



Preclinical evidence supporting the clinical development of central pattern generator-modulating therapies for chronic spinal cord-injured patients

Pierre A. Guertin^{1,2*}

¹ Department of Psychiatry and Neurosciences, Laval University, Quebec City, QC, Canada

² Spinal Cord Injury and Functional Recovery Laboratory, Laval University Medical Center (CHU de Quebec), Quebec City, QC, Canada

Edited by:

Marco Iosa, Fondazione Santa Lucia, Italy

Reviewed by:

Auke Ijspeert, Ecole Polytechnique Federale de Lausanne (EPFL), Switzerland
Yuri P. Ivanenko, IRCCS Fondazione Santa Lucia, Italy

*Correspondence:

Pierre A. Guertin, Spinal Cord Injury and Functional Recovery Laboratory, Laval University Medical Center (CHU de Quebec), 2705 Laurier Boulevard, Room RC-9800 (Neuroscience Unit), Quebec City, QC G1V 4G2, Canada
e-mail: Pierre.Guertin@crchul.ulaval.ca

Ambulation or walking is one of the main gaits of locomotion. In terrestrial animals, it may be defined as a series of rhythmic and bilaterally coordinated movement of the limbs which creates a forward movement of the body. This applies regardless of the number of limbs—from arthropods with six or more limbs to bipedal primates. These fundamental similarities among species may explain why comparable neural systems and cellular properties have been found, thus far, to control in similar ways locomotor rhythm generation in most animal models. The aim of this article is to provide a comprehensive review of the known structural and functional features associated with central nervous system (CNS) networks that are involved in the control of ambulation and other stereotyped motor patterns—specifically Central Pattern Generators (CPGs) that produce basic rhythmic patterned outputs for locomotion, micturition, ejaculation, and defecation. Although there is compelling evidence of their existence in humans, CPGs have been most studied in reduced models including *in vitro* isolated preparations, genetically-engineered mice and spinal cord-transected animals. Compared with other structures of the CNS, the spinal cord is generally considered as being well-preserved phylogenetically. As such, most animal models of spinal cord-injured (SCI) should be considered as valuable tools for the development of novel pharmacological strategies aimed at modulating spinal activity and restoring corresponding functions in chronic SCI patients.

Keywords: CPG, locomotion, SGE, ejaculation, LDC, defecation, SMC, micturition

INTRODUCTION

Locomotion is the act of self-propulsion by an animal (Hugues and Wiersma, 1960; Delcomyn, 1977; Kandel et al., 2000; Hopper and DiCaprio, 2004). Forms of terrestrial locomotion generally include walking, running, and hopping. In vertebrates, its control depends upon several neural systems that ensure propulsion, body orientation (equilibrium or postural control), and steering (goal-direction control) (Ivanenko et al., 2006). Certain areas of the brain have for role, through signals sent via descending neural pathways to the spinal cord, to trigger and modulate basic locomotor outputs generated spinally. The latter are organized essentially by a network localized in the lumbar segments of the spinal cord, generally referred to as the Central Pattern Generator (CPG) for locomotion. That network is responsible for much of the timing and pattern of the complex, rhythmic, coordinated muscle activities that underlie locomotion.

Abbreviations: CPG, Central Pattern Generator; SMC, Sacral or Spinal Micturition Center; LDC, Lumbosacral Defecation Center; SGE, Spinal Generator for Ejaculation; TX, Spinal Cord Transection; SCI, Spinal Cord Injury; NMDA, N-Methyl-D-aspartate; 5-HT, Serotonin; DA, Dopamine; CNS, Central Nervous System; EMG, Electromyogram.

In addition to this direct contribution from central systems, there are also different feedback and feedforward loops that use peripheral cues for proper adaptation of gait under different circumstances. Since most methods and experimental tools needed for studying these systems at the cellular level are rather invasive, it has remained difficult to study them in great details in humans. However, research in different animal models has revealed significant details about the organization and function of each constitutive element of these systems (Buschges et al., 2008). In most cases, great similarities have been found between neuronal circuits that generate rhythmic motor patterns among species. In fact, just examining similarities between various forms of locomotion among species already provide preliminary evidence of well-preserved spinal mechanisms throughout evolution. The neural system controlling locomotion is probably the most studied systems thus far among all spinal systems underlying comparable stereotyped motor behaviors such as micturition, ejaculation or defecation (Guertin and Steuer, 2009). Nonetheless, from compiling data gathered about these other systems more recently, it may be said that great similarities seem to exist among spinal systems that control these fundamental behaviors.

DIFFERENT FORMS OF LOCOMOTION AMONG VERTEBRATE SPECIES

Although some differences exist between forward and backward locomotion in cats, mice humans, salamanders or lampreys, to name a few, most of the basic alternating patterns between flexor-like and extensor-like muscles are generally maintained even in invertebrate species (Székely et al., 1969; Pearson and Duysens, 1976; Kristan and Weeks, 1983; Robertson and Pearson, 1985; Pearson, 1993; Currie and Lee, 1997; Cheng et al., 1998; Currie, 1999; Matsumoto et al., 2007; Liu et al., 2012). For instance, lampreys and salamanders swim by contracting muscles on either side of the body in order to generate waves of flexion that travel the length of the body from nose to tail (rostrocaudal propagation of undulations), generally getting larger as they go along (Getting, 1977). Locomotion on land raises different complications, such as the effects of gravity. Nonetheless, walking—the most common gait in legged animals, shares comparable key features with swimming. Indeed, it is essentially composed of a series of rhythmic contractions between antagonist muscles of the main moving segments, the legs instead of the whole body. More specifically, it is the result of successive coordinated contractions between flexors and extensors of all limbs accompanied of bilateral alternation for bipeds and quadrupeds.

SEVERAL NEURAL NETWORKS INVOLVED

As mentioned earlier, it is now generally accepted that the basic motor patterns and rhythms underlying walking (i.e., the most extensively studied form of locomotion) are organized and generated in the spinal cord by the CPG (Grillner, 1981; Guertin, 2009a). However, many other networks in the CNS have key roles to play for successful locomotion to be achieved. For instance, supraspinal centers in the brainstem and forebrain are essential for initiating and controlling locomotor movements. One of them located in the cerebellum plays an important role in sensorimotor control and in intra- and inter-limb coordination. The vermis and the intermediate region of the cerebellum receive information through the spinocerebellar pathways about the ongoing activities in the CPG and the somatosensory receptors. The information is conveyed to Purkinje neurons, transformed, returned to brainstem descending tract neurons also involved in the initiation of locomotion. Another important center is the mesencephalic locomotor region (MLR), first discovered in cats and later found in all vertebrate species tested to date (Orlovskii et al., 1966; Orlovski et al., 1966; Orlovsky and Shik, 1976; Jordan et al., 2008; Juvin et al., 2012). The reticulospinal tract that originates in the reticular formation has also been found to be essential for the production of locomotor activity evoked by brainstem stimulation in many different species, its activation is necessary for the initiation of locomotion in normal conditions (Grillner, 1981; Cheng and Magnuson, 2011; Le Ray et al., 2011). Along this idea, locomotor-initiating centers in the midbrain/pontine tegmentum and cerebellum converge in the reticular nucleus before descending to the spinal cord.

SPINAL CORD ORGANIZATION

It is impossible to provide an update on neural systems that control locomotion or other stereotyped motor behaviors, specifically

those in the spinal cord, without reviewing first, fundamental details about spinal cord organization (Guertin, 2012). The spinal cord has often been considered as a simple relay between brain cells and effective organs (muscles, skin, etc.). However, it is increasingly recognized that the spinal cord is also a “command center” involved in the control and modulation of several functions (Kandel et al., 2000). With its one billion neurons (Kalat, 1998), it is definitely a significant structure of the CNS that can control both simple motor acts such as reflexes (e.g., monosynaptic excitatory, reciprocal inhibitory, withdrawal and crossed-extension reflexes) as well as more complex motor functions such as locomotion, bladder/bowel control, and sexual function (Guertin, 2012).

GROSS ANATOMY OF THE SPINAL CORD

The spinal cord constitutes the most caudally located structure of the CNS (Netter, 2006). Contained within the vertebral column, it extends from the medulla to the first lumbar vertebra (Patesta and Gartner, 2006). In humans, its long and thin elliptical structure varies in length between 43 and 45 cm comprising 31 segments—8 cervical, 12 thoracic, 5 lumbar, 5 sacral, and 1 coccygeal. Every segment is associated with a pair of nerves (and roots) on each side that comprises sensory nerve roots entering the spinal cord at each level, and motor roots emerging from the cord at each level. In rostral parts, the spinal nerves exit directly from (just above from C1 to C7 or just below from C8 and lower) the vertebra associated numerically with the corresponding spinal cord segment. However, in caudal parts of the spinal cord, the spinal nerves travel further down the column before exiting (also known as the cauda equine). In transverse sections, the spinal cord displays white and gray matter tissues. The white matter, located more peripherally, contains white matter tracts, ascending and descending myelinated fibers, carrying both sensory and motor inputs. The gray matter, more centrally located, is characterized by its butterfly-shape that contains unmyelinated cells and, specifically, simple as well as more complex spinal circuits. In the center, there is the central canal that contains cerebrospinal fluids formed mainly in the ventricular system (choroid plexus). Peripherally, the meninges are layers of tissue surrounding the spinal cord for its protection—the dura, the arachnoid and the pia. The latter is relatively thin and tightly associated with the surface of the spinal cord. The spinal cord is well-vascularized with blood vessels including the anterior spinal artery, the bilateral sulcal branches, the bilateral posterior spinal arteries, the pial arterial plexus as well as the anterior/posterior spinal veins, the anterior/posterior sulcal veins, and the pial venous plexus.

SPINAL CORD WHITE MATTER

Several tracts carry information between supraspinal (MLR, reticular formation, etc.) and spinal structures (e.g., motoneurons, simple reflexes, CPG, etc.). Ascending tracts convey sensory signals associated with the sense of touch, pressure, proprioception, and vibration via relatively large myelinated fibers (e.g., gracile fasciculus, cuneate fasciculus). They travel contralaterally (decussation) in the medulla prior to be redirected toward the thalamus and the sensory cortex. In contrast, the lateral or so-called anterolateral columns convey information about pain and

thermal sensation and decussate instead at the spinal cord level (close primary afferent entry) prior to ascend to the brain via a number of tracts (e.g., Lissauer's tract, dorsospinal tract, ventrospinal tract, spinocerebellar tract, spinothalamic tract, etc.).

The descending motor system is also divided in multiple tracts essentially composing the so-called pyramidal and extrapyramidal tracts. The corticospinal tracts, also called the pyramidal tracts, contain about one million axons on each side involved mainly in skilled movements. The extrapyramidal tracts are involved instead in the control of posture and locomotion (Bretzner and Drew, 2005; Barthélemy et al., 2011). The pyramidal tract originates from the cerebral cortex and from some brainstem motor nuclei. It constitutes the most direct descending motor pathway (e.g., some monosynaptic connections) between the motor cortex (Brodmann's areas 1, 2, 3, 4, and 6) and the final common motor pathway, namely the spinal motoneurons all segmental levels. About 80–90% of these corticospinal axons decussate contralaterally at the pyramid level in the medulla oblongata. From there, they form the lateral corticospinal tract that sends input to spinal motoneurons in the ventral horn. 10–20% descends instead ipsilaterally as the ventral corticospinal tract and decussates in the spinal cord prior to synapsing with spinal motoneurons. A few others form instead the corticobulbar tract that sends input to brainstem motoneurons for face, head and neck muscle control. Extrapyramidal tract neurons (rubrospinal, vestibulospinal, tectospinal and reticulospinal) originate from subcortical nuclei in the pons (reticular formation) and medulla oblongata where they send projections to spinal motoneurons of all segments. The rubrospinal tract receives inputs from the motor and premotor areas 4 and 6 prior to send projections contralaterally (decussation in the brain) to spinal motoneurons cervically. It is generally considered to facilitate flexion and inhibit extension in the upper extremities. The tectospinal tract sends axons in premotor lamina of the spinal cord (VI–VIII) cervically for neck and head motor control whereas the reticulospinal tract sends projections to all levels of the spinal cord for autonomic control (e.g., cardiovascular and respiratory functions, blood pressure, micturition, defecation). The vestibulospinal tract sends projections laterally or ventrally on the ipsilateral side of the spinal cord to laminae VII and VIII for extensor control (English, 1985).

SIMPLE REFLEX PATHWAYS OF THE GRAY MATTER

As described above, white matter tracts are mainly “relays” sending signals up (sensory-related) and down (motor control) between supraspinal and spinal structures. In contrast, gray matter neurons in the spinal cord form reflex pathways and complex neuronal networks. Simple reflex arcs or pathways are normally associated with well-known classical reflexes (Matthews, 1972, 1991). By definition, a simple reflex, in normal condition, typically leads to a rapid, predictable, repeatable, stereotyped and involuntary motor reaction in response to the corresponding specific stimulus. Most reflexes are mediated locally, within one or two spinal segments, by relatively simple neural pathways in the spinal cord—involving generally either one (monosynaptic), two (disynaptic) or more (polysynaptic) synapses and corresponding neurons (i.e., interneurons and motoneurons). Simple reflexes

may be of autonomic (related with inner organs, eyes, blood vessels, etc.) or somatic (related with skeletal muscle responses) origin. The latter has been more extensively studied mainly in the spinal cord-transected or decerebrate cat models. Among the main somatic spinal reflexes, the Ia, Ib, II, and FRA (flexion reflex afferent) reflexes have been particularly well-characterized although their roles in the control of locomotion or other stereotyped motor behaviors remain incompletely understood (Henneman, 1974; Forssberg and Hirschfeld, 1988; Field-Fote et al., 2012). This said, activation of some of these reflexes via electrical stimulation of muscles, nerves or lumbar spinal cord area in spinal cord-injured (SCI) patients was shown to temporarily elicit or promote some waking movements with or without pharmacological aids (see recent work from Edgerton's or Gerasimenko's groups) (Edgerton et al., 2001).

MONOSYNAPTIC REFLEX

The so-called Ia monosynaptic reflex arc is a well-characterized somatic spinal reflex pathway. It is considered the simplest and fastest reflex of all, mediating primary afferent (Ia) inputs from muscle spindles typically activated by muscle stretch (e.g., following a tendon jerk, tendon tap or myotatic reflex). Ia inputs establish monosynaptic connections with homonymous alpha-motoneurons in the ventral horn gray matter. This excitatory reflex increases homonymous muscle contraction in response to muscle elongation while inhibiting antagonist muscle contraction via collateral branching. It is generally considered to play a role in tonus and postural adjustments which is important for terrestrial locomotion. Its level of activity is phase and task-dependent—i.e., larger response during standing compared with walking or running as well as during extension (stance phase) compared with flexion (swing phase) (Stein and Capaday, 1988). Findings in decerebrate and paralyzed cats provided clear evidence of its role also in reflexively-increased extensor activity during stance and in resetting rhythm or cycle probably via direct input upon CPG elements (Guertin et al., 1995; Angel et al., 1996, 2005). Hence, Ia inputs during locomotion may serve to compensate for an unsuspected increase of loading during ambulation as well as to increase tonus in extensor muscles during stance (Guertin et al., 1995; Pearson, 1995; Guertin, 1996).

Ib REFLEX PATHWAYS

Also known as the inverse myotatic reflex or autogenic inhibitory reflex pathway, the Ib reflex arc is associated with peripheral afferent inputs from Ib afferent fibers and Golgi tendon organs. It has been shown to inhibit homonymous and synergistic alpha-motoneurons at rest via a disynaptic arc (two synapses involving one inhibitory neuron called the Ib interneuron), originally believed to serve as a protective mechanism against excessive muscle contraction. This view has eventually changed when discovering that a wide range of muscle activity and load can alter Ib firing. It is also task-dependent since during locomotion, Ib inhibition is replaced by Ib excitation to homonymous and synergistic alpha-motoneurons at all joints of the lower extremities via a rhythmically active candidate excitatory interneuron located in the lumbar enlargement area (lamina VII) (Gossard et al., 1994; Angel et al., 1996, 2005). Its role during locomotion may be also

to enhance muscular contraction of extensors during the stance phase and to reset stepping to extension when activated during the swing phase (Guertin et al., 1995; Pearson, 1995).

