



Measurement of Antioxidant Activity toward Superoxide in Natural Waters

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Antioxidants are a class of molecules that provide a protective function against reactive oxygen species (ROS) in biological systems by out competing physiologically important molecules for ROS oxidation. In natural waters, the reactivity of antioxidants gives an estimate of oxidative stress, and may determine the reactivity and distribution of reactive oxidants. We present an analytical method to measure antioxidant activity in natural waters through the competition between ascorbic acid, an antioxidant, and MCLA, a chemiluminescent probe for superoxide. A numerical kinetic model of the analytical method has been developed to optimize analytical performance. Measurements of antioxidant concentrations in pure and seawater are possible with detection limits below 0.1 nM. Surface seawater samples collected at solar noon contained over 0.4 nM of antioxidants and exhibited first-order decay with a half-life of 3–7 min, consistent with a reactive species capable of scavenging photochemically produced superoxide.

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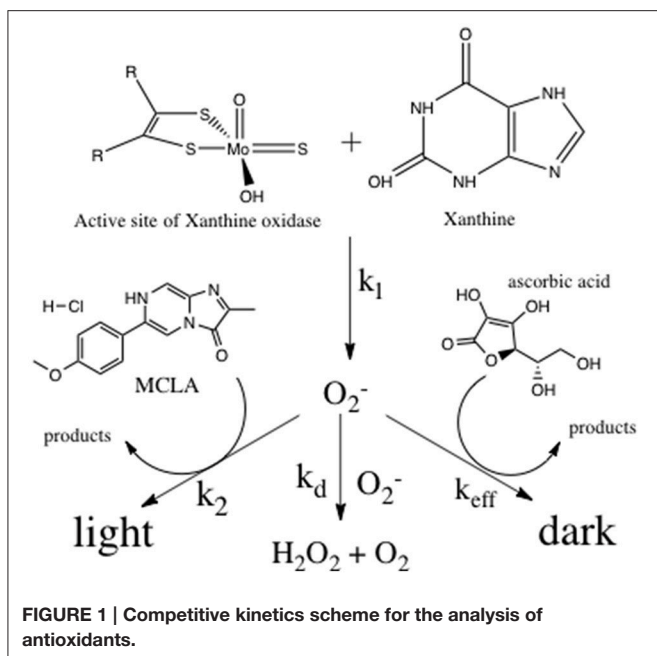
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INTRODUCTION

Reactive oxygen species (ROS) are short lived, oxygen containing molecules including singlet oxygen ($^1\text{O}_2$), superoxide (O_2^-), hydrogen peroxide (H_2O_2), hydroxyl radical ($\bullet\text{OH}$), and the carbonate radical ($\bullet\text{CO}_3^-$). In natural systems, these species are capable of oxidizing a range of molecules including metal complexes, dissolved organic matter, and biomolecules. The reactivity of ROS species has prompted extensive analytical work in ROS analysis (Burns et al., 2012). Antioxidants are a class of molecules that provide a protective function against ROS species in biological systems by out competing physiologically important molecules for ROS oxidation (Schlesier et al., 2002). In natural waters the reactivity of antioxidants gives an estimate of oxidative stress and will determine the reactivity and distribution of ROS (Goldstone and Voelker, 2000; Heller and Croot, 2010; Roe et al., 2016). Methods for quantifying antioxidants include direct analysis using chromatography with UV-VIS (Cai et al., 2003) or MS detection (Lee et al., 2006) and the colorimetric analysis of derivatized analytes (Prior et al., 2005). Direct analysis is desirable, but limited in environmental systems due to poor detection limits and the unknown chemical structure of many natural antioxidants. Indirect antioxidant analysis relies on competitive kinetic reactions between antioxidants and known probe molecules with ROS—often superoxide or hydroxyl radicals (Hosaka et al., 2005; Besco et al., 2007; Giokas et al., 2007; Yu et al., 2008; Anifowose et al., 2015). Indirect measurements do not necessarily measure specific molecule concentrations, but rather weighted antioxidant reactivity defined as the sum of all antioxidant concentrations scaled by their reaction rate constants with specific ROS. In this indirect, competitive kinetics method, experimentally generated superoxide competes with the antioxidants of interest, and an analytically observable probe molecule (**Figure 1**).



Superoxide was produced at a constant, known rate through the enzymatically catalyzed reaction of xanthine with xanthine oxidase (X/XO; Fridovich, 1970; Rose et al., 2008a). Superoxide decay will occur via three bimolecular pathways; reaction with the chemiluminescent probe molecule MCLA (Burns et al., 2012), disproportionation with another superoxide molecule (Bielski, 1978), or reaction with ascorbic acid (Nandi and Chatterjee, 1987)—the reference antioxidant. Antioxidant reactivity was determined by measuring the quenching of the superoxide-MCLA reaction with standard or unknown antioxidants. This method will only detect chemical species that react with superoxide at rates equal or greater than superoxide disproportionation (i.e., the dark pathway must be at least 1% of the superoxide sink). The method does not measure a specific chemical species, but rather the reaction rate-weighted sum of all antioxidants. In this way, the method is a measure of antioxidants capable of perturbing the steady state concentration of superoxide. The method will be insensitive to antioxidants that react with hydrogen peroxide or molecular oxygen since these ROS species do not react with the CL probe MCLA, and these reaction rates are much slower than the superoxide reactions shown in **Figure 1**.

