# Epigenetic: a molecular link between testicular cancer and environmental exposures

Aurelie Vega<sup>1,2,3,4</sup>, Marine Baptissart<sup>1,2,3,4</sup>, Françoise Caira<sup>1,2,3,4</sup>, Florence Brugnon<sup>1,2,3,4</sup>, Jean-Marc A. Lobaccaro<sup>1,2,3,4</sup> and David H. Volle<sup>1,2,3,4</sup>\*

- <sup>1</sup> Génétique Reproduction et Développement, INSERM U 1103, Aubière, France
- <sup>2</sup> Génétique Reproduction et Développement, Clermont Université, Université Blaise Pascal, Clermont-Ferrand, France
- <sup>3</sup> Génétique Reproduction et Développement, CNRS, UMR 6293, Aubière, France
- <sup>4</sup> Centre de Recherche en Nutrition Humaine d'Auvergne, Clermont-Ferrand, France

#### Edited by:

Alberto Ferlin, University of Padova, Italy

#### Reviewed by:

Andrea Garolla, University of Padova, Italy Donatella Paoli, University of Rome La Sapienza, Italy

#### \*Correspondence:

David H. Volle, Génétique Reproduction et Développement, INSERM U 1103, CNRS, UMR 6293, Clermont Université, 24 avenue des Landais, BP 80026, 63171 Aubière Cedex, France. e-mail: david.volle@inserm.fr In the last decades, studies in rodents have highlighted links between *in utero* and/or neonatal exposures to molecules that alter endocrine functions and the development of genital tract abnormalities, such as cryptorchidism, hypospadias, and impaired spermatogenesis. Most of these molecules, called endocrine disrupters exert estrogenic and/or antiandrogenic activities. These data led to the hypothesis of the testicular dysgenesis syndrome which postulates that these disorders are one clinical entity and are linked by epidemiological and pathophysiological relations. Furthermore, infertility has been stated as a risk factor for testicular cancer (TC). The incidence of TC has been increasing over the past decade. Most of testicular germ cell cancers develop through a pre-invasive carcinoma *in situ* from fetal germ cells (primordial germ cell or gonocyte). During their development, fetal germ cells undergo epigenetic modifications. Interestingly, several lines of evidence have shown that gene regulation through epigenetic mechanisms (DNA and histone modifications) plays an important role in normal development as well as in various diseases, including TC. Here we will review chromatin modifications which can affect testicular physiology leading to the development of TC; and highlight potential molecular pathways involved in these alterations

etadata, citation and similar papers at core.ac.uk

#### **TESTICULAR CANCERS**

Testicular cancer (TC) is a rare solid tumor which account for 1% of cancers in men. However, it is the most common cancer in men in their 20s and 30s (Ziglioli et al., 2011). It is a real sanitary problem as it affects men during their reproductive years. Interestingly, the development of TC has been associated with urogenital abnormalities (Olesen et al., 2007). TC development has been associated with cryptorchidism, hypospadia, and hypofertility. Indeed, epidemiological studies argue for an increased risk of testicular germ cell tumor in males who suffer for fertility troubles (Burns et al., 2010).

### **EPIDEMIOLOGY**

In developed countries, the prognosis for TC is excellent, with a 5-year survival rate >95%. Despite it represents the most frequent solid cancer in young men, it seems a quite rare pathology (Richardson et al., 2012). Thus all these data has led to consider testicular tumors as a curable neoplasm. It has to be noted that there are some geographic and racial variation in the risk of TC development (Richiardi et al., 2004). Indeed, in US, Hispanic men had the largest increase in incidence rates for non-seminomas (Townsend et al., 2010). Next to this, the genetic predisposition has been suggested as men with familial history of germ cell TC (TGCC) have a 4- to 10-fold increased risk to develop also such tumor (Greene et al., 2010). The analysis of this genetic risk seems to be associated with an autosomal recessive mode of inheritance that must result

of the combined effects of several alleles. It was estimated that 25% of TC susceptibility account for by genetic effects (Turnbull and Rahman, 2011). If TCs are quite rare in African men, the morbidity and mortality of TC is quite high in developing countries (Ugwumba and Aghaji, 2010). These data highlight the impact of poverty and paucity of resources to detect and treat cancers. In that line, it has been reported that socioeconomic position has an impact on the risk of TC development as well as the survival rate (Richardson et al., 2012).

The main way to identify TC as soon as possible is autopalpation by men to look at increased size or the appearance of "hard" testis (Umeh and Chadwick, 2010). After identification of the anomalies, biopsies will help to characterize the type of TC. Then imaging will also play a key role to determine the lymphatic extension and potential metastasis (Brunereau et al., 2012).

There are several kinds of TCs that need to be characterized by the physician using anatomopathological approach. There are several kinds depending on cell type at the origin of the development of TC (van de Geijn et al., 2009); thus there are intratubular germ cell neoplasia (IGCN), atypical germ cells with maturation arrest (MA), pseudolymphovascular invasion, real lymphovascular invasion in germ cell tumors, macroscopic Sertoli cell nodules, and Sertoli cell tumors. However, we can thus distinct germ cell tumors versus non-germ cell tumors. The most common TC are germ cell one which represent 95% of these tumors. There are two main types described which are seminoma and non-seminomatous

germ cell tumors, with a third one which is the spermatocytic seminoma (Mannuel et al., 2012).

Here we will focus this review on germ cell cancers. Their incidence has been reported to be continuously increasing in the last decades in men of different industrialized countries.

Regarding the two major groups of testicular germ cell cancers which are the seminomas and non-seminomas, each represent approximately 50% of germ cell tumors. It is interesting to note that around 15% of TGCC contain a mix of seminoma and non-seminomatous tumors (Aschim et al., 2006).

Seminomas are managed with orchiectomy and surveillance or radiotherapy in stage I and chemotherapy in more advanced stages (Warde et al., 2011). On the other side, non-seminomas which consist of several histologic types, such as choriocarcinoma, teratoma, yolk sac tumor, and embryonal cell carcinoma, each with different tumor marker profile, are less sensitive to radiation and are frequently treated with chemotherapy (Warde et al., 2011).

#### **GENETIC ASSOCIATION**

The development of TGCC is associated with many chromosomal abnormalities (Summersgill et al., 1998). Indeed, TGCC are aneuploid; with non-seminomatous tumors and seminoma being respectively hypotriploid and hypertriploid. The major association is with the gain of material from the chromosome arm 12p in both seminoma and non-seminomatous tumors (Atkin and Baker, 1982). The analyses of these chromosomal reorganizations lead to the characterization of potential candidates involved in the TGCC pathology. Among them, there is amplification of the KRAS and Cyclin d2 genes (Rodriguez et al., 2003). They are associated with malignant transformation and proliferation. Next to these genes, there are also those implicated in the cell pluripotency such as Stella and Nanog (Clark et al., 2004).

The strongest association for TGCC susceptibility is for single nucleotide polymorphisms (SNPs) at the 12q22 within the *kitligand* gene (Kanetsky et al., 2009). It is correlated with a 2.5-fold increased risk of disease. This gene has been involved in several aspect of primordial germ cell (PGC) development. Indeed, it seems to act on PGC migration and survival (Gu et al., 2009). These impacts might rely on the downstream target KRAS which then activate the p110 catalytic subunit of the PI3K pathway which in turn through AKT pathways will act on proliferation, survival, and migration (Sasaki et al., 2003). KRAS could also mobilize the MAPK pathways reinforcing its impact on proliferation, survival, and migration processes.

Next to this, as testicular physiology is under the control of the endocrine functions mainly through the activity of androgen and estrogen receptors, it was deeply studied if polymorphisms of genes involved in hormonal metabolism could be associated with a higher risk of TCs. Even if some reports are contradictory, the studies focused on the androgen receptor (AR), the estrogen receptors and genes involved in either synthesis or degradation of the hormones.

Regarding the estrogen receptors, it was demonstrated that polymorphisms in ER $\alpha$  are associated with azoospermia (Romerius et al., 2011) and are more likely to be associated with the risk of seminoma and metastasis (Brokken et al., 2012); whereas polymorphisms in ER $\beta$  are more likely to be link to altered

spermatogenesis (Aschim et al., 2005) and with risk of TGCC. New to this, polymorphisms in 17- $\beta$  hydroxydehydrogenase-4 which convert androgen and estrogen to weaker hormones were associated with TGCC (Chia et al., 2010; Ferlin et al., 2010). In these metabolic pathways, polymorphisms in cytochrome P450 Cyp-1A1 gene, encoding a hormone-metabolizing protein, were identified and inversely correlated with TC. Their effects were more or less severe regarding the different polymorphisms suggesting that it may contribute to susceptibility to TGCC development (Figueroa et al., 2008; Kristiansen et al., 2011).

In this hormonal context, one of the most studied genes in regards to polymorphisms is the AR. The AR gene has two polymorphic regions in exon-1 with CAG codon encoding for glutamine and GCN which encode for glycine. Changes in the length of these polymorphic trinucleotide repeats, (CAG) and/or (GGN), lead to altered transactivation of the AR which has been shown to play a role in several forms of endocrine cancer such as prostate cancer. Regarding TC some studies are a bit contradictory showing either or not link with increased risk of TGCC (Rajpert-De Meyts et al., 2002; Garolla et al., 2005). However, it appears that the increased risk of seminoma was associated with the shorter CAG repeat length. This suggests that an increased AR transactivation may be involved in the development of seminoma and/or progression of carcinoma in situ (CIS) to seminoma (Davis-Dao et al., 2011). It is also demonstrated that the combination of altered number in repeat for both CAG and CGC is important for the correlation with TC. Indeed, Garolla et al. (2005) showed that the combination of CAG (20 repeats) and GGC (17 repeats) was more frequent in patient with TGCC (8 versus 1.7% in control patients).

Like most of the cancer pathologies, TCs seem to be the results of either genetic and environmental factors. It has been stated that TCs derive from a precocious lesion, the CIS of the testis, also known as IGCN or testicular intraepithelial neoplasia (TIN; Sonne et al., 2008). This lesion deserves great attention, because the diagnosis of CIS may lead to a precocious diagnosis of TCs. Usually, the diagnosis of CIS is incidental.