FLEXION (WITHDRAWAL) REFLEX PATHWAYS

The FRA pathway is activated specifically by high-threshold afferent fibers (e.g., associated with cutaneous nociceptor A or C fibers, group II, III and IV muscle afferent fibers, etc.) (Knikou et al., 2009). Ipsilaterally, it involves at least two interneurons (three or more synapses) over several segments prior to synapse upon alpha-motoneurons ipsilaterally for extended flexor contraction and extended extensor inhibition. With weaker stimulus, only ipsilateral effects are found. However, with stronger stimulus, opposite contralateral effects may also be found (excitation of extensor muscles and inhibition of flexors)—the crossed-extension reflex. These reflex pathways play a role in reflex withdrawal of a limb (unilateral flexion) from a painful stimulus. Task-dependency has also been found since, during locomotion, a long-lasting burst of activity is unraveled ipsilaterally during pharmacologically-induced fictive locomotion (long-lasting FRA response) probably via CPG elements (specifically the flexor portion of the CPG) since FRA stimulation under experimental conditions was shown to reset the step cycle to flexion (Jankowska et al., 1967a,b; Perreault et al., 1995; Schomburg et al., 1998; Ollivier-Lanvin et al., 2011). Clinically, it is associated with the Babinski sign (i.e., tongue depressor-induced plantar extension) as an indication of neurological problems in adults (e.g., spinal pyramidal tracts-induced injury caused by trauma or tumor). The long-lasting FRA response as well as myoclonus can also be uncovered following FRA stimulation in patients with SCI.

CENTRAL PATTERN GENERATORS—EARLY EVIDENCE AND UNDERLYING CONCEPTS

To date, the best-characterized spinal network is undoubtedly the CPG for locomotion that directly controls the basic motor commands underlying ambulation (Guertin, 2009a; Guertin and Steuer, 2009). Seminal work from Flourens, Phillipson, Sherrington, and Graham Brown supported by subsequent evidence generated largely from the 1960s onwards showed that, across species, rhythmic and stereotyped motor behaviors including walking, flying, and swimming are controlled largely by a neuronal network generally referred to as the CPG for locomotion (Flourens, 1824; Freusberg, 1874; Sherrington, 1910; Graham Brown, 1911, 1914; Lhermitte, 1919; Grillner, 2006; Clarac and Pearlstein, 2007). Early observations from paraplegic dogs revealed the existence of locomotor-like movements that can occur spontaneously after a complete transection (TX) of the spinal cord. That was elicited specifically when dropping one of the limbs from a flexed position. Comparable observations by Phillipson led him to conclude that the spinal cord could control locomotion using both central and reflex mechanisms. Sir Charles Sherrington's work in TX cats and dogs provided additional evidence that such spinal locomotor-like movements were the result of reflex actions from proprioceptors onto some spinal centers (Sherrington, 1910). However, it is Thomas Graham Brown, who described more

directly the existence of a spinal neuronal network as main command center for locomotion (see also Stuart and Hultborn, 2008 for a thorough description of Sherrington and Graham Brown's original contributions) in anesthetized animals lying on one side when stepping movements in the hindlimbs were spontaneously expressed ("narcosis progression") after TX thoracically (Graham Brown, 1911, 1914; Stuart and Hultborn, 2008). More evidence of its existence cellularly (half-center now generally referred to as the CPG) occurred when intracellular recordings became possible in the 1960s. A Swedish group led by Anders Lundberg recorded interneurons located in the lumbar segments of the spinal cord (lamina VII), active following FRA stimulation. Since then, several theories have been proposed to tentatively describe the functional organization of that CPG—the Miller and Scott (1977) hypothesis involving Renshaw cells, the "ring" model to explain more complex locomotor patterns (e.g., backward vs. forward walking), the flexor burst generator with an asymmetrical excitatory drive from a flexor burst generator as well as the unit burst generator model with symmetrically-organized burst generators (for each articulation or sets of muscles) even in absence of peripheral input. More recently, the "synergy model" from Bizzi's group and the bipartite model (or two-level CPGs) from McCrea's group were also proposed (Tresch et al., 1999; Bizzi et al., 2008; McCrea and Rybak, 2008).

CONCEPTUAL ORGANIZATION OF CPG FOR LOCOMOTION

HALF-CENTER MODEL

Sherrington used the term "half-center" to explain the spinal pathway for reciprocal inhibition. That expression was used again by Graham Brown in his model of spinal locomotor control based on his experiments showing that the basic pattern for stepping was generated entirely in the spinal cord even in absence of peripheral afferent contribution (due to narcosis) in spinal cord-transected cats, rabbits and guinea-pigs (Graham Brown, 1911, 1914). The animals, under general anesthesia, were lying on one side when stepping movements in the hindlimbs were spontaneously evoked ("narcosis progression") after a transection of the cord at the lower thoracic level. Since the level of anesthetic used was shown to abolish proprio- and exteroceptive reflexes but not locomotor activity, Graham Brown proposed a "half-center" model made of two groups of spinal neurons reciprocally organized and mutually inhibiting each others that were capable of producing the basic rhythm and pattern for stepping. Activity in the first group of neurons (e.g., extensor half-center) would send motor commands to motoneurons (exciting extensors), and would inhibit simultaneously the reciprocal group of neurons (flexor half-center) preventing the excitation of antagonists (silencing flexors). After a period of "depression" (e.g., fatigue, adaptation, post-inhibitory rebound) of the extensor half-center, the flexor half-drive would predominate for a new phase of activity. Despite these findings, the general opinion of scientists between 1920 and 1960 remained that basic locomotor activity largely depends upon sensory input from the peripheral nervous system. However, it is Elzbieta Jankowska and Anders Lundberg who have provided in the 1960s using intracellular recording techniques the first direct evidence supporting the existence of Graham Brown's model

(Jankowska, 2008). They identified intracellularly interneurons located in the lumbar segments of the cord (specifically in the lamina VII) that are active following FRA stimulation. Specifically, one group of neurons was found to be activated by ipsilateral FRA, a second group by contralateral FRA (coFRA), and a third group by both ipsi- and contralateral stimulation. After injection of L-DOPA and nialamide in spinal cord-transected cats, FRA stimulation evoked a high frequency burst followed by a long-lasting self-sustained series of discharges. Some neurons did not even show any short latency effects during the stimulus train. These interneurons were found to be monosynaptically excited by ventro-lateral funicular stimulation which contains descending fibers from the reticular formation. One of the most important features was the reciprocal organization between these groups of interneurons since coFRA stimulation abolished the long-latency discharges evoked by ipsilateral FRA and vice versa. Finally, it was proposed that Ia interneurons could participate in the production of the locomotor pattern by receiving strong excitatory input from FRA interneurons given their corresponding rhythmic activity in L-DOPA-treated cats (Jankowska et al., 1967a,b). However, in the 1970s, several neuroscientists began to provide evidence suggesting that, as it is, this half-center organization can not fully explain the complex patterns of muscle activation found during terrestrial locomotion (e.g., in quadrupeds) (Grillner, 1981).

MILLER AND SCOTT MODEL

Sharing similarities with the half-center model, the Miller and Scott hypothesis proposed that Renshaw cells rather than fatigue are responsible for the alternation between flexion and extension (Miller and Scott, 1977). Increasing activity in one pool of motoneurons (e.g., extensors) would be gradually inhibited by a corresponding increase of recurrent inhibition. Architecturally, this model takes into account known neuronal connections—it is constituted of a closed chain of neurons to which flexor and extensor motoneurons are connected in different parts. Renshaw cells and Ia inhibitory interneurons which are part of this chain of neurons are mainly responsible for reciprocal activation of the flexor and extensor motoneurons (Bergmans et al., 1969). Simultaneously, Renshaw cells would remove reciprocal inhibition of antagonists (recurrent facilitation) via their spindle Ia monosynaptic inhibitory input onto Ia interneurons allowing the flexor excitatory drive to take over for a new phase of activity (i.e., flexion). Interestingly, by varying the tonic input to the alpha motoneurons and Ia inhibitory interneurons, coactivation of the flexors and extensors may be achieved. Although, the reciprocity between flexors and extensors is nicely explained by this model, the origin of the rhythmicity itself is rather unclear (Kriellaars, 1992; Kriellaars et al., 1994). Other criticisms came from results showing that Renshaw cell activity may be inhibited during locomotor activity although discrepancies were reported during fictive locomotion whereas both classes of neurons were reported not to be essential for the production of a basic locomotor pattern in motoneurons. Other evidence against the Miller and Scott model was provided by Jordan's group who showed that the basic locomotor pattern and Ia interneuron activity remain after i.v. injection of the nicotinic antagonist mecamylamine (MEC),

which greatly reduces Renshaw cell activation (McCrea et al., 1980; reviewed in Guertin, 2009a).

RING MODEL

The highly conceptual “ring” model was one of the models subsequently proposed to explain the existence of complex locomotor patterns (e.g., taking into account differences between backward or forward walking, synchronization and phase coupling between activity of the ring and the cyclic afferent input, etc.) (reviewed in Guertin, 2009a). It is made of a closed chain of at least five groups of neurons (e.g., 2 pure extensors, 2 pure flexors and 1 bifunctional) that project to the motoneurons either directly or through specific interneurons. The sequence of these projections determines the order of activation of various muscles during the step cycle. At rest, a number of ring neurons are tonically inhibited by a certain group of spinal neurons. Activation of the monoaminergic descending system would result in the inhibition of the inhibitory neurons with consequent disinhibition of the ring neurons. Activity within the ring is based on a cyclically propagated inhibitory drive that would travel at different speeds from one group to the other depending on the excitability (e.g., modulated by afferent input) of the path (“ring”) interconnecting them. The activity within the ring starts when the excitability level is raised (through disinhibition), so the neurons would discharge when not inhibited. A slow-propagated drive would activate neurons of a group for a longer period of time (e.g., pure extensor during the stance phase) whereas a fast-propagated one would activate a group of neurons for a brief moment in motoneurons to bifunctional muscles. However, this highly conceptual model has generally failed to convince most scientists in this field.

FLEXOR BURST GENERATOR MODEL

During those same years, other models such as the flexor burst generator were also proposed. Pearson and Duysens introduced this model for insects and cats (Pearson and Duysens, 1976; Duysens, 1977; Duysens et al., 2013). It consists of a rhythmic excitatory drive from the flexor burst generator to populations of flexor motoneurons. The burst generator would inhibit, via an inhibitory interneuron, the activity of extensors otherwise activated during the flexor silence by a tonic excitatory input. This asymmetrical model was abandoned later on in favor of a more symmetrical bipartite organization (i.e., in which both extensor and flexor portions are equally driven). This change in views is likely related to the subsequent description of a powerful feedback system associated with ankle extensor group I afferents that can reset rhythms and strongly excite most pools of hindlimb extensor motoneurons during locomotion. This said, recent data mainly from Brownstone's group provided evidence suggesting that an asymmetrical CPG organization should perhaps be re-considered. They proposed that a rhythm-generating layer, composed of a kernel of heterogeneous and electrotonically-coupled neurons, would project directly to the flexor half-center of the pattern formation layer (Brownstone and Wilson, 2008).

UNIT BURST GENERATOR MODEL

In the 70s and early 80s, the unit burst generator model contributed to the demonstration of a symmetrically-organized

generator in the spinal cord that can produce the basic pattern of motor commands for walking even in absence of peripheral input (Edgerton et al., 1976; Grillner, 1981). It was proposed essentially to explain that locomotion is not only a strictly alternating pattern of flexor and extensor activity (requiring all motoneurons to belong to one of these two groups) as proposed by the half-center model (initially by Graham Brown and subsequently by Lundberg et al.). Patterns of locomotor activity are often complex and may include some motoneuron pools that display activity during both the flexion and extension phases of the step cycle or that display differences in the onset and offset of activity in individual flexor and extensor pools. The persistence of such complex activity patterns following bilateral deafferentation of the hindlimbs in decerebrate cats, led Grillner and Zangger (1975) to conclude that the locomotor CPG does not simply generate an alternating activation of flexors and extensors but a more complex pattern that will sequentially start and terminate distinctively, the activity in the appropriate muscles (Grillner and Zangger, 1974). Their idea was further developed in a proposal for a CPG architecture in which separate “modules” or unit burst generators would control subsets of motoneurons. First, Székely et al. showed that the locomotor pattern (in the forelimbs) in freely moving newts was similar before and after a bilateral section of the dorsal roots (Székely et al., 1969). This was also shown in decerebrate cats leading then to the suggestion that a CPG could exist for each joint of each limb. Activity from these “units” would be tightly coupled during “normal” walking but individually controlled by supraspinal input to produce different types of motor patterns. This model emerged after analyzing more complex patterns of locomotor activity such as backward walking, climbing, etc. For instance, it was occasionally observed during fictive locomotion that one hindlimb motor nerve can display tonic activity while the others display a normal rhythmic pattern. Also, the activity of pluriarticular muscle nerves such as *semitendinosus* is sometimes in phase with extensors, or flexors, or both, which some authors found difficult to explain with a half-center type of model. Along this idea, Bizzi’s group provided experimental and analytical results suggesting instead that sensory-dependent linear combinations of a small number of muscle synergies may generate diverse motor and locomotor patterns. Some of the cellular components of Grillner’s CPG model were identified in the 80s using a simpler non-mammalian vertebrate nervous system preparation—the *in vitro* isolated lamprey preparation (Grillner and Wallén, 1985; Grillner, 2006). However, despite the attractiveness of this proposal, the unit burst generator model has not generally explained the existence of other complex patterns of motoneuronal activity such as those found during spontaneous deletions (see section below).

TWO-LEVEL-HALF-CENTER MODELS

Thus far, most models had failed to entirely explain the many patterns that can occur in the generally alternating activity of flexors and extensors during locomotion. In particular, unpredictable changes called “deletions” which refer to periods of silenced activity in some populations of motoneurons (e.g., extensors such as the *soleus*) accompanied of sustained or rhythmic activity in antagonist motoneurons (e.g., flexors such as the

tibialis anterior) while post-deletion rhythm is generally maintained. This is essentially why attempts to explain these other types of changes have first been proposed based on bipartite CPG levels. Indeed, a detailed analysis of nerve and muscle activity during spontaneous walking in non-paralyzed cats or during fictive locomotor activity in paralyzed decorticated cats let Perret and Cabelguen to initially propose a bipartite or two-level (half-center-like) model that explains the complex biphasic activity in so-called bifunctional motoneurons (e.g., *semitendinosus*) (Perret and Cabelguen, 1980; McCrea and Rybak, 2008; reviewed in Guertin, 2009a). They proposed that not only one half-center but both half-centers (extensor and flexor ones) would send motor commands to bifunctional motoneurons. A variety of motoneuron patterns could be produced by modulating the half-centers output “en route” to these motoneurons. They also suggested that a rhythm generator would be functionally separated from a pattern generator since the rhythm and the amplitude of the locomotor drive potentials appear to be two distinct characteristics that can be independently and spontaneously changing. This paved the way to other studies from Kriellaars and Jordan who proposed a functional separation of pattern and amplitude (Kriellaars, 1992; Kriellaars et al., 1994). They showed that locomotor drive potentials monitored simultaneously (by dual intracellular recordings *in vivo!*) in pairs of motoneurons generally covary in amplitude in homonymous motoneurons while antagonist motoneurons inversely covary. The complex locomotor pattern in bifunctional motoneurons receiving input from both half-centers would be sculpted by controlling the amplitude of the flexor and extensor locomotor drive “en route” to these motoneurons. A similar separation of CPG function (rhythm vs. pattern and amplitude) was suggested in other studies to explain how sensory stimulation can also alter locomotor cycle timing without altering the level of motoneuron activity which has served in the 90s as basis to the elaboration of the most recently proposed multi-level models (2+ and 3 levels, see Rybak’s work). Additional evidence from non-resetting deletions reported by McCrea et al. during fictive locomotion and scratch in the decerebrate cat strongly supported also this two-level CPG organization (Lafreniere-Roula and McCrea, 2005). Finally, mathematical models recently developed by McCrea and Rybak have constituted additional supports for the existence of multi-level CPG (half-center-like) organizations that can explain spontaneous deletions and other complex patterns of activity during locomotion (McCrea and Rybak, 2008; Knusel et al., 2013).