There are four reactions describing the change in superoxide concentration in this indirect analytical method. First, the production rate of superoxide is given by

$$\frac{d[O_2^-]}{dt} = k_1 \quad (1)$$

where k_1 is the enzyme limited rate of reaction between xanthine and xanthine oxidase to produce superoxide (Rose et al., 2008a). Second, the loss of superoxide through the reaction with MCLA is given by

$$\frac{d[O_2^-]}{dt} = -k_2 [O_2^-] [MCLA] = \text{Chemiluminescence Flux} \quad (2)$$

where k_2 is the pH dependent rate constant for the reaction between superoxide and MCLA (Goto and Takagi, 1980; Fujimori et al., 1993). This reaction also describes the rate of light production from the chemiluminescence reaction, CL. Third, the loss of superoxide via disproportionation is

$$\frac{d[O_2^-]}{dt} = -k_d [O_2^-]^2 \quad (3)$$

where k_d is the pH dependent rate constant for the disproportionation reaction and $[O_2^-]$ represents the total $[O_2^-]$ and $[HO_2]$ in solution (Bielski, 1978; Burns et al., 2012). Finally, the loss of superoxide through the reaction with antioxidants (AO) is

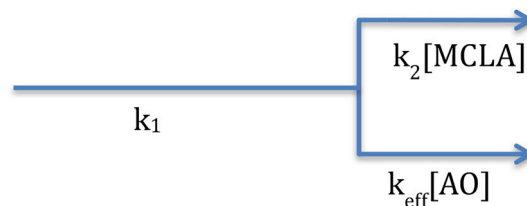
$$\frac{d[O_2^-]}{dt} = -k_{eff} [O_2^-] [AO] \quad (4)$$

where k_{eff} is the effective rate constant for the reaction between all antioxidants and superoxide (Bielski et al., 1985; Nandi and Chatterjee, 1987). The rate of change of superoxide concentration as a function of time is defined by Equation (5).

$$\frac{d[O_2^-]}{dt} = k_1 - k_d [O_2^-]^2 - k_2 [O_2^-] [MCLA] - k_{eff} [O_2^-] [AO] \quad (5)$$

Under steady state conditions ($d[O_2^-]/dt = 0$) with slow superoxide disproportionation, we can simplify Equations (5) to

$$k_1 = k_2 [O_2^-] [MCLA] + k_{eff} [O_2^-] [AO] \quad (6)$$



with the relative rate of each superoxide loss pathway independent of superoxide (Equation 7). Since only the superoxide-MCLA reaction, top path, will result in chemiluminescence, the fraction of unquenched CL signal can be computed from mass balance.

$$\begin{aligned} \text{Fraction of CL signal} &= \frac{k_2 [MCLA] [O_2^-]}{k_2 [MCLA] [O_2^-] + k_{eff} [AO] [O_2^-]} \\ &= \frac{k_2 [MCLA]}{k_2 [MCLA] + k_{eff} [AO]} \quad (7) \end{aligned}$$

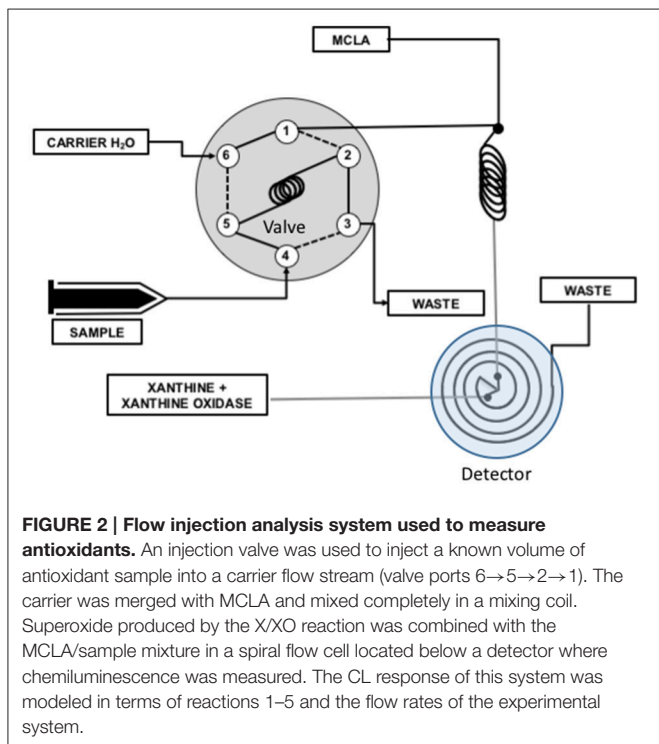
The change in CL signal as a function of antioxidant concentration is the basis of this analytical method.

