



Opposing roles for heat and heat shock proteins in macrophage functions during inflammation: a function of cell activation state?

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Macrophages function both under normothermia and during periods of body temperature elevation (fever). Whether macrophages sense and respond to thermal signals in a manner which regulates their function in a specific manner is still not clear. In this brief review, we highlight recent studies which have analyzed the effects of mild heating on macrophage cytokine production, and summarize thermally sensitive molecular mechanisms, such as heat shock protein (HSP) expression, which have been identified. Mild, physiologically achievable, hyperthermia has been shown to have *both pro- and anti-inflammatory effects on macrophage inflammatory cytokine production* and overall it is not clear how hyperthermia or HSPs can exert opposing roles on macrophage function. We propose here that the stage of activation of macrophages predicts how they respond to mild heating and the specific manner in which HSPs function. Continuing research in this area is needed which will help us to better understand the immunological role of body temperature shifts. Such studies could provide a scientific basis for the use of heat in treatment of inflammatory diseases.

Keywords: inflammation, hyperthermia, heat shock protein, fever, heat shock factor, cytokines, arthritis

Inflammation is usually considered a beneficial event which helps to heighten the immune response initiated following tissue injury or infection. Molecular alarm signals generated from the damaged tissues or invading pathogens are recognized by antigen-presenting cells (APCs) such as macrophages. Macrophages are key players in innate immunity and respond rapidly to danger signals generated from inflamed sites. Recognition of these danger signals through specific cell surface pattern recognition receptors (PRRs) on macrophages results in the production of anti-microbial products as well as pro-inflammatory mediators (Depraetere, 2000; Fujiwara and Kobayashi, 2005).

Inflammation is a tightly regulated process with production of alarm signals sharply diminishing after the pathogens or cellular debris have been eliminated and tissue homeostasis restored. Many “stop signals” at appropriate checkpoints prevent further leukocyte infiltration into tissues if they are no longer needed (Serhan et al., 2007). If the balance between inflammation and resolution becomes dysregulated, excess macrophage responses will lead to chronic inflammation, and resultant disease states, which include atherosclerosis, rheumatoid arthritis (RA), asthma (Nathan, 2002), and cancer (Grivennikov et al., 2010). Therefore, understanding the regulation of inflammation and its resolution could help in the development of new therapeutic approaches for inflammatory disease management.

Macrophages are optimally activated by a combination of two macromolecular signals in their environment: antigen recognition by PRPs and IFN- γ (Mosser, 2003; O’Shea and Murray, 2008) and

These pro-inflammatory cytokines are important pyrogens capable of inducing the formation of the fever response and elevation of body temperature through complex neurological and behavioral changes (Kluger, 1991a,b; Cooper et al., 1994; Leon et al., 1997; Romanovsky et al., 1998, 2005). Fever is a highly conserved phenomenon in evolution (Kluger, 1979a, 1986; Mackowiak, 1981; Hasday et al., 2000). Although numerous human and animal studies reveal a positive correlation between the febrile response and a better survival rate, the mechanism by which increased core temperature improves host defense is still not clear. In addition, new studies suggest additional levels of complex interaction between macrophage function and body temperature. Nguyen et al. (2011) have shown that exposure to a cold environment induces alternative activation of adipose tissue macrophages. Thus, while it is likely that their function is responsive to thermal signals, the specific mechanisms by which macrophages sense and respond to temperature change in a way that specifically regulates their function has not been clarified.

To investigate the effect of febrile temperatures on macrophage activity, researchers often simply increase temperature in cell cultures, or elevate body temperature by external heat application in murine models. While these procedures do not reproduce critical features of actual fevers, several studies reveal both *positive* (Jiang et al., 2000b; Ostberg et al., 2000; Lee et al., 2012) and *negative* (Ensor et al., 1994; Fairchild et al., 2000; Hagiwara et al., 2007) effects on macrophage pro-inflammatory cytokine production. Both positive and negative effects of hyperthermia

have been correlated with an induction of heat shock proteins (HSPs). The overall aim of this brief review is to present previous studies that have studied the impact of hyperthermia and HSPs on macrophage functions and to summarize available information regarding the underlying molecular mechanisms which may mediate this complex interaction of thermal signals.