If there is a consensus on the fact that the process of the TC pathology may found its origin during embryonic life of the individual, it can also be suspected that there might also be some other events participating to its appearance. Indeed, it seems quite a long process that TC occurs in the 20s or 30s of life when first event took place in fetal life. Thus it could be hypothesized that there might be a second hit at puberty, like hormone burst that could provoke the definitive occurrence of TC. This clearly highlights the importance of the microenvironment. A study on patients with testicular maldescent shows that there is around a two-fold increased risk to develop TGCC if the orchidopexy is performed after the age of 13, compare to men who had the operation before their 13 (Walsh et al., 2007). Thus placing the testes in the scrotum before puberty decreases the incidence of TGCC; suggesting that intra-abdominal location at puberty promotes testicular tumorigenesis. Gene expression profiling carried out on TGCC samples demonstrate marked differences between the histological subtypes of TGCC. This reflects stages and patterns of differentiation. It also supports a model of differentiation from IGCN to seminomas or embryonal carcinomas. Gene expression patterns and genomic imprinting analyses in TGCC show similarity to PGCs and gonocytes (Ziglioli et al., 2011).

#### **ENVIRONMENTAL ISSUES**

#### THE TESTICULAR DYSGENESIS SYNDROME HYPOTHESIS

In 1993, Skakkebaek and colleagues proposed that various disorders of male reproductive health, namely cryptorchidism, hypospadias, subfertility, and testicular germ cell tumor, derived from perturbations of embryonal programing and gonadal development during fetal life (Sharpe and Skakkebaek, 1993). Thus they defined it as testicular dysgenesis syndrome (TDS; Skakkebaek et al., 2001). The anomalies may lead to early symptoms, such as hypospadias and undescended testis, as well as late effects such as testis cancer and infertility. The most frequent abnormality due to TDS may be impaired spermatogenesis.

A fetal origin is obvious with regard to two symptoms of TDS: hypospadias and cryptorchidism. Moreover, studies suggested that the precursor cells of testis cancer, CIS testis, are similar to fetal gonocytes. The accepted etiology for germ cell cancer suggests that developmental arrest of fetal germ cell differentiation is a main event leading to persistence of gonocytes, which in turn develop into CIS. The causes remain unknown, although disturbances in the microenvironment provided by the Sertoli and Leydig cells may play a major role. Moreover, spermatogenesis is strictly controlled and depends on a succession of signals provided by the local environment (Skinner et al., 1991; Verhoeven, 1992; Jégou, 1993).

Among all the studies on testicular functions, there are several windows of time that must be critical for its development. Next to the importance of the fetal development, it appears that puberty must be an important timing. This is when hormone levels will reach optimal concentrations leading to the secondary sexual characters. This will also coincide with the appearance of the TGCC, as men are affected between 15 and 35 years old.

Hormones play major impacts on testicular functions throughout the life of the individuals. Indeed, testis is a key target for androgen and estrogen actions. These hormonal sensitivities have been studied for decades (Verhoeven et al., 2010). The role of testosterone is evident in patients with complete androgen insensitivity syndrome (Sultan et al., 1993).

Moreover, testicular descent occurs in two phases: the transabdominal phase, which animal studies suggest depends on the insulin-like hormone 3 (INSL3) produced by interstitial Leydig cells (Kaleva and Toppari, 2005; Ferlin et al., 2006, 2009). It has been demonstrated that the expression of the Insl3 gene is under the control of the estrogen signaling pathway (Cederroth et al., 2007). Then, the inguinoscrotal phase is dependent on androgens.

Androgens play a crucial role in the control of spermatogenesis. Molecular details have been discovered using transgenic mice invalidated (knock-out, KO) for AR either in the testis or in different testicular compartments. Such mice have low testosterone levels with altered expression of steroidogenic enzymes, even Leydig cell mass is altered (Wang et al., 2009). AR is involved in autocrine action of testosterone on Leydig cells. Testosterone deficiency is responsible for spermatogenesis arrest due to altered Sertoli functions (Wang et al., 2009).

In mouse ER $\alpha$  have been demonstrated to be involved in the maturation of the spermatozoa (Lubahn et al., 1993). These KO mice present an excess of fluid which increases the pressure within the seminiferous tubules and leads to the germ cell loss (Eddy et al., 1996; Hess et al., 1997). Surprisingly, the ER $\beta$ -KO mice show no testicular phenotype (Couse and Korach, 1999).

The deficient male mice for Cyp19 (Cyp19-KO), encoding for the enzyme responsible for the aromatization of testosterone into estrogens, develop normally and their genital tract is anatomically similar to that of the wild-type (Robertson et al., 1999). Males are fertile; however, some of Cyp19-KO males exhibit an altered spermatogenesis by the age of 5 months (Robertson et al., 1999). By the age of 1 year, all males develop abnormal spermatogenesis with a blockage of germ cell maturation at the spermatid stage. This is due to an increase in apoptotic rates when compared to the wild-type animals. The observation of abnormal acrosome development in the Cyp19-KO mouse suggests that acrosome biogenesis could be estrogen-dependent (Robertson et al., 1999). Interestingly, estradiol have been demonstrated to play a role as a survival factor for the human germ cells (Pentikäinen et al., 2000), and also is beneficial for sperm motility (Carreau and Hess, 2010). Moreover, next to these data, deleterious effects of numerous endocrine disruptors on sperm count and male genital tract (cryptorchidism, hypospadia, and infertility) have been documented (Iguchi et al., 2001; Sikka and Wang, 2008) particularly in the context of in utero and/or neonatal exposures.

Imbalanced equilibrium between the estrogen and androgen levels *in utero* is hypothesized to influence TC risk. Thus, alterations in genes involved in the action of sex hormones may contribute to variability of an individual's susceptibility to TC. Mutations in testosterone pathway genes may alter the level of testosterone *in vivo* and hypothetically the risk of developing TC (Kristiansen et al., 2012). In regard with the hypothesis of the TDS and the known impact of steroids on testicular development and functions, it has been hypothesized that endocrine-disrupting chemicals could play a role in these pathologies.

## **ENDOCRINE DISRUPTERS**

The endocrine disrupters (EDs) are compounds which may be of industrial or natural origin and which act to dysregulate steroid function and metabolism. They produce their effects by mimicking, antagonizing, or altering endogenous steroid levels (androgens or estradiol, E2) *via* changing rates of their synthesis or metabolism and/or expression or action at receptor targets.

The question of environmental endocrine disruption has been a topic of public concern for many years and remains high on the scientific agenda. Indeed the number of chemical used is constantly increasing in developed countries, it is supposed that humans and animals can be exposed to a growing number of contaminants which can accumulate in their bodies and may have adverse consequences for health. Large range of chemicals (banned or still in use) have been characterized as EDs. Among these EDs, highly produced, Bisphenol A (BPA; Rubin, 2011) is present in plastics, including beverage and food storage containers and in the ink used for thermal paper receipts. Some individuals have also been exposed to contaminants with adverse effects originating from medical use (diethylstilbestrol, DES; Bullock et al., 1988),

or dietary habits (phytoestrogens; Naciff and Daston, 2004). More surprisingly, in China, human excretions were suspected to be the major contributor of estrogens in municipal wastewater (Zhou et al., 2012).

Moreover, studies have made correlations between elevated levels of phthalates in urines of pregnant women and lower masculinization of their progeny (Suzuki et al., 2012). EDs have also been detected in the maternal milk (Hines et al., 2009).

Regarding the urogenital development and more particularly testis, the impacts of EDs have been quite well described on human and rodent models (Volle et al., 2009; Toppari et al., 2010; Desdoits-Lethimonier et al., 2012). This is mainly true for hypospadia, cryptorchidism, and infertility; but the link with TGCC remain to be defined.

It has been demonstrated that EDs with estrogenic activity lead to a decrease of expression of steroidogenic genes which by the end result in lower testosterone concentrations (Joensen et al., 2008). However, the mechanisms seem to differ between species. This highlights the difficulties to find good experimental models. Indeed, one will claim that human cells will not react as murine cells; and other may assume that cell lines or explants tissues might not totally react as in the body. Indeed, endocrine systems are really complex and need to be analyzed in regard of the complexity of integrative physiology.

The effect of such EDs in human pathology is quite difficult to establish. It is really difficult to have standardized cohort between the studies, which is due to the history of patients (environmental and genetic factors). This heterogeneity leads to inconsistent studies which do not allow definitive conclusions on the potential involvement of EDs in TGCC.

It has been demonstrated that there was a six-fold increase in the risk of seminoma among plastic workers exposed to polyvinyl chloride (PVC; Ohlson and Hardell, 2000). However, Hardell et al. (2004a) did not found association between PVC exposure and TC.

Another study from Hardell et al. (2006) show that chlorinated biphenyls (PCBs) could play a role during fetal exposure in the etiology of TC as case mothers were identified to have significantly higher concentrations of these PCBs. However, regarding PCBs, no different risk pattern could be demonstrated for seminoma and non-seminomatous TC (Hardell et al., 2003, 2004b).

However, several studies have demonstrated some effects of these molecules on risk of TGCC (Hardell et al., 2003, 2004b, 2006; McGlynn et al., 2009). In their study McGlynn et al. (2009) associated PCBs with a decrease risk of TGCC development. Different PCBs are either associated with decrease risk of either seminoma or non-seminoma. In the same line of evidence, it was demonstrated that in patients with seminoma there was differences in concentrations of different PCBs (Purdue et al., 2009). Some PCBs (44, 49, and 52) were found to be lower accumulated in patients with seminoma compare to congeners; whereas other PCBs (99, 138, 153, 167, 183, and 195) were significantly higher accumulated in these patients. This suggest that rather than specific concentration of one or another PCB, it may be the relative concentrations of multiple one that might be relevant to established clear correlation with the risk of TC.

Moreover, evidences suggest that exposures to pesticides could be risk factor of TC development. Indeed, a recent

study reported a significant association between TC development and use of insecticides such as organochlorine pesticides, namely dichlorodiphenyldichloroethylene (p,p'-DDE) isomer and hexachlorobenzene (Giannandrea et al., 2011).

