

# The effects of psychostimulant drugs on blood brain barrier function and neuroinflammation

Sharanya M. Kousik<sup>1,2\*</sup>, T. Celeste Napier<sup>1,2</sup> and Paul M. Carvey<sup>1,2,3</sup>

<sup>1</sup> Department of Pharmacology, Rush University Medical Center, Chicago, IL, USA

<sup>2</sup> Center for Compulsive Behavior and Addiction, Rush University Medical Center, Chicago, IL, USA

<sup>3</sup> Department of Neurological Sciences, Rush University Medical Center, Chicago, IL, USA

## Edited by:

Joana A. Palha, University of Minho, Portugal

## Reviewed by:

Raja S. Settivari, The Dow Chemical Company, USA

James Haorah, University of Nebraska Medical Center, USA

## \*Correspondence:

Sharanya M. Kousik, Department of Pharmacology and the Center for Compulsive Behaviors and Addiction, Rush University, Robert H. and Terri Cohn Research Building, 1735 West Harrison Street, Chicago, IL 60612, USA.

e-mail: [sharanya\\_m\\_kousik@rush.edu](mailto:sharanya_m_kousik@rush.edu)

The blood brain barrier (BBB) is a highly dynamic interface between the central nervous system (CNS) and periphery. The BBB is comprised of a number of components and is part of the larger neuro(glio)vascular unit. Current literature suggests that psychostimulant drugs of abuse alter the function of the BBB which likely contributes to the neurotoxicities associated with these drugs. In both preclinical and clinical studies, psychostimulants including methamphetamine, MDMA, cocaine, and nicotine, produce BBB dysfunction through alterations in tight junction protein expression and conformation, increased glial activation, increased enzyme activation related to BBB cytoskeleton remodeling, and induction of neuroinflammatory pathways. These detrimental changes lead to increased permeability of the BBB and subsequent vulnerability of the brain to peripheral toxins. In fact, abuse of these psychostimulants, notably methamphetamine and cocaine, has been shown to increase the invasion of peripheral bacteria and viruses into the brain. Much work in this field has focused on the co-morbidity of psychostimulant abuse and human immunodeficiency virus (HIV) infection. As psychostimulants alter BBB permeability, it is likely that this BBB dysfunction results in increased penetration of the HIV virus into the brain thus increasing the risk of and severity of neuro AIDS. This review will provide an overview of the specific changes in components within the BBB associated with psychostimulant abuse as well as the implications of these changes in exacerbating the neuropathology associated with psychostimulant drugs and HIV co-morbidity.

**Keywords:** blood brain barrier, cocaine, HIV, mdma, methamphetamine, neuroinflammation, nicotine

## INTRODUCTION

The blood brain barrier (BBB) is a specialized structure formed by brain endothelial cells (BECs) that are tightly interconnected to form a boundary between the central nervous system (CNS) and periphery (Abbott et al., 2010). Once thought to be static, the BBB is now known to be an active and dynamic interface that responds to signals from both the brain parenchyma and vasculature. BBB dysfunction contributes to the pathophysiology of numerous CNS disorders including Alzheimer's disease (AD), Parkinson's disease (PD), multiple sclerosis, ischemia, and neuroAIDS (Stewart et al., 1992; Nath et al., 2001; Samii et al., 2004; Compston and Coles, 2008; Yenari and Han, 2012). Recent evidence points to the ability of drugs of abuse (e.g., stimulants) to disrupt BBB function. This review will provide an overview of normal BBB structure and function, focusing on the regulation of neuroinflammation, and will provide examples of BBB disruption specifically associated with stimulant drug abuse. As neuroAIDS is exacerbated by stimulant abuse, we will also discuss the implications of stimulant-induced BBB dysfunction as it relates to neuroAIDS disease progression.

## COMPONENTS OF THE BBB

The structure of the BBB has been thoroughly described in other reviews (Sandoval and Witt, 2008; Carvey et al., 2009). In brief, the

BBB is comprised of highly specialized BECs which interact with pericytes, the vascular basement membrane, and astrocytes (Diaz-Flores et al., 1991; Krueger and Bechmann, 2010). BECs are closely associated through cell-cell complexes of tight junction proteins: claudins, occludins, and junctional adhesion molecules (JAMs; Hawkins and Davis, 2005). Adherens junctions also form between BECs. Accessory proteins like zona occludins (ZO), cingulin, and afadin (AF-6) provide structural support and stability to both tight and adherens junctions (Yamamoto et al., 1999; Mark and Davis, 2002). Though little is known about pericyte involvement within the BBB, these cells likely regulate cerebral blood flow (Bandyopadhyay et al., 2001) and stabilize forming vessels (Ramsauer et al., 2002). BECs and pericytes are enveloped by extracellular matrix proteins that comprise the basement membrane (Farkas and Luiten, 2001). Astrocytes interact with components of the BBB through their foot processes that make contact with BECs to form the *glia limitans perivascularis* as well as the basement membrane (Krueger and Bechmann, 2010). Astrocytic foot process in the margin of the brain form a thin membrane called the *glia limitans superficialis* in the subarachnoid space. These two *glia limitans* are classically viewed as continuous, starting at the subarachnoid space, and merging with the parenchymal *limitans* as vessels penetrate into brain. Astrocytes facilitate BBB development by inducing tight junction formation, providing microvascular support, and

allowing molecule (e.g., water, sugar, ions) diffusion into the brain (Janzer and Raff, 1987; Madsen and Hirschberg, 2010).

### NEUROVASCULAR COUPLING AT THE BBB INTERFACE

There is a complex relationship between vascular and neuronal systems within the CNS. These interactions are promoted by the BBB, neurons, and glia, which together make up the neuro(glio)vascular unit (Hawkins and Davis, 2005; Koehler et al., 2009). This neurovascular coupling regulates cerebral blood flow enabling efficient oxygen and nutrient supply to various cell types and brain regions (Iadecola, 2004). Moreover, intracellular communication within the neuro(glio)vascular unit allows for localized control of cerebral blood flow to match the needs of specific brain regions. These interactions are important for blood flow to the neurons, as well as for the development, and maintenance of the neurovasculature.

### FUNCTIONS OF THE BBB

The main function of the BBB is to maintain homeostasis of the brain. This barrier protects the CNS from peripheral toxins within the circulation. These toxins may be endogenous factors or exogenous xenobiotics (Abbott et al., 2010). The impenetrability of the BBB results from physical restriction due to tight junction connectivity of BECs, transport regulation limiting transcellular migration, and enzymatic activities of BECs that metabolize harmful substances at the vascular face of the BBB (Madsen and Hirschberg, 2010).

Transport of molecules across the BBB is strictly regulated. A wide range of lipid-soluble molecules passively diffuse through BECs and certain molecules and cells are capable of paracellular migration between adjacent BECs. Most molecules are restricted either by lipid-solubility, polarity, or size, and thus require transporters to cross the BBB. Transporters found within the BBB are numerous, but in general, transporters can be categorized as being ion transporters, active transporters (e.g., *P*-glycoprotein), sodium-dependent transporters, or sodium-independent transporters (Carvey et al., 2009). These transport systems supply nutrients to the brain from the blood and remove metabolic byproducts from the brain to the blood for elimination (Ohtsuki, 2004).