FEVER-RANGE TEMPERATURES CAN ENHANCE MACROPHAGE PRO-INFLAMMATORY CYTOKINE PRODUCTION

Inducing hyperthermia has been used to study the role of febrile temperatures on the host defense system (Hasday et al., 2000; Jiang et al., 2000a). It was shown that increasing core body temperature has a protective role in the outcome of infection; the improved survival seen was determined not to be simply due to thermal suppression of bacterial growth. Data published by our laboratory (Ostberg et al., 2000) as well as by others (Jiang et al., 1999b) shows that mild, systemic heating at a target temperature of 39.5°C significantly enhances the concentration of TNF- α (3-fold increase in sera; 2.5-fold increase in liver) and IL-6 (4-fold increase in sera; 2.6-fold increase in spleen, 3.4-fold in lung, and 15-fold in liver) in the blood and tissues of BALB/c mice challenged with bacteria endotoxin (LPS). In addition, Jiang et al. (1999a) identified hepatic Kupffer cells as the predominant source of excess TNF- α secretion while multiple organs including lung, spleen, and liver could produce excess IL-6 in the warmer animals. These temperature-induced changes in cytokine expression are associated with an induction of HSP72 in the liver.

Our laboratory has recently observed that mild elevation of body temperature not only significantly enhances subsequent LPS-induced release of TNF- α (3-fold increase), but also reprograms macrophages, resulting in sustained subsequent responsiveness to LPS, i.e., this treatment reduces “endotoxin tolerance” *in vitro* and *in vivo* (Lee et al., 2012). Heat treatment results in an increase in LPS-induced downstream signaling including enhanced phosphorylation of I κ B kinase (IKK) and I κ B, NF- κ B nuclear translocation and binding to the TNF- α promoter in macrophages upon secondary stimulation. The induction of HSP70 is important for mediation of thermal effects on macrophage function (Lee et al., 2012). Our *in vitro* experiments also show that the production of nitric oxide (NO) and inducible NO synthase (iNOS) by peritoneal macrophages is increased by exposure to febrile temperature together with LPS and IFN- γ stimulation. This result is correlated with the presence of HSP70 in the heat-treated macrophages (Pritchard et al., 2005). Collectively, these data suggest that fever-range hyperthermia can enhance macrophage cytokine expression and HSP70 expression, which in turn may help to improve host defense in response to infection.

FEVER-RANGE TEMPERATURES CAN ALSO SUPPRESS MACROPHAGE PRO-INFLAMMATORY CYTOKINE PRODUCTION

In contrast to the research summarized above, other studies using the macrophage cell lines RAW264.7 or human

monocyte-derived macrophages have shown that hyperthermia has anti-inflammatory effects and suppresses activated macrophage pro-inflammatory cytokine expression (TNF- α , reduced by 50–98%; IL-6, reduced by 83–87%; and IL-1 β , reduced by 50–94%; Ensor et al., 1994; Fairchild et al., 2000; Hagiwara et al., 2007). This inhibition is linked to a marked reduction in cytokine gene transcription and mRNA stability (Ensor et al., 1995). In addition, this thermally suppressed cytokine production is mediated by the binding of heat shock factor (HSF)-1, a transcriptional repressor, to the heat shock response element in the cytokine promoter region, including IL-1 β and TNF- α (Cahill et al., 1996; Singh et al., 2002). Recently, Cooper et al. (2010a,b) have shown that fever-range temperatures selectively reduce LPS-induced recruitment of NF- κ B and Sp-1 transcription factors to the TNF- α promoter regions.