Next to this, as TC development seems to be either associated with genetic predisposition and or environmental exposure, it might be of interest to analyze the potential combination of such anomalies. Consistent with this hypothesis, as polymorphisms in AR and some organochlorine pesticides have been associated to risk of TGCC development, and that some of these organochlorine pesticides present anti-androgenic activities, Biggs et al. (2008) have studied the potential interaction of AR polymorphisms and exposure to p,p'-DDE and the association with TC risk. According to their study, they were not able to demonstrate any association between p,p'-DDE exposure and TC risk, either or not in combination with (CAG)n length. However, a point to take in consideration is that p,p'-DDE concentration was measured in adult patients, but as TC is supposed to found its origin in fetal life, this must be the exposition in this period of life that must be significant to define such association. However, it will be difficult for instance to establish such correlation as we do not have access to fetal/neonatal blood of these patients. However, it could be though to perform some clinical trials to collect blood of new borns and make assay for pesticides concentrations. Then it will be necessary to follow these boys and see if they will further develop TC, and perform correlation with fetal/neonatal concentrations of pesticides.

#### ED and insulin-like-3

Next to their steroidogenic function, Leydig cells during development express the insulin-like-3 gene which is responsible for gubernaculum maturation (Ivell and Anand-Ivell, 2011). In human, INSL3 is produced by prenatal, neonatal, and adult Leydig cells to various extents (Ivell and Anand-Ivell, 2011). INSL3 production seems to be dependent on the state of Leydig cell differentiation, and is stimulated by the long-term trophic effects mediated by luteinizing hormone (LH; Toppari et al., 2007). This finding clearly helps in understanding the complete process of testis descent. The role of the INSL3 on testis descent was highlighted by the fact that mouse KO for the gene encoding Insl3 results in cryptorchidism (Adham and Agoulnik, 2004). Moreover, animal model of ED exposures demonstrated that INSL3 production is sensitive to estrogenic or anti-androgenic compounds. This clearly suggests that maternal exposure to EDs during pregnancy can result in cryptorchidism, a factor that predispose to TC development.

## ED and steroidogenesis

This link is of particular importance. Indeed, if CIS have been described in boys at birth (Jacobsen and Henriques, 1992), TCs appear in young men between 15 and 35 years old, suggesting that puberty and probably the increase of hormone concentrations must be key events.

The impact of EDs on steroidogenesis has been demonstrated for several decades now. EDs have been described to inhibit critical cellular functions involved in steroidogenesis, such as transport of cholesterol into the mitochondria, the expression of steroidogenic genes or the activity of these enzymes (Vanparys et al., 2012). Indeed, it has been demonstrated that exposure of adult rats to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCCD) led to lower production of testosterone in response to human choriogonadotropin (hCG), relative to testes from control rats (Kleeman et al., 1990). The addition of pregnenolone is able to restore normal testosterone secretion. This highlights that the impact on steroid synthesis might results from altered cholesterol synthesis or mobilization to the mitochondria (Moore et al., 1991). TCDD was also observed to reduce the number and size of the Leydig cells (Johnson et al., 1994). Moreover, the use of primary culture of Leydig cells also demonstrate that TCDD represses *Cyp11a1* expression, through the alteration of the ability of hCG to increase intracellular cAMP levels (Lai et al., 2005).

The impact on the animal phenotypes will depend on the age at exposure. Thus fetal/neonatal exposure will lead to altered internal genital organs. Indeed, experimental models have shown that disruption of the androgen signaling results in feminization of external genitalia (Nagahama et al., 2004). In animal models, exposures to low concentrations of EDs are able to alter testosterone synthesis without any effect of feminization. However, the most interesting thing is that even after neonatal exposure, the adult testosterone concentrations are decreased suggesting long-term impact that is specific to this particular window of exposure (Volle et al., 2009). This has been clearly highlighted as the exposition of adult animal lead to transitory testosterone decrease, associated with temporary germ cell death. In human, majority of the results relies on epidemiological studies and correlative data (Woodruff, 2011). However, several studies using testicular explant models have tried to characterize the impact of EDs on human Leydig cells (Desdoits-Lethimonier et al., 2012). By the end, all these data led to the idea that fetal cells must be more sensitive than adult Leydig cells.

Regarding the signaling pathways involved, and as main regulators of the expression of steroidogenic genes, a reduced activity of the cAMP-mediated PKA pathway would also be expected to reduce the mobilization of cholesterol by cholesterol hydrolases. However, at the molecular level, it has been demonstrated in mouse that neonatal exposure to EDs with estrogenic activity leads to a decrease of testosterone synthesis through ER receptors and that the orphan nuclear receptor SHP must be a key intermediary for this effect (Volle et al., 2009). Indeed, the SHP KO mice seem to be less sensitive to estrogenic EDs than their wild-type littermates. It was also demonstrated that the impact of SHP on steroidogenesis is due to the repression of either expression and/or activity of the nuclear receptors Sf-1 and Lrh-1 (Volle et al., 2009). It is to note that SHP was not involved in early post-natal decrease of testosterone production induced by estrogenic EDs. This highlights that the involved molecular pathways affected by EDs are multiple and complex.

### ED and germ cell differentiation

Spermatogenesis, leading to spermatozoa formation, is a complex process with multiple steps involving mitosis, meiosis, and spermiogenesis. It takes place in the seminiferous tubules. There, germ cells are organized from the base of the tubule to the lumen. Germ cells are supported by the nursing Sertoli cells, which extend from the base to the lumen of the seminiferous tubules. Efficient

spermatogenesis also relies on the integrity of tight junctions between the Sertoli cells which form the blood–testis barrier (BTB; Cheng and Mruk, 2012). BTB has many functions in the testis such as maintaining a particular immune context and also the control of the flow of nutrients and growth factors that are required for the development of germ cells.

The key role of retinoids in the differentiation process of germ cells was highlighted by two studies demonstrating stra8 as a key factor (Bowles et al., 2006; Koubova et al., 2006). It also involved the timely regulation of the gene encoding for cyp26b1, an enzyme responsible for retinoid degradation. Indeed, CYP26B1 in Sertoli cells acts as a masculinizing factor to arrest male germ cells in the G0 phase of the cell cycle and prevents them from entering meiosis (Li et al., 2009). This induction of retinoid pathway to induced entry in meiosis seems to be inhibited by the nuclear receptor SHP, a co-repressor of the retinoid acid receptors RAR (Volle et al., 2007). In the opposite manner, the FGF9 signaling pathway acts to determine germ cell fate to enter meiosis (Boisen et al., 2001).

## **EPIGENETIC AND TESTICULAR CANCERS**

## TRANSGENERATIONAL EFFECTS: ROLE OF EPIGENETIC MODIFICATIONS

Epigenetic refers to changes of DNA information without any change of the sequences. It relies on histones post-transductional modification (acetylation, methylation...) or of DNA methylation levels. Thus, inheritance of these information requires germline transmission of epigenetic patterns between generations. The epigenetic programing of the germline occurs during embryonic development in a sex-specific manner (Western, 2009). These processes are crucial for reproductive functions, as most of mouse models with specific germline invalidation for gene responsible of DNA or histones modifications led to sterile animals (Peters et al., 2001; La Salle et al., 2007). Regarding the male germline, it becomes imprinted following sex determination. Thus after puberty when spermatogenesis is fully functional, there are also specific epigenetic modifications along the different steps of germ cell differentiation from spermatogonial precursors up to testicular spermatozoa. Indeed, it has been demonstrated that throughout the different steps of spermatogenesis, germ cells have a dynamic of the epigenetic modifications. This is highlighted by the changes in the expression levels of the enzymes involved in these modifications. If DNA methyltransferases (Dnmts) are mainly expressed in the spermatogonia, histone methyltransferases (HMTs) are mainly expressed at the spermatocyte levels (Godmann et al., 2009). Then other histone modifications such as hyperacetylation of the H4 histone play a key role in the removal of histones and their replacement by protamines during spermatogenesis (Dhar et al., 2012). Such epigenetic modifications have also been demonstrated to be associated with infertility as for example reduced expression of Dnmt3b in patients with spermatogenic arrest (Adiga et al., 2011).

If the EDs cannot alter the DNA sequence, there are numerous of studies demonstrating that they can impact the epigenome. Indeed, environmental factors can alter the epigenetic programing which will impact the development of the offsprings. Interestingly, it has been demonstrated in the last decade that these effects could also potentially impact the subsequent generations

(Huang et al., 2011; Schoevers et al., 2012). Such effect was first demonstrated for the exposure with vinclozolin, a compound with anti-androgenic activities (Anway et al., 2005). Thus exposure during embryonic gonadal sex determination can alter the male germline epigenetics (e.g., DNA methylation). The epigenetic mechanism involves the alteration of DNA methylation in the germline that appears to transmit transgenerational adult onset disease, including spermatogenic defects and cancer (Anway et al., 2005). At the molecular level, the authors identify by global transcriptomic approach that the expression of genes involved in these processes such as the Dnmt or responsible for histone modifications was affected by vinclozolin treatment (Anway et al., 2005). All these data suggest the main involvement in transgenerational transmission.

## Potential link between epigenetic and TGCT?

These modifications in epigenetic programing by EDs are of particular interest as they have been suspected to be responsible of the increased rate of pathologies related to the TDS such as TCs. This potential association between ED and epigenetic in TGCT development is highlighted by the fact that recently, number of studies have demonstrated the modified epigenetic profile of tumor cells compare to "normal" cells (Okamoto and Kawakami, 2007; Brait et al., 2012). Indeed, it appears that tumor cells show an epigenetic pattern similar to undifferentiated spermatogonial stem cells (Almstrup et al., 2010). Indeed, CIS cells have a permissive and fetal-like chromatin structure (Almstrup et al., 2010). This is consistent with the fact that CIS, precursor of TGCT have already been identify in testis of embryos.

**DNA methylation.** DNA methylation is an important event during germ cell development. These enzymatic modifications of DNA rely on Dnmts. Among them, expression profiles of DNMT3a and DNMT3l suggest that they might act during prenatal germ cell development for the establishment of *de novo* methylation. On the other side, DNMT1 and DNMT3b rise shortly after birth in the male (Godmann et al., 2009). It was thus hypothesized that these two Dnmts must be involved in the maintenance of methylation patterns in proliferating spermatogonia.

The use of mouse models invalidated for genes encoding these enzymes highlights their crucial involvement during spermatogenesis. Dnmt3a<sup>-/-</sup> males show a greatly reduced number of spermatocytes (Yaman and Grandjean, 2006; La Salle et al., 2007). This suggests a major involvement for progression through meiosis. In the same line, the males invalidated for Dnmt3L participate to the acquisition of DNA methylation at paternally imprinted regions, unique non-pericentric heterochromatic sequences, and interspersed repeats, including transposable elements. Moreover, Dnmt3L<sup>-/-</sup> males present alterations of meiotic process leading to spermatogenesis arrest, and spermatocytes apoptosis (Webster et al., 2005). As Dnmt3L expression is restricted to gonocytes, the presence of defects in later stages suggests alteration of processes required for completion of spermatogenesis.