Through neurovascular coupling, the BBB regulates cerebral blood flow. Contractile proteins identified in pericytes are thought to alter vascular diameter and cerebrovascular blood flow (Bandopadhyay et al., 2001). Transport systems bring in vasoactive factors from the periphery that interact with smooth muscle cells and pericytes surrounding the BBB endothelium to allow for vasodilation or constriction (Peppiatt et al., 2006; Bell et al., 2010). Additionally, cells within the neuro(glio)vascular unit can secrete local vasoregulators like norepinephrine, nitric oxide, and endothelin which regulate regional cerebral blood flow (Lecrux and Hamel, 2011).

Inflammatory cells and signaling factors from the periphery into the CNS are trafficked across the BBB. The neuroinflammatory responses produced by these cells and factors can also alter BBB function and stability. While it is yet to be determined whether BBB dysfunction occurs prior to neuroinflammation or *vice versa*, studies have clearly shown that the inflammatory response by resident neuroglia perpetuates BBB dysfunction and this BBB dysfunction increases the neuroinflammatory

response. Under normal physiological conditions, the BBB limits the entry of most leukocytes into the CNS. However, select leukocytes like neutrophils, can cross the BBB during normal immune surveillance with little disruption to BBB tight junctions (Petty and Lo, 2002). During a neuroinflammatory response, immune factors, and endothelial adhesion molecules signal to increase immune cell migration through the BBB (Petty and Lo, 2002). Signals resulting in increased immune cell migration can alter the structural organization of tight junction proteins and lead to actin cytoskeleton remodeling in the BBB basement membrane (Deli et al., 1995; Couraud, 1998; Ransohoff et al., 2003). Migrating peripheral immune cells disrupt normal BBB function, and produce cytokines and chemokines within the CNS which furthers BBB disruption (Cartier et al., 2005). Cytokines [e.g., tumor necrosis factor alpha (TNF $\alpha$ ), interleukin (IL)-1 $\beta$  and IL-6] and the CCL2 chemokine are increased in serum, neuronal tissue, and cerebrospinal fluid in several CNS disorders including traumatic brain injury (Morganti-Kossmann et al., 1997), HIV-associated encephalitis (Cartier et al., 2005), and Huntington disease (Stolp and Dziegielewska, 2009). These neuroinflammatory factors likely contribute to BBB breakdown occurring in these disorders through activity at their receptors on BECs and other BBB cell types (Buckner et al., 2006). Peripheral immune cells can also trigger increased production of inflammatory factors (Verma and Szmítko, 2006; Fletcher et al., 2009), neurotransmitters (e.g., glutamate), neurotrophic factors (e.g., vascular endothelial growth factor), and proteases (e.g., matrix metalloproteinase (MMP-9; Petty and Lo, 2002) from BECs and astrocytes. These factors perpetuate neuroinflammation, and contribute to continuing alterations to the BBB (Petty and Lo, 2002). Alterations in BBB integrity resulting from neuroinflammation produce a cascade resulting in further BBB disruption and increased penetration of immune cells into the CNS. This feed-forward cycle can lead to disruptions in physiological functions of the BBB normally in place to protect against peripheral toxins and xenobiotics, allow for cross-barrier transport, and regulate cerebral blood flow.

Many abused drugs, including stimulants, are pro-inflammatory which leads to BBB disruption and propagates the cyclic relationship between neuroinflammation and BBB damage. This may accelerate the onset of the neurotoxicities associated with chronic use of these potent drugs (Czub et al., 2001; Nath et al., 2002). This review focuses on the effects of psychostimulant drugs though many of the molecular and cellular mechanisms of BBB dysfunction and neuroinflammation have been well-characterized in studies focused on the CNS depressant alcohol (see reviews: de la Monte et al., 2009; Perdisky et al., 2011; Strazza et al., 2011).

## STIMULANT DRUGS AND THE BBB

### METHAMPHETAMINE

Current surveys estimate that approximately 13 million people ages 12 and older have abused methamphetamine (meth) in their lifetimes, making it the second most widely abused illicit drug in the United States after cannabis (NSDUH 2011; United Nations Office on Drugs and Crime 2007). Meth reverses the transport of norepinephrine, dopamine (DA), and serotonin (5-HT) leading to excess release of these monoamines from the cytoplasm and storage vesicles into the synapse (Rothman et al., 2001). Meth

also prevents monoamine reuptake causing them to remain in the synaptic cleft to increase post-synaptic receptor stimulation. Chronic meth abuse in humans is associated with neurotoxicities resulting in damage to both DA and 5-HT terminals in a variety of brain regions (see review: Krasnova and Cadet, 2009). Proposed mechanisms underlying meth-induced neurotoxicity include increased reactive oxygen species (ROS) and dopamine-quinone (DAQ) production (Kuhn et al., 2006), hyperthermia (Kiyatkin et al., 2007), neuroinflammation (Guilarte et al., 2003), and BBB dysfunction (Sharma and Kiyatkin, 2009). It is likely that more than one mechanism underlies the neuronal maladaptations and damage associated with meth abuse, and that this damage is associated with the interaction of multiple mechanisms. For example, oxidative stress and hyperthermia can increase BBB dysfunction which in turn, can exacerbate neuroinflammation after meth exposure. While some studies show direct neurotoxicity (Ricaurte et al., 1980; McCann et al., 2008) after meth exposure without BBB disruption or neuroinflammation, these studies did not assess changes in the BBB or in the inflammatory response so it is uncertain whether neurotoxicity can occur independent of BBB disruption and/or neuroinflammation.

Preclinical studies have reported BBB dysfunction in both whole animal and *in vitro* models. Our laboratory recently showed that a single intraperitoneal injection of either 3 or 9 mg/kg meth in rats resulted in BBB damage in the prefrontal cortex and nucleus accumbens shell, identified by punctate areas of fluorescein isothiocyanate (FITC)-labeled albumin leakage (Kousik et al., 2011). Other rodent studies also showed BBB dysfunction following acute meth treatment marked by leakage of Evans blue into the parenchyma or through increased IgG immunostaining (Bowyer and Ali, 2006; Bowyer et al., 2008; Kiyatkin and Sharma, 2009, 2011; Sharma and Kiyatkin, 2009; Kuroda et al., 2010; Martins et al., 2011). The treatment regimens varied greatly from single doses ranging from 3–40 mg/kg to several acute doses over 24 h, all of which compromised BBB integrity. Alterations to the BBB occurred where meth-induced changes in DA or 5-HT levels and terminal damage were most pronounced, including the striatum, cortex, and hippocampus, but not in areas where monoamine levels were unaffected by meth (e.g., the substantia nigra; Sharma and Ali, 2006; Bowyer et al., 2008; Kousik et al., 2011; Martins et al., 2011). Increased permeability across BECs after meth treatment was also observed *in vitro* through dose-dependent decreases in transendothelial electrical resistance, a method commonly used to measure paracellular permeability (Mahajan et al., 2008; Zhang et al., 2009; Abdul Muneer et al., 2011). Clinical studies report increased levels of peripheral toxins, like HIV and hepatitis C virus, in the brain parenchyma of meth-abusing humans (Nath et al., 2001; Letendre et al., 2005, 2007; Schep et al., 2010). In fact, 70% of individuals undergoing treatment for hepatitis C in one study were self-reported chronic meth abusers (Letendre et al., 2007). These individuals had a higher hepatitis C viral load in the brain than meth-naïve individuals diagnosed with hepatitis C suggesting that meth-induced BBB dysfunction can increase viral penetration into the CNS (Letendre et al., 2007).