Modeled CL Reaction

We used National Instruments LabView to numerically model reactions 1–5 and evaluate the steady state assumptions that are the basis for Equation (7). The physical system is shown in **Figure 2**.

The reaction was modeled using initial concentrations of MCLA, antioxidant, and XO. It was assumed that the xanthine substrate saturated the XO active site and superoxide production was only a function of XO activity, $0.25 \text{ nM s}^{-1} \text{ unit}^{-1}$ (Rose et al., 2008a). Reaction rates were corrected to pH 6 with $k_d = 5.4 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ (Bielski et al., 1985), $k_2 = 6.2 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ (Fujimori et al., 1993), and k_{eff} at $2.7 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ (Bielski et al., 1985) to $8.2 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ (Nandi and Chatterjee, 1987). Reaction progress was computed using a simple finite difference calculation with a simulation time step of 10 ms. The valve timing was simulated by turning on the antioxidant concentration 10 s into the simulation. Time was used to simulate the flow of reactants in the analytical system, **Figure 2**, and the CL flux was modeled as a function of flow cell position since only light produced in the flow cell will be detected by the PMT. **Figure 3** shows a typical simulation run with 0, 100, and 1000 nM of antioxidant.

At low initial steady state levels of superoxide, the MCLA (1 μM) and AO concentrations are essentially constant over the entire analytical run. Therefore, the CL signal is a linear function of superoxide concentration and the curves in **Figure 3B** are proportional to PMT counts. When AO concentrations increase, superoxide concentrations decrease due to Equation (4) and we observe a decrease in the CL signal (**Figure 3A**). It is recognized that the maximum CL signal is observed when $[\text{AO}] = \text{zero}$



and the minimum CL signal is observed when $[\text{AO}] \gg [\text{MCLA}]$. Rewriting Equation (7) in terms of the decrease in CL intensity, S (**Figure 3A**), we obtain

$$\begin{aligned} \text{Fraction of CL signal (F)} &= \frac{k_2 [\text{MCLA}]}{k_2 [\text{MCLA}] + k_{eff} [\text{AO}]} \\ &= \{S - S_{\max}\} / \{S_{\min} - S_{\max}\} \quad (8) \end{aligned}$$

where S_{\max} is the maximum decrease in CL signal and S_{\min} is the decrease in signal for a blank (typically zero). When simulations were run with varying AO concentrations a calibration curve was generated and depicted in **Figure 4**.

Notice that the fit to the calculated signal is not as perfect as expected for a numerical fit to simulated data. The discrepancy is due to limitations in the steady state assumptions used to derive Equation (7), and the real detection window that integrates a changing CL signal as analytical solution mixes and flows outward in the detector spiral. Nevertheless, Equation (8) provides a remarkably good description of CL quenching as a function of AO concentration, which will be shown for pure water and natural water systems below.

Calibration of this method requires measuring CL quenching, S , for a range of AO concentrations. Quenching measurements

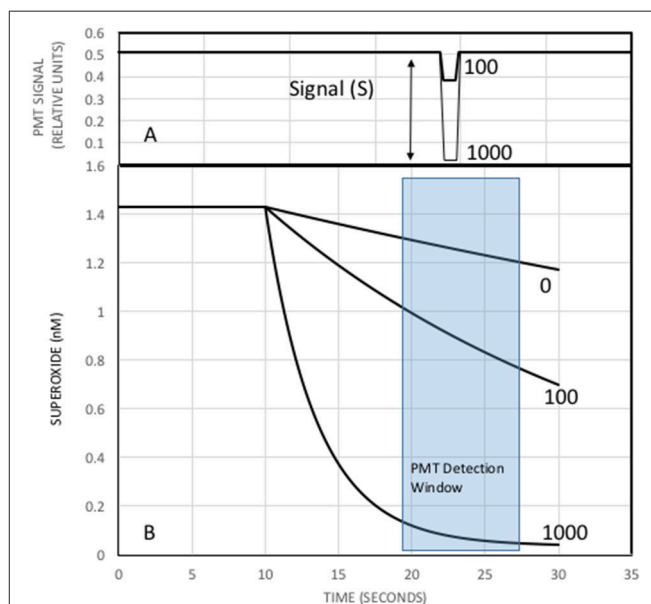


FIGURE 3 | Simulation of the analytical system for three different antioxidant concentrations. (B) Superoxide from the X/XO reaction starts at a steady state of 1.4 nM. The MCLA-antioxidant solution is mixed with superoxide at time 10 s causing a decrease in superoxide concentrations and production of CL. Numbers next to each curve are $[\text{AO}]$ in nM. Less superoxide will produce less light. **(A)** CL signal as a function of time. The zero AO addition would produce a flat line signal and is not shown. Since the sample loop is a finite size, CL signals begin high (carrier only), decrease when the sample passes through the detector, and returns to initial values as the sample bolus exits to waste. The PMT detection window (blue box) in **(B)** will change the CL response in **(A)**. The optimal detection window will produce the largest differences in the integrated superoxide concentrations and thus the biggest changes in CL.