The major role of these epigenetic alterations has been demonstrated in carcinogenesis. Indeed, it has been shown that DNA methylation is associated with repression of tumor suppressor gene expression. This epigenetic process is one of the most studied

in the research field, and has been recognized as a major mechanism during TGCC progression (Manton et al., 2005; Ellinger et al., 2009). Moreover, the DNA methylation pattern seems to correlate with histological features of the different types of TGCC. Undifferentiated TGCC (seminomas, IGCN unclassified, and gonadoblastomas) are hypomethylated, whereas more differentiated TGCC (teratomas, yolk sac tumors, and choriocarcinomas) show a higher degree of methylation. Embryonal carcinomas show an intermediate pattern. Thus such parameters could be used to discriminate between seminoma and non-seminoma (Brait et al., 2012).

Such impact involved modifications of genes encoding for enzymes involved in DNA methylation. Consistent with cell type origin of TGCC, the Dnmts are mainly expressed in fetal testis and in the more undifferentiated cell type (spermatogonia) during adult normal spermatogenesis.

Regarding cancers, Dnmt1 was not expressed in seminoma, but upregulated in embryonal carcinoma (Omisanjo et al., 2007). In contrast, the expression of *Dnmt3a* was found up-regulated in TGCC compared to non-tumor testicular tissues (Yamada et al., 2004). The expression pattern of *Dnmt3b* has been deeply studied and show that it could be used as a predictive marker for relapse of stage I seminomas (Arai et al., 2012). Lastly, *Dnmt3l* was overexpressed in the non-seminoma tumors (Minami et al., 2010).

These changes in Dnmts expression will lead to main alteration of transcriptomic profile between tumors and normal tissues (Alagaratnam et al., 2011). Among them, two targets are of particular interest as they are genes of pluripotency. Indeed, early fetal germ cells and undifferentiated germ cell tumors have in common the expression of pluripotency markers such as the transcription factors *Nanog* and *Oct3/4*. Regarding *Nanog* promoter, it was found hypomethylated in spermatogonia and hypermethylated in sperm (Nettersheim et al., 2011). This selective repression might reflect that the cells need to suppress pluripotency in order to prevent malignant transformation. Finally, methylation of CpGs in the *Nanog* promoter in germ cell tumors and derived cell lines correlated to differentiation state. In the same line, the study *Oct3/4* showed that seminoma and embryonal carcinoma were hypomethylated (De Jong et al., 2007).

Histone methylation. Many enzymes are involved in histone methylation with specific or redundant level of methylation. These differences rely on the modified histone, targeted amino-acid residues and the number of methyl groups that are added to histone. These modifications are performed by several members of the HMT family. On those involved in spermatogenesis, the Suv39h1 and suv39h2 mediate histone H3 di and/or trimethylation at lysine 9 (Schotta et al., 2003). They are involved in meiosis as double KO mice present defects in male meiosis and highly pronounced apoptosis of stage IV spermatocytes during the transition from mid to late pachytene (O'Carroll et al., 2000; Peters et al., 2001).

G9a, a mammalian HMTase, is a candidate for H3-K9 methylation in non-heterochromatic loci (Tachibana et al., 2007). G9a is essential for early embryonic development and plays a dominant role in H3-K9 methylation of euchromatin. Its role was highlighted as mice lacking G9a are sterile, with germ cells undergoing

apoptosis during the pachytene stage (Tachibana et al., 2007). Interestingly, it was demonstrated that G9a was a target of retinoid signaling pathway, a key regulator of germ cell differentiation, and that it was inhibited in the context of estrogenic exposure leading to the increased germ cell apoptosis induced by EDs (Volle et al., 2009).

Another HMT is the enhancer of zeste 2 (EZH2) which trimethylate histone H3 at the lysine 27 (Chang and Hung, 2012). It was demonstrate that during spermatogenesis, EZH2 is restricted to round spermatids in the perinuclear acrosome region (Lambrot et al., 2012). This localization is concomitant with the dramatic epigenetic reorganization that occurs during spermiogenesis leading to an extreme compaction of the chromatin.

DNA methylation, histone methylation are epigenetic modifications functioning in transcriptional control and have been implicated in the deregulation of gene expression in cancer. As mentioned above, the CIS cells present epigenetic profile similar to ES cells. Such properties of chromatin have been associated with a high transcriptional and proliferative activity. This is due to lower levels of DNA methylation and of histone methylation (H3K9me2 and H3K27me3; Almstrup et al., 2010). This is consistent with a lower expression of the gene encoding for the EZH2 in the TGCC compared to normal testis tissues (Hinz et al., 2010). This suggests that in TGCC EZH2 does not exert its often assumed oncogenic properties during malignant transformation and progression. High EZH2 levels in normal testicular tissue and the inverse association of its expression levels with the severity of spermatogenic failure point to its potential value as a molecular marker for spermatogenic defects and may indicate an important physiological role of EZH2 during intact spermatogenesis.

It has been suggested that histone H2A and H4 arginine 3 dimethylation might be a mechanism by which IGCNU and seminoma maintain the undifferentiated state; while loss of these histone modifications (Eckert et al., 2008) could be involved in the somatic differentiation observed in non-seminomatous tumors.

In healthy testis, the distribution of histone H3 methylation was dependent on the developmental stage of spermatogenic cells and in non-seminoma, histone H3-K4 and K9 methylation was detected in all histological subtypes (Lambrot and Kimmins, 2011). This suggested that histone H3-K4 and K9 methylation could be associated with abnormal gene expression in non-seminoma. Histone modifications determine epigenetic patterns of gene expression with methylation of histone H3 at lysine 4 (H3K4), and are often associated with active promoters. LSD1/KDM1 is a histone demethylase that suppresses gene expression by converting dimethylated H3K4 to mono- and unmethylated H3K4 (Wang et al., 2011). Interestingly, LSD1 protein level is highly elevated in pluripotent cancer cells and in human testicular seminoma tissues that express *Oct3/4* (Wang et al., 2011).

Histone acetylation. This is another histone modification that must also lead to gene repression. These modifications are performed by specific enzymes so-called histone deacetylase (HDCA). Such post-translation modifications are implicated in normal spermatogenesis (Fenic et al., 2008). Indeed, H3K9ac was

shown in spermatogonia, spermatocytes, elongating spermatids, and ejaculated spermatozoa of fertile and infertile men (Steilmann et al., 2011). In spermatogonia, the stainings for H3K9ac, H3K18ac, and H3K23ac were strong. Then spermatocytes, the stainings for H3K9ac, H3K18ac, H3K23ac, and H3K4me3 were reduced in the preleptotene to pachytene stage, but in diplotene stage the stainings for H3K18ac, H3K23ac, and H3K4me3 seemed to become intense in later stages (Song et al., 2011a).

The main involvement of histone acetylation during spermatogenesis is the hyperacetylation of histone H4 during spermiogenesis (Dhar et al., 2012). This signal seems to play crucial role for removal of histones and their replacement by protamines, which is key feature for nucleus condensation, and thus formation of spermatozoa. There are multiple members classified in different subfamilies. Interestingly, choriocarcinomas showed generally high expression for all three class I HDAC isoforms (Gryder et al., 2012). However in contrast with other types of tumors, no diagnostic or prognostic values for HDAC1–3 in TGCC could be inferred (Fritzsche et al., 2011).

Impact of TGCC treatment. Some testis tumors are treated using cisplatin (Koychev et al., 2011). Recent report analyzed the impact of such treatment on the integrity of spermatozoa chromatin in rats (Maselli et al., 2012). As expected, the cisplatin treatment lead to susceptibility of DNA to denaturation and the number of strand breaks were significantly increased in mature sperm. After a recovery period, it was noted that mature sperm did not show significant DNA damages. However, the protamination level of the sperm of these animals was significantly decreased. This was associated with an up-regulation of the histones H1.2, H4, H2A1, and H2B1A. This suggests long-term effect of cisplatin treatment that could have consequences for progenies even after the arrest of cancer cure.

Small non-coding RNA. Next to these well studied epigenetic processes, it also appears that microRNA and small RNA play important roles in both germ cell differentiation and transmission to subsequent generations (He et al., 2009; Suh and Blelloch, 2011; Buckley et al., 2012). Indeed, achieving the correct spatial and temporal expression of germ cell-specific genes is fundamental to the production of spermatozoa (Song et al., 2011b). Notably, for the regulation of genes involved in the repression of protein translation is central to many embryonic processes, and is particularly active during spermatogenesis.

The miRNA and siRNA are generated by the nuclear RNase III enzyme Drosha and the cytoplasmic RNase III enzyme Dicer (Papaioannou and Nef, 2010). The involvement of these particular RNA has been highlighted by the generation of mouse models invalidated for genes encoding Drosha or Dicer (Korhonen et al., 2011; Wu et al., 2012). These invalidations led to sterility due to disrupted spermatogenesis characterized by depletion of spermatocytes and spermatids leading to oligoteratozoospermia or azoospermia. miRNAs mostly act by destabilizing target mRNAs or inhibiting their translation. Next to this, the PIWI-interacting RNAs (piRNAs) are predominantly expressed in the germ cell lineage (Kibanov et al., 2012). The analyses on this particular class of RNA suggest that they have a potential role in epigenetic

regulation of cell polarization. Moreover piRNA seem to be involved in silencing of transposon expression.

Many miRNA and siRNA have been implicated in the different steps of spermatogenesis. Among them, miR-449 and miR-34 seems to have common targets on the E2F signaling pathway which is mainly involved in the regulation of male germ cell development (Bao et al., 2012). Mir-17-92 (Mirc1) cluster and Mir-106b-25 (Mirc3) cluster miRNAs were suggested to cooperate in regulating spermatogonial development (Tong et al., 2012). The overexpression of miR-184 was demonstrated to promote the proliferation of a germ cell line, GC-1spg (Wu et al., 2011). It was also shown that transient inhibition of miR-21 in SSC-enriched germ cell cultures increased the number of germ cells undergoing apoptosis (Niu et al., 2011).