All and all, none of the above models have been refuted and yet, according to some, none of the conceptual models is capable to explain the wide variety and diversity of locomotor patterns that exist in real life.

CELLULAR CONSTITUENTS OF THE CPG FOR LOCOMOTION

Beyond these conceptual considerations about its organization, the CPG for locomotion has been characterized in part as a group of interneurons localized mainly in the lumbar area of the spinal cord (Giszter et al., 2007; Guertin, 2009a; Guertin and Steuer, 2009). With an *in vitro* isolated spinal cord preparation from neonatal rats, Kjaerulff et al. used sulforhodamine-101, an

activity-dependent marker/dye, to identify CPG neuron candidates in L1–L6 near the central canal as well as near the medial intermediate zone (Barajon et al., 1992; Kjaerulff et al., 1994). A comparable approach used by Cina and Hochman showed the existence of a restricted number of labeled cells (presumably CPG neuron candidates) more specifically in L1–L5 segmental areas (Hochman et al., 1994; Cina and Hochman, 2000). These findings are supported also by other studies that showed, using electrical stimulation or selective lesions, key rhythmogenic CPG elements specifically in L1 and L2 in mice (Nishimaru et al., 2000). This is also supported by findings from Dimitrijevic et al. (1998) in SCI patients following epidural stimulation near L1–L2 that triggered locomotor-like movements in the lower extremities (Dimitrijevic et al., 1998; Shapkova and Schomburg, 2001; Selionova et al., 2009). Observations in one patient with a complete spinal cord transection mid-thoracically displaying spontaneous episodes of locomotor-like movements when lying in a bed could be interpreted as valuable evidence of a CPG in the lumbar cord of humans (Nadeau et al., 2010). Discrepancies may exist in other species regarding the exact localization of the CPG (e.g., in mid-lumbar segments in cats) (Langlet et al., 2005). In primitive vertebrate species such as lampreys, cellular components have been identified and even recorded from electrophysiologically nearly 30 years ago (LC cells, CC interneurons, etc.). Indeed, with its simpler neural system, it has been easier to investigate extensively neuronal activity from single spinal cells using the *in vitro* isolated spinal cord preparation from lampreys. The quest for dissecting further the CPGs has been supported also by findings obtained in parallel (or before) from non-mammalian and invertebrate species. In fact, studies on sea slugs, leeches, cockroaches, stick insects and crustacean locomotor (swimmeret) and motor (e.g., somatogastric system) pattern-generating networks have played a pivotal role in understanding further the cellular and network bases of rhythmic motor and locomotor patterns in both invertebrate and vertebrate species (Hugues and Wiersma, 1960; Pearson, 1993; Hopper and DiCaprio, 2004; Clarac and Pearlstein, 2007; Buschges et al., 2008; Harris-Warrick, 2011).

PHARMACOLOGICAL AND ELECTROPHYSIOLOGICAL PROPERTIES OF THE LOCOMOTOR CPG

Since the 1980s, pharmacological manipulations using a plethora of newly available receptor ligands (agonists and antagonists) have largely contributed to understand and further describe the detailed organization of the locomotor CPG (Rossignol et al., 2001; Harris-Warrick, 2011). Experiments in *in vitro* isolated rodents (rat, mouse and turtle preparations) as well as, subsequently, in *in vivo* TX rodents (adult rat and mouse models) led to substantial advances in cellular target identification (receptors and channels) associated with locomotor rhythm generation and modulation (Guertin and Hounsgaard, 1998a,b; Guertin, 2009b). Blood brain barrier (BBB) permeable ligands have served to pharmacologically “dissect” *in vivo* the contribution of specific receptors and channels to locomotor rhythm-generation (Guertin, 2008). Clear CPG-activating effects induced by specific drugs have also been found in various *in vitro* preparations from invertebrate and vertebrate species, although it is beyond

the scope of this review to report on all of them (Grillner and Wallén, 1985; Sigvardt et al., 1985; Cazalets et al., 1990; Cowley and Schmidt, 1994; Kiehn and Kjaerulff, 1996; Schmidt et al., 1998; Schmidt and Jordan, 2000; Bonnot et al., 2002; Kiehn et al., 2008). As mentioned earlier, the noradrenergic system and specifically L-DOPA has been among the first systems to be associated with CPG activation in acutely TX cats and rabbits (Jankowska et al., 1967a,b; Viala and Buser, 1969; Grillner and Zangger, 1974). Clonidine, an alpha-2 adrenergic receptor agonist, administered during sensory stimulation (e.g., tail or sexual organ pinching) was also reported to enhance the effects of training and/or sensory stimulation (remains unclear) on locomotor rhythmogenesis in TX cats (Forssberg and Grillner, 1973; Barbeau and Rossignol, 1990, 1991; Pearson and Rossignol, 1991; Chau et al., 1998a,b; Rossignol et al., 2001). However, it has been difficult to determine site-specific actions (e.g., on motoneurons, CPG neurons or primary afferents) from results in many of these earlier studies that were not designed to specifically assess drug-induced CPG activation *per se* (i.e., given the use of additional stimuli including tail stimulation, sexual organ pinching, regular training or weight support assistance, see Lovely et al., 1986; Bélanger et al., 1996; Zhang et al., 2010). More recently, experiments conducted in our laboratory in a mouse model of paraplegia (a complete low-thoracic TX) with no assistance or additional stimuli (e.g., no training, no tail stimulation, no sexual organ pinching, and no weight-support assistance to avoid unspecific non-drug induced effects) have contributed to identify clearly a subset of transmembrane receptors involved in pharmacologically-elicited, CPG-mediated locomotor-like movements in the lower extremities (Guertin, 2008, 2009a,b). For instance, L-DOPA, serotonin (5-HT) or dopamine (DA) receptor ligands such as 8-OH-DPAT, buspirone (5-HT_{1A/7} agonists), quipazine (5-HT_{2A/2C} agonist) or SKF-81297 (D₁-like agonist) were found to trigger significant locomotor-like movements (i.e., rhythmic bilaterally alternating flexions and extensions involving one or several hindlimb joints) whereas others ligands such as 3-Trifluoromethylphenylpiperazine (TFMPP) (5-HT_{1B}), m-CPP (5-HT_{2B/2C}), SR57227A (5-HT₃) or clonidine (adrenergic alpha-2 agonist) were shown to elicit mainly non-locomotor movements (i.e., non-bilaterally alternating movements, twitches, cramps, etc.) in TX mice (Guertin, 2004, 2005; Landry and Guertin, 2004; Guertin and Steuer, 2005; in rats, see Antri et al., 2005; Lapointe et al., 2008; Ung et al., 2008). Using selective antagonists and genetically-manipulated animals (e.g., 5-HT₇KO mice), it has been clearly established that N-Methyl-D-aspartate (NMDA), 5-HT₁, 5-HT₇, 5-HT_{2A} and D₁ receptors were specifically involved in mediating such CPG-activating locomotor-like effects (Landry et al., 2006a; Lapointe et al., 2009). For instance, endogenous glutamate release and NMDA receptor activation were reported as critically important for quipazine-induced effects since a complete loss of induced movement was found in NMDA antagonist (MK-801)-treated animals previously pretreated with NMDA (Guertin, 2004). Regarding DA receptors, administration of D₂, D₃ or D₄ agonists was found not to generate significant hindlimb locomotor-like movements whereas D₁/D₅ agonists such as SKF-81297 can potentially elicit locomotor-like movements that

are lost in selective D1-like (D1/D5) antagonist-pretreated TX mice but not in D5 $-/-$ paraplegic TX mice suggesting a specific contribution of the D1 subtype to CPG activation (Lapointe et al., 2009). All and all, pharmacological approaches in *in vivo*, untrained and non-assisted TX animals have contributed to identify a subset of CPG-activating compounds (and corresponding receptors confirmed with selective antagonists and knockout animal models) (Guertin, 2009a, 2012). However, none of these molecules were found to generate large amplitude weight bearing stepping movements *per se* in untrained, non-assisted and non-sensory stimulated TX animals suggesting that only partial CPG activating effects can be achieved using these ligands administered separately. Subsequent studies conducted in our laboratory have shown that only simultaneous activation of some of these candidate receptors can, using similar pharmacological approaches and animal models, induce full locomotor-inducing effects. Partial CPG-activating effects (i.e., associated with crawling rather than full weight bearing stepping) induced by some ligands, as mentioned above, were found indeed to turn into full CPG-activating effects (i.e., weight bearing stepping in non-assisted, untrained and non-stimulated paraplegic animals) by simultaneously administering 8-OH-DPAT, quipazine, L-DOPA, or SKF-81297 (Guertin, 2009b; Guertin et al., 2010, 2011). Prior to these findings, numerous pharmacological studies aimed at improving locomotor function recovery after SCI have been also conducted in other *in vivo* models of SCI (e.g., in TX cats). However, it has remained difficult to consider those results as evidence of cellular targets associated with CPG activation or modulation given the used paradigm (experiments conducted during tail stimulation, sex organ pinching, body-weight supported, movement-assisted manually, etc.) (Forssberg and Grillner, 1973; Lovely et al., 1986; Barbeau and Rossignol, 1991; Bélanger et al., 1996). Some of our findings, reproduced subsequently in other laboratories, have thus been independently validated (Courtine et al., 2011; van den Brand et al., 2012).

This said, the extent to which the cellular network activated pharmacologically *in vivo* corresponds to already identified CPG neuron candidates (electrophysiologically or genetically) remains unclear (see section below). However, dual immunohistochemical experiments recently showed locomotor activity-labeled (*c-fos*) 5-HT1A-, 5-HT2A- or 5-HT7- positive neurons in the cat lumbar cord (specifically in laminae VII-VIII, Noga et al., 2009) suggesting that some of the locomotor activity-related receptors identified recently in *in vivo* models may indeed be located on CPG neurons (Hochman et al., 2012).

SPECIFIC IONIC CONDUCTANCES AND CHANNELS

This is also an aspect of the CPG that remains incompletely understood. It is generally accepted that one of the main cellular features expected to be found in CPG neuron candidates is its capacity to express endogenously rhythmic activity during fictive or real locomotion (Eken et al., 1989; MacLean et al., 1997; Brownstone and Wilson, 2008; Brocard et al., 2010). For several decades, it was also generally accepted that some neurons of the CPG were expected to express specifically autorhythmic or pacemaker-like properties given that locomotion is, by definition,

a rhythmic motor behavior. There have been indeed several cell populations in the corresponding area of the CPG (i.e., lumbar segments in mammalian species) shown in the last few decades to be capable of expressing pacemaker-like properties in specific conditions such as in the presence of NMDA alone or combined, at lower dose, with serotonin and DA (e.g., bath-applied in the case of *in vitro* isolated spinal cord preparations) (Wang et al., 2006; Grob and Guertin, 2007). Plateau potential is another intrinsic property believed by some researchers to be associated with endogenous (tetrodotoxin-resistant) pacemaker-like activity (Eken et al., 1989). Transmembrane currents and channels underlying both of these properties (voltage oscillations, NMDA ionophore, L-type (CaV1.3) Ca²⁺ channel, persistent Na²⁺ current, I_h current, I_A current, I_{CAN}, I_{NaP} current, I_{K(Ca)} current, post-inhibitory rebound have been relatively well-characterized in lamprey, turtle, tadpole, rodent and cat preparations with some species-dependent specificities (Grillner and Wallén, 1985; Reith and Sillar, 1998; Harris-Warrick, 2002, 2011; Grillner, 2006; Wang et al., 2006; Grob and Guertin, 2007). Nonetheless, the specific contribution of these properties (in motoneurons and some interneurons) to real life locomotion remains speculative and a source of debate.

GENETICALLY-IDENTIFIED NEURONS OF THE CPG FOR LOCOMOTION

In recent years, advances in genetics and transgenic murine models have largely contributed to the identification of specific CPG neuron candidates *per se* (Ginty et al., 1992; Lanuza et al., 2004; Goulding, 2009). Targets initially identified in the developing neural tube (i.e., 11 distinct populations of spinal neurons (dI1–dI6, V0–V3, VMN) based on the expression of transcription factors (e.g., Jessell's work) have largely contributed to these findings. Several populations of interneurons involved in locomotor activity were indeed characterized genetically (e.g., V0–V3). One of them is the population of V0 interneurons that was associated with left-right alternation since mice without V0 interneurons (lacking the transcriptional factor *Dbx1*) displayed bilateral synchrony rather than bilateral alternation during locomotion. Another population referred to as the V1 inhibitory interneurons (expressing the transcription factor *Engrailed 1*) was associated with high locomotor frequencies since slow rhythms were found in *En1-DTA* mice (Gosgnach et al., 2006). Other genetically-identified populations include the *Chx10*-expressing cells (V2a glutamatergic and V2b gabaergic interneurons) located in the intermediate zone of the gray matter in the lumbar spinal cord (Lundfald et al., 2007; Crone et al., 2008). These neurons were associated with frequency, amplitude and bilateral coordination since all of these parameters were affected in *Chx10-DTA* mice (lacking V2a interneurons). V3 interneurons (*Sim-1* expressing cells) constitute another recently identified population shown to participate in the production of a robust and balanced rhythm during locomotion. Indeed, rhythmic activity was found to be partially disrupted in mice lacking V3 interneurons (Zhang et al., 2008). Genetically-engineered animals were also utilized to show that hopping instead of normal walking is displayed in mice lacking the *ephA4* receptor-expressing lumbar interneurons. Finally, another population of CPG neuron candidate

called HB9 neurons was reported to provide neuronal excitation during locomotion. Although, no corresponding knockout model has been tested, compelling evidence suggests that the HB9 excitatory interneuron belongs to an asymmetrically-organized rhythm-generating network (Brownstone's and Jessell's work). Taken altogether, results from these exciting new studies in mice suggest that these genetically-characterized interneurons (V0–V3, EphA4, HB9) may constitute different cellular components of the CPG (Kullander et al., 2003; Wilson et al., 2005; Brownstone and Wilson, 2008). For instance, V0 interneurons form many cell types including commissural neurons which could establish inhibitory reciprocal connections between the two sides of the spinal cord. In turn, V1 interneurons may be associated with inhibitory interneurons such as the Ia inhibitory and Renshaw cells whereas HB9s may be excitatory interneurons constituting at least part of the rhythm generating layer. However, additional studies need to be conducted in order to fully characterize these promising new CPG neuron candidates (Butt et al., 2002; Wilson et al., 2007). It is important to mention also in most cases of gene deletions using these KO models, elimination of rhythmic motoneuron excitation or rhythmic inhibition is rarely found suggesting that several CPG secrets remain to be described.