Alterations in tight junction complexes can contribute to loss of BBB integrity after meth treatment. Meth treatment decreases the expression of ZO-1, occludin, and claudin-5 in both *in vivo*

and *in vitro* studies (Mahajan et al., 2008; Ramirez et al., 2009; Banerjee et al., 2010; Abdul Muneer et al., 2011; Martins et al., 2011). Additionally, meth can also alter other BBB cell types and interactions. For example, meth increases the expression of peptidases, like MMP-1 and MMP-9, which are involved in degrading certain tight junction proteins to produce structural changes to the BBB basement membrane (Conant et al., 2004). Meth also activates microglia and astrocytes within the neuro(glio)vascular unit which may contribute to meth-induced neurotoxicity, potentially through the secretion of inflammatory cytokines, chemokines, and vasoactive factors (Kiyatkin et al., 2007; Wisor et al., 2011). These structural and functional changes within the BBB/neuro(glio)vascular unit can also result in brain edema and disruption of proper ion flow across the barrier (Sharma and Kiyatkin, 2009).

Neuroinflammation is a proposed mechanism underlying meth-induced neurotoxicity and may also contribute to BBB damage produced by meth. A single 30 mg/kg intraperitoneal injection of meth increases the expression TNF $\alpha$  and IL-6 in the hippocampus, frontal cortex, and striatum in mice (Goncalves et al., 2008). Similar increases in TNF $\alpha$ , IL-1 $\beta$ , and IL-6 are reported after multiple meth treatments and are potentially linked to meth-induced microglial activation (Cadet et al., 1994; Lai et al., 2009; Goncalves et al., 2010). Mahajan et al. (2008) showed that exposure to 50 nM meth produced a 47% increase in the transmigration of peripheral blood mononuclear cells using an *in vitro* BBB model. Meth-induced increases in glial activation as well as TNF $\alpha$  and TNF receptor expression were attenuated with pre-treatment of indomethacin, an anti-inflammatory agent, suggesting a potential therapeutic target for impeding meth-neurotoxicity (Goncalves et al., 2010). In summary, neuroinflammation contributes to the neurotoxicities seen after meth exposure and potentiates the feed-forward cycle between neuroinflammation and BBB damage to produce prolonged BBB damage (see Table 1 for a summary of the effects of meth on BBB function).

**Table 1 | Summary of the preclinical and clinical literature on meth-induced BBB dysfunction.**

<b><i>In vitro</i> BBB model</b>	Decreased transendothelial resistance (TEER) <sup>1,2,3</sup> Increased leakage of FITC-Dextran <sup>2,4</sup> Increased transmigration of PBMCs <sup>1</sup> Increased MMP-1 activation <sup>5</sup>
<b>Rodent models</b>	Increased leakage of FITC-labeled albumin (FITC-LA) and Evans Blue <sup>6,7,8,9</sup> Increased IgG penetration <sup>10,11,12</sup> Increased edema <sup>8,13</sup> Decreased expression of zona occludin 1 (ZO-1), occludin, and claudin-5 <sup>3,4,14</sup>
<b>Clinical studies</b>	Increased penetration of peripheral viruses like HIV and Hepatitis C <sup>15,16</sup>

<sup>1</sup>Mahajan et al. (2008); <sup>2</sup>Zhang et al. (2009); <sup>3</sup>Abdul Muneer et al. (2011); <sup>4</sup>Ramirez et al. (2009); <sup>5</sup>Conant et al. (2004); <sup>6</sup>Kousik et al. (2011); <sup>7</sup>Martins et al. (2011); <sup>8</sup>Sharma and Kiyatkin (2009); <sup>9</sup>Kiyatkin and Sharma (2011); <sup>10</sup>Sharma and Ali (2006); <sup>11</sup>Bowyer et al. (2008); <sup>12</sup>Kuroda et al. (2010); <sup>13</sup>Kiyatkin et al. (2007); <sup>14</sup>Banerjee et al. (2010); <sup>15</sup>Nath et al. (2001); <sup>16</sup>Letendre et al. (2005, 2007).

## MDMA (ECSTASY)

3,4-Methylenedioxymethamphetamine (MDMA or “Ecstasy”) is a synthetic derivative of amphetamine. There is a growing population of MDMA abusers, particularly among young adults who attend “raves” or private clubs (NIDA 2008). Like other amphetamines, MDMA increases the release of monoamines from nerve terminals through interactions with the serotonin transporter (SERT) and dopamine transporter (DAT; Sulzer et al., 2005). MDMA has a higher affinity for SERT than DAT resulting in a greater release of 5-HT than DA (Rothman and Baumann, 2003). Like meth, secondary release of glutamate may contribute to the neurotoxic effects of MDMA (Nash and Yamamoto, 1992). Though acute consequences of MDMA use have been well-studied, long-term effects and potential neurotoxicities are not well known. MDMA appears to be selectively toxic to 5-HT terminals and produces long-lasting depletions of 5-HT in the hippocampus, prefrontal cortex, amygdala, and striatum (Morgan and Gibb, 1980; Ricaurte et al., 1985, 1988). Moreover, thioester metabolites of MDMA, including 3,4-dihydroxyamphetamine (DHA), also produce long-term depletion of 5-HT (Monks et al., 2004). Though 5-HT toxicities have been identified in multiple brain areas, the mechanisms underlying this selective toxicity remain unknown.

Recent studies suggest that BBB dysfunction may contribute to MDMA-induced neurotoxicities. Increased leakage of Evans Blue or trypan blue as well as increased IgG immunostaining were reported in several cortical areas, hippocampus, cerebellum, and striatum of rats treated acutely with MDMA (Yamamoto and Bankson, 2005; Sharma and Ali, 2008; Torres et al., 2011). BBB dysfunction was observed immediately following acute MDMA treatment and up to 10 weeks following an acute injection. Increased BBB permeability after MDMA treatment was associated with increased parenchymal penetration of endogenous albumin (Sharma and Ali, 2008), increased activation of astrocytes, and microglia (Monks et al., 2004), and increased brain water content suggesting edema (Sharma and Ali, 2008).

In addition to altering BBB permeability, MDMA increases expression of pro-inflammatory cytokines, including IL-1 $\beta$ , in brain tissue (Torres et al., 2011). These cytokines can contribute to the feed-forward cycle between BBB dysfunction and neuroinflammation. MDMA abuse also leads to oxidative stress (Yamamoto and Bankson, 2005). MDMA-induced excess release of DA and 5-HT results in the formation of ROS, DAQ, and toxic metabolites from 5-HT oxidation (Quinton and Yamamoto, 2006). The accumulation of toxic free radicals increases the susceptibility of brain tissue to ischemic injury and triggers a variety of signaling cascades leading to BBB dysfunction, brain edema, and neuroinflammation (Gu et al., 2012). It is likely that the oxidative stress and neuroinflammation produced by MDMA results in BBB dysfunction, though further research is needed to verify if MDMA has a direct detrimental effect on the BBB (see Table 2 for a summary of the effects of MDMA on BBB function).