Interestingly, several miRNAs are unique to testis. Regarding the involvement of miR199a-5p was highlighted by the study showing an inverse relationship between miR-199a-5p and embryonal carcinoma antigen podocalyxin-like protein 1 (PODXL) expression (Cheung et al., 2011). This suggests that PODXL must be a downstream effector mediating the action of miR199a-5p. This is of particular interest as PODXL, an anti-adhesive protein, is expressed in aggressive tumors.

Next to this, miR-371-373 and miR-302 clusters are overexpressed in malignant TGCC (Novotny et al., 2012). It downregulates mRNAs involved in biologically significant pathways involved in cellular senescence induced by oncogenic stress.

Among other examples, miRNA-383 expression is downregulated in the testes of infertile men with MA (Lian et al., 2009). These results suggest that it functions as a negative regulator of proliferation, in part, through inactivation of the pRb pathway. Thus an abnormal expression of miRNA-383 may potentiate the connections between male infertility and testicular germ cell tumor.

Therapy issues. A percentage of tumors are resistant to cisplatin treatment. It seems to be associated with the high cytoplasmic expression of p21. Interestingly, it was demonstrated that there is an inverse association between cisplatin resistance and the expression of Oct4 and miR-106b (Koster et al., 2010). Thus, it was suggested that modulation of the Oct4/miR-106b/p21 pathway

## **REFERENCES**

Adham, I. M., and Agoulnik, A. I. (2004). Insulin-like 3 signalling in testicular descent. *Int. J. Androl.* 27, 257–265.

Adiga, S. K., Ehmcke, J., Schlatt, S., Kliesch, S., Westernströer, B., Luetjens, C. M., et al. (2011). Reduced expression of DNMT3B in the germ cells of patients with bilateral spermatogenic arrest does not lead to changes in the global methylation status. Mol. Hum. Reprod. 17, 545–549

Alagaratnam, S., Lind, G. E., Kraggerud, S. M., Lothe, R. A., and Skotheim, R. I. (2011). The testicular germ cell tumour transcriptome. *Int. J. Androl.* 34(Pt 2), e133–e150. Almstrup, K., Nielsen, J. E., Mlynarska, O., Jansen, M. T., Jørgensen, A., Skakkebæk, N. E., et al. (2010). Carcinoma in situ testis displays permissive chromatin modifications similar to immature foetal germ cells. *Br. J. Cancer* 103, 1269–1276.

Anway, M. D., Cupp, A. S., Uzumcu, M., and Skinner, M. K. (2005). Epigenetic transgenerational actions of endocrine disruptors and male fertility. *Science* 308, 1466– 1469.

Arai, E., Nakagawa, T., Wakai-Ushijima, S., Fujimoto, H., and Kanai, Y. (2012). DNA methyltransferase 3B expression is associated with poor outcome of stage I testicular seminoma. Histopathology 60, E12–E18. could open new perspectives in the treatment of chemoresistant TC.

### **CONCLUSIONS/PERSPECTIVES**

The increasing incidence of reproductive tract diseases and particularly TC, during the last decades, is of concern. Indeed, even if it is a well curable disease with a good 5-year survival rate, it affects men during the time of their reproductive life (between 15 and 40), suggesting that it may affect both fertility and also health of the progeny. It cannot be exclude that germ cells generated at the beginning of carcinogenesis could transmit altered DNA material, due to genetic, epigenetic perturbation. This higher rate of appearance for TC cases is supposed to be associated with exposure to environmental chemicals. This also need to be deeply studied as in our modern society, men are exposed to an increasing amount of chemicals. This suggests that the incidence of TC could be even more important in the future decades. All these potential consequences point out the importance to study the involved mechanisms in appearance, and progression of TGCC. This means that we have to better understand the etiology for such cancers.

In long-term perspective, an increased knowledge of genetic, epigenetic, and gene expression patterns correlated with data of anatomopathology will lead to a better definition and understanding of the pathology. It also suggests that there is a need to analyze patients in a case by case approach in order to identify genetic, epigenetic alterations, and modifications of gene expression patterns. This will help to propose personalized therapy that would probably help in improving survival rate and life quality of survivors and avoid relapse.

## **ACKNOWLEDGMENTS**

Lobaccaro's lab is supported by Association de Recherche sur les Tumeurs Prostatiques, Ligue contre le Cancer (Comité Allier), Fondation pour la Recherche Médicale (FRM), Fondation BNP-Paribas and Association de Recherche contre le Cancer (ARC), Grant from Ministère de l'Enseignement Supérieur et de la recherche (to Marine Baptissart), Nouveau Chercheur Auvergne (#R12087CC to David H. Volle), ANR Jeune Chercheur (#1103, to David H. Volle).

Aschim, E. L., Haugen, T. B., Tretli, S., Daltveit, A. K., and Grotmol, T. (2006). Risk factors for testicular cancer – differences between pure non-seminoma and mixed seminoma/non-seminoma? *Int. J. Androl.* 29, 458–467.

Aschim, E. L., Giwercman, A., Ståhl, O., Eberhard, J., Cwikiel, M., et al. (2005). The RsaI polymorphism in the estrogen receptor-beta gene is associated with male infertility. J. Clin. Endocrinol. Metab. 90, 5343–5348.

Atkin, N. B., and Baker, M. C. (1982). Specific chromosome change, i(12p), in testicular tumours? *Lancet* 2, 1349. Bao, J., Li, D., Wang, L., Wu, J., Hu, Y., Wang, Z., et al. (2012). MicroRNA-449 and microRNA-34b/c function redundantly in murine testes by targeting E2F transcription factor-retinoblastoma protein (E2F-pRb) pathway. J. Biol. Chem. 287, 21686–21698.

Biggs, M. L., Davis, M. D., Eaton, D. L., Weiss, N. S., Barr, D. B., Doody, D. R., et al. (2008). Serum organochlorine pesticide residues and risk of testicular germ cell carcinoma: a population-based case—control study. Cancer Epidemiol. Biomarkers Prev. 17, 2012–2018.

Boisen, K. A., Main, K. M., Rajpert-De Meyts, E., and Skakkebaek, N. E. (2001). Are male reproductive disorders a common entity? The testicular

- dysgenesis syndrome. Ann. N. Y. Acad. Sci. 948, 90–99.
- Bowles, J., Knight, D., Smith, C., Wilhelm, D., Richman, J., Mamiya, S., et al. (2006). Retinoid signaling determines germ cell fate in mice. *Science* 312, 596–600.
- Brait, M., Maldonado, L., Begum, S., Loyo, M., Wehle, D., Tavora, F. F., et al. (2012). DNA methylation profiles delineate epigenetic heterogeneity in seminoma and non-seminoma. *Br. J. Cancer* 106, 414–423.
- Brokken, L. J. S., Lundberg-Giwercman, Y., De-Meyts, E. R., Eberhard, J., Ståhl, O., Cohn-Cedermark, G., et al. (2012). Association of polymorphisms in genes encoding hormone receptors ESR1, ESR2 and LHCGR with the risk and clinical features of testicular germ cell cancer. *Mol. Cell. Endocrinol.* 351, 279–285.
- Brunereau, L., Bruyère, F., Linassier, C., and Baulieu, J.-L. (2012). The role of imaging in staging and monitoring testicular cancer. *Diagn. Interv. Imaging* 93, 310–318.
- Buckley, B. A., Burkhart, K. B., Gu, S. G., Spracklin, G., Kershner, A., Fritz, H., et al. (2012). A nuclear Argonaute promotes multigenerational epigenetic inheritance and germline immortality. *Nature* 489, 447–451.
- Bullock, B. C., Newbold, R. R., and McLachlan, J. A. (1988). Lesions of testis and epididymis associated with prenatal diethylstilbestrol exposure. *Environ. Health Perspect.* 77, 29–31.
- Burns, W. R., Sabanegh, E., Dada, R., Rein, B., and Agarwal, A. (2010). Is male infertility a forerunner to cancer? *Int. Braz. J. Urol.* 36, 527–536.
- Carreau, S., and Hess, R. A. (2010).

  Oestrogens and spermatogenesis.

  Philos. Trans. R. Soc. Lond. B Biol. Sci.
  365. 1517–1535.
- Cederroth, C. R., Schaad, O., Descombes, P., Chambon, P., Vassalli, J.-D., and Nef, S. (2007). Estrogen receptor alpha is a major contributor to estrogen-mediated fetal testis dysgenesis and cryptorchidism. *Endocrinology* 148, 5507–5519.
- Chang, C.-J., and Hung, M.-C. (2012). The role of EZH2 in tumour progression. *Br. J. Cancer* 106, 243–247.
- Cheng, C. Y., and Mruk, D. D. (2012). The blood–testis barrier and its implications for male contraception. *Pharmacol. Rev.* 64, 16–64.
- Cheung, H.-H., Davis, A. J., Lee, T.-L., Pang, A. L., Nagrani, S., Rennert, O. M., et al. (2011). Methylation of an intronic region regulates miR-199a in testicular tumor malignancy. Oncogene 30, 3404–3415.
- Chia, V. M., Li, Y., Quraishi, S. M., Graubard, B. I., Figueroa, J. D., Weber,