CHARACTERISTICS OF OTHER CPGs

SPINAL GENERATOR FOR EJACULATION

A breakthrough finding in 2002 has drastically changed our view of the neurobiology of sexual function (Truitt and Coolen, 2002; Coolen et al., 2003). Indeed, Truitt and Coolen have shown that ejaculation critically depends upon a spinal generator or network of neurons referred to as the Spinal Generator for Ejaculation (SGE; Truitt and Coolen, 2002). Located in the lumbar segments L3 and L4, the SGE is defined as a circuit capable of producing self-sustained rhythmic output to pudendal motoneurons. The SGE was found to contain a key population of neurons (lumbar spino-thalamic neurons also called LSt cells) that (1) project to forebrain; (2) project to pudendal motoneurons (correspond to those located in the Onuf's nucleus in men); and (3) receive input from sexual organs via the pudendal and dorsal nerve of the penis. In fact, ejaculation is completely lost in animals undergoing LSt cell-lesioned procedures (using SSP-saporin). LSt cells are found in the vicinity of the central canal (lamina X and medial portion of lamina VII, and most of them specifically contain galanin, CCK, enkephalin and NK-1 receptors. Electrical stimulation of the pudendal or dorsal nerve of the penis nerves was shown to elicit ejaculatory responses in low-thoracic Tx rats. Comparable data were found in SCI men supporting the existence of LSt cells (or at least of a SGE) in humans as in other species. However, the type(s) of neurotransmitters involved and the subset of post-synaptic receptors associated with SGE activation and ejaculation remain largely unknown (Courtois et al., 2008, 2013).

This said, only a few families of compounds are known to modulate sexual function (either erection, ejaculation or both) (McKenna et al., 1991; Pomerantz et al., 1993; Vargas et al., 2004; Moreland and Makela, 2005; García-Bravo et al., 2006). For instance, Guttman and Walsh showed (earlier case reports also

exist) that the cholinesterase inhibitor prostigmine administered intrathecally (i.t.) could elicit spontaneous erection accompanied sometimes with ejaculation in SCI men (Guttman and Walsh, 1971). Midodrine (alpha-1 agonist) was also shown to induce either normal (anterograde—i.e., outside the urethral meatus) or abnormal (retrograde—i.e., back inside into the bladder) ejaculation in patients with SCI but, as with cholinesterase inhibitors, it was also found to increase blood pressure and autonomic dysreflexia (Jonas et al., 1979; Riley and Riley, 1982; Staerman et al., 2001; Blanchard-Dauphin et al., 2005; Courtois et al., 2008). In animal models, muscarine (i.p. or i.t.) was found to promote erection and sometimes ejaculation in sensory-stimulated Tx rats (i.e., penile sheath retraction known as the “penile reflex” model) (Durán et al., 2000). Alpha-1 agonists such as methoxamine (which, unlike midodrine, crosses the BBB) were reported to promote reflexively-induced ejaculation (either immediately after Tx or by urethral stimulation with fluid injection) in freely moving Tx rats. P-chloroamphetamine (amphetamine derivative that increases 5-HT release) was shown to elicit fictive ejaculation (Electromyogram (EMG) correlates) in anesthetized Tx rats (Stafford et al., 2006). Supporting a role for subclasses of spinal 5-HT receptors, 5-HT receptor agonists such as m-CPP (5-HT₂) or 8-OH-DPAT (5-HT_{1A}) were also found to promote sometimes ejaculation in sensory-stimulated Tx (self-grooming) or non-Tx (copulating) rats (Camacho et al., 2007). However, mixed 5-HT-induced effects have also been reported suggesting that serotonergic modulation may depend upon the site of action and route of delivery (inhibitory effects in brain vs. excitatory effects in spinal cord) (Yonezawa et al., 2008). Regarding DA agonists, none have been tested in Tx animals although D2-like agonists i.c.v. injected was reported to promote ejaculation in intact rats.

SPINAL MICTURITION CENTER

Micturition essentially depends, for the storage and periodic elimination of urine, on the coordinated activity of smooth and striated muscles in the several functional units of the lower urinary tract, namely the urinary bladder, the bladder neck, the urethra and the urethral sphincter (Mallory et al., 1991; de Groat et al., 1993; Andersson and Pehrson, 2003; Sugaya et al., 2005). The coordination between these organs is mediated by a complex neural control system that is located partly in the spinal cord (Schröder, 1985; Birder and de Groat, 1993; Birder et al., 1999; Dolber et al., 2007). Central interneurons retrogradely labeled by injection of pseudorabies virus into the urinary bladder of the rat were found in regions receiving afferent input from the bladder (Nadelhaft and Vera, 1995; Sugaya et al., 1997). A comparable distribution was shown following injections of virus into the urethra (Vizzard's work) or the external urethral sphincter (EUS), indicating a prominent overlap of the interneuronal pathways controlling the various target organs of the lower urinary tract (Vizzard et al., 1995). In addition, spinal interneurons (located near dorsal commissure, superficial dorsal horn and sacral parasympathic nucleus) involved in processing afferent input from the lower urinary tract have been identified by c-fos expression following noxious or non-noxious stimulation of the bladder and urethra in rats (de Groat's work). Some of these interneurons send long

projections to the brain, whereas others make local connections in the spinal cord and participate in segmental spinal reflexes. These results provide strong evidence of a CPG for EUS control both in the thoracolumbar (T8-9 for storage of urine) and lumbosacral (L3-L4, L6-S1 for expulsion of urine) spinal cord. Various neurotransmitters have been associated with the control of the lower urinary tract including glutamate, tachykinins, pituitary-adenylate-cyclase-activating polypeptide, NO and ATP. Glutamate, acting on NMDA and non-NMDA receptors, seems to be critically involved in spinal and supraspinal reflex pathways that control the bladder and the EUS (Yoshiyama and de Groat, 2005). In contrast, GABA (γ -aminobutyric acid), glycine and enkephalins were reported to exert a tonic inhibitory control in the pontine micturition center (PMC) and regulate bladder capacity. Other neurotransmitters including DA, 5-HT, NA, and acetylcholine have either inhibitory or excitatory effects, depending on the type and location of activated receptors (de Groat et al., 1993; de Groat and Yoshimura, 2001; Chang et al., 2006; Dolber et al., 2007). For example, DA elicits inhibitory effects on micturition through D₁-like receptors and facilitatory effects through D₂-like receptors. Also, much like what has been shown with 5-HT_{1A/7} receptor agonists on CPG activation (Landry et al., 2006a), 5-HT_{1A/7} receptor agonist 8-OH-DPAT was found to induce rhythmic EUS relaxation during voiding in urethane-anesthetized chronic Tx rats. Other potential pharmacological treatments to facilitate micturition for instance after SCI; are (1) intravesical administration of drugs such as vanilloids which is aimed at desensitizing bladder afferents (Fowler et al., 1994); (2) injection of BoNT/A into the detrusor to temporarily block the pre-synaptic release of ACh from the parasympathetic innervation and produce a paralysis of the detrusor smooth muscle (Schurch et al., 2000); and (3) administration of NO donors such as isosorbide dinitrate to produce a significant reduction in striated sphincter pressure at rest and during dyssynergic contraction (Reitz et al., 2004).

SPINAL DEFECATION CENTER

Bowel control involves a complex interactions between the autonomic (sympathetic and parasympathetic), central (brain and spinal cord) and muscular systems (sphincters and smooth muscles) (Rostad, 1973). Experiments in spinal cord-transected rats (cervical level) have clearly shown the existence of a reflex defecation center (also known as the lumbosacral defecation center) located by retrograde labeling at the low lumbar and upper sacral level (i.e., L6-S1 in rats) (Shimizu et al., 2006). Evidence of increased defecation in humans caused by Lumbosacral Defecation Center (LDC)-mediated effects, induced by ghrelin receptor agonists, has been shown whereas direct evidence using intrathecal injections have been reported in animal models (Shimizu et al., 2006). Indeed, defecation induced pharmacologically was prevented by cutting the nerve pathways that connect the lumbosacral spinal cord and the colorectum, but not by severing the thoracic spinal cord (Ferens et al., 2011; Pustovit et al., 2014). Unfortunately, Ghrelin or its known agonists (e.g., ulimorelin) do not elicit defecation when administered orally and may increase blood pressure when delivered i.t.

CPGS IN HUMANS: EVIDENCE FROM DIRECT STIMULATION AND SPONTANEOUS PLASTICITY AND ROLE OF FEEDBACK AND FEEDFORWARD

Some people are reluctant to believe that spinal pattern generators or CPGs exist in humans given the lack of direct evidence. It is true that most data have been obtained from animal models, as described in this review. Experiments and tools used to explore CPG properties in animals are simply too invasive for comparable exploration in humans. But again, given that the spinal cord, among all structures of the CNS, is particularly well-preserved phylogenetically, it may be more reasonable to argue the opposite—are there evidence that humans do not have spinal CPGs given that all vertebrate species examined thus far strongly suggest the existence of comparable neural control systems for locomotion and other stereotyped motor behaviors?

The answer is “no”, we do not have evidence against the existence of spinal CPGs in humans (Bussel et al., 1989, 1992). There is increasing evidence suggesting the existence of comparable spinal networks in primates including humans—at least for locomotion (Tan, 2006a,b, 2008). For instance, epidural or intraspinal stimulation near L1–L2 to successfully trigger locomotor-like movements in the lower extremities of completely SCI patients (Dimitrijevic et al., 1998; Holinski et al., 2011; Moshonkina et al., 2012). Given that electrical stimulation of CPG does not generally lead to full weight bearing stepping (e.g., often only to air-stepping or locomotor-like movements lying in bed, this suggests that only partial CPG activation may be achieved electrically. In turn, it has been possible to activate the locomotor CPG by administration i.t., of a variety of pharmacological agents to acutely spinalized marmoset monkeys (*Callithrix jacchus*) in the absence of phasic afferent input to the spinal cord (Fedirchuk et al., 1998). Spontaneous expression, within weeks or months post-SCI, of rhythmic movements (myoclonus or locomotor-like movements) in humans have also been reported (Pozos and Iuzzo, 1991; Bussel et al., 1992, 1988; Calancie et al., 1994; Chervin et al., 2003; Calancie, 2006; Consentino et al., 2006; Gerasimenko et al., 2008, 2010; Nadeau et al., 2010; Field-Fote et al., 2012; Gorodnichev et al., 2012; Rye and Trotti, 2012).

How could such spontaneous movements occur (Clemens et al., 2006; Harkema, 2008; Nadeau et al., 2010)? No clear answer exists in humans (Calancie et al., 1994; Byrnes et al., 2006). However, related spontaneous plasticity events have clearly been shown in animal models to be associated with some immediate early gene expression (IEG) in CPG-corresponding areas of the spinal cord. Changes in IEG expression levels post-SCI were associated also with a cascade of subsequent changes (TNF- α , NOS; C1qb, Galectin-3, and p22(phox), glycine receptors, GAD-67, 5-HT_{1A/7} receptors; NGF, BDNF, and NT-3) and spontaneous spinal locomotor-like activity (Chi et al., 1993; Yakovlev and Faden, 1994; Giroux et al., 1999; Tillakaratne et al., 2002; Gómez-Pinilla et al., 2004; Landry et al., 2006b; Lukacova et al., 2006; Peng et al., 2006; Schmitt et al., 2006; Giszter et al., 2007; Li et al., 2007; Ung et al., 2007).

It is clear though that despite direct stimulation electrically or pharmacologically, spinal-mediated stereotyped motor functions such as locomotion requires at least some feedforward and

feedback mechanisms for goal-oriented functional recovery. Interestingly, some feedback mechanisms may remain operational despite severe spinal cord lesions at the thoracic or cervical level. Indeed, it has clearly been shown in Tx cats, displaying some restored spinal-mediated locomotor activity on a motorized treadmill, that adaptation to different speeds remains possible (Edgerton's or Rossignol's work). However, to reach more complex levels of adaptation such as in climbing steps, feedforward mechanisms are probably required. Recent experiments in partially Tx rats showed that at least some descending fibers need to be intact or functional for animals, driven to walk by CPG-activating drug cocktails (such as those identified in Guertin's laboratory), for displaying complex patterns such as walking up stairs and changing directions during walking (SŁawinska et al., 2012; van den Brand et al., 2012). It is unclear which descending systems are most important for such key feedforward mechanisms, however, cells from the parietal cortex and cerebellar systems are key to visually-guided locomotion whereas brainstem circuits and corresponding descending tracts (e.g., vestibulo-spinal tracts) are known for significant roles in equilibrium and tonus (Gurfinkel and Shik, 1973; Aydogdu et al., 2011).

All in all, future drug treatments identified in animal models to modulate spinal network activity are thus likely to be associated with comparable efficacy data in humans given significant similarities in spinal cord structures and cellular properties from lampreys to primates. However, the role of feedback and feedforward mechanisms in patients with SCI that will be given CPG-modulating and reactivating-drug treatments remains to be explored.

FURTHER EVIDENCE FROM PHARMACOLOGICAL REACTIVATION OF THE CORRESPONDING SPINAL NETWORKS

As mentioned above and in different sections throughout this review, it is clear that different types of CPGs exist and a wide variety of responses may be triggered pharmacologically pending upon the classes and subclasses of transmembrane receptors activated or blocked by specific ligands or exogenous neurotransmitter administration (Johnston et al., 2003; Guertin and Steuer, 2009).

In my laboratory, an extended series of drug screening experiments has been conducted in recent years. Using an experimental model of complete SCI (a low-thoracic spinal cord transected mouse) that eases and accelerates the identification of CPG-activating compounds (i.e., given the completeness and high reproducibility of this exact type of injury), we identified families of ligands capable of inducing, within minutes post-administration (s.c., p.o. or i.p.), either episodes of locomotor movements in the hindlimbs, micturition, defecation or ejaculation. Given that spinal networks controlling these functions are located below injury level in our model and that brain-mediated effects through descending inputs are unlikely in Tx mice, then such well-coordinated and stereotyped motor behaviors must be induced, at least partly, by corresponding spinal network reactivation following administration of specific drug(s).

Reactivation of spinal catecholaminergic receptors using L-DOPA/carbidopa or DA receptor agonists and 5-HT_{1A} receptors using buspirone or 8-OH-DPAT was found to trigger 45-min episodes of basic straightforward walking in chronic Tx mice (see Guertin, 2004 work since 2004). This patent-protected technologically, now referred to as SpinalonTM, was shown to elicit comparable effects in other species (patent application WO 2006026850) whereas repeated administration was found to prevent muscular atrophy and anemia among other biological functions and systems. L-DOPA/carbidopa and buspirone were shown not to have drug-drug interactions *in vitro* and *in vivo*, providing evidence of safety in animals (Guertin, 2009b; Guertin et al., 2010, 2011). Clinical trials (Phase I/IIa) in paraplegic and tetraplegic patients that began recently (December 2013) may provide further evidence of safety in patients while possibly reporting the first evidence ever of efficacy in humans for a CPG-activating drug therapy.

Given that no sensory stimulation, no harness and no electrical stimulation are used for these tests in motor-complete SCI patients, if efficacy data in ASIA-As are to be found, this could be considered as clear evidence of a CPG in humans that remains functional and reactivable with drugs despite a SCI at higher (rostral) levels. Upon positive results, a Phase IIb and eventually a pivotal Phase III shall be undertaken to eventually meet criteria for new drug approval (NDA) in the U.S., Europe and Canada expected for the end of 2018.

Given the successes obtained with this approach, additional drug screening studies were conducted in our laboratory to identify also Sacral or Spinal Micturition Center (SMC), LDC and SGE activating compounds (Guertin, 2005, 2008, 2009b). We recently identified leads and provided proof-of-concept data *in vivo* clearly demonstrating that pharmacological reactivation of these other networks upon systemic drug delivery is possible at least in animal models of complete SCI.

A combination of transmembrane receptor agonists (that can not yet be revealed for intellectual property reasons) found to elicit superior effects was selected as our final candidate product suitable for full preclinical and clinical development. Now referred to as MicselonTM, this drug combination was shown to elicit, within 15 to 30 min, episodes of micturition in SCI mice suffering of chronic urinary retention (patent application US 61/946,097). Again, the well-coordinated motor responses triggered by this therapy in completely SCI animals that induce in absence of sensory stimulation, providing strong evidence of SMC-activating effects. Lack of drug-drug interactions and evidence of efficacy in SCI patients remain to be shown prior to IND and NDA approval of this new experimental therapy against urinary retention.