Heat-induced suppression may involve high mobility group box 1 (HMGB1), an intra-nuclear protein that can be released by activated macrophages, necrotic or damaged cells during inflammation. HMGB1 helps immune cells to recognize damaged tissues and initiates intracellular signaling to activate NF- κ B and pro-inflammatory cytokine production (Fiuza et al., 2003). Hyperthermia has been shown to inhibit macrophage HMGB1 secretion following LPS stimulation (Fairchild et al., 2000) and Hagiwara et al. (2007) found that high fever temperature (40°C) enhances HSF-1 and HSP70 expression. These increased levels of HSF-1 and HSP70 may reduce HMGB1 secretion and subsequent NF- κ B activation and cytokine production. In general, these data support the concept that febrile temperatures can also have inhibitory effects on macrophage cytokine gene expression, an effect correlated with the activation of HSF-1 or induction of HSP70.

REGULATION OF HEAT SHOCK PROTEINS

Heat shock proteins are evolutionarily conserved proteins that can be induced by stress signals, including environmental stresses (e.g., heat shock), and pathophysiological states (e.g., fever, inflammation, and infection) as well as those induced by normal development stresses (Morimoto, 1993, 1998; Wu, 1995).

Heat shock proteins function as chaperones to assist with protein folding in order to protect cells from protein denaturation or cell death under stress conditions (Fink, 1999; Jaattela, 1999). Although HSPs are considered to be intracellular proteins, they can be mobilized to the plasma membrane or released into the extracellular environment and have immunomodulatory functions (Johnson and Fleshner, 2006). HSPs (e.g., HSP60, HSP70, HSP90, gp96, etc.) can be released from various cells through either a passive (during cell injury) or an active (translocation to the plasma membrane and then secretion) pathway. Previous studies have shown that HSP70 is released into the extracellular environment in a membrane-associated form after heat stress (Multhoff, 2007; Vega et al., 2008). HSPs are known to have both positive and negative effects in regulating macrophage function and this may depend on the cellular location of these HSPs. It is proposed that extracellular HSPs might serve as a danger signal to stimulate the immune response, whereas intracellular HSPs could serve as a negative regulator to control the inflammation (Schmitt et al., 2007).

PRO-INFLAMMATORY ROLE OF HEAT SHOCK PROTEINS ON MACROPHAGE FUNCTION

Previous studies have shown that extracellular HSPs exert immunostimulatory effects (Johnson and Fleshner, 2006). Wang et al. (2006) have shown that extracellular HSP70 binds to the lipid raft microdomain on the plasma membrane of macrophages and enhances their phagocytic ability. This HSP70-mediated phagocytosis enhances the processing and presentation of internalized antigens to CD4 T cells. Furthermore, extracellular HSPs can robustly stimulate the release of TNF- α , IL-6, IL-1 β , IL-12, NO, as well as chemokines by monocytes/macrophages (Lehner et al., 2000; Asea et al., 2002; Panjwani et al., 2002; Vega et al., 2008). This effect is mediated through the CD14/TLR (both TLR2 and TLR4) complexes which lead to the activation of downstream NF- κ B and MAPK pathway (Asea et al., 2000; Kol et al., 2000; Vabulas et al., 2002). In addition, HSPs are actively synthesized in peritoneal macrophages in mice which are administrated with LPS (Zhang et al., 1994). HSP70 and HSP90 have been shown to be involved in the innate recognition of bacterial products. These HSPs are able to bind LPS and form a cluster with TLR4–MD2 within lipid raft to deliver LPS to the complex. Following stimulation, these HSPs further assist the trafficking and targeting of this complex to the Golgi apparatus (Triantafilou et al., 2001; Triantafilou and Triantafilou, 2004). These results indicate that elevation of extracellular HSPs may serve as endogenous danger signals to alert the host defense system through their cytokine-like function.