- J.-P., et al. (2010). Effect modification of endocrine disruptors and testicular germ cell tumour risk by hormonemetabolizing genes. *Int. J. Androl.* 33, 588–596.
- Clark, A. T., Rodriguez, R. T., Bodnar, M. S., Abeyta, M. J., Cedars, M. I., Turek, P. J., et al. (2004). Human STELLAR, NANOG, and GDF3 genes are expressed in pluripotent cells and map to chromosome 12p13, a hotspot for teratocarcinoma. Stem Cells 22, 169–179.
- Couse, J. F., and Korach, K. S. (1999). Estrogen receptor null mice: what have we learned and where will they lead us? *Endocr. Rev.* 20, 358–417.
- Davis-Dao, C. A., Siegmund, K. D., Vandenberg, D. J., Skinner, E. C., Coetzee, G. A., Thomas, D. C., et al. (2011). Heterogenous effect of androgen receptor CAG tract length on testicular germ cell tumor risk: shorter repeats associated with seminoma but not other histologic types. *Carcinogenesis* 32, 1238–1243.
- De Jong, J., Weeda, S., Gillis, A. J. M., Oosterhuis, J. W., and Looijenga, L. H. J. (2007). Differential methylation of the OCT3/4 upstream region in primary human testicular germ cell tumors. Oncol. Rep. 18, 127–132.
- Desdoits-Lethimonier, C., Albert, O., Le Bizec, B., Perdu, E., Zalko, D., Courant, F., et al. (2012). Human testis steroidogenesis is inhibited by phthalates. *Hum. Reprod.* 27, 1451– 1459.
- Dhar, S., Thota, A., and Rao, M. R. S. (2012). Insights into role of bromodomain, testis-specific (Brdt) in acetylated histone H4-dependent chromatin remodeling in mammalian spermiogenesis. J. Biol. Chem. 287, 6387–6405
- Eckert, D., Biermann, K., Nettersheim, D., Gillis, A. J. M., Steger, K., Jäck, H.-M., et al. (2008). Expression of BLIMP1/PRMT5 and concurrent histone H2A/H4 arginine 3 dimethylation in fetal germ cells, CIS/IGCNU and germ cell tumors. *BMC Dev. Biol.* 8, 106. doi: 10.1186/1471-213X-8-106
- Eddy, E. M., Washburn, T. F., Bunch, D. O., Goulding, E. H., Gladen, B. C., Lubahn, D. B., et al. (1996). Targeted disruption of the estrogen receptor gene in male mice causes alteration of spermatogenesis and infertility. Endocrinology 137, 4796–4805.
- Ellinger, J., Albers, P., Perabo, F. G., Müller, S. C., von Ruecker, A., and Bastian, P. J. (2009). CpG island hypermethylation of cell-free circulating serum DNA in patients with testicular cancer. *J. Urol.* 182, 324–329.

- Fenic, I., Hossain, H. M., Sonnack, V., Tchatalbachev, S., Thierer, F., Trapp, J., et al. (2008). In vivo application of histone deacetylase inhibitor trichostatin-a impairs murine male meiosis. J. Androl. 29, 172–185.
- Ferlin, A., Bogatcheva, N. V., Gianesello, L., Pepe, A., Vinanzi, C., Agoulnik, A. I., et al. (2006). Insulin-like factor 3 gene mutations in testicular dysgenesis syndrome: clinical and functional characterization. *Mol. Hum. Reprod.* 12, 401–406.
- Ferlin, A., Ganz, F., Pengo, M., Selice, R., Frigo, A. C., and Foresta, C. (2010). Association of testicular germ cell tumor with polymorphisms in estrogen receptor and steroid metabolism genes. *Endocr. Relat. Cancer* 17, 17–25.
- Ferlin, A., Zuccarello, D., Garolla, A., Selice, R., Vinanzi, C., Ganz, F., et al. (2009). Mutations in INSL3 and RXFP2 genes in cryptorchid boys. *Ann. N. Y. Acad. Sci.* 1160, 213–214.
- Figueroa, J. D., Sakoda, L. C., Graubard, B. I., Chanock, S., Rubertone, M. V., Erickson, R. L., et al. (2008). Genetic variation in hormone metabolizing genes and risk of testicular germ cell tumors. Cancer Causes Control 19, 917–929.
- Fritzsche, F. R., Hasler, A., Bode, P. K., Adams, H., Seifert, H. H., Sulser, T., et al. (2011). Expression of histone deacetylases 1, 2 and 3 in histological subtypes of testicular germ cell tumours. *Histol. Histopathol.* 26, 1555–1561.
- Garolla, A., Ferlin, A., Vinanzi, C., Roverato, A., Sotti, G., Artibani, W., et al. (2005). Molecular analysis of the androgen receptor gene in testicular cancer. *Endocr. Relat. Cancer* 12, 645–655.
- Giannandrea, F., Gandini, L., Paoli, D., Turci, R., and Figà-Talamanca, I. (2011). Pesticide exposure and serum organochlorine residuals among testicular cancer patients and healthy controls. J. Environ. Sci. Health B 46, 780–787.
- Godmann, M., Lambrot, R., and Kimmins, S. (2009). The dynamic epigenetic program in male germ cells: its role in spermatogenesis, testis cancer, and its response to the environment. *Microsc. Res. Tech.* 72, 603–619.
- Greene, M. H., Kratz, C. P., Mai, P. L., Mueller, C., Peters, J. A., Bratslavsky, G., et al. (2010). Familial testicular germ cell tumors in adults: 2010 summary of genetic risk factors and clinical phenotype. *Endocr. Relat. Cancer* 17, R109–R121.
- Gryder, B. E., Sodji, Q. H., and Oyelere, A. K. (2012). Targeted cancer therapy: giving histone deacetylase inhibitors

- all they need to succeed. Future Med. Chem. 4, 505–524.
- Gu, Y., Runyan, C., Shoemaker, A., Surani, A., and Wylie, C. (2009). Steel factor controls primordial germ cell survival and motility from the time of their specification in the allantois, and provides a continuous niche throughout their migration. Development 136, 1295–1303.
- Hardell, L., Bavel, B., Lindström, G., Eriksson, M., and Carlberg, M. (2006). In utero exposure to persistent organic pollutants in relation to testicular cancer risk. *Int. J. Androl.* 29, 228–234.
- Hardell, L., Malmqvist, N., Ohlson, C.-G., Westberg, H., and Eriksson, M. (2004a). Testicular cancer and occupational exposure to polyvinyl chloride plastics: a case–control study. *Int. J. Cancer* 109, 425–429.
- Hardell, L., van Bavel, B., Lindström, G., Carlberg, M., Eriksson, M., Dreifaldt, A. C., et al. (2004b). Concentrations of polychlorinated biphenyls in blood and the risk for testicular cancer. *Int. J. Androl.* 27, 282–290.
- Hardell, L., van Bavel, B., Lindström, G., Carlberg, M., Dreifaldt, A. C., Wijkström, H., et al. (2003). Increased concentrations of polychlorinated biphenyls, hexachlorobenzene, and chlordanes in mothers of men with testicular cancer. *Environ. Health Per*spect. 111, 930–934.
- He, Z., Kokkinaki, M., Pant, D., Gallicano, G. I., and Dym, M. (2009).Small RNA molecules in the regulation of spermatogenesis. *Reproduction* 137, 901–911.
- Hess, R. A., Bunick, D., Lee, K. H., Bahr, J., Taylor, J. A., Korach, K. S., and Lubahn, D. B. (1997). A role for oestrogens in the male reproductive system. *Nature* 390, 509–512.
- Hines, E. P., Calafat, A. M., Silva, M. J.,
  Mendola, P., and Fenton, S. E. (2009).
  Concentrations of phthalate metabolites in milk, urine, saliva, and serum of lactating North Carolina women.
  Environ. Health Perspect. 117, 86–92.
- Hinz, S., Magheli, A., Weikert, S., Schulze, W., Krause, H., Schrader, M., et al. (2010). Deregulation of EZH2 expression in human spermatogenic disorders and testicular germ cell tumors. World J. Urol. 28, 631–635.
- Huang, L., Huang, R., Ran, X.-R., Liu, H.-Y., Zhang, Y., Dai, L.-J., et al. (2011). Three-generation experiment showed female C57BL/6J mice drink drainage canal water containing low level of TCDD-like activity causing high pup mortality. *J. Toxicol. Sci.* 36, 713–724.
- Iguchi, T., Watanabe, H., and Katsu, Y. (2001). Developmental effects of

- estrogenic agents on mice, fish, and frogs: a mini-review. *Horm. Behav.* 40, 248–251.
- Ivell, R., and Anand-Ivell, R. (2011). Biological role and clinical significance of insulin-like peptide 3. Curr. Opin. Endocrinol. Diabetes Obes. 18, 210–216.
- Jacobsen, G. K., and Henriques, U. V. (1992). A fetal testis with intratubular germ cell neoplasia (ITGCN). Mod. Pathol. 5, 547–549.
- Jégou, B. (1993). The Sertoli-germ cell communication network in mammals. Int. Rev. Cytol. 147, 25–96.
- Joensen, U. N., Jørgensen, N., Rajpert-De Meyts, E., and Skakkebaek, N. E. (2008). Testicular dysgenesis syndrome and Leydig cell function. *Basic Clin. Pharmacol. Toxicol.* 102, 155–161.
- Johnson, L., Wilker, C. E., Safe, S. H., Scott, B., Dean, D. D., and White, P. H. (1994). 2,3,7,8-Tetrachlorodibenzop-dioxin reduces the number, size, and organelle content of Leydig cells in adult rat testes. *Toxicology* 89, 49–65.
- Kaleva, M., and Toppari, J. (2005). Cryptorchidism: an indicator of testicular dysgenesis? *Cell Tissue Res.* 322, 167–172.
- Kanetsky, P. A., Mitra, N., Vardhanabhuti, S., Li, M., Vaughn, D. J., Letrero, R., et al. (2009). Common variation in KITLG and at 5q31.3 predisposes to testicular germ cell cancer. *Nat. Genet.* 41, 811–815.
- Kibanov, M. V., Gvozdev, V. A., and Olenina, L. V. (2012). Germ granules in spermatogenesis of *Drosophila*: evidences of contribution to the piRNA silencing. *Commun. Integr. Biol.* 5, 130–133.
- Kleeman, J. M., Moore, R. W., and Peterson, R. E. (1990). Inhibition of testicular steroidogenesis in 2,3,7,8-tetrachlorodibenzo-pdioxin-treated rats: evidence that the key lesion occurs prior to or during pregnenolone formation. *Toxicol.* Appl. Pharmacol. 106, 112–125.
- Korhonen, H. M., Meikar, O., Yadav, R. P., Papaioannou, M. D., Romero, Y., Da Ros, M., et al. (2011). Dicer is required for haploid male germ cell differentiation in mice. *PLoS ONE* 6, e24821. doi: 10.1371/journal.pone.0024821
- Koster, R., di Pietro, A., Timmer-Bosscha, H., Gibcus, J. H., van den Berg, A., Suurmeijer, A. J., et al. (2010). Cytoplasmic p21 expression levels determine cisplatin resistance in human testicular cancer. *J. Clin. Invest.* 120, 3594–3605.
- Koubova, J., Menke, D. B., Zhou, Q., Capel, B., Griswold, M. D., and Page,

