary fiber, and incidence of type 2 diabetes in older women. *Am. J. Clin. Nutr.* 71, 921–930.
- Mine, Y., Wong, A. H. K., and Jiang, B. (2005). Fibrinolytic enzymes in Asian traditional fermented foods. *Food Res. Int.* 38, 243–250. doi: 10.1016/j.foodres.2004.04.008
- Mitra, S., Chakrabarty, P. K., and Biswas, S. R. (2010). Potential production and preservation of dahi by *Lactococcus lactis* W8, a nisin-producing strain. *LWT-Food Sci. Technol.* 43, 337–342. doi: 10.1016/j.lwt.2009.08.013
- Mo, H., Zhu, Y., and Chen, Z. (2008). Review. Microbial fermented tea – a potential source of natural food preservatives. *Trends Food Sci. Technol.* 19, 124–130. doi: 10.1016/j.tifs.2007.10.001
- Mogensen, G., Salminen, S., O'Brien, J., Ouwehand, A., Holzapfel, W., Shortt, C., et al. (2002). Inventory of micro-organisms with a documented history of use in food. *Bulletin* 377, 10–19.
- Mohania, D., Kansal, V. K., Sagwal, R., and Shah, D. (2013). Anticarcinogenic effect of probiotic dahi and piroxicam on DMH-induced colorectal carcinogenesis in Wister rats. *Am. J. Cancer Ther. Pharmacol.* 1, 8–24.

- Moktan, B., Saha, J., and Sarkar, P. K. (2008). Antioxidant activities of soybean as affected by *Bacillus*-fermentation to Kinema. *Food Res. Int.* 4, 586–593.
- Monteagudo-Mera, A., Rodríguez-Aparicio, L., Rúa, J., Martínez-Blanco, H., Navasa, N., García-Armesto, M. R., et al. (2012). In vitro evaluation of physiological probiotic properties of different lactic acid bacteria strains of dairy and human origin. *J. Funct. Foods* 4, 531–541. doi: 10.1016/j.jff.2012.02.014
- Montriwong, A., Kaewphuak, S., Rodtong, S., and Roytrakul, S. (2012). Novel fibrinolytic enzymes from *Virgibacillus halodenitrificans* SK1-3-7 isolated from fish sauce fermentation. *Process. Biochem.* 47, 2379–2387. doi: 10.1007/s12010-015-1591-5
- Moslehshad, M., Ehsani, M. R., and Salami, M. (2013). The comparative assessment of ACE-inhibitory and antioxidant activities of peptide fractions obtained from fermented camel and bovine milk by *Lactobacillus rhamnosus* PTCC1637. *Int. Dairy Res.* 29, 82–87. doi: 10.1016/j.idairyj.2012.10.015
- Murata, M., Houdai, T., Yamamoto, H., Matsumori, M., and Oishi, T. (2006). Membrane interaction of soyasaponins in association with their antioxidation effect – analysis of biomembrane interaction. *Soy Protein Res.* 9, 82–86.
- Nagai, T. (2015). “Health benefits of Natto,” in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York, NY: CRC Press), 433–453.
- Nagai, T., and Tamang, J. P. (2010). “Fermented soybeans and non-soybeans legume foods,” in *Fermented Foods and Beverages of the World*, eds J. P. Tamang and K. Kailasapathy (New York, NY: CRC Press), 191–224.