A comparable technology was also found to restore some defecatory activities in SCI animals. Leads were identified using our technological platform adapted to assess defecation in paraplegic mice. Proof-of-concept data *in vivo* were obtained recently and Optimization phase studies with drug combinations revealed superior defecation-eliciting effects with a bi-therapy referred to as TransilonTM. It was found to elicit within 30 min episodes of defecation in chronic SCI mice suffering of constipation (patent application US 61/946,113). Again, the well-coordinated motor

responses triggered by this therapy in animals in absence of sensory stimulation provide strong evidence of LDC-activating effects. Lack of drug-drug interactions and evidence of efficacy in SCI patients remain to be shown prior to regulatory approval of this new experimental therapy against chronic constipation (expected by 2020).

Finally, we found a 4th class of drug treatments capable of spinal network reactivation in completely SCI subjects. Lead compounds were identified although Optimization phase studies have not been completed yet. Nonetheless, we found compounds and specifically drug combinations (which specific identity can not be revealed yet for intellectual property reasons) that can elicit unconditionally (with or without erection) seminal emission alone or seminal emission accompanied of seminal expulsion (full ejaculatory motor pattern) in male Tx mice (patent application US 60/996,535). These results provided proof-of-concept evidence of efficacy (SGE activation) in *in vivo* animal models of SCI. Again, additional tests remain to be performed to show comparable efficacy in humans and thus to demonstrate the existence of SGE neurons remaining functional and reactivable pharmacologically in people with SCI suffering of anejaculation.

In conclusion, CPGs involved in the control of several stereotyped motor behaviors such as locomotion, micturition, defecation and ejaculation have clearly been shown in a wide variety of species (Vizzard et al., 1995; Dimitrijevic et al., 1998; Truitt and Coolen, 2002; Shimizu et al., 2006; Clarac and Pearlstein, 2007; Guertin, 2009a; Guertin and Steuer, 2009). The one underlying pattern and rhythm generation for locomotion is by the far the most extensively studied although cellular characterization remains incomplete. Other CPGs, although poorly characterized cellularly, have also been clearly shown in different species. Nonetheless, this has not prevented research from identifying drug treatments capable of reactivating these networks separately after SCI at higher (rostral) levels. In fact, several experimental drugs (drug combinations composed of known and rather safe molecules) are currently in development clinically or preclinically. These new pharmacological aids may provide both, further evidence of spinal CPGs in humans and, useful noninvasive approaches to temporarily restore some key biological functions in people with SCI or comparable neurological disorders (e.g., multiple sclerosis) (Holmes, 1915; Fung et al., 1990; Wainberg et al., 1990; Rémy-Néris et al., 1999; Edgerton et al., 2001; Guertin and Steuer, 2009; Thorpe et al., 2011; Dietz and Sinkjaer, 2012; Guertin, 2012; Harkema et al., 2012; Jones and Cavanna, 2012; Schürks and Bussfeld, 2013).

REFERENCES

- Andersson, K. E., and Pehrson, R. (2003). CNS involvement in overactive bladder: pathophysiology and opportunities for pharmacological intervention. *Drugs* 63, 2595–2611. doi: 10.2165/00003495-200363230-00003
- Angel, M. J., Guertin, P., Jiménez, I., and McCrea, D. A. (1996). Group I extensor afferents evoke disynaptic EPSPs in cat hindlimb extensor motoneurons during fictive locomotion. *J. Physiol.* 493, 851–861.
- Angel, M. J., Jankowska, E., and McCrea, D. A. (2005). Candidate interneurons mediating group I disynaptic EPSPs in extensor motoneurons during fictive locomotion in the cat. *J. Physiol.* 563(Pt. 2), 597–610. doi: 10.1113/jphysiol.2004.076034
- Antri, M., Barthe, J. Y., Mouffle, C., and Orsal, D. (2005). Long-lasting recovery of locomotor function in chronic spinal rat following chronic combined pharmacological stimulation of serotonergic receptors with 8-OHDPAT and quipazine. *Neurosci. Lett.* 384, 162–167. doi: 10.1016/j.neulet.2005.04.062
- Aydogdu, I., Tanriverdi, Z., and Ertekin, C. (2011). Dysfunction of bulbar central pattern generator in ALS patients with dysphagia during sequential deglutition. *Clin. Neurophysiol.* 122, 1219–1228. doi: 10.1016/j.clinph.2010.11.002
- Barajon, I., Gossard, J. P., and Hultborn, H. (1992). Induction of fos expression by activity in the spinal rhythm generator for scratching. *Brain Res.* 588, 168–172. doi: 10.1016/0006-8993(92)91359-m
- Barbeau, H., and Rossignol, S. (1990). The effects of serotonergic drugs on the locomotor pattern and on cutaneous reflexes of the adult chronic spinal cat. *Brain Res.* 514, 55–67. doi: 10.1016/0006-8993(90)90435-e
- Barbeau, H., and Rossignol, S. (1991). Initiation and modulation of the locomotor pattern in the adult chronic spinal cat by noradrenergic, serotonergic and dopaminergic drugs. *Brain Res.* 546, 250–260. doi: 10.1016/0006-8993(91)91489-n
- Barthélemy, D., Grey, M. J., Nielsen, J. B., and Bouyer, L. (2011). Involvement of the corticospinal tract in the control of human gait. *Prog. Brain Res.* 192, 181–197. doi: 10.1016/b978-0-444-53355-5.00012-9
- Bélanger, M., Drew, T., Provencher, J., and Rossignol, S. (1996). A comparison of treadmill locomotion in adult cats before and after spinal transection. *J. Neurophysiol.* 76, 471–491.
- Bergmans, J., Burke, R., and Lundberg, A. (1969). Inhibition of transmission in the recurrent inhibitory pathway to motoneurons. *Brain Res.* 13, 600–602. doi: 10.1016/0006-8993(69)90270-4
- Birder, L. A., and de Groat, W. C. (1993). Induction of c-fos expression in spinal neurons by nociceptive and nonnociceptive stimulation of LUT. *Am. J. Physiol.* 265, R326–R333.
- Birder, L. A., Roppolo, J. R., Erickson, V. L., and de Groat, W. C. (1999). Increased c-fos expression in spinal lumbosacral projection neurons and preganglionic neurons after irritation of the lower urinary tract in the rat. *Brain Res.* 834, 55–65. doi: 10.1016/s0006-8993(99)01546-2
- Bizzi, E., Cheung, V. C., d'Avella, A., Saltiel, P., and Tresch, M. (2008). Combining modules for movement. *Brain Res. Rev.* 57, 125–133. doi: 10.1016/j.brainresrev.2007.08.004
- Blanchard-Dauphin, A., Rigot, J. M., and Thevenon, A. (2005). Treatment of ejaculation disorders by midodrine (Gutron) per os retrospective study of about 16 subjects. *Ann. Readapt. Med. Phys.* 48, 34–40. doi: 10.1016/j.anrmp.2004.09.004
- Bonnot, A., Whelan, P. J., Mentis, G. Z., and O'Donovan, M. J. (2002). Locomotor-like activity generated by the neonatal mouse spinal cord. *Brain Res. Brain Res. Rev.* 40, 141–151. doi: 10.1016/s0165-0173(02)00197-2
- Bretzner, F., and Drew, T. (2005). Contribution of the motor cortex to the structure and the timing of hindlimb locomotion in the cat: a microstimulation study. *J. Neurophysiol.* 94, 657–672. doi: 10.1152/jn.01245.2004
- Brocard, F., Tazerart, S., and Vinay, L. (2010). Do pacemakers drive the central pattern generator for locomotion in mammals? *Neuroscientist* 16, 139–155. doi: 10.1177/1073858409346339
- Brownstone, R. M., and Wilson, J. M. (2008). Strategies for delineating spinal locomotor-rhythm generating networks and the possible role of Hb9 interneurons in rhythmogenesis. *Brain Res. Rev.* 57, 64–76. doi: 10.1016/j.brainresrev.2007.06.025
- Buschges, A., Akay, T., Gabriel, J. P., and Schmidt, J. (2008). Organizing network action for locomotion: insights from studying insect walking. *Brain Res. Rev.* 57, 162–171. doi: 10.1016/j.brainresrev.2007.06.028
- Bussel, B., Roby-Brami, A., and Azouvi, P. (1992). "Organization of reflexes elicited by flexor reflex afferents in paraplegic man: evidence for a stepping generator," in *Muscle Afferents and Spinal Control of Movement*, eds L. Jami, E. Pierrot-Deseilligny and D. Zytnicki, 1st Edn. (Oxford, UK: Pergamon), 427–432.
- Bussel, B., Roby-Brami, A., Yakovlev, A., and Bennis, N. (1989). Late flexion reflex in paraplegic patients. Evidence for a spinal stepping generator. *Brain Res. Bull.* 22, 53–56. doi: 10.1016/0361-9230(89)90127-5
- Bussel, B. C., Roby-Brami, A., Yakovlev, A., and Bennis, N. (1988). "Evidences for the presence of a spinal stepping generator in patients with a spinal cord section," in *Posture and Gait: Development, Adaptation and Modulation*, eds B. Amblard, A. Berthoz and F. Clarac (Amsterdam, the Netherlands: Elsevier), 273–278.

- Butt, S. J., Harris-Warrick, R. M., and Kiehn, O. (2002). Firing properties of identified interneuron populations in the mammalian hindlimb central pattern generator. *J. Neurosci.* 22, 9961–9971.
- Byrnes, K. R., Garay, J., Di Giovanni, S., De Biase, A., Knobloch, S. M., Hoffman, E. P., et al. (2006). Expression of two temporally distinct microglia-related gene clusters after spinal cord injury. *Glia* 53, 420–433. doi: 10.1002/glia.20295
- Calancie, B. (2006). Spinal myoclonus after spinal cord injury. *J. Spinal Cord Med.* 29, 413–424.
- Calancie, B., Needham-Shropshire, B., Jacobs, P., Willer, K., Zych, G., and Green, B. A. (1994). Involuntary stepping after chronic spinal cord injury. Evidence for a central rhythm generator for locomotion in man. *Brain* 117(Pt. 5), 1143–1159. doi: 10.1093/brain/117.5.1143
- Camacho, F. J., Castro, M., Hernandez, V., and Paredes, R. G. (2007). Facilitation of ejaculation induced by 8-OH-DPAT does not produce conditioned place preference in male rats. *Behav. Neurosci.* 121, 579–585. doi: 10.1037/0735-7044.121.3.579
- Cazalets, J. R., Grillner, P., Menard, I., Cremieux, J., and Clarac, F. (1990). Two types of motor rhythm induced by NMDA and amines in an in vitro spinal cord preparation of neonatal rat. *Neurosci. Lett.* 111, 116–121. doi: 10.1016/0304-3940(90)90354-c
- Chang, H. Y., Cheng, C. L., Chen, J. J., and de Groat, W. C. (2006). Roles of glutamatergic and serotonergic mechanisms in reflex control of the external urethral sphincter in urethane-anesthetized female rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 291, R224–R234. doi: 10.1152/ajpregu.00780.2005
- Chau, C., Barbeau, H., and Rossignol, S. (1998a). Effects of intrathecal alpha1- and alpha2-noradrenergic agonists and norepinephrine on locomotion in chronic spinal cats. *J. Neurophysiol.* 79, 2941–2963.
- Chau, C., Barbeau, H., and Rossignol, S. (1998b). Early locomotor training with clonidine in spinal cats. *J. Neurophysiol.* 79, 392–409.
- Cheng, J., Stein, R. B., Jovanovic, K., Yoshida, K., Bennett, D. J., and Han, Y. (1998). Identification, localization and modulation of neural networks for walking in mudpuppy (*Necturus maculatus*) spinal cord. *J. Neurosci.* 18, 4295–4305.
- Cheng, J., and Magnuson, D. S. (2011). Initiation of segmental locomotor-like activities by stimulation of ventrolateral funiculus in the neonatal rat. *Exp. Brain Res.* 214, 151–161. doi: 10.1007/s00221-011-2816-7
- Chervin, R. D., Consens, F. B., and Kutluay, E. (2003). Alternating leg muscle activation during sleep and arousals: a new sleep-related motor phenomenon? *Mov. Disord.* 18, 551–559. doi: 10.1002/mds.10397
- Chi, S. L., Levine, J. D., and Basbaum, A. I. (1993). Peripheral and central contributions to the persistent expression of spinal cord fos-like immunoreactivity produced by sciatic nerve transection in the rat. *Brain Res.* 617, 225–237. doi: 10.1016/0006-8993(93)91090-f
- Cina, C., and Hochman, S. (2000). Diffuse distribution of sulforhodamine-labeled neurons during serotonin-evoked locomotion in the neonatal rat thoracolumbar spinal cord. *J. Comp. Neurol.* 423, 590–602. doi: 10.1002/1096-9861(20000807)423:4<590::aid-cne5>3.0.co;2-l
- Clarac, F., and Pearlstein, E. (2007). Invertebrate preparations and their contribution to neurobiology in the second half of the 20th century. *Brain Res. Rev.* 54, 113–161. doi: 10.1016/j.brainresrev.2006.12.007
- Clemens, S., Rye, D., and Hochman, S. (2006). Restless legs syndrome: revisiting the dopamine hypothesis from the spinal cord perspective. *Neurology* 67, 125–130. doi: 10.1212/01.wnl.0000223316.53428.c9
- Consentino, F. L., Iero, I., Lanuzza, B., Tripodi, M., and Ferri, R. (2006). The neurophysiology of the alternating leg muscle activation (ALMA) during sleep: study of one patient before and after treatment with pramipexole. *Sleep Med.* 7, 63–71. doi: 10.1016/j.sleep.2005.06.007
- Coolen, L. M., Veening, J. G., Wells, A. B., and Shirpley, M. T. (2003). Afferent connections of the parvocellular subparafascicular thalamic nucleus in the rat: evidence for functional subdivisions. *J. Comp. Neurol.* 463, 132–156. doi: 10.1002/cne.10739
- Courtine, G., van den Brand, R., and Musienko, P. (2011). Spinal cord injury: time to move. *Lancet* 377, 1896–1898. doi: 10.1016/s0140-6736(11)60711-3
- Courtois, F., Carrier, S., Charvier, K., Guertin, P. A., and Journel, M. (2013). The control of male sexual responses. *Curr. Pharm. Des.* 19, 4341–4356. doi: 10.2174/13816128113199990333
- Courtois, F. J., Charvier, K. F., Leriche, A., Vezina, J. G., Côté, M., and Bélanger, M. (2008). Blood pressure changes during sexual stimulation, ejaculation and midodrine treatment in men with spinal cord injury. *BJU Int.* 101, 331–337. doi: 10.1111/j.1464-410x.2007.07254.x
- Cowley, K. C., and Schmidt, B. J. (1994). A comparison of motor patterns induced by N-methyl-D-aspartate, acetylcholine and serotonin in the in vitro neonatal rat spinal cord. *Neurosci. Lett.* 171, 147–150. doi: 10.1016/0304-3940(94)90626-2
- Crone, S. A., Quinlan, K. A., Zagoraiou, L., Droho, S., Restrepo, C. E., Lundfald, L., et al. (2008). Genetic ablation of V2a ipsilateral interneurons disrupts left-right locomotor coordination in mammalian spinal cord. *Neuron* 60, 70–83. doi: 10.1016/j.neuron.2008.08.009
- Currie, S. N. (1999). Fictive hindlimb motor patterns evoked by AMPA and NMDA in turtle spinal cord-hindlimb nerve preparations. *J. Physiol. Paris* 93, 199–211. doi: 10.1016/s0928-4257(99)80152-1
- Currie, S. N., and Lee, S. (1997). Glycinergic inhibition contributes to the generation of rostral scratch motor patterns in the turtle spinal cord. *J. Neurosci.* 17, 3322–3333.
- de Groat, W. C., and Yoshimura, N. (2001). Pharmacology of the lower urinary tract. *Ann. Rev. Pharmacol. Toxicol.* 41, 691–721. doi: 10.1146/annurev.pharmtox.41.1.691
- de Groat, W. C., Booth, A. M., and Yoshimura, N. (1993). *Nervous Control of the Urogenital System (Autonomic Nervous System)* (Vol. 3), ed C. A. Maggi (London: Harwood Academic Publishers), 227–289.
- Delcomyn, F. (1977). “Co-ordination of invertebrate locomotion,” in *Mechanics and Energetics of Locomotion*, eds R. M. Alexander and G. Gold Spink (London: Chapman and Hall), 82–114.
- Dietz, V., and Sinkjaer, T. (2012). Spasticity. *Handb. Clin. Neurol.* 109, 197–211. doi: 10.1016/B978-0-444-52137-8.00012-7
- Dimitrijevic, M. R., Gerasimenko, Y., and Pinter, M. M. (1998). Evidence for a spinal central pattern generator in humans. *Ann. N Y Acad. Sci.* 860, 360–376. doi: 10.1111/j.1749-6632.1998.tb09062.x
- Dolber, P. C., Gu, B., Zhang, X., Fraser, M. O., Thor, K. B., and Reiter, J. P. (2007). Activation of the external urethral sphincter central pattern generator by a 5-HT(1A) receptor agonist in rats with chronic spinal cord injury. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 292, R1699–R1706. doi: 10.1152/ajpregu.00142.2006
- Durán, I., Gil, L., and Cueva-Rolon, R. (2000). Masculine copulatory behavior is facilitated by intrathecally administered muscarine. *Exp. Brain Res.* 134, 490–496. doi: 10.1007/s002210000488
- Duysens, J., De Groote, F., and Jonkers, I. (2013). The flexion synergy, mother of all synergies and father of new models of gait. *Front. Comput. Neurosci.* 7:14. doi: 10.3389/fncom.2013.00014
- Duysens, J. (1977). Reflex control locomotion as revealed by stimulation of cutaneous afferents in spontaneously walking pre-mammillary cats. *J. Neurophysiol.* 40, 737–751.
- Edgerton, V. R., Grillner, S., Sjöström, A., and Zangger, P. (1976). “Central generation of locomotion in vertebrates,” in *Neural Control of Locomotion*, eds R. Herman, S. Grillner and P. Zangger (New York: Plenum Press), 439–464.
- Edgerton, V. R., Leon, R. D., Harkema, S. J., Hodgson, J. A., London, N., Reinkensmeyer, D. J., et al. (2001). Retraining the injured spinal cord. *J. Physiol.* 533, 15–22. doi: 10.1111/j.1469-7793.2001.0015b.x
- Eken, T., Hultborn, H., and Kiehn, O. (1989). Possible functions of transmitter-controlled plateau potentials in alpha motoneurons. *Prog. Brain Res.* 80, 257–267; discussion 239–242. doi: 10.1016/s0079-6123(08)62219-0
- English, A. W. (1985). Interlimb coordination during stepping in the cat: the role of the dorsal spinocerebellar tract. *Exp. Neurol.* 87, 96–108. doi: 10.1016/0014-4886(85)90136-0
- Fedirchuk, B., Nielsen, J., Petersen, N., and Hultborn, H. (1998). Pharmacologically evoked fictive motor patterns in the acutely spinalized marmoset monkey (*Callithrix jacchus*). *Exp. Brain Res.* 122, 351–361. doi: 10.1007/s002210050523
- Ferens, D. M., Habgood, M. D., Saunders, N. R., Tan, Y. H., Brown, D. J., Brock, J. A., et al. (2011). Stimulation of defecation in spinal cord-injured rats by a centrally acting ghrelin receptor agonist. *Spinal Cord* 49, 1036–1041. doi: 10.1038/sc.2011.60
- Field-Fote, E., Ness, L. L., and Ionno, M. (2012). Vibration elicits involuntary, step-like behavior in individuals with spinal cord injury. *Neurorehabil. Neural Repair* 26, 861–869. doi: 10.1177/1545968311433603
- Flourens, M.-J.-P. (1824). *Recherches expérimentales sur les propriétés et les Fonctions du Système Nerveux, dans les Animaux Vertébrés, Experimental Studies on the Properties and Functions of the Nervous System in Vertebrate Animals*. Paris: Chez Crevot.