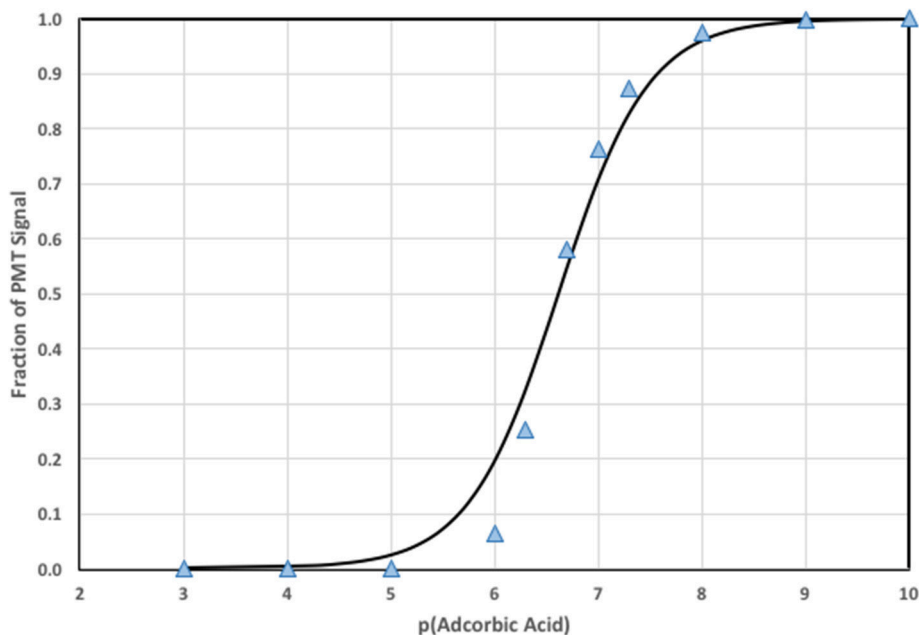


FIGURE 4 | Calibration curve using the numerical model to estimate S and F (points) and the fit to the data using Equation (7) (line).

at $[AO] = 0$ and $[AO] \gg [MCLA]$ yields values for S_{\min} and S_{\max} respectively. F -values are computed using Equation (8) and a calibration curve is generated using Equation (7) with k_{eff} as the only fitting parameter. F -values for unknown samples yield effective AO concentrations through reorganization of Equation (7).

$$\frac{k_2[MCLA] - k_2[MCLA]}{k_{eff}} = [AO] \quad (9)$$

Ascorbic acid was selected as a model antioxidant because it is a commercially available solid that serves as a useful analytical reference standard. Ascorbic acid is not expected to be present in natural waters. Natural antioxidants will have faster and slower k_{eff} compared to ascorbic acid. Natural antioxidants with slow reaction rates for superoxide will have lower effective concentrations, while natural antioxidants with faster reaction rates for superoxide will appear to have higher effective concentrations. Since the goal of this work was to determine the ability of natural waters to respond to rapid oxidative stress, using a reactivity scaled measure of antioxidant concentration allows intercomparison of data from different water types where the chemical identity of specific antioxidants is currently not known.

METHODS

The analytical system used was built by Waterville Analytical and is shown in **Figure 2**. Flow rates in the analytical lines were 1 ml/min. The PMT was operated at the default high

voltage with an integration time of 800 ms. Sample load time was 125 s and analysis time was 60 s with an acquisition frequency of 1 Hz.

All solutions were prepared in high purity water. Diethylenetriaminepentaacetic acid (DTPA), xanthine, xanthine oxidase from bovine milk, disodium phosphate, sodium bicarbonate, and ascorbic acid were reagent grade and obtained from Sigma-Aldrich. 2-methyl-6-[p-methoxyphenyl]-3,7-dihydroimidazo[1,2-a]pyrazin-3-one (MCLA) was obtained from Thermo Fisher. Five milliliter vials of 0.4 mM MCLA solution were prepared by adding 15 mg of solid MCLA to 130 mL of high purity water. The MCLA vials were stored at -80°C until used to prepare fresh solutions.