- Nakajima, N., Nozaki, N., Ishihara, K., Ishikawa, A., and Tsuji, H. (2005). Analysis of isoflavone content in tempeh: a fermented soybean product, and preparation of a new isoflavone-enriched tempeh. *J. Biosci. Bioeng.* 100, 685–687. doi: 10.1263/jbb.100.685
- Nakamura, Y., Masuda, O., and Takano, T. (1996). Decrease of tissue angiotensin I-converting enzyme activity upon feeding sour milk in spontaneously hypertensive rats. *Biosci. Biotechnol. Biochem.* 60, 488–489. doi: 10.1271/bbb.60.488
- Nikkuni, S., Karki, T. B., Vilku, K. S., Suzuki, T., Shindoh, K., Suzuki, C., et al. (1995). Mineral and amino acid contents of kinema, a fermented soybean food prepared in Nepal. *Food Sci. Technol. Int.* 1, 107–111. doi: 10.3136/fsti9596t9798.1.107
- Nishito, Y., Osana, Y., Hachiya, T., Popendorf, K., Toyoda, A., Fujiyama, A., et al. (2010). Whole genome assembly of a natto production strain *Bacillus subtilis* natto from very short read data. *BMC Genomics* 11:243. doi: 10.1186/1471-2164-11-243
- Nout, M. J. R. (1994). Fermented foods and food safety. *Food Res. Int.* 27, 291–298. doi: 10.1016/0963-9969(94)90097-3
- Nout, M. J. R., and Aidoo, K. E. (2002). “Asian fungal fermented food,” in *The Mycota*, ed. H. D. Osiewacz (New York: Springer-Verlag), 23–47.
- Nurrahman, Astuti, M., Suparmo, M., and Soesatyo, H. N. E. (2013). The role of black soybean tempe in increasing antioxidant enzyme activity and human lymphocyte proliferation in vivo. *Int. J. Curr. Microbiol. Appl. Sci.* 2, 316–327.
- Okada, N. (1989). Role of microorganism in tempeh manufacture. Isolation of vitamin B12 producing bacteria. *Japan Agric. Res. Q.* 22, 310–316.
- Omizu, Y., Tsukamoto, C., Chettri, R., and Tamang, J. P. (2011). Determination of saponin contents in raw soybean and fermented soybean foods of India. *J. Sci. Indus. Res.* 70, 533–538.
- Oppermann-Sanio, F. B., and Steinbüchel, A. (2002). Occurrence, functions and biosynthesis of polyamides in microorganisms and biotechnological productions. *Naturwissenschaften* 89, 11–22. doi: 10.1007/s00114-001-0280-0
- Orel, R., and Trop, T. K. (2014). Intestinal microbiota, probiotics and prebiotics in inflammatory bowel disease. *World J. Gastroenterol.* 20, 11505–11524. doi: 10.3748/wjg.v20.i33.11505
- Otes, S., and Cagindi, O. (2003). Kefir: a probiotic dairy-composition, nutritional and therapeutic aspects. *Pakistan J. Nutri.* 2, 54–59. doi: 10.3923/pjn.2003.54.59
- Ouwehand, A. C., Salminen, S., and Isolauri, E. (2002). Probiotics: an overview of beneficial effects. *Antonie Van Leeuwen* 82, 279–289. doi: 10.1023/A:1020620607611
- Papadimitriou, C. G., Vafopoulou-Mastrojiannaki, A., Silva, S. V., Gomes, A. M., Malcata, F. X., and Alichanidis, E. (2007). Identification of peptides in traditional and probiotic sheep milk yoghurt with angiotensin I-converting enzyme (ACE)-inhibitory activity. *Food Chem.* 105, 647–656. doi: 10.1016/j.foodchem.2007.07.011
- Park, J. A., Tirupathi Pichiah, P. B., Yu, J. J., Oh, S. H., Daily, J. W. III, and Cha, Y. S. (2012). Anti-obesity effect of kimchi fermented with *Weissella koreensis* OK1-6 as starter in high-fat diet-induced obese C57BL/6J mice. *J. Appl. Microbiol.* 113, 1507–1516. doi: 10.1111/jam.12017
- Park, J. E., Moon, Y. J., and Cha, Y. S. (2008). Effect of functional materials producing microbial strains isolated from Kimchi on antiobesity and inflammatory cytokines in 3T3-L1 preadipocytes. *FASEB J.* 23:111.