## COCAINE

Cocaine inhibits monoamine reuptake, particularly through binding DAT (Barnett et al., 1981). While cocaine abuse was at its highest levels in the 1970–1980s, the United States is still the world’s number one importer and user with approximately 1.9

**Table 2 | Summary of the preclinical and clinical literature on MDMA-induced BBB dysfunction.**

Rodent models	Findings
	Increased leakage of Evans Blue and Trypan Blue <sup>1</sup>
	Increased penetration of IgG and endogenous albumin <sup>2</sup>
	Increased edema <sup>1</sup>
	Increased activation of astrocytes and other neuroglial cells <sup>1,3</sup>
	Increased expression of pro-inflammatory cytokines <sup>2,4,5</sup>
	Increased production of reactive oxygen species <sup>6,7</sup>

<sup>1</sup>Sharma and Ali (2008); <sup>2</sup>Torres et al. (2011); <sup>3</sup>Monks et al. (2004); <sup>4</sup>Orio et al. (2004); <sup>5</sup>O’Shea et al. (2005); <sup>6</sup>Yamamoto and Bankson (2005); <sup>7</sup>Yamamoto and Raudensky (2008).

million current cocaine abusers (NSDUH 2008). Cocaine abuse is not linked to DA or 5-HT terminal damage (Bennett et al., 1993) though cocaine can produce oxidative stress and neuroinflammation. These potentially toxic effects likely contribute to cocaine-induced BBB dysfunction. Chronic cocaine administration (30 mg/kg/day intraperitoneal injection) ruptures the neurovascular capillaries and basement membranes of rats (Barroso-Moguel et al., 1997). Increased BBB permeability marked by approximately 50% more Evans Blue or sodium fluorescein leakage from blood to brain after cocaine exposure has also been observed in rodents (Sharma et al., 2009; Yang et al., 2010; Yao et al., 2011). Whole animal studies are more recent in this field as most studies investigating the effects of cocaine on BBB integrity use an *in vitro* BBB model comprised of brain microvascular endothelial cells and C6 astrocytes. Decreases in transendothelial electrical resistance and increases in FITC-Dextran leakage across an endothelial monolayer indicates increased permeability across BECs after cocaine treatment (Fiala et al., 1998, 2005; Zhang et al., 1998; Gandhi et al., 2010; Yao et al., 2011). Clinical studies reveal similar BBB breakdown in the basal ganglia and increased HIV penetration into the CNS of cocaine-abusing humans (Nath et al., 2001).

Cocaine-induced BBB dysfunction is partially characterized by loss of, or alterations in, tight junction protein complexes (Fiala et al., 2005, 2008). Significant decreases in ZO-1 and JAM-2 occur in both rodents and *in vitro* BBB preparations (Dhillon et al., 2008; Gandhi et al., 2010; Yao et al., 2011). Cocaine treatment also increases gene expression of factors, including MMP-1, that contribute to basement membrane actin rearrangement resulting in stress fiber formation around cerebral vessels (Nair et al., 2004; Fiala et al., 2005; Dhillon et al., 2008). Persistent loss of, or conformation changes in tight junction proteins, and reorganization of basement membrane fibers leave the brain open to peripheral toxin penetration leading to CNS disorders linked with cocaine abuse (see Table 3 for a summary of the effects of cocaine on BBB function).

Neuroinflammation induced by cocaine (through increases in endothelial adhesion molecules and inflammatory factors) potentially plays an important role in BBB dysfunction. Increases in intracellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), endothelial-leukocyte adhesion molecule (ELAM or selectin-1), and platelet endothelial cell

**Table 3 | Summary of the preclinical and clinical literature on cocaine-induced BBB dysfunction.**

<b>In vitro BBB model</b>	Decreased transendothelial resistance (TEER) <sup>1,2</sup> Increased leakage of FITC-Dextran, Evans Blue, and toxins <sup>1,2,3,4,5,6</sup> Increased transmigration of peripheral blood mononuclear cells (PBMCs) and expression of adhesion molecules <sup>4,7,8,9,10</sup> Decreased expression of zona occludin 1 (ZO-1), junctional adhesion molecule 2 (JAM-2), and other tight junction proteins <sup>1,2,5,7,8</sup> Increased expression of pro-inflammatory cytokines <sup>3,10,11</sup>
<b>Rodent models</b>	Increased leakage of Evans Blue and NaFl <sup>5,12</sup> Increased edema <sup>13,14</sup> Increased transmigration of leukocytes and expression of adhesion molecules <sup>13,15</sup> Decreased expression of zona occludin 1 (ZO-1) <sup>5</sup>
<b>Clinical studies</b>	Increased penetration of peripheral viruses like HIV <sup>16</sup> Increased transmigration of leukocytes and expression of adhesion molecules <sup>17</sup>

<sup>1</sup>Fiala et al. (2005); <sup>2</sup>Gandhi et al. (2010); <sup>3</sup>Zhang et al. (1998); <sup>4</sup>Fiala et al. (1998); <sup>5</sup>Yao et al. (2011); <sup>6</sup>Fiala et al. (1996); <sup>7</sup>Fiala et al. (2008); <sup>8</sup>Dhillon et al. (2008); <sup>9</sup>Nair et al. (2004); <sup>10</sup>Gan et al. (1999); <sup>11</sup>Lee et al. (2001); <sup>12</sup>Yao et al. (2011); <sup>13</sup>Barroso-Moguel et al. (1997); <sup>14</sup>Sharma et al. (2009); <sup>15</sup>Chang et al. (2000); <sup>16</sup>Nath et al. (2001); <sup>17</sup>Buch et al. (2011).

adhesion molecule 1 (PECAM1), have been indentified in both preclinical studies (Fiala et al., 1998, 2005; Gan et al., 1999; Chang et al., 2000) and in cocaine-abusing humans (Buch et al., 2011). Increases in circulating leukocytes and cell adhesion molecules on leukocytes have also been reported with cocaine treatment in rodents and *in vitro* (Chang et al., 2000; Nair et al., 2004). Increases in cell adhesion molecules on the BBB endothelium and on leukocytes promote more leukocyte extravasation from blood to brain (Barroso-Moguel et al., 1997; Gan et al., 1999). Indeed, cocaine treatment *in vitro* results in a 100% increase in the number of peripheral blood mononuclear cells migrating across BECs (Fiala et al., 1998, 2008; Zhang et al., 1998). Cocaine-induced increases in leukocyte-endothelial adhesion are accompanied by elevated levels of pro-inflammatory cytokines and chemokines. Western blotting and immunostaining reveal increases in the expression of TNF $\alpha$ , IL-6, IL-8, nuclear factor kappa B (NF $\kappa$ B), activator protein 1 (AP-1), and CCR2 (Zhang et al., 1998; Gan et al., 1999; Lee et al., 2001; Dhillon et al., 2008). These inflammatory signals are also associated with increased viral invasion of macrophage-tropic HIV across an *in vitro* BBB (Fiala et al., 1996; Zhang et al., 1998) which may explain the greater neuropathology observed in patients co-morbid for HIV infection and cocaine abuse (Fiala et al., 2008). The effects of cocaine on the BBB are likely exacerbated by cocaine-induced neuroinflammation.

## NICOTINE

Though the number of cigarette smokers in the United States has been steadily on the decline since the 1960s, an estimated

45.3 million Americans are still considered daily cigarette smokers (CDC 2010). An average cigarette yields approximately 1 mg of absorbed nicotine, a stimulant alkaloid responsible for the addictive-properties of cigarette smoking (Connolly et al., 2007). Nicotine is a soluble small molecule that rapidly diffuses through the BBB and can interact directly with nicotinic acetylcholine receptors (nAChRs) on BECs (Le, 2003). Preclinical studies using rodent models of acute and chronic nicotine exposure reported compromised BBB integrity, marked by Evans blue or [<sup>14</sup>C] sucrose leakage into brain parenchyma. This BBB disruption occurred as early as 2 h after acute nicotine and after 6 weeks of chronic exposure (Lin et al., 1992; Uzum et al., 1999; Venisnik, 2000; Hawkins et al., 2004, 2005). *In vitro* studies using either brain microvascular endothelial cells alone or in combination with C6 astrocytes revealed increases of FITC-Dextran and [<sup>14</sup>C]sucrose leakage across the cells after nicotine treatment (Schilling et al., 1992; Abbruscato et al., 2002). Nicotine also decreases transendothelial electrical resistance in an *in vitro* BBB model (Hutamekalin et al., 2008; Rodriguez-Gaztelumendi et al., 2011). This decrease in resistance is dose-dependent (0.1–100  $\mu$ M) and sufficient enough to produce a 60–150% increased invasion of *Escherichia coli* across brain microvascular endothelial cells (Chen et al., 2002). Whole animal studies also revealed nicotine-induced BBB dysfunction leading to increased transport from blood to brain of xenobiotics like squalinivir, a protease inhibitor included in anti-retroviral therapy for HIV infection (Manda et al., 2010a). These studies offer added insight into the consequences of nicotine use relating to uptake of therapeutics into the CNS. Finally, both *in vitro* and *in vivo* studies report significant loss, or alterations in, tight junction proteins including ZO-1, claudin-3, JAMs, and occludin (Abbruscato et al., 2002; Hawkins et al., 2004; Hutamekalin et al., 2008; Manda et al., 2010b).