ANTI-INFLAMMATORY ROLE OF HEAT SHOCK PROTEINS ON MACROPHAGE CYTOKINE EXPRESSION

On the other hand, intracellular HSPs have been shown to have anti-inflammatory roles in suppressing macrophage cytokine production. Intracellular HSPs are involved in protecting the organism from a variety of insults by directly interfering with cell death pathway and suppressing the expression of inflammatory genes (Yenari et al., 2005). The protect roles of stress-inducible intracellular HSPs (such as HSP72) in lethal sepsis and infection are well known. Induction of HSP70 *in vitro* by heat shock response or through overexpression can reduce mortality in experimental models of septic shock and endotoxemia as well as down-regulate inflammatory gene expression (Snyder et al., 1992; Hotchkiss et al., 1993; Villar et al., 1994; Van Molle et al., 2002; Shi et al., 2006). It has been shown that intracellular HSP70 can interact with IKK, prevent I κ B phosphorylation, and NF- κ B activation (Ran et al., 2004). In addition, HSP70 is actively synthesized in macrophages after exposure to endotoxin. Intracellular HSP70 inhibits LPS-induced NF- κ B activation by binding with TRAF6, the important adaptor protein downstream of TLR4 and preventing its ubiquitination (Chen et al., 2006). These results suggest that intracellular HSP70 may act as a suppressor to interfere NF- κ B signaling and downstream inflammatory cytokine production. Furthermore, HSP72 has been shown to inhibit LPS- and TNF- α -induced HMGB1 release and subsequent pro-inflammatory cytokine production in macrophages (Tang et al., 2007).

On the other hand, there is only one study by Bouchama et al. (2000) showing that hyperthermia (using target temperature 39, 41, and 43°C) inhibits LPS-stimulated IL-10 production

by mononuclear cells isolated from healthy donor as compared to cells maintained at 43°C. In addition, by over-expressing HSP70 in human monocyte-derived macrophages, Ding et al. (2001) found that LPS-induced IL-10 production was significantly inhibited. In our unpublished data, we tried to address whether fever-range temperature affects macrophage IL-10 production at different activation stages. However, we found that temperature did not have any effect on IL-10 expression by either naïve macrophages or previously activated macrophages. Taken together, these studies suggest that intracellular HSPs may exert negative regulatory effects to dampen the inflammatory response in order to prevent tissue damage.

HEAT SHOCK PROTEINS AND INFLAMMATORY DISORDERS

There is some evidence that HSPs may play a role as immunological targets in chronic inflammatory diseases, such as RA (van Eden et al., 2005) and atherosclerosis (Xu, 2002). Increased HSP60 and HSP70 expression have been identified in the synovial tissues from animals with experimental induced arthritis and patients with RA (de Graeff-Meeder et al., 1990; Boog et al., 1992; Martin et al., 2003). Importantly, arthritis disease severity depends on the ability of human HSP60 and HSP70 to induce IL-10 production by monocytes/macrophages and fibroblast-like synoviocytes (MacHt et al., 2000; Luo et al., 2008). The production of IL-10 is negatively correlated with arthritis progression and joint destruction (Isomaki et al., 1996; Tanaka et al., 1996). Several clinical trials in RA patients using immunotherapies involving HSPs have shown to promote anti-inflammatory cytokine production and ameliorate disease severity, indicating that HSPs have immunoregulatory potential (Vischer, 1990; Rosenthal et al., 1991; Prakken et al., 2004).

On the other hand, HSP60 and HSP70 are also found in atherosclerotic lesions (Berberian et al., 1990; Kleindienst et al., 1993; Johnson et al., 1995). During the past decade, it has been noted that infections (e.g., chlamydiae infection) might contribute to the pathogenesis of atherosclerosis to directly stimulate cells of the arterial wall and/or other tissues to express high levels of HSPs (Kiechl et al., 2001). Chlamydial HSP60 (cHSP60) colocalizes with human HSP60 within macrophages in atherosclerotic lesions (Kol et al., 1998). Both chlamydial and human HSPs stimulate the expression of pro-inflammatory cytokines and adhesion molecules by macrophages through TLR4/MD2-dependent pathway (Bulut et al., 2002). Because of the high homology between chlamydial and human HSPs, it is possible that cross-reactions of antibodies and T cells against HSPs between microbes and humans contribute to the development of atherosclerosis (Xu, 2002). Taken together, a better understanding of the role of HSPs in the inflammatory process may lead to the development of new therapies to modify aberrant immune responses.