- Forsberg, H., and Grillner, S. (1973). The locomotion of the acute spinal cat injected with clonidine i.v. *Brain Res.* 50, 184–186. doi: 10.1016/0006-8993(73)90606-9
- Forsberg, H., and Hirschfeld, H. (1988). Phasic modulation of postural activation patterns during human walking. *Prog. Brain Res.* 76, 221–227. doi: 10.1016/s0079-6123(08)64508-2
- Fowler, C. J., Beck, R. O., Gerrard, S., Betts, C. D., and Fowler, C. G. (1994). Intravascular capsaicin for treatment of detrusor hyperreflexia. *J. Neurol. Neurosurg. Psych.* 57, 169–173. doi: 10.1136/jnnp.57.2.169
- Freusberg, A. (1874). Reflexbewegungen beim hunde. *Pflügers Arch* 9, 358–391. doi: 10.1007/bf01612347
- Fung, J., Stewart, J. E., and Barbeau, H. (1990). The combined effects of clonidine and cyproheptadine with interactive training on the modulation of locomotion in spinal cord injured subjects. *J. Neurol. Sci.* 100, 85–93. doi: 10.1016/0022-510x(90)90017-h
- García-Bravo, A. M., Suarez-Hernandez, D., Ruiz-Fernandez, M. A., Silva Gonzalez, O., Barbara-Bataller, E., and Méndez Suarez, J. L. (2006). Determination of changes in blood pressure during administration of Viagra in patients with spinal cord injury and erectile dysfunction. *Spinal Cord* 44, 301–308. doi: 10.1038/sj.sc.3101846
- Gerasimenko, Y., Gorodnichev, R., Machueva, E., Pivovarova, E., Semyenov, D., Savochin, A., et al. (2010). Novel and direct access to the human locomotor spinal circuitry. *J. Neurosci.* 30, 3700–3708. doi: 10.1523/jneurosci.4751-09.2010
- Gorodnichev, R. M., Pivovarova, E. A., Pukhov, A., Moiseev, S. A., Savokhin, A. A., and Moshonkina, T. R. (2012). [Transcutaneous electrical stimulation of the spinal cord: non-invasive tool for activation of locomotor circuitry in human]. *Fiziol. Cheloveka* 38, 46–56.
- Gerasimenko, Y., Roy, R. R., and Edgerton, V. R. (2008). Epidural stimulation: comparison of the spinal circuits that generate and control locomotion in rats, cats and humans. *Exp. Neurol.* 209, 417–425. doi: 10.1016/j.expneurol.2007.07.015
- Getting, P. A. (1977). Neuronal organization of escape swimming in Tritonia. *J. Comp. Physiol.* 121, 325–342. doi: 10.1007/bf00613012
- Ginty, D. D., Bading, H., and Greenberg, M. E. (1992). Trans-synaptic regulation of gene expression. *Curr. Opin. Neurobiol.* 2, 312–316. doi: 10.1016/0959-4388(92)90121-z
- Giroux, N., Rossignol, S., and Reader, T. A. (1999). Autoradiographic study of alpha1- and alpha2-noradrenergic and serotonin1A receptors in the spinal cord of normal and chronically transected cats. *J. Comp. Neurol.* 406, 402–414. doi: 10.1002/(sici)1096-9861(19990412)406:3<402::aid-cne8>3.3.co;2-6
- Giszter, S., Patil, V., and Hart, C. (2007). Primitives, premotor drives and pattern generation: a combined computational and neuroethological perspective. *Prog. Brain Res.* 165, 323–346. doi: 10.1016/s0079-6123(06)65020-6
- Gómez-Pinilla, F., Ying, Z., Roy, R. R., Hodgson, J., and Edgerton, V. R. (2004). Afferent input modulates neurotrophins and synaptic plasticity in the spinal cord. *J. Neurophysiol.* 92, 3423–3432. doi: 10.1152/jn.00432.2004
- Gosgnach, S., Lanuza, G. M., Butt, S. J., Saueressig, H., Zhang, Y., Velasquez, T., et al. (2006). V1 spinal neurons regulate the speed of vertebrate locomotor outputs. *Nature* 440, 215–219. doi: 10.1038/nature04545
- Gossard, J. P., Brownstone, R. M., Barajon, I., and Hultborn, H. (1994). Transmission in a locomotor related group Ib pathway from hindlimb extensor muscles in the cat. *Exp. Brain Res.* 98, 213–228. doi: 10.1007/bf00228410
- Goulding, M. (2009). Circuits controlling vertebrate locomotion: moving in a new direction. *Nat. Rev. Neurosci.* 10, 507–518. doi: 10.1038/nrn2608
- Graham Brown, T. (1911). The intrinsic factors in the act of progression in the mammal. *Proc. R. Soc. Lond.* 84, 309–319. doi: 10.1098/rspb.1911.0077
- Graham Brown, T. (1914). On the nature of the fundamental activity of the nervous centres; together with an analysis of the conditioning of rhythmic activity in progression and a theory of the evolution of function in the nervous system. *J. Physiol.* 48, 18–46.
- Grillner, S. (1981). “Control of locomotion in bipeds, tetrapods and fish,” in *Handbook of Physiology. The Nervous System II*, eds J. M. Brookhart and V. B. Mountcastle (Bethesda: Am. Physiol. Soc.), 1179–1236.
- Grillner, S. (2006). Neuronal networks in motion from ion channels to behaviour. *An. R. Acad. Nac. Med. (Madr)*. 123, 297–298.
- Grillner, S., and Wallén, P. (1985). The ionic mechanisms underlying N-methyl-D-aspartate receptor-induced, tetrodotoxin-resistant membrane potential oscillations in lamprey neurons active during locomotion. *Neurosci. Lett.* 60, 289–294. doi: 10.1016/0304-3940(85)90592-0
- Grillner, S., and Zangger, P. (1974). Locomotor movements generated by the deafferented spinal cord. *Acta Physiol. Scand.* 91, 38–39.
- Grillner, S., and Zangger, P. (1975). How detailed is the central pattern generation for locomotion? *Brain Res.* 88, 367–371. doi: 10.1016/0006-8993(75)90401-1
- Grob, M., and Guertin, P. A. (2007). [Role of Ca(2+) in the pacemaker-like property of spinal motoneurons]. *Med. Sci. (Paris)* 23, 64–66. doi: 10.1051/medsci/200723164
- Guertin, P. (1996). “Central mediation of group I muscle afferent evoked adaptation of the locomotor step cycle in decerebrate cats”. Thesis, (University of Manitoba).
- Guertin, P., Angel, M. J., Perreault, M.-C., and McCrea, D. A. (1995). Ankle extensor group I afferents excite extensors throughout the hindlimb during fictive locomotion in the cat. *J. Physiol.* 487, 197–209.
- Guertin, P. A. (2004). Role of NMDA receptor activation in serotonin agonist-induced air-stepping in paraplegic mice. *Spinal Cord* 42, 185–190. doi: 10.1038/sj.sc.3101580
- Guertin, P. A. (2005). Semiquantitative assessment of hindlimb movement recovery without intervention in adult paraplegic mice. *Spinal Cord* 43, 162–166. doi: 10.1038/sj.sc.3101701
- Guertin, P. A. (2008). A technological platform to optimize combinatorial treatment design and discovery for chronic spinal cord injury. *J. Neurosci. Res.* 86, 3039–3051. doi: 10.1002/jnr.21761
- Guertin, P. A. (2009a). The mammalian central pattern generator for locomotion. *Brain Res. Rev.* 62, 45–56. doi: 10.1016/j.brainresrev.2009.08.002
- Guertin, P. A. (2009b). Recovery of locomotor function with combinatory drug treatments designed to synergistically activate specific neuronal networks. *Curr. Med. Chem.* 16, 1366–1371. doi: 10.2174/092986709787846541
- Guertin, P. A. (2012). “The spinal cord: functional organization, diseases and dysfunctions,” in *Neuromethods: Animal Models in Spinal Cord Repair*, ed H. Aldskogius (New York, NY: Humana Press), 1–23.
- Guertin, P. A., and Hounsgaard, J. (1998a). Chemical and electrical stimulation induce rhythmic motor activity in an in vitro preparation of the spinal cord from adult turtles. *Neurosci. Lett.* 245, 5–8. doi: 10.1016/s0304-3940(98)00164-5
- Guertin, P. A., and Hounsgaard, J. (1998b). NMDA-Induced intrinsic voltage oscillations depend on L-type calcium channels in spinal motoneurons of adult turtles. *J. Neurophysiol.* 80, 3380–3382.
- Guertin, P. A., and Steuer, I. (2005). Ionotropic 5-HT₃ receptor agonist-induced motor responses in the hindlimbs of paraplegic mice. *J. Neurophysiol.* 94, 3397–3405. doi: 10.1152/jn.00587.2005
- Guertin, P. A., and Steuer, I. (2009). Key central pattern generators of the spinal cord. *J. Neurosci. Res.* 87, 2399–2405. doi: 10.1002/jnr.22067
- Guertin, P. A., Ung, R. V., and Rouleau, P. (2010). Oral administration of a tri-therapy for central pattern generator activation in paraplegic mice: proof-of-concept of efficacy. *Biotechnol. J.* 5, 421–426. doi: 10.1002/biot.200900278
- Guertin, P. A., Ung, R. V., Rouleau, P., and Steuer, I. (2011). Effects on locomotion, muscle, bone and blood induced by a combination therapy eliciting weight-bearing stepping in nonassisted spinal cord-transected mice. *Neurorehabil. Neural Repair* 25, 234–242. doi: 10.1177/1545968310378753
- Gurfinkel, V. S., and Shik, M. L. (1973). “The control of posture and locomotion,” in *Motor Control*, eds A. A. Gydkov, N. T. Tankov and D. S. Kosarov (New York: Plenum Press), 217–234.
- Guttman, L., and Walsh, J. J. (1971). Prostagmine assessment test of fertility in spinal man. *Paraplegia* 9, 39–51. doi: 10.1038/sc.1971.7
- Harkema, S., Behrman, A., and Barbeau, H. (2012). Evidence-based therapy for recovery of function after spinal cord injury. *Handb. Clin. Neurol.* 109, 259–274. doi: 10.1016/B978-0-444-52137-8.00016-4
- Harkema, S. J. (2008). Plasticity of interneuronal networks of the functionally isolated human spinal cord. *Brain Res. Rev.* 57, 255–264. doi: 10.1016/j.brainresrev.2007.07.012
- Harris-Warrick, R. M. (2002). Voltage-sensitive ion channels in rhythmic motor systems. *Curr. Opin. Neurobiol.* 2, 646–651. doi: 10.1016/s0959-4388(02)00377-x
- Harris-Warrick, R. M. (2011). Neuromodulation and flexibility in central pattern generator networks. *Curr. Opin. Neurobiol.* 21, 685–692. doi: 10.1016/j.conb.2011.05.011
- Henneman, E. (1974). “Spinal reflexes and the control of movement,” in *Medical Physiology* (Vol. 1), ed V. B. Mountcastle, 13th Edn. (Mosby, MO: St Louis), 651–667.