In addition to altering BBB tight junctions, nicotine affects transport and receptor systems involved in normal BBB function. Nicotine increases the activity of monocarboxylase and organic cation transporters while decreasing the functional activity of ion transporters like Na<sup>+</sup>, K<sup>+</sup>, 2Cl<sup>-</sup> co- transporter, and Na<sup>+</sup>, K<sup>+</sup>-ATPase on BECs (Wang et al., 1994; Abbruscato et al., 2004; Lockman et al., 2005; Paulson et al., 2006; Liou et al., 2007). Nicotine treatment produced a 22 and 17% decrease in Na<sup>+</sup> and K<sup>+</sup>-ATPase activity in the microvasculature and brain, respectively (Wang et al., 1994). Nicotine also inhibited the activity of P-glycoprotein, an important efflux transporter responsible for impeding the entry of a range of compounds into the CNS (Manda et al., 2010a). Changes in transport mechanisms across the BBB can produce detrimental alterations in ion gradients and in the nutrients available to the brain (Paulson et al., 2006; Yang et al., 2006). Nicotine-induced alterations in ion transport are especially injurious as changes in ion gradients lead to brain edema lasting for up to 3 weeks after nicotine exposure as well as increased cerebral ischemic injury (Wang et al., 1997; Paulson et al., 2010). nAChRs on cells within the BBB are also affected by nicotine. Chronic nicotine decreases nAChR surface expression as well as expression of certain nAChR subunits, particularly the  $\alpha$ 2 isoform, on BECs (Wang et al., 1994; Abbruscato et al., 2004; Lockman et al., 2005). While nicotine activity at nAChRs on BECs alters the BBB, treatment with nAChRs antagonists (e.g.,



mecamylamine,  $\alpha$ -bungarotoxin, and hexamethonium) decreases nicotine-induced BBB dysfunction. This decrease in BBB disruption was identified by attenuation of [ $^{14}$ C]sucrose leakage into the brain parenchyma, reversal of ZO-1 loss, and decreased *Escherichia coli* invasion in an *in vitro* BBB model (Abbruscato et al., 2002; Chen et al., 2002; Conklin et al., 2002; Hawkins et al., 2005; see **Table 4** for a summary of the effects of nicotine on BBB function).

Nicotine can activate inflammatory signals that directly affect BBB integrity. Studies have highlighted the effects of nicotine on adhesion molecules necessary for immune cell extravasation from the blood into the brain. Nicotine enhances peripheral blood mononuclear cell migration across cerebral vessels through a cascade promoting increased expression of adhesion molecules and inflammatory cytokines (Yong et al., 1997; Albaugh et al., 2004; Bradford et al., 2011). The expression of ICAM-1, VCAM-1, and P-selectin is increased with chronic nicotine treatment (Yong et al., 1997; Albaugh et al., 2004). Furthermore, nicotine increases gene expression of TNF $\alpha$ , IL-18, IL-1 $\beta$ , and chemokines including CCL2, CCL8, and CXCL1 (Bradford et al., 2011). Nicotine also decreases gene expression of anti-inflammatory factors (e.g., Bcl6, IL-10, and CCL25; Bradford et al., 2011). These studies indicate that nicotine increases the expression of inflammatory cytokines and endothelial adhesion molecules and can enhance leukocyte migration across the BBB, particularly during neuroinflammation.

**Table 4 | Summary of the preclinical and clinical literature on nicotine-induced BBB dysfunction.**

<b><i>In vitro</i> BBB model</b>	Decreased transendothelial resistance (TEER) <sup>1,2</sup> Increased leakage of FITC-Dextran, <sup>14</sup> Sucrose, and toxins <sup>3,4,5</sup> Increased transmigration of peripheral blood mononuclear cells (PBMCs) and expression of adhesion molecules <sup>2,6,7</sup> Decreased expression of zona occludin 1 (ZO-1), occludin, and cadherins <sup>2,4,8</sup> Changes in nicotinic acetylcholine receptors (nAChR), ion channels, and transporter systems <sup>9,10,11,12</sup>
<b>Rodent models</b>	Increased leakage of Evans Blue, <sup>14</sup> Sucrose, and peripheral toxins <sup>8,12,13,14,15</sup> Increased edema <sup>16,17</sup> Decreased expression of zona occludin 1 (ZO-1), occludin, and claudin-5 <sup>8,12</sup> Increased expression of pro-inflammatory cytokines <sup>18</sup> Changes in nAChRs, ion channels, and transporter systems <sup>19,20,21</sup> Increased production of reactive oxygen species (ROS) <sup>8,22,23</sup>

<sup>1</sup>Rodriguez-Gaztelumendi et al. (2011); <sup>2</sup>Hutamekalin et al. (2008); <sup>3</sup>Schilling et al. (1992); <sup>4</sup>Abbruscato et al. (2002); <sup>5</sup>Chen et al. (2002); <sup>6</sup>Albaugh et al. (2004); <sup>7</sup>Speer et al. (2002); <sup>8</sup>Manda et al. (2010a,b); <sup>9</sup>Liou et al. (2007); <sup>10</sup>Yang et al. (2006); <sup>11</sup>Paulson et al. (2006); <sup>12</sup>Abbruscato et al. (2004); <sup>13</sup>Hawkins et al. (2005); <sup>14</sup>Uzum et al. (1999); <sup>15</sup>Lin et al. (1992); <sup>16</sup>Wang et al. (1997); <sup>17</sup>Paulson et al. (2010); <sup>18</sup>Bradford et al. (2011); <sup>19</sup>Canis et al. (2009); <sup>20</sup>Lockman et al. (2005); <sup>21</sup>Wang et al. (1994); <sup>22</sup>Gerzanich et al. (2003); <sup>23</sup>Yong et al. (1997).

## STIMULANT CO-MORBIDITY WITH HIV INFECTION

Mechanisms thought to underlie stimulant-mediated toxicities parallel those associated with HIV infection. Indeed, the co-morbidity of stimulant abuse and HIV is well documented. Studies report that meth abuse is linked with enhanced frequency of unprotected sex and increased HIV transmission (Semple et al., 2004; Buchacz et al., 2005). Moreover, the morbidity and mortality of meth-abusing HIV-positive individuals far exceeds that seen with either pathology alone.