- D. C. (2006). Retinoic acid regulates sex-specific timing of meiotic initiation in mice. *Proc. Natl. Acad. Sci. U.S.A.* 103, 2474–2479.
- Koychev, D., Oechsle, K., Bokemeyer, C., and Honecker, F. (2011). Treatment of patients with relapsed and/or cisplatin-refractory metastatic germ cell tumours: an update. *Int. J. Androl.* 34(Pt 2), e266–e273.
- Kristiansen, W., Aschim, E. L., Andersen, J. M., Witczak, O., Fosså, S. D., and Haugen, T. B. (2012). Variations in testosterone pathway genes and susceptibility to testicular cancer in Norwegian men. *Int. J. Androl.* doi: 10.1111/j.1365-2605.2012.01297.x [Epub ahead of print].
- Kristiansen, W., Haugen, T. B., Witczak, O., Andersen, J. M., Fosså, S. D., and Aschim, E. L. (2011). CYP1A1, CYP3A5 and CYP3A7 polymorphisms and testicular cancer susceptibility. *Int. J. Androl.* 34, 77–83.
- Lai, K. P., Wong, M. H., and Wong, C. K. C. (2005). Inhibition of CYP450scc expression in dioxin-exposed rat Leydig cells. J. Endocrinol. 185, 519–527.
- Lambrot, R., Jones, S., Saint-Phar, S., and Kimmins, S. (2012). Specialized distribution of the histone methyltransferase ezh2 in the nuclear apical region of round spermatids and its interaction with the histone variant h1t2. *J. Androl.* 33, 1058–1066.
- Lambrot, R., and Kimmins, S. (2011). Histone methylation is a critical regulator of the abnormal expression of POU5F1 and RASSF1A in testis cancer cell lines. *Int. J. Androl.* 34, 110–123.
- La Salle, S., Oakes, C. C., Neaga, O. R., Bourc'his, D., Bestor, T. H., and Trasler, J. M. (2007). Loss of spermatogonia and wide-spread DNA methylation defects in newborn male mice deficient in DNMT3L. BMC Dev. Biol. 7, 104. doi: 10.1186/1471-213X-7-104
- Li, H., MacLean, G., Cameron, D., Clagett-Dame, M., and Petkovich, M. (2009). Cyp26b1 expression in murine Sertoli cells is required to maintain male germ cells in an undifferentiated state during embryogenesis. PLoS ONE 4, e7501. doi: 10.1371/journal.pone.0007501
- Lian, J., Zhang, X., Tian, H., Liang, N., Wang, Y., Liang, C., et al. (2009). Altered microRNA expression in patients with nonobstructive azoospermia. Reprod. Biol. Endocrinol. 7, 13.
- Lubahn, D. B., Moyer, J. S., Golding, T. S., Couse, J. F., Korach, K. S., and Smithies, O. (1993). Alteration of reproductive function but

- not prenatal sexual development after insertional disruption of the mouse estrogen receptor gene. *Proc. Natl. Acad. Sci. U.S.A.* 90, 11162–11166.
- Mannuel, H. D., Mitikiri, N., Khan, M., and Hussain, A. (2012). Testicular germ cell tumors: biology and clinical update. *Curr. Opin. Oncol.* 24, 266–271.
- Manton, K. J., Douglas, M. L., Netzel-Arnett, S., Fitzpatrick, D. R., Nicol, D. L., Boyd, A. W., et al. (2005). Hypermethylation of the 5' CpG island of the gene encoding the serine protesse testisin promotes its loss in testicular tumorigenesis. *Br. J. Cancer* 92, 760–769.
- Maselli, J., Hales, B. F., Chan, P., and Robaire, B. (2012). Exposure to bleomycin, etoposide, and *cis*-platinum alters rat sperm chromatin integrity and sperm head protein profile. *Biol. Reprod.* 86, 166, 1–10.
- McGlynn, K. A., Quraishi, S. M., Graubard, B. I., Weber, J.-P., Rubertone, M. V., and Erickson, R. L. (2009). Polychlorinated biphenyls and risk of testicular germ cell tumors. *Cancer Res.* 69, 1901–1909.
- Minami, K., Chano, T., Kawakami, T., Ushida, H., Kushima, R., Okabe, H., et al. (2010). DNMT3L is a novel marker and is essential for the growth of human embryonal carcinoma. *Clin. Cancer Res.* 16, 2751–2759.
- Moore, R. W., Jefcoate, C. R., and Peterson, R. E. (1991). 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin inhibits steroidogenesis in the rat testis by inhibiting the mobilization of cholesterol to cytochrome P450scc. *Toxicol. Appl. Pharmacol.* 109, 85–97.
- Naciff, J. M., and Daston, G. P. (2004). Toxicogenomic approach to endocrine disrupters: identification of a transcript profile characteristic of chemicals with estrogenic activity. *Toxicol. Pathol.* 32(Suppl. 2), 59–70.
- Nagahama, Y., Nakamura, M., Kitano, T., and Tokumoto, T. (2004). Sexual plasticity in fish: a possible target of endocrine disruptor action. *Environ*. Sci. 11, 73–82
- Nettersheim, D., Biermann, K., Gillis, A. J. M., Steger, K., Looijenga, L. H. J., and Schorle, H. (2011). NANOG promoter methylation and expression correlation during normal and malignant human germ cell development. *Epigenetics* 6, 114–122.
- Niu, Z., Goodyear, S. M., Rao, S., Wu, X., Tobias, J. W., Avarbock, M. R., et al. (2011). MicroRNA-21 regulates the self-renewal of mouse spermatogonial stem cells. *Proc. Natl. Acad. Sci.* U.S.A. 108, 12740–12745.

- Novotny, G. W., Belling, K. C., Bramsen, J. B., Nielsen, J. E., Bork-Jensen, J., Almstrup, K., et al. (2012). MicroRNA expression profiling of carcinoma in situ cells of the testis. *Endocr. Relat. Cancer* 19, 365–379.
- O'Carroll, D., Scherthan, H., Peters, A. H., Opravil, S., Haynes, A. R., Laible, G., et al. (2000). Isolation and characterization of Suv39h2, a second histone H3 methyltransferase gene that displays testis-specific expression. *Mol. Cell. Biol.* 20, 9423–9433.
- Ohlson, C. G., and Hardell, L. (2000). Testicular cancer and occupational exposures with a focus on xenoestrogens in polyvinyl chloride plastics. *Chemosphere* 40, 1277–1282.
- Okamoto, K., and Kawakami, T. (2007). Epigenetic profile of testicular germ cell tumours. *Int. J. Androl.* 30, 385–392.
- Olesen, I. A., Sonne, S. B., Hoei-Hansen, C. E., Rajpert-DeMeyts, E., and Skakkebaek, N. E. (2007). Environment, testicular dysgenesis and carcinoma in situ testis. *Best Pract. Res. Clin. Endocrinol. Metab.* 21, 462–478.
- Omisanjo, O. A., Biermann, K., Hartmann, S., Heukamp, L. C., Sonnack, V., Hild, A., et al. (2007). DNMT1 and HDAC1 gene expression in impaired spermatogenesis and testicular cancer. *Histochem. Cell Biol.* 127, 175–181.
- Papaioannou, M. D., and Nef, S. (2010). microRNAs in the testis: building up male fertility. *J. Androl.* 31, 26–33.
- Pentikäinen, V., Erkkilä, K., Suomalainen, L., Parvinen, M., and Dunkel, L. (2000). Estradiol acts as a germ cell survival factor in the human testis in vitro. J. Clin. Endocrinol. Metab. 85, 2057–2067.
- Peters, A. H., O'Carroll, D., Scherthan, H., Mechtler, K., Sauer, S., Schöfer, C., Weipoltshammer, K., et al. (2001). Loss of the Suv39h histone methyltransferases impairs mammalian heterochromatin and genome stability. *Cell* 107, 323–337.
- Purdue, M. P., Engel, L. S., Langseth, H., Needham, L. L., Andersen, A., Barr, D. B., et al. (2009). Prediagnostic serum concentrations of organochlorine compounds and risk of testicular germ cell tumors. *Environ. Health Perspect.* 117, 1514–1519.
- Rajpert-De Meyts, E., Leffers, H., Daugaard, G., Andersen, C. B., Petersen, P. M., Hinrichsen, J., et al. (2002). Analysis of the polymorphic CAG repeat length in the androgen receptor gene in patients with testicular germ cell cancer. *Intl. J. Cancer* 102, 201–204.
- Richardson, L. C., Neri, A. J., Tai, E., and Glenn, J. D. (2012). Testicular

- cancer: a narrative review of the role of socioeconomic position from risk to survivorship. *Urol. Oncol.* 30, 95–101.
- Richiardi, L., Bellocco, R., Adami, H.-O., Torrång, A., Barlow, L., Hakulinen, T., et al. (2004). Testicular cancer incidence in eight northern European countries: secular and recent trends. Cancer Epidemiol. Biomarkers Prev. 13, 2157–2166.
- Robertson, K. M., O'Donnell, L., Jones, M. E., Meachem, S. J., Boon, W. C., Fisher, C. R., et al. (1999). Impairment of spermatogenesis in mice lacking a functional aromatase (cyp 19) gene. Proc. Natl. Acad. Sci. U.S.A. 96, 7986–7991.
- Rodriguez, S., Jafer, O., Goker, H., Summersgill, B. M., Zafarana, G., Gillis, A. J. M., et al. (2003). Expression profile of genes from 12p in testicular germ cell tumors of adolescents and adults associated with i(12p) and amplification at 12p11.2-p12.1. Oncogene 22, 1880–1891.
- Romerius, P., Giwercman, A., Moëll, C., Relander, T., Cavallin-Ståhl, E., Wiebe, T., et al. (2011). Estrogen receptor α single nucleotide polymorphism modifies the risk of azoospermia in childhood cancer survivors. *Pharmacogenet. Genomics* 21, 263–269.
- Rubin, B. S. (2011). Bisphenol A: an endocrine disruptor with widespread exposure and multiple effects. J. Steroid Biochem. Mol. Biol. 127, 27–34.
- Sasaki, A., Taketomi, T., Kato, R., Saeki, K., Nonami, A., Sasaki, M., et al. (2003). Mammalian Sprouty4 suppresses Ras-independent ERK activation by binding to Raf1. Nat. Cell Biol. 5, 427–432.
- Schoevers, E. J., Santos, R. R., Colenbrander, B., Fink-Gremmels, J., and Roelen, B. A. J. (2012). Transgenerational toxicity of Zearalenone in pigs. *Reprod. Toxicol.* 34, 110–119.
- Schotta, G., Ebert, A., and Reuter, G. (2003). SU(VAR)3-9 is a conserved key function in heterochromatic gene silencing. *Genetica* 117, 149–158.
- Sharpe, R. M., and Skakkebaek, N. E. (1993). Are oestrogens involved in falling sperm counts and disorders of the male reproductive tract? *Lancet* 341, 1392–1395.
- Sikka, S. C., and Wang, R. (2008). Endocrine disruptors and estrogenic effects on male reproductive axis. Asian J. Androl. 10, 134–145.
- Skakkebaek, N. E., Rajpert-De Meyts, E., and Main, K. M. (2001). Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental

- aspects. *Hum. Reprod.* 16 972–978.
- Skinner, M. K., Norton, J. N., Mullaney, B. P., Rosselli, M., Whaley, P. D., and Anthony, C. T. (1991). Cell–cell interactions and the regulation of testis function. *Ann. N. Y. Acad. Sci.* 637, 354–363.
- Song, N., Liu, J., An, S., Nishino, T., Hishikawa, Y., and Koji, T. (2011a). Immunohistochemical analysis of histone H3 modifications in germ cells during mouse spermatogenesis. Acta Histochem. Cytochem. 44, 183–190.
- Song, R., Hennig, G. W., Wu, Q., Jose, C., Zheng, H., and Yan, W. (2011b). Male germ cells express abundant endogenous siRNAs. *Proc. Natl. Acad. Sci. U.S.A.* 108, 13159–13164.
- Sonne, S. B., Kristensen, D. M., Novotny, G. W., Olesen, I. A., Nielsen, J. E., Skakkebaek, N. E., et al. (2008). Testicular dysgenesis syndrome and the origin of carcinoma in situ testis. *Int. J. Androl.* 31, 275–287.
- Steilmann, C., Paradowska, A., Bartkuhn, M., Vieweg, M., Schuppe, H.-C., Bergmann, M., et al. (2011). Presence of histone H3 acetylated at lysine 9 in male germ cells and its distribution pattern in the genome of human spermatozoa. Reprod. Fertil. Dev. 23, 997–1011.
- Suh, N., and Blelloch, R. (2011). Small RNAs in early mammalian development: from gametes to gastrulation. *Development* 138, 1653–1661.
- Sultan, C., Lumbroso, S., Poujol, N., Belon, C., Boudon, C., and Lobaccaro, J. M. (1993). Mutations of androgen receptor gene in androgen insensitivity syndromes. J. Steroid Biochem. Mol. Biol. 46, 519–530.
- Summersgill, B., Goker, H., Weber-Hall, S., Huddart, R., Horwich, A., and Shipley, J. (1998). Molecular cytogenetic analysis of adult testicular germ cell tumours and identification of regions of consensus copy number change. Br. J. Cancer 77, 305–313.
- Suzuki, Y., Yoshinaga, J., Mizumoto, Y., Serizawa, S., and Shiraishi, H. (2012). Foetal exposure to phthalate esters and anogenital distance in male newborns. *Int. J. Androl.* 35, 236–244.
- Tachibana, M., Nozaki, M., Takeda, N., and Shinkai, Y. (2007). Functional dynamics of H3K9 methylation during meiotic prophase progression. *EMBO J.* 26, 3346–3359.
- Tong, M.-H., Mitchell, D. A., McGowan,
  S. D., Evanoff, R., and Griswold, M. D.
  (2012). Two miRNA clusters, Mir-1792 (Mirc1) and Mir-106b-25 (Mirc3),
  are involved in the regulation of spermatogonial differentiation in mice.
  Biol. Reprod. 86, 72.

- Toppari, J., Kaleva, M., Virtanen, H. E., Main, K. M., and Skakkebaek, N. E. (2007). Luteinizing hormone in testicular descent. Mol. Cell. Endocrinol. 269, 34–37.
- Toppari, J., Virtanen, H. E., Main, K. M., and Skakkebaek, N. E. (2010). Cryptorchidism and hypospadias as a sign of testicular dysgenesis syndrome (TDS): environmental connection. *Birth Defects Res. A Clin. Mol. Teratol.* 88, 910–919.
- Townsend, J. S., Richardson, L. C., and German, R. R. (2010). Incidence of testicular cancer in the United States, 1999–2004. Am. J. Mens Health 4, 353–360.
- Turnbull, C., and Rahman, N. (2011). Genome-wide association studies provide new insights into the genetic basis of testicular germ-cell tumour. *Int. J. Androl.* 34(Pt 2), e86–e96.
- Ugwumba, F. O., and Aghaji, A. E. (2010). Testicular cancer: management challenges in an African developing country. S. Afr. Med. J. 100, 452–455.
- Umeh, K., and Chadwick, R. (2010). Early detection of testicular cancer: revisiting the role of self-efficacy in testicular self-examination among young asymptomatic males. *J. Behav. Med.* doi: 10.1007/s10865-010-9262-z [Epub ahead of print].
- van de Geijn, G.-J. M., Hersmus, R., and Looijenga, L. H. J. (2009). Recent developments in testicular germ cell tumor research. *Birth Defects Res. C Embryo Today* 87, 96–113.
- Vanparys, C., Hectors, T. L. M., Blust, R., and De Coen, W. (2012). Mechanistic profiling of the cAMPdependent steroidogenic pathway in the H295R endocrine disrupter screening system: new endpoints for toxicity testing. *Toxicol. Lett.* 208, 174–184.
- Verhoeven, G. (1992). Local control systems within the testis. *Baillieres Clin. Endocrinol. Metab.* 6, 313–333.
- Verhoeven, G., Willems, A., Denolet, E., Swinnen, J. V., and De Gendt, K. (2010). Androgens and spermatogenesis: lessons from transgenic mouse models. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 365, 1537–1556.
- Volle, D. H., Decourteix, M., Garo, E., McNeilly, J., Fenichel, P., Auwerx, J., et al. (2009). The orphan nuclear receptor small heterodimer partner mediates male infertility induced by diethylstilbestrol in mice. J. Clin. Invest. 119, 3752–3764.
- Volle, D. H., Duggavathi, R., Magnier, B. C., Houten, S. M., Cummins, C. L., Lobaccaro, J.-M. A., et al. (2007). The small heterodimer partner is a gonadal gatekeeper of sexual

- maturation in male mice. *Genes Dev.* 21, 303–315
- Walsh, T. J., Dall'Era, M. A., Croughan, M. S., Carroll, P. R., and Turek, P. J. (2007). Prepubertal orchiopexy for cryptorchidism may be associated with lower risk of testicular cancer. *J. Urol.* 178(Pt 1), 1440–1446.
- Wang, J., Lu, F., Ren, Q., Sun, H., Xu, Z., Lan, R., et al. (2011). Novel histone demethylase LSD1 inhibitors selectively target cancer cells with pluripotent stem cell properties. *Cancer Res.* 71, 7238–7249.
- Wang, R.-S., Yeh, S., Tzeng, C.-R., and Chang, C. (2009). Androgen receptor roles in spermatogenesis and fertility: lessons from testicular cell-specific androgen receptor knockout mice. *Endocr. Rev.* 30, 119–132.
- Warde, P., Huddart, R., Bolton, D., Heidenreich, A., Gilligan, T., and Fossa, S. (2011). Management of localized seminoma, stage I-II: SIU/ICUD Consensus Meeting on Germ Cell Tumors (GCT), Shanghai 2009. *Urology* 78(Suppl.), S435–S443.
- Webster, K. E., O'Bryan, M. K., Fletcher, S., Crewther, P. E., Aapola, U., Craig, J., et al. (2005). Meiotic and epigenetic defects in Dnmt3L-knockout mouse spermatogenesis. *Proc. Natl. Acad. Sci. U.S.A.* 102, 4068–4073.
- Western, P. (2009). Foetal germ cells: striking the balance between pluripotency and differentiation. *Int. J. Dev. Biol.* 53, 393–409.
- Woodruff, T. J. (2011). Bridging epidemiology and model organisms to increase understanding of endocrine disrupting chemicals and human health effects. J. Steroid Biochem. Mol. Biol. 127, 108–117.
- Wu, J., Bao, J., Wang, L., Hu, Y., and Xu, C. (2011). MicroRNA-184 downregulates nuclear receptor corepressor 2 in mouse spermatogenesis. BMC Dev. Biol. 11, 64. doi: 10.1186/1471-213X-11-64
- Wu, Q., Song, R., Ortogero, N., Zheng, H., Evanoff, R., Small, C. L., et al. (2012). The RNase III enzyme DROSHA is essential for microRNA production and spermatogenesis. *J. Biol. Chem.* 287, 25173–25190.
- Yamada, S., Kohu, K., Ishii, T., Ishi-doya, S., Ishidoya, S., Hiramatsu, M., et al. (2004). Gene expression profiling identifies a set of transcripts that are up-regulated inhuman testicular seminoma. DNA Res. 11, 335–344.
- Yaman, R., and Grandjean, V. (2006). Timing of entry of meiosis depends on a mark generated by DNA methyltransferase 3a in testis. *Mol. Reprod. Dev.* 73, 390–397.
- Zhou, Y., Zha, J., Xu, Y., Lei, B., and Wang, Z. (2012). Occurrences

of six steroid estrogens from different effluents in Beijing, China. *Environ. Monit. Assess.* 184, 1719–1729.

Ziglioli, F., Maestroni, U., Dinale, F., Ciuffreda, M., and Cortellini, P. (2011). Carcinoma in situ (CIS) of the testis. Acta Biomed. 82, 162–169.

Conflict of Interest Statement: The authors declare that the research was

conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 27 September 2012; paper pending published: 24 October 2012; accepted: 13 November 2012; published online: 29 November 2012. Citation: Vega A, Baptissart M, Caira F, Brugnon F, Lobaccaro J-MA and Volle DH (2012) Epigenetic: a molecular link between testicular cancer and environmental exposures. Front. Endocrin. 3:150. doi: 10.3389/fendo.2012.00150

This article was submitted to Frontiers in Cancer Endocrinology, a specialty of Frontiers in Endocrinology. Copyright © 2012 Vega, Baptissart, Caira, Brugnon, Lobaccaro and Volle. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.