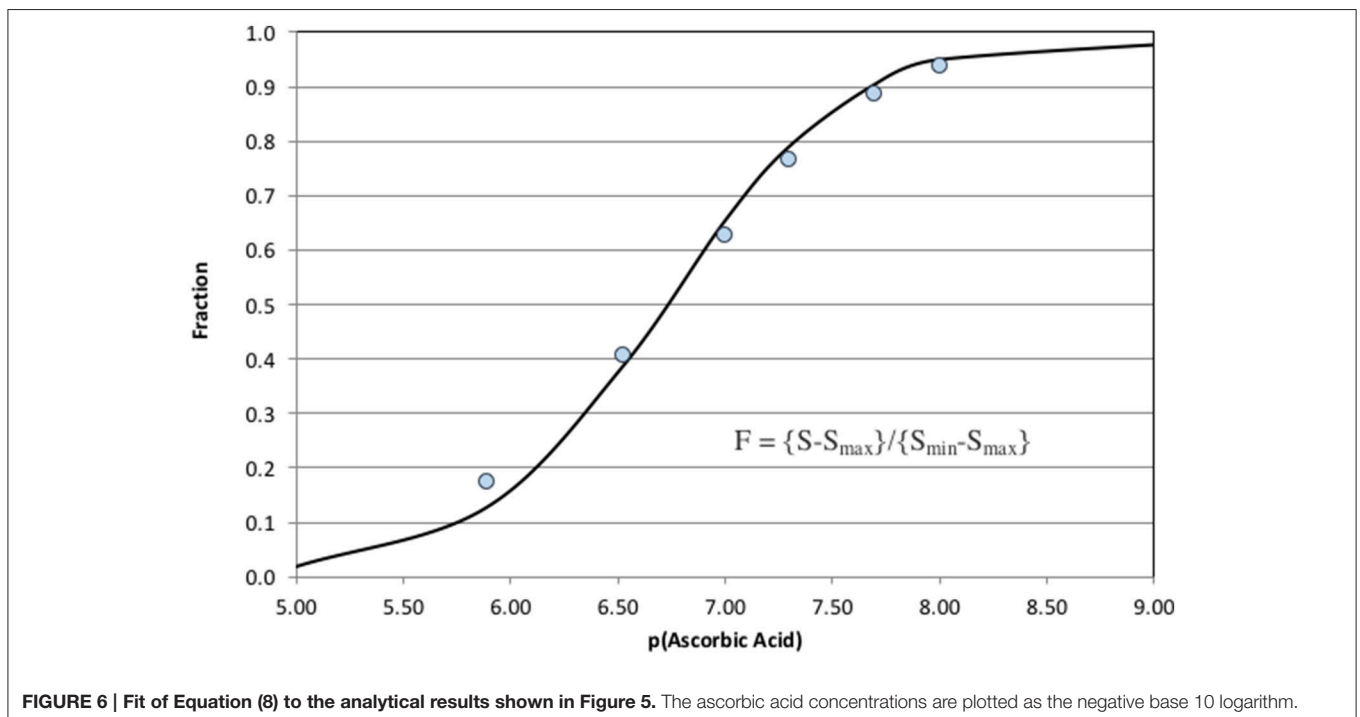
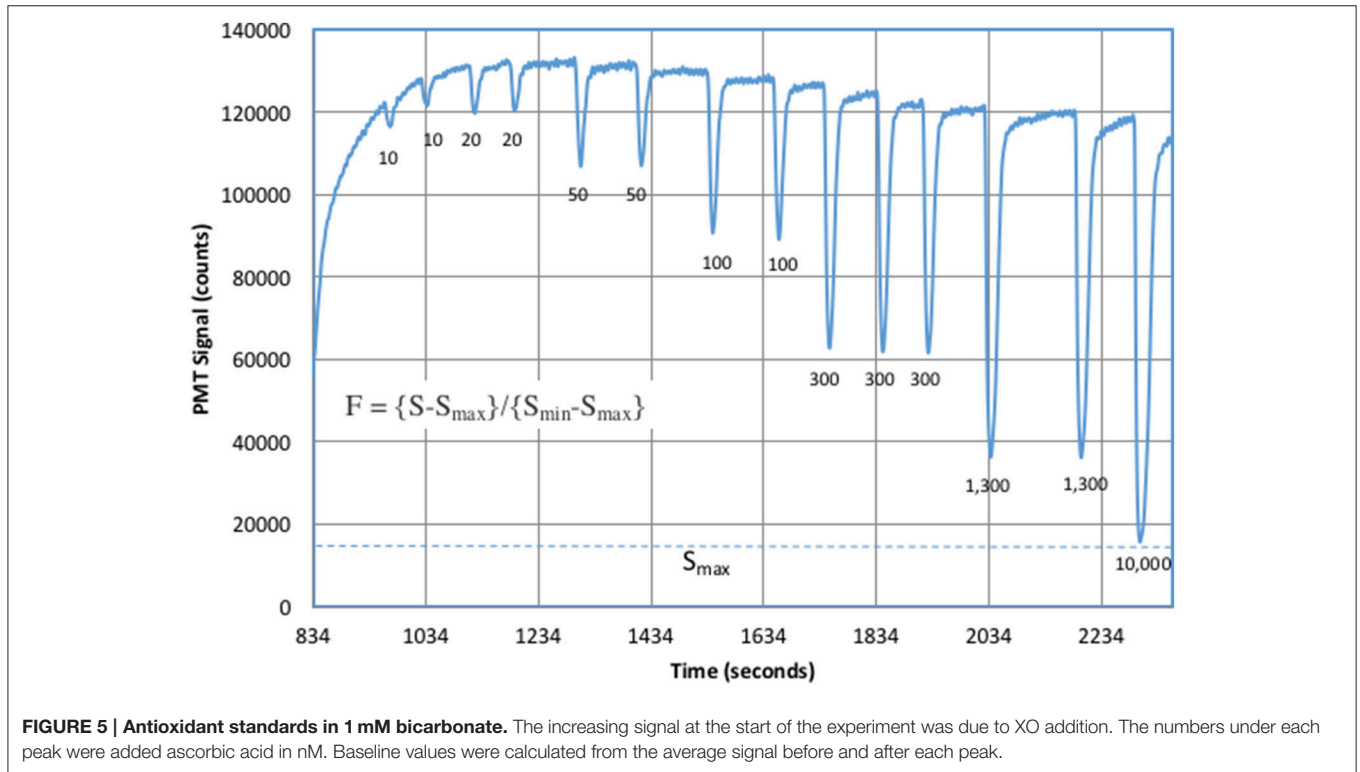
MCLA solutions (1–4 μM) in 0.1 M phosphate buffer were made by adding MCLA to pH 6, 0.01 M phosphate buffer containing 1 μM DTPA. The xanthine solution was 1 μM in pH 6, 0.001 M phosphate buffer containing 1 μM DTPA. XO was added directly to the xanthine solution at 5 μL increments until nM levels of antioxidant resulted in a 30% quenching of the CL signal. Typical XO activities were 0.01 nM/s. Alternatively, XO solutions were prepared in 0.001 M phosphate buffer containing 1 μM DTPA and merged with xanthine in a reaction spiral for 1 min. Higher XO activities were used in this alternative configuration due to shorter reaction times. Separating the X and XO reagents results in improved baseline stability. The carrier for laboratory experiments was pH 7, 1 mM sodium bicarbonate. The carrier for ocean samples was oligotrophic seawater collected at 400 m and stored in the dark at room temperature.