HIV infects peripheral leukocytes which migrate across the BBB, thus allowing the virus access to the CNS (An et al., 1999). This migration occurs early in HIV infection (Risdaal et al., 1998; Romanelli et al., 2006) though HIV-induced toxicity of BECs occurs throughout infection and parallels HIV toxicity of neurons (Price et al., 2005). Once HIV-infected cells enter the brain, HIV-produced proteins trigger neuroinflammation which can exacerbate the neuroinflammatory effects of stimulant drugs. As overviewed above, stimulant drug abuse compromises BBB integrity through decreases in tight junction proteins and increases in endothelial adhesion molecules which promote leukocyte extravasation from the blood into the brain.

Meth increases the expression of HIV co-receptors, CXCR4 and CCR5 (Nair et al., 2009), and can significantly increase HIV replication in astrocytes (Gavrilin et al., 2002). In HIV-positive meth abusers, meth-induced BBB dysfunction facilitates increased transport of HIV-infected leukocytes into the brain (Liang et al., 2008). Moreover, HIV-induced BBB damage might increase the concentration of meth within the CNS contributing to further meth-neurotoxicity. Co-treatment of meth and HIV protein gp120 decreases transendothelial electrical resistance across BECs more than treatment with meth or gp120 alone (Mahajan et al., 2008). Administration of meth and gp120 or the HIV protein Tat in combination, further decreased the expression of ZO-1, claudin-3, claudin-5, and JAM-2 than either treatment alone (Mahajan et al., 2008; Banerjee et al., 2010). In addition to the effects of meth and HIV on tight junction proteins, increased levels of MMP-2 and MMP-9 were reported with meth and Tat treatment in an *in vitro* BBB model (Conant et al., 2004) and in the cerebrospinal fluid of HIV-positive individuals (Conant et al., 1999; Liuzzi et al., 2004). These peptidases can produce persistent BBB damage, leaving the brain open to prolonged infiltration of HIV and other peripheral toxins.

Clinical investigations revealed potential additive and/or synergistic effects of meth and HIV within the CNS. Chang et al. (2005) found increased neuronal loss in frontal gray matter (−5.7%), frontal white matter (−6.1%), and basal ganglia (−6%) when compared with HIV-positive meth-naïve individuals and HIV-negative meth abusers. These studies also reported increased glial activation and changes in cellular metabolism in frontal white matter and basal ganglia in HIV-positive meth abusers (Chang et al., 2005). Meth-induced glial activation may lead to increased progression of neuroAIDS in HIV-positive meth abusers (Garden, 2002). DA neurotoxicity is associated with both meth abuse, and HIV infection appears worsened by the combination (Czub et al., 2001; Maragos et al., 2002; Theodore et al., 2006). Increased DA neurotoxicity with meth abuse and HIV/AIDS co-morbidity may increase the risk of DA-associated disorders like PD. In fact, meth abusers have a twofold increase in developing PD compared with

healthy controls or cocaine abusers (Callaghan et al., 2012) and as many as 10% of patients with HAND also display Parkinsonian features (Berger and Nath, 1997). In summary, the neuropathological features associated with both meth abuse and HIV/AIDS may be exacerbated through meth-induced BBB dysfunction resulting in, or worsened by, neuroinflammation.

Similar to meth abuse, individuals abusing MDMA are more likely to engage in risky sexual behaviors and injection drug use (Dunn et al., 2010). Indeed, 77% of participants in a clinical investigation reported not using condoms when under the influence of MDMA, and of this group, 54% reported having two or more sexual partners during this time (Dunn et al., 2010). In a similar clinical study, 9.3% of MDMA users surveyed reported having unprotected sex with partners who were either HIV-positive or of unknown HIV status (Klitzman et al., 2002). These behaviors likely increase HIV transmission within this population (Mitcheson et al., 2008). Little is currently known about the neurotoxic implications of MDMA and HIV co-morbidity. As MDMA produces BBB dysfunction as well as neuroinflammation and oxidative stress, which both perpetuate BBB damage, MDMA-induced BBB dysfunction would be expected to increase HIV-infected leukocyte infiltration into the CNS. The interaction between MDMA and HIV has not yet been characterized and may represent an unknown population at risk for HIV and MDMA-associated neurotoxicities. These toxicities may be exacerbated by MDMA-induced BBB dysfunction leading to increased HIV penetration into the brain.

Early investigations reported cocaine abuse as a risk factor for increased HIV transmission and a more rapid progression to AIDS (Fiala et al., 1998; Webber et al., 1999). Cocaine increases HIV infiltration into the CNS through increased HIV-infected leukocyte adhesion and transmigration across the BBB endothelium. BBB dysfunction observed following cocaine exposure can increase the capacity of other immune cells and inflammatory factors to enter into the brain from the periphery. These features of cocaine likely promote the neuroinflammation associated neuroAIDS. Cocaine also increases HIV replication in monocytes, macrophages, and astrocytes (Peterson et al., 1991; Roth et al., 2002; Reynolds et al., 2006). This is especially important as astrocytes may act as a reservoir for HIV within the brain (Nath, 2010). Preclinical studies revealed that cocaine enhances oxidative stress, neuronal dysfunction, and apoptosis in gp120 and Tat-treated cells *in vitro* and in rodents (Koutsilieri et al., 1997; Nath et al., 2000; Bagetta et al., 2004). Recent findings from Napier et al. (2010) revealed that cocaine and Tat synergize to over activate cortical neurons. Like cocaine, Tat can inhibit DAT which may contribute to HIV-induced DA neurotoxicity (Aksenov et al., 2008). When combined, cocaine plus Tat may result in DA neurotoxicities which exacerbate HIV neuropathology. Cocaine-induced BBB dysfunction leading to increased HIV transmission into the CNS is apt to contribute to the resulting neurotoxicities associated with cocaine and HIV co-morbidity.

There are a number of clinical reports documenting the effects of cocaine and HIV co-morbidity. Cocaine abuse accelerates HIV disease progression by decreasing CD4+ T-cell counts and increasing HIV viral load, independent of anti-retroviral therapies (Baum et al., 2009). Chaisson et al. (1989) found that daily cocaine use increased the risk of HIV infection up to sixfold in African

American and Hispanic populations. Persistent cocaine abusers are also three times more likely to die from AIDS-related complications than cocaine-naïve individuals (Cook et al., 2008). This suggests a synergistic interaction between cocaine and HIV which exacerbates HIV disease progression. Interactions between cocaine and HIV within the brain parenchyma may be enhanced through cocaine-induced increases in HIV-infected leukocyte trafficking across the BBB.

Studies report a 51% or higher cigarette smoking rate among nationally surveyed HIV-positive individuals (Collins et al., 2001; Gritz et al., 2004; Burkhalter et al., 2005). Though little is known about the effects of nicotine on HIV disease progression, smoking increases the likelihood of HIV-related complications (e.g., bacterial pneumonia and HIV-associated dementia) and increases the mortality rate of HIV-positive individuals (Hirschtick et al., 1995; Conley et al., 1996; Feldman et al., 2006). Furthermore, nicotine can act as a major immunosuppressive agent by attenuating the immune and virological response to anti-retroviral therapies by up to 40% (Humfleet et al., 2009). Nicotine has both pro- and anti-inflammatory properties. The immunosuppressive effects of nicotine include increasing T-cell unresponsiveness, increasing IL-4 production, and inhibiting the production of certain pro-inflammatory cytokines (Sopori and Kozak, 1998). Immunosuppressive functions can add to the already detrimental immune suppression produced by HIV infection leading to an accelerated progression of HIV-associated complications. The pro-inflammatory effects of nicotine within the CNS include increasing the release of pro-inflammatory cytokines and increasing leukocyte transmigration through BECs from blood to brain. Nicotine may increase HIV viral load within the CNS through increasing BBB permeability and promoting leukocyte migration into the brain parenchyma. Nicotine enhances the production of HIV in macrophages (Abbud et al., 1995) and in activated microglia (Rock et al., 2008). Similar increases in HIV replication are also seen with opiate and cocaine abuse which are reported to promote HIV neuropathology (Nath et al., 2002; Roth et al., 2005). The scope of what is currently known about the effects of nicotine on HIV disease progression is limited, though HIV-positive individuals who smoke cigarettes show a decreased immune response, poorer responses to anti-retroviral therapies, greater risk of viral rebound, and an increased probability for HIV-related complications (Feldman et al., 2006). Nicotine may increase HIV infiltration into the CNS through nicotine-induced BBB dysfunction and increase HIV replication within microglia in the brain to accelerate progression to neuroAIDS.