- Hochman, S., Gozal, E. A., Hayes, H. B., Anderson, J. T., DeWeerth, S. P., and Chang, Y. H. (2012). Enabling techniques for in vitro studies on mammalian spinal locomotor mechanisms. *Front. Biosci. (Landmark Ed.)* 17, 2158–2180. doi: 10.2741/4043
- Hochman, S., Jordan, L. M., and MacDonald, J. F. (1994). N-methyl-D-aspartate receptor-mediated voltage oscillations in neurons surrounding the central canal in slices of rat spinal cord. *J. Neurophysiol.* 72, 565–577.
- Holinski, B. J., Mazurek, K. A., Everaert, D. G., Stein, R. B., and Mushahwar, V. K. (2011). Restoring stepping after spinal cord injury using intraspinal microstimulation and novel control strategies. *Conf. Proc. IEEE Eng. Med. Biol. Soc.* 2011, 5798–5801. doi: 10.1109/IEMBS.2011.6091435
- Holmes, G. (1915). Spinal injuries of warfare. *Br. Med. J.* 2, 815–821. doi: 10.1136/bmj.2.2866.815
- Hopper, S. L., and DiCaprio, R. A. (2004). Crustacean motor pattern generator networks. *Invertebrate neural networks. Neurosignals* 13, 50–69. doi: 10.1159/000076158
- Huges, G. M., and Wiersma, C. A. G. (1960). The coordination of swimmeret movements in the crayfish, *procambarus clarkii* (girard). *J. Exptl. Biol.* 39, 657–670.
- Ivanenko, Y. P., Poppele, R. E., and Lacquantini, F. (2006). Motor control programs and walking. *Neuroscientist* 12, 339–348. doi: 10.1177/1073858406287987
- Jankowska, E. (2008). Spinal interneuronal networks in the cat: elementary components. *Brain Res. Rev.* 57, 46–55. doi: 10.1016/j.brainresrev.2007.06.022
- Jankowska, E., Jukes, M. G., Lund, S., and Lundberg, A. (1967a). The effect of DOPA on the spinal cord. 5. Reciprocal organization of pathways transmitting excitatory action to alpha motoneurons of flexors and extensors. *Acta Physiol. Scand.* 70, 369–388. doi: 10.1111/j.1748-1716.1967.tb03636.x
- Jankowska, E., Jukes, M. G., Lund, S., and Lundberg, A. (1967b). The effect of DOPA on the spinal cord. 6. Half-centre organization of interneurons transmitting effects from the flexor reflex afferents. *Acta Physiol. Scand.* 70, 389–402. doi: 10.1111/j.1748-1716.1967.tb03637.x
- Johnston, M. V., Mullaney, B., and Blue, M. E. (2003). Neurobiology of rett syndrome. *J. Child Neurol.* 18, 688–692. doi: 10.1177/08830738030180100501
- Jonas, D., Linzbach, P., and Weber, W. (1979). The use of Midodrin in the treatment of ejaculation disorders following retroperitoneal lymphadenectomy. *Eur. Urol.* 5, 184–187.
- Jones, R., and Cavanna, A. E. (2012). The neurobiology and treatment of restless legs syndrome. *Behav. Neurol.* 26, 283–292. doi: 10.1155/2013/585439
- Jordan, L. M., Liu, J., Hedlund, P. B., Akay, T., and Pearson, K. G. (2008). Descending command systems for the initiation of locomotion in mammals. *Brain Res. Rev.* 57, 183–191. doi: 10.1016/j.brainresrev.2007.07.019
- Juvin, L., Le Gal, J. P., Simmers, J., and Morin, D. (2012). Cervicolumbar coordination in mammalian quadrupedal locomotion: role of spinal thoracic circuitry and limb sensory inputs. *J. Neurosci.* 32, 953–965. doi: 10.1523/jneurosci.4640-11.2012
- Kalat, J. W. (1998). *Biological Psychology*. 6th Edn. Pacific Grove, CA: Brooks/Cole Publishing Company.
- Kandel, E. R., Schwartz, J. H., and Jessell, T. M. (2000). *Principles of Neural Science*. 4th Edn. New York: McGraw-Hill.
- Kiehn, O., and Kjaerulff, O. (1996). Spatiotemporal characteristics of 5-HT and dopamine-induced rhythmic hindlimb activity in the in vitro neonatal rat. *J. Neurophysiol.* 75, 1472–1482.
- Kiehn, O., Quinlan, K. A., Restrepo, C. E., Lundfald, L., Borgius, L., Talpalar, A. E., et al. (2008). Excitatory components of the mammalian locomotor CPG. *Brain Res. Rev.* 57, 56–63. doi: 10.1016/j.brainresrev.2007.07.002
- Kjaerulff, O., Barajon, I., and Kiehn, O. (1994). Sulphorhodamine-labelled cells in the neonatal rat spinal cord following chemically induced locomotor activity in vitro. *J. Physiol.* 478, 265–273.
- Knikou, M., Angeli, C. A., Ferreira, C. K., and Harkema, S. J. (2009). Flexion reflex modulation during stepping in human spinal cord injury. *Exp. Brain Res.* 196, 341–351. doi: 10.1007/s00221-009-1854-x
- Knusel, J., Bicanski, A., Ryczko, D., Cabelguen, J. M., and Ljsspeert, A. J. (2013). A salamander's flexible spinal network for locomotion, modeled at two levels of abstraction. *Integr. Comp. Biol.* 53, 269–282. doi: 10.1093/icb/ict067
- Kriellaars, D. J., Brownstone, R. M., Noga, B. R., and Jordan, L. M. (1994). Mechanical entrainment of fictive locomotion in the decerebrate cat. *J. Neurophysiol.* 71, 2074–2086.
- Kriellaars, D. J. (1992). *Generation and Peripheral Control of Locomotor Rhythm*. Ph.D. thesis. Manitoba: University of Manitoba.
- Kristan, W. B. Jr., and Weeks, J. C. (1983). Neurons controlling the initiation, generation and modulation of leech swimming. *Symp. Soc. Exp. Biol.* 37, 243–260.
- Kullander, K., Butt, S. J., Le Bret, J. M., Lundfald, L., Restrepo, C. E., Rydstrom, A., et al. (2003). Role of EphA4 and EphrinB3 in local neuronal circuits that control walking. *Science* 299, 1889–1892. doi: 10.1126/science.1079641
- Lafreniere-Roula, M., and McCrear, D. A. (2005). Deletions of rhythmic motoneuron activity during fictive locomotion and scratch provide clues to the organization of the mammalian central pattern generator. *J. Neurophysiol.* 94, 1120–1132. doi: 10.1152/jn.00216.2005
- Landry, E. S., and Guertin, P. A. (2004). Differential effects of 5-HT1 and 5-HT2 receptor agonists on hindlimb movements in paraplegic mice. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 28, 1053–1060. doi: 10.1016/j.pnpb.2004.05.001
- Landry, E. S., Lapointe, N. P., Rouillard, C., Levesque, D., Hedlund, P. B., and Guertin, P. A. (2006a). Contribution of spinal 5-HT1A and 5-HT7 receptors to locomotor-like movement induced by 8-OH-DPAT in spinal cord-transected mice. *Eur. J. Neurosci.* 24, 535–546. doi: 10.1111/j.1460-9568.2006.04917.x
- Landry, E. S., Rouillard, C., Levesque, D., and Guertin, P. A. (2006b). Profile of immediate early gene expression in the lumbar spinal cord of low-thoracic paraplegic mice. *Behav. Neurosci.* 120, 1384–1388. doi: 10.1037/0735-7044.120.6.1384
- Langlet, C., Leblond, H., and Rossignol, S. (2005). Mid-lumbar segments are needed for the expression of locomotion in chronic spinal cats. *J. Neurophysiol.* 93, 2474–2488. doi: 10.1152/jn.00909.2004
- Lanuzza, G. M., Gosgnach, S., Pierani, A., Jessell, T. M., and Goulding, M. (2004). Genetic identification of spinal interneurons that coordinate left-right locomotor activity necessary for walking movements. *Neuron* 42, 375–386. doi: 10.1016/s0896-6273(04)00249-1
- Lapointe, N. P., Ung, R. V., Rouleau, P., and Guertin, P. A. (2008). Effects of spinal alpha(2)-adrenoceptor and I(1)-imidazoline receptor activation on hindlimb movement induction in spinal cord-injured mice. *J. Pharmacol. Exp. Ther.* 325, 994–1006. doi: 10.1124/jpet.107.134874
- Lapointe, N. P., Rouleau, P., Ung, R. V., and Guertin, P. A. (2009). Specific role of D1 receptors in spinal network activation and rhythmic movement induction in vertebrates. *J. Physiol.* 587, 1499–1511. doi: 10.1113/jphysiol.2008.166314
- Le Ray, D., Juvin, L., Ryczko, D., and Dubuc, R. (2011). Chapter 4—supraspinal control of locomotion: the mesencephalic locomotor region. *Prog. Brain Res.* 188, 51–70. doi: 10.1016/B978-0-444-53825-3.00009-7
- Lhermitte, J. (1919). *La section totale de la Moelle Dorsale*. Bourges, France: Tardy Pigelet.
- Li, X. L., Zhang, W., Zhou, X., Wang, X. Y., Zhang, H. T., Qin, D. X., et al. (2007). Temporal changes in the expression of some neurotrophins in spinal cord transected adult rats. *Neuropeptides* 41, 135–143. doi: 10.1016/j.npep.2007.02.001
- Liu, Y. C., Bailey, I., and Hale, M. E. (2012). Alternative startle motor patterns and behaviors in the larval zebrafish (*Danio rerio*). *J. Comp. Physiol. A Neuroethol. Sens. Neural Behav. Physiol.* 198, 11–24. doi: 10.1007/s00359-011-0682-1
- Lovely, R. G., Gregor, R. J., Roy, R. R., and Edgerton, V. R. (1986). Effects of training on the recovery of full-weight-bearing stepping in the adult spinal cat. *Exp. Neurol.* 92, 421–435. doi: 10.1016/0014-4886(86)90094-4
- Lukacova, N., Kolesarova, M., Kucharova, K., Pavel, J., Kolesar, D., Radonak, J., et al. (2006). The effect of a spinal cord hemisection on changes in nitric oxide synthase pools in the site of injury and in regions located far away from the injured site. *Cell. Mol. Neurobiol.* 26, 1365–1383. doi: 10.1007/s10571-006-9092-2
- Lundfald, L., Restrepo, C. E., Butt, S. J., Peng, C. Y., Droho, S., Endo, T., et al. (2007). Phenotype of V2-derived interneurons and their relationship to the axon guidance molecule EphA4 in the developing mouse spinal cord. *Eur. J. Neurosci.* 26, 2989–3002. doi: 10.1111/j.1460-9568.2007.05906.x
- MacLean, J. N., Schmidt, B. J., and Hochman, S. (1997). NMDA receptor activation triggers voltage oscillations, plateau potentials and bursting in neonatal rat lumbar motoneurons in vitro. *Eur. J. Neurosci.* 9, 2702–2711. doi: 10.1111/j.1460-9568.1997.tb01699.x
- Mallory, B. S., Roppolo, J. R., and de Groat, W. C. (1991). Pharmacological modulation of the pontine micturition center. *Brain Res.* 546, 310–320.
- Matsumoto, N., Yoshida, M., and Uematsu, K. (2007). Effects of partial ablation of the cerebellum on sustained swimming in goldfish. *Brain Behav. Evol.* 70, 105–114. doi: 10.1159/000102972