- Park, J. M., Shin, J. H., Gu, J. G., Yoon, S. J., Song, J. C., Jeon, W. M., et al. (2011). Effect of antioxidant activity in kimchi during a short-term and over-ripening fermentation period. *J. Biosci. Bioeng.* 112, 356–359. doi: 10.1016/j.jbiosc.2011.06.003
- Park, K. Y., Jeong, J. K., Lee, Y. E., and Daily, J. W. III (2014). Health benefits of kimchi (Korean fermented vegetables) as a probiotic food. *J. Med. Foods* 17, 6–20. doi: 10.1089/jmf.2013.3083
- Patel, A., Prajapati, J. B., Holst, O., and Ljungh, A. (2014). Determining probiotic potential of exopolysaccharide producing LAB isolated from vegetables and traditional Indian fermented food products. *Food Biosci.* 5, 27–33. doi: 10.1016/j.foodbi.2013.10.002
- Pattanagul, P., Pinthong, R., Phianmongkhol, A., and Tharatha, S. (2008). Mevinolin, citrinin and pigments of adlay angkak fermented by *Monascus* sp. *Int. J. Food Microbiol.* 126, 20–23. doi: 10.1016/j.ijfoodmicro.2008.04.019
- Paucar-Menacho, L. M., Amaya-Farfan, J., Berhow, M. A., Mandarino, J. M. G., de Mejia, E., and Chang, Y. K. (2010). A high-protein soybean cultivar contains lower isoflavones and saponins but higher minerals and bioactive peptides than a low-protein cultivar. *Food Chem.* 120, 15–21. doi: 10.1016/j.foodchem.2009.09.062
- Pedone, C. A., Arnaud, C. C., Postaire, E. R., Bouley, C. F., and Reinert, P. (2000). Multicentric study of the effect of milk fermented by *Lactobacillus casei* on the incidence of diarrhoea. *Int. J. Clin. Pract.* 54, 568–571.
- Peñas, E., Limón, R. I., Vidal-Valverde, C., and Frias, J. (2013). Effect of storage on the content of indole-glucosinolate breakdown products and vitamin C of sauerkrauts treated by high hydrostatic pressure. *LWT-Food Sci. Technol.* 53, 285–289. doi: 10.1016/j.lwt.2013.01.015
- Perna, A., Intaglietta, I., Simonetti, A., and Gambacorta, E. (2013). Effect of genetic type and casein halotype on antioxidant activity of yogurts during storage. *J. Dairy Sci.* 96, 1–7. doi: 10.3168/jds.2012-5859
- Phelan, M., and Kerins, D. (2011). The potential role of milk derived peptides in cardiovascular diseases. *Food Funct.* 2, 153–167. doi: 10.1039/c1fo10017c
- Ping, S. P., Shih, S. C., Rong, C. T., and King, W. Q. (2012). Effect of isoflavone aglycone content and antioxidation activity in natto by various cultures of *Bacillus subtilis* during the fermentation period. *J. Nutri. Food Sci.* 2:153. doi: 10.4172/2155-9600.1000153
- Qian, B., Xing, M., Cui, L., Deng, Y., Xu, Y., Huang, M., et al. (2011). Antioxidant, antihypertensive, and immunomodulatory activities of peptide fraction from fermented skim milk with *Lactobacillus delbrueckii* ssp. *bulgaricus* LB340. *J. Dairy Res.* 78, 72–79. doi: 10.1017/S0022029910000889
- Quiros, A., Hernandez-Ledesma, B., Ramos, M., Amigo, L., and Recio, I. (2005). Angiotensin-converting enzyme inhibitory activity of peptides derived from caprine kefir. *J. Dairy Sci.* 88, 3480–3487. doi: 10.3168/jds.S0022-0302(05)73032-0
- Rabie, M. A., Siliha, H., El-Saidy, S., El-Badawy, A. A., and Malcata, F. X. (2011). Reduced biogenic amine contents in sauerkraut via addition of selected LAB. *Food Chem.* 129, 1778–1782. doi: 10.1016/j.foodchem.2011.05.106
- Ramadori, G., Gautron, L., Fujikawa, T., Claudia, R., Vianna, J., Elmquist, E., et al. (2009). Central administration of resveratrol improves diet-induced diabetes. *Endocrinology* 150, 5326–5333. doi: 10.1210/en.2009-0528
- Ramrez, J. F., Sanchez-Marroquin, A., Alvarez, M. M., and Valyasebi, R. (2004). “Industrialization of Mexican pulque,” in *Industrialization of Indigenous Fermented Foods*, 2nd Edn, ed. K. Steinkraus (New York, NY: Marcel Dekker), 547–586.