## CONCLUSION

The BBB/neuro(glio)vascular unit plays an important function in the neurotoxicity of stimulant abuse; both as a factor involved in the development of neurotoxicities and also in its response to these toxic effects. This suggests that components within the BBB and neuro(glio)vascular unit may be potential therapeutic sites targeted to impede the progression of stimulant-induced neurotoxicity. Most stimulants dysregulate the BBB through alterations in tight junction complexes or through oxidative stress and neuroinflammation. Neuroinflammation plays a particularly important role as it contributes to a feed-forward process between

BBB dysfunction and neuroinflammation which likely results in the CNS being more vulnerable to infiltration of pathological mediators from the periphery. This is especially important in comorbid situations of stimulant abuse and immune disorders, like HIV/AIDS. Stimulants not only increase the migration of peripheral HIV-infected leukocytes into the CNS, but once inside the brain, can accelerate the progression to neuroAIDS.

The effects of stimulant drugs have been extensively researched, though further studies are required to advance the field toward identification of therapeutic targets for medication development. Preclinical studies thus far have revealed a great deal about the direct and indirect effects of stimulants on the BBB. Future studies must build upon what is already known using a model of stimulant abuse and co-occurring morbidities, like HIV infection. Such

models of co-morbidity would better emulate the human condition and would allow more direct assessments of the dynamic interactions between all cells within the BBB/neuro(glio)vascular unit in these clinically important human scenarios.

## ACKNOWLEDGMENTS

The authors gratefully acknowledge support from the Kenneth Douglas Foundation, the Daniel F. and Ada L. Rice Foundation, and the Center for Compulsive Behavior and Addiction-Division for Blood Brain Barrier and Substance Abuse at Rush University, Chicago, IL, USA. The authors also acknowledge the thoughtful insight on the link between stimulant abuse and HIV/AIDS neuropathology offered by Dr. Linda Baum, Division of Immunology and Microbiology, Rush University, Chicago, IL, USA.

## REFERENCES

- Abbott, N. J., Patabendige, A. A., Dolman, D. E., Yusof, S. R., and Begley, D. J. (2010). Structure and function of the blood-brain barrier. *Neurobiol. Dis.* 37, 13–25.
- Abbruscato, T. J., Lopez, S. P., Mark, K. S., Hawkins, B. T., and Davis, T. P. (2002). Nicotine and cotinine modulate cerebral microvascular permeability and protein expression of ZO-1 through nicotinic acetylcholine receptors expressed on brain endothelial cells. *J. Pharm. Sci.* 91, 2525–2538.
- Abbruscato, T. J., Lopez, S. P., Roder, K., and Paulson, J. R. (2004). Regulation of blood-brain barrier Na,K,2Cl<sup>-</sup> cotransporter through phosphorylation during in vitro stroke conditions and nicotine exposure. *J. Pharmacol. Exp. Ther.* 310, 459–468.
- Abbud, R. A., Finegan, C. K., Guay, L. A., and Rich, E. A. (1995). Enhanced production of human immunodeficiency virus type 1 by in vitro infected alveolar macrophages from otherwise healthy cigarette smokers. *J. Infect. Dis.* 172, 859–863.
- Abdul Muneer, P. M., Alikunju, S., Szlachetka, A. M., Murrin, L. C., and Haorah, J. (2011). Impairment of brain endothelial glucose transporter by methamphetamine causes blood-brain barrier dysfunction. *Mol. Neurodegener.* 6, 23.
- Aksenov, M. Y., Aksenova, M. V., Silvers, J. M., Mactutus, C. F., and Booze, R. M. (2008). Different effects of selective dopamine uptake inhibitors, GBR 12909 and WIN 35428, on HIV-1 Tat toxicity in rat fetal mid-brain neurons. *Neurotoxicology* 29, 971–977.
- Albaugh, G., Bellavance, E., Strande, L., Heinburger, S., Hewitt, C. W., and Alexander, J. B. (2004). Nicotine induces mononuclear leukocyte adhesion and expression of adhesion molecules, VCAM and ICAM, in endothelial cells in vitro. *Ann. Vasc. Surg.* 18, 302–307.
- An, S. F., Groves, M., Gray, F., and Scaravilli, F. (1999). Early entry and widespread cellular involvement of HIV-1 DNA in brains of HIV-1 positive asymptomatic individuals. *J. Neuropathol. Exp. Neurol.* 58, 1156–1162.
- Bagetta, G., Piccirilli, S., Del Duca, C., Morrone, L. A., Rombola, L., Nappi, G., De Alba, J., Knowles, R. G., and Corasaniti, M. T. (2004). Inducible nitric oxide synthase is involved in the mechanisms of cocaine enhanced neuronal apoptosis induced by HIV-1 gp120 in the neocortex of rat. *Neurosci. Lett.* 356, 183–186.
- Bandopadhyay, R., Orte, C., Lawrenson, J. G., Reid, A. R., De, S. S., and Allt, G. (2001). Contractile proteins in pericytes at the blood-brain and blood-retinal barriers. *J. Neurocytol.* 30, 35–44.
- Banerjee, A., Zhang, X., Manda, K. R., Banks, W. A., and Ercal, N. (2010). HIV proteins (gp120 and Tat) and methamphetamine in oxidative stress-induced damage in the brain: potential role of the thiol antioxidant N-acetylcysteine amide. *Free Radic. Biol. Med.* 48, 1388–1398.
- Barnett, G., Hawks, R., and Resnick, R. (1981). Cocaine pharmacokinetics in humans. *J. Ethnopharmacol.* 3, 353–366.
- Barroso-Moguel, R., Villeda-Hernández, J., Méndez-Armenta, M., and Rios, C. (1997). Brain capillary lesions produced by cocaine in rats. *Toxicol. Lett.* 92, 9–14.
- Baum, M. K., Rafie, C., Lai, S., Sales, S., Page, B., and Campa, A. (2009). Crack-cocaine use accelerates HIV disease progression in a cohort of HIV-positive drug users. *J. Acquir. Immune Defic. Syndr.* 50, 93–99.
- Bell, R. D., Winkler, E. A., Sagare, A. P., Singh, I., LaRue, B., Deane, R., and Zlokovic, B. V. (2010). Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* 68, 409–427.
- Bennett, B. A., Hyde, C. E., Pecora, J. R., and Clodfelter, J. E. (1993). Differing neurotoxic potencies of methamphetamine, mazindol, and cocaine in mesencephalic cultures. *J. Neurochem.* 60, 1444–1452.
- Berger, J. R., and Nath, A. (1997). HIV dementia and the basal ganglia. *Intervirolgy* 40, 122–131.
- Bowyer, J. F., and Ali, S. F. (2006). High doses of methamphetamine that cause disruption of the blood-brain barrier in limbic regions produce extensive neuronal degeneration in mouse hippocampus. *Synapse* 60, 521–532.
- Bowyer, J. F., Thomas, M., Schmued, L. C., and Ali, S. F. (2008). Brain region-specific neurodegenerative profiles showing the relative importance of amphetamine dose, hyperthermia, seizures, and the blood-brain barrier. *Ann. N. Y. Acad. Sci.* 1139, 127–139.
- Bradford, S. T., Stamatovic, S. M., Dondeti, R. S., Keep, R. F., and Andjelkovic, A. V. (2011). Nicotine aggravates the brain postischemic inflammatory response. *Am. J. Physiol. Heart Circ. Physiol.* 300, H1518–H1529.
- Buch, S., Yao, H., Guo, M., Mori, T., Su, T. P., and Wang, J. (2011). Cocaine and HIV-1 interplay: molecular mechanisms of action and addiction. *J. Neuroimmune Pharmacol.* 6, 503–515.
- Buchacz, K., McFarland, W., Kellogg, T. A., Loeb, L., Holmberg, S. D., Dille, J., and Klausner, J. D. (2005). Amphetamine use is associated with increased HIV incidence among men who have sex with men in San Francisco. *AIDS* 19, 1423–1424.
- Buckner, D., Wilson, S., Kurk, S., Hardy, M., Miessner, N., and Jutila, M. A. (2006). Use of early passage fetal intestinal epithelial cells in semi-high-throughput screening assays: an approach to identify new innate immune system adjuvants. *J. Biomol. Screen* 11, 664–671.
- Burkhalter, J. E., Springer, C. M., Chhabra, R., Ostroff, J. S., and Rapkin, B. D. (2005). Tobacco use and readiness to quit smoking in low-income HIV-infected persons. *Nicotine Tob. Res.* 7, 511–522.
- Cadet, J. L., Sheng, P., Ali, S. F., Rothman, R., Carlson, E., and Epstein, C. (1994). Attenuation of methamphetamine-induced neurotoxicity in copper/zinc superoxide dismutase transgenic mice. *J. Neurochem.* 62, 380–383.
- Callaghan, R. C., Cunningham, J. K., Sykes, J., and Kish, S. J. (2012). Increased risk of Parkinson's disease in individuals hospitalized with conditions related to the use of methamphetamine or other amphetamine-type drugs. *Drug Alcohol Depend.* 120, 35–40.
- Canis, M., Mack, B., Gires, O., Maurer, M. H., Kuschinsky, W., Duembgen, L., and Duelli, R. (2009). Increased densities of monocarboxylate transport protein MCT1 after chronic administration of nicotine in rat brain. *Neurosci. Res.* 64, 429–435.
- Cartier, L., Hartley, O., Dubois-Dauphin, M., and Krause, K. H. (2005). Chemokine receptors in the central nervous system: role in brain inflammation and neurodegenerative diseases. *Brain Res. Brain Res. Rev.* 48, 16–42.
- Carvey, P. M., Hendey, B., and Monahan, A. J. (2009). The blood-brain barrier in neurodegenerative disease: a rhetorical perspective. *J. Neurochem.* 111, 291–314.
- Chaisson, R. E., Bacchetti, P., Osmond, D., Brodie, B., Sande, M. A., and Moss, A. R. (1989). Cocaine use and HIV infection in intravenous drug users in San Francisco. *JAMA* 261, 561–565.