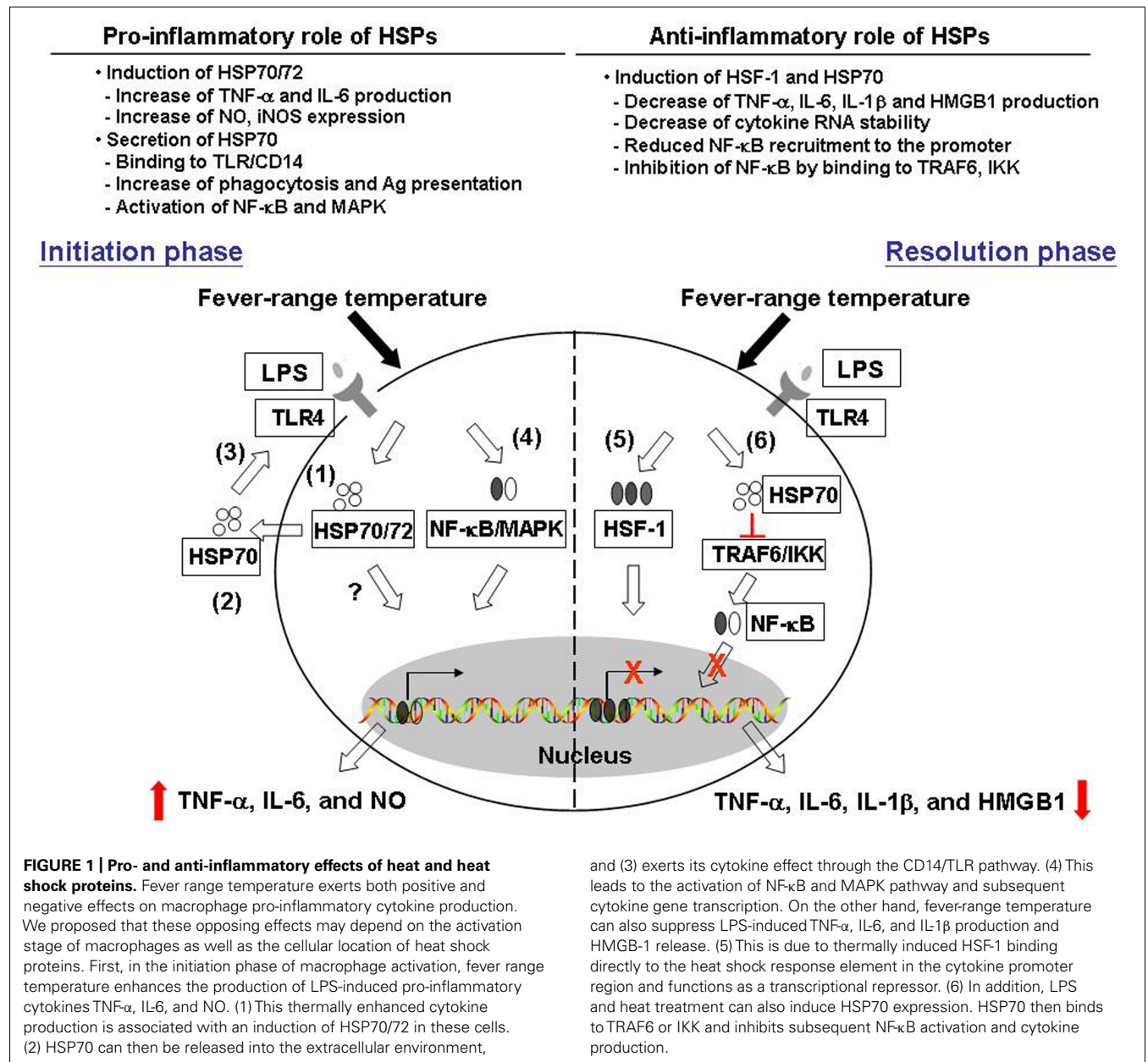
DOES THE MATURATION STATE OF MACROPHAGES INFLUENCE THE EFFECT OF HEAT AND HEAT SHOCK PROTEINS?