- Ranadheera, R., Baines, S., and Adams, M. (2010). Importance of food in probiotic efficacy. *Food Res. Int.* 43, 1–7. doi: 10.1016/j.foodres.2009.09.009
- Rauscher-Gabernig, E., Grossgut, R., Bauer, F., and Paulsen, P. (2009). Assessment of alimentary histamine exposure of consumers in Austria and development of tolerable levels in typical foods. *Food Control* 20, 423–429. doi: 10.1016/j.foodcont.2008.07.011

- Reddy, N. R., and Salunkhe, D. K. (1980). Effect of fermentation on phytate phosphorus, and mineral content in black gram, rice, and black gram and rice blends. *J. Food Sci.* 45, 1708–1712. doi: 10.1111/j.1365-2621.1980.tb07594.x
- Saad, N., Delattre, C., Urdaci, M., Schmitter, J. M., and Bressollier, P. (2013). An overview of the last advances in probiotic and prebiotic field. *LWT Food Sci. Technol.* 50, 1–16. doi: 10.1016/j.lwt.2012.05.014
- Sabeena, F. K. H., Baron, C. P., Nielsen, N. S., and Jacobsen, C. (2010). Antioxidant activity of yoghurt peptides: part 1-in vitro assays and evaluation in ω -3 enriched milk. *Food Chem.* 123, 1081–1089. doi: 10.1016/j.foodchem.2010.05.067
- Sanchez, P. C. (2008). *Philippine Fermented Foods: Principles and Technology*. (Quezon City: University of the Philippines Press), 511.
- Sarkar, P. K., Jones, L. J., Craven, G. S., and Somerset, S. M. (1997). Oligosaccharides profile of soybeans during kinema production. *Letts. Appl. Microbiol.* 24, 337–339. doi: 10.1046/j.1472-765X.1997.00035.x
- Sarkar, P. K., Morrison, E., Tingii, U., Somerset, S. M., and Craven, G. S. (1998). B-group vitamin and mineral contents of soybeans during kinema production. *J. Sci. Food Agric.* 78, 498–502. doi: 10.1002/(SICI)1097-0010(199812)78:4<498::AID-JSFA145>3.3.CO;2-3
- Sarkar, P. K., and Tamang, J. P. (1995). Changes in the microbial profile and proximate composition during natural and controlled fermentations of soybeans to produce kinema. *Food Microbiol.* 12, 317–325. doi: 10.1016/S0740-0020(95)80112-X
- Seppo, L., Kerokjoki, O., Suomalainen, T., and Korpela, R. (2002). The effect of a *Lactobacillus helveticus* LBK-16 H fermented milk on hypertension — a pilot study on humans. *Milchwissen* 57, 124–127.
- Shah, N. P. (2015). “Functional properties of fermented milks,” in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York, NY: CRC Press), 261–274.
- Shah, N. P., da Cruz, A. G., and Faria, J. D. A. F. (2013). *Probiotics and Probiotic Foods: Technology, Stability and Benefits to Human Health*. New York, NY: Nova Science Publishers.