- Matthews, P. B. C. (1972). *Mammalian Muscle Receptors and Their Central Actions*. Baltimore, MD: Williams and Wilkins.
- Matthews, P. B. C. (1991). The human stretch reflex and the motor cortex. *Trends Neurosci.* 14, 87–91. doi: 10.1016/0166-2236(91)90064-2
- McCrea, D. A., and Rybak, I. A. (2008). Organization of mammalian locomotor rhythm and pattern generation. *Brain Res. Rev.* 57, 134–146. doi: 10.1016/j.brainresrev.2007.08.006
- McCrea, D. A., Pratt, C. A., and Jordan, L. M. (1980). Renshaw cell activity and recurrent effects on motoneurons during fictive locomotion. *J. Neurophys.* 44, 475–488.
- McKenna, K. E., Chung, S. K., and McVary, K. T. (1991). A model for the study of sexual function in anesthetized male and female rats. *Am. J. Physiol.* 261, 1276–1285.
- Miller, S., and Scott, P. D. (1977). The spinal locomotor generator. *Exp. Brain Res.* 30, 387–403. doi: 10.1007/bf00237264
- Moreland, A. J., and Makela, E. H. (2005). SSRIs in the treatment of premature ejaculation. *Ann. Pharmacother.* 39, 1296–1301.
- Moshonkina, T. R., Makarovski, A. N., Bogacheva, I. N., Scherbakova, N. A., Savohin, A. A., and Gerasimenko, Y. P. (2012). Effects of spinal cord electrical stimulation in patients with vertebrospinal pathology. *Bull. Exp. Biol. Med.* 153, 16–20. doi: 10.1007/s10517-012-1632-9
- Nadeau, S., Jacquemin, G., Fournier, C., Lamarre, Y., and Rossignol, S. (2010). Spontaneous motor rhythms of the back and legs in a patient with a complete spinal cord transection. *Neurorehabil. Neural Repair* 24, 377–383. doi: 10.1177/1545968309349945
- Nadelhaft, I., and Vera, P. L. (1995). Central nervous system neurons infected by pseudorabies virus injected into the rat urinary bladder following unilateral transection of the pelvic nerve. *J. Comp. Neurol.* 359, 443–456. doi: 10.1002/cne.903590307
- Netter, F. H. (2006). *Atlas of Human Anatomy*. Philadelphia, PA: Saunders/Elsevier.
- Nishimaru, H., Takizawa, H., and Kudo, N. (2000). 5-Hydroxytryptamine-induced locomotor rhythm in the neonatal mouse spinal cord in vitro. *Neurosci. Lett.* 280, 187–190. doi: 10.1016/s0304-3940(00)00805-3
- Noga, B. R., Johnson, D. M., Riesgo, M. I., and Pinzon, A. (2009). Locomotor-activated neurons of the cat. I. Serotonergic innervation and co-localization of 5-HT7, 5-HT2A, and 5-HT1A receptors in the thoraco-lumbar spinal cord. *J. Neurophysiol.* 102, 1560–1576. doi: 10.1152/jn.91179.2008
- Ollivier-Lanvin, K., Krupka, A. J., AuYong, N., Miller, K., Prilutsky, B. I., and Lemay, M. A. (2011). Electrical stimulation of the sural cutaneous afferent nerve controls the amplitude and onset of the swing phase of locomotion in the spinal cat. *J. Neurophysiol.* 105, 2297–2308. doi: 10.1152/jn.00385.2010
- Orlovski, G. N., Severin, F. V., and Shik, M. L. (1966). Effect of speed and load on coordination of movements during running of the dog. *Biophysica* 11, 414–417.
- Orlovskii, G. N., Severin, F. V., and Shik, M. L. (1966). Locomotion induced by stimulation of the mesencephalon. *Dokl. Akad. Nauk. SSSR* 169, 1223–1226.
- Orlovsky, G., and Shik, M. (1976). Control of locomotion: a neurophysiological analysis of the cat locomotor system. *Intern. Rev. Physiol. Ser. II* 10, 281–317.
- Pateatas, M. A., and Gartner, L. P. (2006). “Ascending sensory pathways,” in *A Textbook of Neuroanatomy*, eds M. A. Pateatas and L. P. Gartner (Etobicoke, ON: Wiley-Blackwell), 137–170.
- Pearson, K., and Duysens, J. (1976). “Function of segmental reflexes in the control of stepping in cockroaches and cats,” in *Neural Control of Locomotion*, eds R. Herman, S. Grillner, P. S. G. Stein and D. Stuart (New York: Plenum Press), 519–538.
- Pearson, K. G. (1993). Common principles of motor control in vertebrates and invertebrates. *Annu. Rev. Neurosci.* 16, 265–297. doi: 10.1146/annurev.neuro.16.1.265
- Pearson, K. G. (1995). Proprioceptive regulation of locomotion. *Curr. Opin. Neurobiol.* 5, 786–791. doi: 10.1016/0959-4388(95)80107-3
- Pearson, K. G., and Rossignol, S. (1991). Fictive motor patterns in chronic spinal cats. *J. Neurophysiol.* 66, 1874–1887.
- Peng, X. M., Zhou, Z. G., Glorioso, J. C., Fink, D. J., and Mata, M. (2006). Tumor necrosis factor- α contributes to below-level neuropathic pain after spinal cord injury. *Ann. Neurol.* 59, 843–851. doi: 10.1002/ana.20855
- Perreault, M. C., Angel, M. J., Guertin, P., and McCrea, D. A. (1995). Effects of stimulation of hindlimb flexor group II afferents during fictive locomotion in the cat. *J. Physiol.* 487(Pt. 1), 211–220.
- Perret, C., and Cabelguen, J. M. (1980). Main characteristics of the hindlimb locomotor cycle in the decorticate cat with special reference to bifunctional muscles. *Brain Res.* 187, 333–352. doi: 10.1016/0006-8993(80)90207-3
- Pomerantz, S. M., Hepner, B. C., and Wertz, J. M. (1993). Serotonergic influences on male sexual behavior of rhesus monkeys: effects of serotonin agonists. *Psychopharmacology (Berl)* 111, 47–54. doi: 10.1007/bf02257406
- Pozos, R. S., and Iaizzo, P. A. (1991). Shivering and pathological and physiological clonic oscillations of the human ankle. *J. Appl. Physiol.* (1985) 71, 1929–1932.
- Pustovit, R. V., Callaghan, B., Kosari, S., Rivera, L. R., Thomas, H., Brock, J. A., et al. (2014). The mechanism of enhanced defecation caused by the ghrelin receptor agonist, ulimorelin. *Neurogastroenterol Motil.* 26, 264–271. doi: 10.1111/nmo.12259
- Reith, C. A., and Sillar, K. T. (1998). A role for slow NMDA receptor-mediated, intrinsic neuronal oscillations in the control of fast fictive swimming in *Xenopus laevis* larvae. *Eur. J. Neurosci.* 10, 1329–1340. doi: 10.1046/j.1460-9568.1998.00144.x
- Reitz, A., Knapp, P. A., Müntener, M., and Schurch, B. (2004). Oral nitric oxide donors: a new pharmacological approach to detrusor-sphincter dyssynergia in spinal cord injured patients? *Eur. Urol.* 45, 516–520. doi: 10.1016/j.eururo.2003.11.006
- Rémy-Néris, O., Barbeau, H., Daniel, O., Boiteau, F., and Bussel, B. (1999). Effects of intrathecal clonidine injection on spinal reflexes and human locomotion in incomplete paraplegic subjects. *Exp. Brain Res.* 129, 433–440. doi: 10.1007/s002210050910
- Riley, A. J., and Riley, E. J. (1982). Partial ejaculatory incompetence: the therapeutic effect of midodrine, an orally active selective alpha-adrenoceptor agonist. *Eur. Urol.* 8, p155–p160.
- Robertson, R. M., and Pearson, K. G. (1985). Neural circuits in the flight system of the locust. *J. Neurophysiol.* 53, 110–128.
- Rossignol, S., Giroux, N., Chau, C., Marcoux, J., Brustein, E., and Reader, T. A. (2001). Pharmacological aids to locomotor training after spinal injury in the cat. *J. Physiol.* 533(Pt. 1), 65–74. doi: 10.1111/j.1469-7793.2001.0065b.x
- Rostad, H. (1973). Extrinsic and central nervous control of colonic motility. A survey of previous and new concepts based on experimental results. *J. Oslo City Hosp.* 23, 65–75.
- Rye, D. B., and Trotti, L. M. (2012). Restless legs syndrome and periodic leg movements of sleep. *Neurol. Clin.* 30, 1137–1166. doi: 10.1016/j.ncl.2012.08.004
- Schmidt, B. J., and Jordan, L. M. (2000). The role of serotonin in reflex modulation and locomotor rhythm production in the mammalian spinal cord. *Brain Res. Bull.* 53, 689–710. doi: 10.1016/s0361-9230(00)00402-0
- Schmidt, B. J., Hochman, S., and MacLean, J. N. (1998). NMDA receptor-mediated oscillatory properties: potential role in rhythm generation in the mammalian spinal cord. *Ann. N Y Acad. Sci.* 860, 189–202. doi: 10.1111/j.1749-6632.1998.tb09049.x
- Schmitt, C., Miranpuri, G. S., Dhodda, V. K., Isaacson, J., Vemuganti, R., and Resnick, D. K. (2006). Changes in spinal cord injury-induced gene expression in rat are strain-dependent. *Spine J.* 6, 113–119. doi: 10.1016/j.spinee.2005.05.379
- Schomburg, E. D., Petersen, N., Barajon, I., and Hultborn, H. (1998). Flexor reflex afferents reset the step cycle during fictive locomotion in the cat. *Exp. Brain Res.* 122, 339–350. doi: 10.1007/s002210050522
- Schröder, H. D. (1985). Anatomical and pathoanatomical studies on the spinal efferent systems innervating pelvic structures. 1. Organization of spinal nuclei in animals. 2. The nucleus X-pelvic motor system in man. *J. Auton. Nerv. Syst.* 14, 23–48. doi: 10.1016/0165-1838(85)90123-7
- Schurch, B., Stöhrer, M., Kramer, G., Schmid, D. M., Gaul, G., and Hauri, D. (2000). Botulinum-A toxin for treating detrusor hyperreflexia in spinal cord injured patients: a new alternative to anticholinergic drugs? Preliminary results. *J. Urol.* 164, 692–697. doi: 10.1016/S0022-5347(05)67283-7
- Schürks, M., and Bussfeld, P. (2013). Multiple sclerosis and restless legs syndrome: a systematic review and meta-analysis. *Eur. J. Neurol.* 20, 605–615. doi: 10.1111/j.1468-1331.2012.03873.x
- Selionova, V. A., Ivanenko, Y. P., Solopova, I. A., and Gurfinkel, V. S. (2009). Tonic central and sensory stimuli facilitate involuntary air-stepping in humans. *J. Neurophysiol.* 101, 2847–2858. doi: 10.1152/jn.90895.2008

- Shapkova, E. Y., and Schomburg, E. D. (2001). Two types of motor modulation underlying human stepping by spinal cord electrical stimulation (SCES). *Acta Physiol. Pharmacol. Bulg.* 26, 155–157.
- Sherrington, C. S. (1910). Flexion-reflex of the limb, crossed extension-reflex and reflex stepping and standing. *J. Physiol.* 40, 28–121.
- Shimizu, Y., Chang, E. C., Shafton, A. D., Ferens, D. M., Sanger, G. J., Witherington, J., et al. (2006). Evidence that stimulation of ghrelin receptors in the spinal cord initiates propulsive activity in the colon of the rat. *J. Physiol.* 576(Pt. 1), 329–338. doi: 10.1113/jphysiol.2006.116160
- Sigvardt, K. A., Grillner, S., Wallen, P., and Van Dongen, P. A. (1985). Activation of NMDA receptors elicits fictive locomotion and bistable membrane properties in the lamprey spinal cord. *Brain Res.* 336, 390–395. doi: 10.1016/0006-8993(85)90676-6
- SŁawinska, U., Rossignol, S., Bennett, D. J., Schmidt, B. J., Frigon, A., Fouad, K., et al. (2012). Comment on “Restoring voluntary control of locomotion after paralyzing spinal cord injury”. *Science* 338:328. doi: 10.1126/science.1226082
- Staerman, F., Bryckaert, P. E., Youinou, Y., Colin, J., Brandt, B., and Lardennois, B. (2001). Pharmacologic stimulation of ejaculation with midodrine hydrochloride (Gutron) for medically assisted reproduction in spinal injury. *Prog. Urol.* 11, 1264–1268.
- Stafford, S. A., Bowery, N. G., Tang, K., and Coote, J. H. (2006). Activation by p-chloroamphetamine of the spinal ejaculatory pattern generator in anaesthetized male rats. *Neuroscience* 140, 1031–1040. doi: 10.1016/j.neuroscience.2006.02.039
- Stein, R. B., and Capaday, C. (1988). The modulation of human reflexes during functional motor tasks. *Trends Neurosci.* 11, 328–332. doi: 10.1016/0166-2236(88)90097-5
- Stuart, D. G., and Hultborn, H. (2008). Thomas Graham Brown (1882–1965), Anders Lundberg (1920–) and the neural control of stepping. *Brain Res. Rev.* 59, 74–95. doi: 10.1016/j.brainresrev.2008.06.001
- Sugaya, K., Nishijima, S., Miyazato, M., and Ogawa, Y. (2005). Central nervous control of micturition and urine storage. *J. Smooth Muscle Res.* 41, 117–132. doi: 10.1540/jsmr.41.117
- Sugaya, K., Roppolo, J. R., Yoshimura, N., Card, J. P., and de Groat, W. C. (1997). The central neural pathways involved in micturition in the neonatal rat as revealed by the injection of pseudorabies virus into the urinary bladder. *Neurosci. Lett.* 223, 197–200. doi: 10.1016/s0304-3940(97)13433-4
- Székely, G., Czéh, G., and Voros, G. (1969). The activity pattern of limb muscles in freely moving normal and deafferented newts. *Exp. Brain Res.* 9, 53–72. doi: 10.1007/bf00235451
- Tan, U. (2006a). A new syndrome with quadrupedal gait, primitive speech, and severe mental retardation as a live model for human evolution. *Int. J. Neurosci.* 116, 361–369. doi: 10.1080/00207450500455330
- Tan, U. (2006b). Evidence for “Uner Tan Syndrome” and the evolution of the human mind. *Int. J. Neurosci.* 116, 763–774. doi: 10.1080/00207450600588733
- Tan, U. (2008). Uner Tan syndrome: review and report of four new cases. *Int. J. Neurosci.* 118, 211–225. doi: 10.1080/00207450701667808
- Thorpe, A. J., Clair, A., Hochman, S., and Clemens, S. (2011). Possible sites of therapeutic action in restless legs syndrome: focus on dopamine and $\alpha 2\delta$ ligands. *Eur. Neurol.* 66, 18–29. doi: 10.1159/000328431
- Tillakaratne, N. J., de Leon, R. D., Hoang, T. X., Roy, R. R., Edgerton, V. R., and Tobin, A. J. (2002). Use-dependent modulation of inhibitory capacity in the feline lumbar spinal cord. *J. Neurosci.* 22, 3130–3143.
- Tresch, M. C., Saltiel, P., and Bizzi, E. (1999). The construction of movement by the spinal cord. *Nat. Neurosci.* 2, 162–167. doi: 10.1038/5721
- Truitt, W. A., and Coolen, L. M. (2002). Identification of a potential ejaculation generator in the spinal cord. *Science* 297, 1566–1569. doi: 10.1126/science.1073885
- Ung, R. V., Landry, E. S., Rouleau, P., Lapointe, N. P., Rouillard, C., and Guertin, P. A. (2008). Role of spinal 5-HT2 receptor subtypes in quipazine-induced hindlimb movements after a low-thoracic spinal cord transection. *Eur. J. Neurosci.* 28, 2231–2242. doi: 10.1111/j.1460-9568.2008.06508.x
- Ung, R. V., Lapointe, N. P., Tremblay, C., Larouche, A., and Guertin, P. A. (2007). Spontaneous recovery of hindlimb movement in completely spinal cord transected mice: a comparison of assessment methods and conditions. *Spinal Cord* 45, 367–379.
- van den Brand, R., Heutschi, J., Barraud, Q., DiGiovanna, J., Bartholdi, K., Huerlimann, M., et al. (2012). Restoring voluntary control of locomotion after paralyzing spinal cord injury. *Science* 336, 1182–1185. doi: 10.1126/science.1217416
- Vargas, V. M., Torres, D., Corona, F., Vergara, M., Gomez, L. E., Delgado-Lezama, R., et al. (2004). Cholinergic facilitation of erection and ejaculation in spinal cord-transected rats. *Int. J. Impot Res.* 16, 86–90. doi: 10.1038/sj.ijir.3901169
- Viala, D., and Buser, P. (1969). The effects of DOPA and 5-HTP on rhythmic efferent discharges in hind limb nerves in the rabbit. *Brain Res.* 12, 437–443. doi: 10.1016/0006-8993(69)90011-0
- Vizzard, M. A., Erickson, V. L., Card, J. P., Roppolo, J. R., and de Groat, W. C. (1995). Transneuronal labeling of neurons in the adult rat brainstem and spinal cord after injection of pseudorabies virus into the urethra. *J. Comp. Neurol.* 355, 629–640. doi: 10.1002/cne.903550411
- Wainberg, M., Barbeau, H., and Gauthier, S. (1990). The effects of cyproheptadine on locomotion and on spasticity in patients with spinal cord injuries. *J. Neurol. Neurosurg. Psychiatry* 53, 754–763. doi: 10.1136/jnnp.53.9.754
- Wang, D., Grillner, S., and Wallén, P. (2006). Effects of flufenamic acid on fictive locomotion, plateau potentials, calcium channels and NMDA receptors in the lamprey spinal cord. *Neuropharmacology* 51, 1038–1046. doi: 10.1016/j.neuropharm.2006.06.012
- Wilson, J. M., Cowan, A. I., and Brownstone, R. M. (2007). Heterogeneous electronic coupling and synchronization of rhythmic bursting activity in mouse HB9 interneurons. *J. Neurophysiol.* 98, 2370–2381. doi: 10.1152/jn.00338.2007
- Wilson, J. M., Hartley, R., Maxwell, D. J., Todd, A. J., Lieberam, I., Kaltschmidt, J. A., et al. (2005). Conditional rhythmicity of ventral spinal interneurons defined by expression of the Hb9 homeodomain protein. *J. Neurosci.* 25, 5710–5719. doi: 10.1523/jneurosci.0274-05.2005
- Yakovlev, A. G., and Faden, A. I. (1994). Sequential expression of c-fos protooncogene, TNF-alpha and dynorphin genes in spinal cord following experimental traumatic injury. *Mol. Chem. Neurobiol.* 23, 179–190. doi: 10.1007/BF02815410
- Yonezawa, A., Yoshizumi, M., Ebiko, M., Ise, S. N., Watanabe, C., Mizoguchi, H., et al. (2008). Ejaculatory response induced by a 5-HT2 receptor agonist m-CPP in rats: differential roles of 5-HT2 receptor subtypes. *Pharmacol. Biochem. Behav.* 88, 367–373. doi: 10.1016/j.pbb.2007.09.009
- Yoshiyama, M., and de Groat, W. C. (2005). Supraspinal and spinal α -amino-3-hydroxy-5-methylisoxazole-4-propionic acid and N-methyl-D-aspartate glutamatergic control of the micturition reflex in the urethane anesthetized rat. *Neuroscience* 132, 1017–1026. doi: 10.1016/j.neuroscience.2005.01.041
- Zhang, S. X., Huang, F., Gates, M., White, J., and Holmberg, E. G. (2010). Tail nerve electrical stimulation induces body weight-supported stepping in rats with spinal cord injury. *J. Neurosci. Methods* 187, 183–189. doi: 10.1016/j.jneumeth.2010.01.008
- Zhang, Y., Narayan, S., Geiman, E., Lanuza, G. M., Velasquez, T., Shanks, B., et al. (2008). V3 spinal neurons establish a robust and balanced locomotor rhythm during walking. *Neuron* 60, 84–96. doi: 10.1016/j.neuron.2008.09.027

Conflict of Interest Statement: Pierre A. Guertin is professor in neurosciences at Laval University as well as CEO of a biopharmaceutical company, Nordic Life Science Pipeline Inc.

Received: 20 December 2013; paper pending published: 07 February 2014; accepted: 11 April 2014; published online: 30 May 2014.

Citation: Guertin PA (2014) Preclinical evidence supporting the clinical development of central pattern generator-modulating therapies for chronic spinal cord-injured patients. *Front. Hum. Neurosci.* 8:272. doi: 10.3389/fnhum.2014.00272

This article was submitted to the journal *Frontiers in Human Neuroscience*.

Copyright © 2014 Guertin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.