In this paper, we described previous studies revealing contrasting effects of febrile temperatures and HSPs on macrophage functions. Mild hyperthermia *enhances* macrophage TNF- α IL-6 as well as NO production in a HSP70/72-dependent mechanism.

On the other hand, mild hyperthermia *suppresses* macrophage pro-inflammatory cytokine production and this may involve an increase in HSF-1 and HSP70 expression. Thermally induced HSF-1 and HSP70 also play a role in blocking the transcription factors NF-κB and Sp-1 recruitment to the cytokine promoter and subsequent cytokine secretion (see **Figure 1**, for possible pathways by which thermal stress may result in both pro- and anti-inflammatory effects).

One possible explanation for these opposing results is that the resultant effect of heat is determined by the specific activation state of macrophages used for the analysis. For example, enhancing effects of heating were observed using cells freshly isolated from mice early after LPS injection (Jiang et al., 1999a,b; Lee et al., 2012). These cells express a naïve phenotype. On the

other hand, the suppressing effects were obtained using RAW264.7 cells which express an activated phenotype (Ensor et al., 1994; Fairchild et al., 2000; Hagiwara et al., 2007). Therefore, we propose that in cells associated with the early activation stage of inflammation, febrile temperature may stimulate macrophage cytokine production, which helps to eliminate invading pathogens, whereas if heat is applied to cells associated with the resolution phase, a suppression of macrophage cytokine production occurs to prevent further tissue damage. This idea, if correct, would suggest that one of the functions of natural fever may be to help resolve inflammation. Furthermore, to our knowledge, there is no previous study addressing the role of heat/HSP on alternatively activated macrophage functions. This indicates that more research is needed to identify the mechanistic



basis of the contrasting effects of hyperthermia on macrophage function.

CONCLUSION

The host inflammatory response to injury or infection is clearly a complex process. Pro-inflammatory cytokines secreted by activated macrophages are essential for successful host defense. However, uncontrolled inflammatory cytokine production will also cause tissue damage. The febrile response is a well recognized component of inflammation and has been shown to provide a beneficial effect for the host but whether the thermal element of fever has a direct role in regulating macrophage function is still not clear. By using hyperthermia, many studies have reported that febrile temperatures can regulate macrophage pro-inflammatory cytokine production both *in vitro* and *in vivo*. These effects are associated with the induction of HSP. However, these effects can be either positive or negative and much more research is needed to understand how these two contrasting effects coincide.

In addition, we also summarized research which shows both the pro- and anti-inflammatory roles of HSPs. Extracellular HSPs might serve as a danger signal to stimulate the immune response, whereas intracellular HSPs could serve as a negative regulator to control the inflammation. Studies of the roles and mechanisms

of heat as well as HSPs in inflammatory disorders could provide more information for therapeutic strategies targeting the molecular levels.

Finally, it is important to remember that most of the research on the role of elevated temperature on macrophage function does not actually use a physiological fever (i.e., induced by endotoxin or other pyrogens), but instead, simply forces the elevation of body temperature using external heating. While some studies described above do include the addition of LPS, or use heat in the presence of infectious antigens, fever involves many biochemical, neurological, and vascular changes not seen in conditions of forced hyperthermia (Kluger, 1979b, 1991a; Gordon, 1993; Romanovsky et al., 1998, 2005). Therefore, it is possible that the elevated temperature associated with actual fever may have additional and more complex regulatory effects on macrophage function.

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