- Shin, D. H., and Jeong, D. (2015). Korean traditional fermented soybean products: *Jang*. *J. Ethnic Foods* 2, 2–7. doi: 10.1016/j.jef.2015.02.002
- Shin, D. H., Jung, S. J., and Chae, S. W. (2015). “Health benefits of Korean fermented soybean products,” in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York, NY: CRC Press), 395–431.
- Shin, M. S., Han, S. K., Ryu, J. S., Kim, K. S., and Lee, W. K. (2008). Isolation and partial characterization of a bacteriocin produced by *Pediococcus pentosaceus* K23-2 isolated from kimchi. *J. Appl. Microbiol.* 105, 331–339. doi: 10.1111/j.1365-2672.2008.03770.x
- Shin, S. K., Kwon, J. H., Jeon, M., Choi, J., and Choi, M. S. (2011). Supplementation of Cheonggukjang and Red Ginseng Cheonggukjang can improve plasma lipid profile and fasting blood glucose concentration in subjects with impaired fasting glucose. *J. Med. Food* 14, 108–113. doi: 10.1089/jmf.2009.1366
- Shon, M. Y., Lee, J., Choi, J. H., Choi, S. Y., Nam, S. H., Seo, K. I., et al. (2007). Antioxidant and free radical scavenging activity of methanol extract of chungkukjang. *J. Food Compos. Anal.* 20, 113–118. doi: 10.1016/j.jfca.2006.08.003
- Singh, T. A., Devi, K. R., Ahmed, G., and Jeyaram, K. (2014). Microbial and endogenous origin of fibrinolytic activity in traditional fermented foods of Northeast India. *Food Res. Int.* 55, 356–362. doi: 10.1016/j.foodres.2013.11.028
- Sipola, M., Finckenberg, P., Korpela, R., Vapaatalo, H., and Nurminen, M. (2002). Effect of long-term intake of milk products on blood pressure in hypertensive rats. *J. Dairy Res.* 69, 103–111. doi: 10.1017/S002202990100526X
- Spano, G., Russo, P., Lonvaud-Funel, A., Lucas, P., Alexandre, H., Grandvalet, C., et al. (2010). Biogenic amine in fermented foods. *Eur. J. Clin. Nutr.* 64, 95–100. doi: 10.1038/ejcn.2010.218
- Stanley, N. R., and Lazazzera, B. A. (2005). Defining the genetic differences between wild and domestic strains of *Bacillus subtilis* that affect poly- γ -D-glutamic acid production and biofilm formation. *Mol. Microbiol.* 57, 1143–1158. doi: 10.1111/j.1365-2958.2005.04746.x
- Steinkraus, K. H. (1996). *Handbook of Indigenous Fermented Food*, 2nd Edn. New York, NY: Marcel Dekker, Inc.
- Suganuma, T., Fujita, K., and Kitahara, K. (2007). Some Distinguishable properties between acid-stable and neutral types of α -amylases from acid-producing koji. *J. Biosci. Bioeng.* 104, 353–362. doi: 10.1263/jbb.104.353
- Suzzi, G., and Gardini, F. (2003). Biogenic amines in dry fermented sausages: a review. *Int. J. Food Microbiol.* 88, 41–54. doi: 10.1016/S0168-1605(03)00080-1
- Syal, P., and Vohra, A. (2013). Probiotic potential of yeasts isolated from traditional Indian fermented foods. *Int. J. Microbiol. Res.* 5, 390–398. doi: 10.9735/0975-5276.5.2.390-398
- Szajewska, H., Skorka, A., Ruszczynski, M., and Gieruszczak-bialek, D. (2007). Meta-analysis: *Lactobacillus* GG for treating acute diarrhoea in children. *Aliment. Pharm. Therapeut.* 25, 871–881. doi: 10.1111/j.1365-2036.2007.03282.x
- Tamang, J. P. (2015). Naturally fermented ethnic soybean foods of India. *J. Ethnic Foods* 2, 8–17. doi: 10.1007/s12275-012-1409-x
- Tamang, J. P., Dewan, S., Tamang, B., Rai, A., Schillinger, U., and Holzapfel, W. H. (2007). Lactic acid bacteria in Hamei and Marcha of North East India. *Indian J. Microbiol.* 47, 119–125. doi: 10.1007/s12088-007-0024-8
- Tamang, J. P., and Fleet, G. H. (2009). “Yeasts diversity in fermented foods and beverages,” in *Yeasts Biotechnology: Diversity and Applications*, eds T. Satyanarayana and G. Kunze (New York: Springer), 169–198.