Oligotrophic seawater samples were collected January 2011 during the Great Belt cruise in the South Atlantic (latitude

W 47.976 and longitude S 39.250) from a flowing stream of surface seawater provided from the sea chest of the ship. Residence time from the surface ocean to a sample bottle was <1 min.

RESULTS AND DISCUSSION

Figure 5 shows typical laboratory results for ascorbic acid standards added to 1 mM bicarbonate solutions with 1 μ M



DTPA. DTPA is a good ligand for metals and significantly reduces metal-superoxide reaction rates (Rose et al., 2008b). CL signals were significantly higher in the presence of DTPA, which suggested that metal contamination in our reagents provided an alternative sink for superoxide.

Increasing ascorbic acid additions cause a significant increase in CL quenching. The signal, S , was defined as the change in CL from the upper baseline to the bottom of each negative peak. Peak height was used instead of peak area because the analytical response was non-linear. The instrumental response

half way down a peak shoulder does not correspond to half the sample concentration. The result of a simple integration was a complicated integral of an exponentially changing response over time. Using peak height avoids awkward integration problems. **Figure 6** shows the fit to the computed F -values (Equation 8) for each sample.

The analytical response was exponential so a direct estimation of analytical detection limits is difficult. The system was clearly capable of detecting of 10 nM of ascorbic acid, AA, in 1 mM bicarbonate. Maximum sensitivity occurred when $F = 0.5$ or

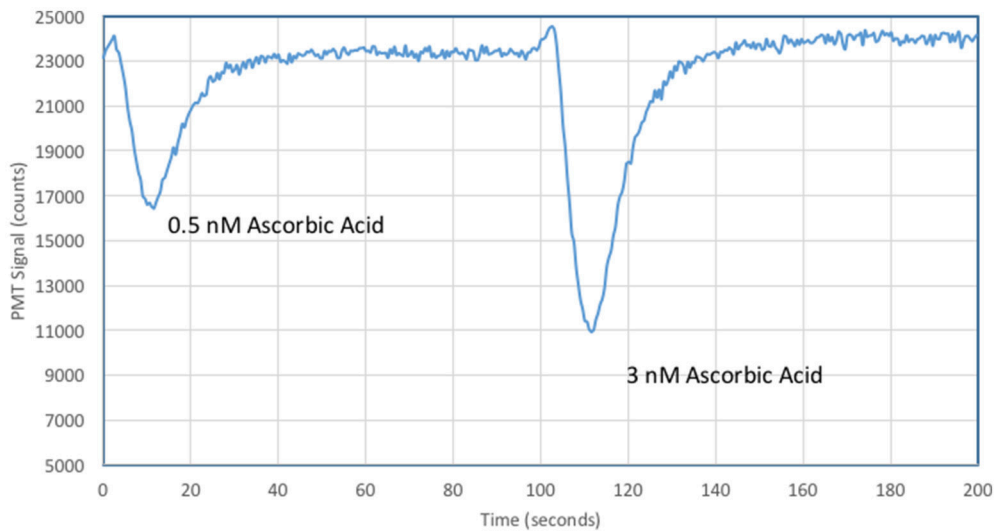


FIGURE 7 | Typical analytical results for ascorbic acid additions to oligotrophic seawater.