- Tamang, J. P., and Nikkuni, S. (1996). Selection of starter culture for production of kinema, fermented soybean food of the Himalaya. *World J. Microbiol. Biotechnol.* 12, 629–635. doi: 10.1007/BF00327727
- Tamang, J. P., and Nikkuni, S. (1998). Effect of temperatures during pure culture fermentation of Kinema. *World J. Microbiol. Biotechnol.* 14, 847–850. doi: 10.1023/A:1008867511369
- Tamang, J. P., Tamang, B., Schillinger, U., Guigas, C., and Holzapfel, W. H. (2009). Functional properties of lactic acid bacteria isolated from ethnic fermented vegetables of the Himalayas. *Int. J. Food Microbiol.* 135, 28–33. doi: 10.1016/j.ijfoodmicro.2009.07.016
- Tamang, J. P., Watanabe, K., and Holzapfel, W. H. (2016). Review: Diversity of microorganisms in global fermented foods and beverages. *Front. Microbiol.* 7:377. doi: 10.3389/fmicb.2016.00377
- Thapa, N., and Tamang, J. P. (2015). “Functionality and therapeutic values of fermented foods,” in *Health Benefits of Fermented Foods*, ed. J. P. Tamang (New York: CRC Press), 111–168.
- Tolhurst, G., Heffron, H., Lam, Y. S., Parker, H. E., Habib, A. M., Diakogiannaki, E., et al. (2012). Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. *Diabetes Metab. Res. Rev.* 61, 364–371. doi: 10.2337/db11-1019
- Tsubura, S. (2012). Anti-periodontitis effect of *Bacillus subtilis* (natto). *Shigaku (Odontol.)* 99, 160–164.
- Tsuyoshi, N., Fudou, R., Yamanaka, S., Kozaki, M., Tamang, N., Thapa, S., et al. (2005). Identification of yeast strains isolated from marcha in Sikkim, a microbial starter for amyolytic fermentation. *Int. J. Food Microbiol.* 99, 135–146. doi: 10.1016/j.ijfoodmicro.2004.08.011
- Urushibata, Y., Tokuyama, S., and Tahara, Y. (2002). Characterization of the *Bacillus subtilis* ywC gene, involved in (–)polyglutamic acid production. *J. Bacteriol.* 184, 337–343. doi: 10.1128/JB.184.2.337-343.2002
- US Probiotics Home (2011). Available at: www.usprobiotics.org
- Varankovich, N. V., Nickerson, M. T., and Korber, D. R. (2015). Probiotic-based strategies for therapeutic and prophylactic use against multiple gastrointestinal diseases. *Front. Microbiol.* 6:685. doi: 10.3389/fmicb.2015.00685
- Verna, E. C., and Lucak, S. (2010). Use of probiotics in gastrointestinal disorders: what to recommend? *Ther. Adv Gastroenterol.* 3, 307–319. doi: 10.1177/1756283X10373814
- Visciano, P., Schirone, N., Tofalo, R., and Suzzi, G. (2014). Histamine poisoning and control measures in fish and fishery products. *Front. Microbiol.* 5:500. doi: 10.3389/fmicb.2014.00500
- Walker, G. M. (2014). “Microbiology of winemaking,” in *Encyclopaedia of Food Microbiology*, 2 Edn, eds C. Batt and M. A. Tortorello (Oxford: Elsevier Ltd.), 787–792.
- Wang, L. J., Li, D., Zou, L., Chen, X. D., Cheng, Y. Q., Yamaki, K., et al. (2007a). Antioxidative activity of douchi (a Chinese traditional salt-fermented soybean food) extracts during its processing. *Int. J. Food Propert.* 10, 1–12. doi: 10.1080/10942910601052715
- Wang, L.-J., Yin, L.-J., Li, D., Zou, L., Saito, M., Tatsumi, E., et al. (2007b). Influences of processing and NaCl supplementation on isoflavone contents and composition during douchi manufacturing. *Food Chem.* 101, 1247–1253. doi: 10.1016/j.foodchem.2006.03.029
- Weill, F. S., Cela, E. M., Paz, M. L., Ferrari, A., Leoni, J., and Gonzalez Maglio, D. H. (2013). Lipoteichoic acid from *Lactobacillus rhamnosus* GG as an oral

- photoprotective agent against UV-induced carcinogenesis. *Br. J. Nutri.* 109, 457–466. doi: 10.1017/S0007114512001225
- Willcox, B. J., Willcox, D. C., and Suzuki, M. (2004). *The Okinawa Diet Plan*. New York, NY: Three Rivers Press.
- Won, T. J., Kim, B., Song, D. S., Lim, Y. T., Oh, E. S., Lee, D. I., et al. (2011). Modulation of Th1/Th2 balance by *Lactobacillus* strains isolated from kimchi via stimulation of macrophage cell line J774A.1 *in vitro*. *J. Food Sci.* 76, H55–H61. doi: 10.1111/j.1750-3841.2010.02031.x
- Yadav, H., Jain, S., and Sinha, P. R. (2007). Antidiabetic effect of probiotic dahi containing *Lactobacillus acidophilus* and *Lactobacillus casei* in high fructose fed rats. *Nutrition* 23, 62–68. doi: 10.1016/j.nut.2006.09.002
- Yanagisawa, Y., and Sumi, H. (2005). Natto bacillus contains a large amount of water-soluble vitamin K (menaquinone-7). *J. Food Biochem.* 29, 267–277. doi: 10.1111/j.1745-4514.2005.00016.x
- Yanping, W., Nv, X., Aodeng, X., Zaheer, A., Bin, Z., and Xiaojia, B. (2009). Effects of *Lactobacillus plantarum* MA2 isolated from Tibet kefir on lipid metabolism and intestinal microflora of rats fed on high-cholesterol diet. *Appl. Microbiol. Biotechnol.* 84, 341–347. doi: 10.1007/s00253-009-2012-x
- Yin, L. J., Li, D., Zou, L., Saito, M., Tatsumi, E., and Li, L. T. (2007). Influences of processing and NaCl supplementation on isoflavone contents and composition during douchi manufacturing. *Food Chem.* 101, 1247–1253.
- Yoon, S., Do, J., Lee, S., and Chag, H. (2000). Production of poly- δ -glutamic acid by fed-batch culture of *Bacillus licheniformis*. *Biotechnol. Lett.* 22, 585–588. doi: 10.1023/A:1005625026623
- Zeng, W., Li, W., Shu, L., Yi, J., Chen, G., and Liang, Z. (2013). Non-sterilized fermentative co-production of poly (γ -glutamic acid) and fibrinolytic enzyme by a thermophilic *Bacillus subtilis* GXA-28. *Bioresour. Technol.* 142, 697–700. doi: 10.1016/j.biortech.2013.05.020
- Zhai, H., Yang, X., Li, L., Xia, G., Cen, J., Huang, H., et al. (2012). Biogenic amines in commercial fish and fish products sold in southern china. *Food Control* 25, 303–308. doi: 10.1016/j.foodcont.2011.10.057
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