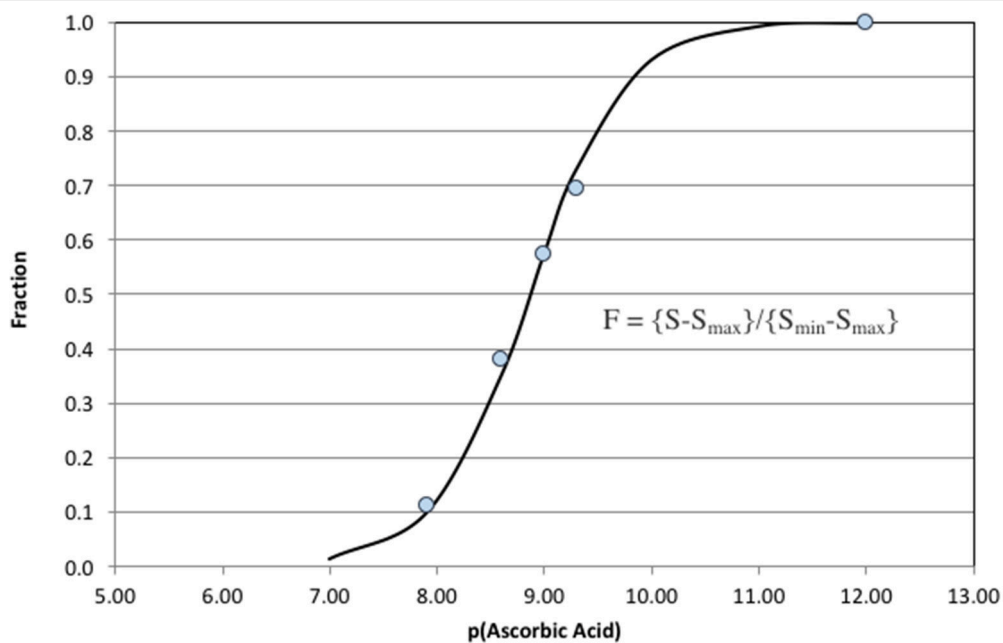


FIGURE 8 | Ascorbic acid additions to oligotrophic seawater. The analytical window in these clean seawater samples was 100–0.01 nM.

$dF/d(pAA)$ was at a maximum. Good quantification of AO is possible at $p[AO] \pm$ two units of $F = 0.5$. This quantification range can be considered the analytical window of the method. It was possible to shift the analytical window to higher or lower concentrations of AO (shifting the point where $F = 0.5$) by decreasing or increasing the MCLA and XO concentrations (Equation 8). The numerical model was very helpful in defining initial analytical conditions for a specific analytical window that were then optimized by the analyst.

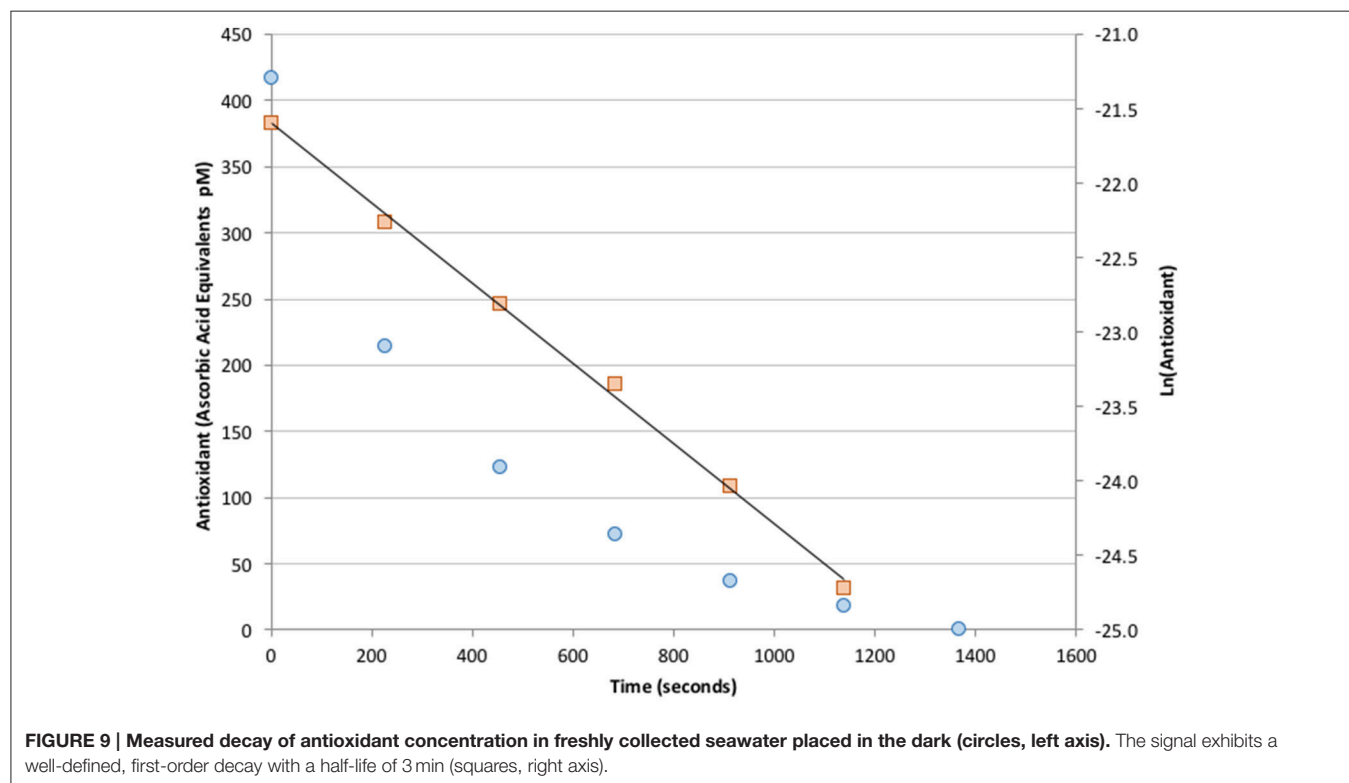
Figure 7 shows the instrumental response for ascorbic acid additions to oligotrophic seawater. Five-fold lower concentrations of MCLA and XO allowed a significant improvement in analytical sensitivity with a 0.5 nM ascorbic acid standard quenching 30% of the CL signal.

The analytical response over a range of ascorbic acid concentrations is shown in Figure 8. The analytical window was $10^{-7} \text{ M} > [AO] > 10^{-11} \text{ M}$. The point at $[AO] = 10^{-12} \text{ M}$ represents the pure carrier solution (deep seawater) and is included in the fit to define the zero quenching condition. This calibration curve was used to measure antioxidant activity in oligotrophic seawater. The analysis does not imply that ascorbic acid is in seawater, but rather a natural antioxidant exists with the same effective reactivity (k_{eff} [ascorbic acid]).

Seawater samples collected at depths below 25 m showed no AO activity above the analytical blank. Additions of Fe(II) at nM levels also did not produce a measurable response (data not shown). However, fresh samples from the ship's uncontaminated seawater system consistently contained detectable levels of antioxidants at $\sim 100\text{--}400 \text{ pM}$ in ascorbic acid units. Antioxidant

activity was only observed during daylight and decayed with a half-life of 3–7 min after being placed in the dark. A typical antioxidant decay curve is shown in Figure 9. The measured antioxidant decay of a few minutes was consistent with non-detectable antioxidant activity in deep samples because sample collection from depth took at least 15 min. It is therefore an open question of whether antioxidant species capable of reacting with superoxide exist below the surface euphotic zone.

The identity of the antioxidant species in the seawater samples is also an open question. If it is assumed that observed antioxidant activity is due to molecules with a reactivity similar to ascorbic acid, then the calculated pseudo-first order rate for superoxide reduction is on the order of 10^{-2} s^{-1} [$0.4 \text{ nM} * k_{eff}$ where $k_{eff} = 8.2 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ (Nandi and Chatterjee, 1987)]. Goldstone and Voelker (2000) have shown that copper complexes are capable of fast bimolecular ($10^9 \text{ M}^{-1} \text{ s}^{-1}$) reactions with superoxide in seawater. Rose et al. (2008b) observed pseudo-first order decay of natural superoxide with a rate of 10^{-3} s^{-1} in samples collected off the Costa Rica Dome. Heller and Croot (2010) have measured the pseudo-first order decay rates for 10 nM superoxide additions to Cape Verde seawater on the order of 10^{-2} s^{-1} , and suggested that this rate could be attributed to the catalytic reaction with quinones or other reactive organic species. However, in these measurements the rates of superoxide reduction were remarkably constant, suggesting catalysis, or a reductant with concentrations well above ambient superoxide to produce pseudo-first order decay. It is possible that photochemical activation of quinone-type species could produce the effective antioxidant activity described in this work. However,



the different pH and lower steady state superoxide concentrations used in this work make direct comparisons to these previously published superoxide decay rates in pH 8 seawater difficult. Future work will probe the antioxidant activity in the surface waters of lakes and the coastal ocean to determine the range of antioxidant potential as a function of sunlight intensity, dissolved organic carbon, and salinity.

AUTHOR CONTRIBUTIONS

DK: Primary Author, field measurements. EB: Significant contributions to the kinetic model, and AO literature. ZH:

Preliminary investigation of the method. EI: Laboratory calibration of AO method and background literature reviews. KM: Significant collaborator during initial method development and insights on the role of pollutants as stressors to generate antioxidant response.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

DK is the owner of Waterville Analytical, an instrument company based in Waterville Maine, that makes the apparatus used in this work.

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