- Chang, L., Ernst, T., Speck, O., and Grob, C. S. (2005). Additive effects of HIV and chronic methamphetamine use on brain metabolite abnormalities. *Am. J. Psychiatry* 162, 361–369.
- Chang, S. L., Bersig, J., Felix, B., Fiala, M., and House, S. D. (2000). Chronic cocaine alters hemodynamics and leukocyte-endothelial interactions in rat mesenteric venules. *Life Sci.* 66, 2357–2369.
- Chen, Y. H., Chen, S. H., Jong, A., Zhou, Z. Y., Li, W., Suzuki, K., and Huang, S. H. (2002). Enhanced *Escherichia coli* invasion of human brain microvascular endothelial cells is associated with alternations in cytoskeleton induced by nicotine. *Cell. Microbiol.* 4, 503–514.
- Collins, R. L., Kanouse, D. E., Gifford, A. L., Senterfitt, J. W., Schuster, M. A., McCaffrey, D. F., Shapiro, M. F., and Wenger, N. S. (2001). Changes in health-promoting behavior following diagnosis with HIV: prevalence and correlates in a national probability sample. *Health Psychol.* 20, 351–360.
- Compston, A., and Coles, A. (2008). Multiple sclerosis. *Lancet* 372, 1502–1517.
- Conant, K., McArthur, J. C., Griffin, D. E., Sjulson, L., Wahl, L. M., and Irani, D. N. (1999). Cerebrospinal fluid levels of MMP-2, 7, and 9 are elevated in association with human immunodeficiency virus dementia. *Ann. Neurol.* 46, 391–398.
- Conant, K., St Hillaire, C., Anderson, C., Galey, D., Wang, J., and Nath, A. (2004). Human immunodeficiency virus type 1 Tat and methamphetamine affect the release and activation of matrix-degrading proteinases. *J. Neurovirol.* 10, 21–28.
- Conklin, B. S., Zhao, W., Zhong, D. S., and Chen, C. (2002). Nicotine and cotinine up-regulate vascular endothelial growth factor expression in endothelial cells. *Am. J. Pathol.* 160, 413–418.
- Conley, L. J., Bush, T. J., Buchbinder, S. P., Penley, K. A., Judson, F. N., and Holmberg, S. D. (1996). The association between cigarette smoking and selected HIV-related medical conditions. *AIDS* 10, 1121–1126.
- Connolly, G. N., Alpert, H. R., Wayne, G. E., and Koh, H. (2007). Trends in nicotine yield in smoke and its relationship with design characteristics among popular US cigarette brands, 1997–2005. *Tob. Control.* 16, e5.
- Cook, J. A., Burke-Miller, J. K., Cohen, M. H., Cook, R. L., Vlahov, D., Wilson, T. E., Golub, E. T., Schwartz, R. M., Howard, A. A., Ponath, C., Plankey, M. W., Levine, A., and Grey, D. D. (2008). Crack cocaine, disease progression, and mortality in a multicenter cohort of HIV-1 positive women. *AIDS* 22, 1355–1363.
- Couraud, P. O. (1998). Infiltration of inflammatory cells through brain endothelium. *Pathol. Biol.* 46, 176–180.
- Czub, S., Koutsilieris, E., Sopper, S., Czub, M., Stahl-Hennig, C., Muller, J. G., Pedersen, V., Gsell, W., Heeney, J. L., Gerlach, M., Gosztonyi, G., Riederer, P., and ter Meulen, V. (2001). Enhancement of central nervous system pathology in early simian immunodeficiency virus infection by dopaminergic drugs. *Acta Neuropathol.* 101, 85–91.
- de la Monte, S. M., Longato, L., Tong, M., DeNucci, S., and Wands, J. R. (2009). The liver-brain axis of alcohol-mediated neurodegeneration: role of toxic lipids. *Int. J. Environ. Res. Public Health* 6, 2055–2075.
- Deli, M. A., Dehouck, M. P., Cecchelli, R., Abraham, C. S., and Joo, F. (1995). Histamine induces a selective albumin permeation through the blood-brain barrier in vitro. *Inflamm. Res.* 44(Suppl. 1), S56–S57.
- Dhillon, N. K., Peng, F., Bokhari, S., Callen, S., Shin, S. H., Zhu, X., Kim, K. J., and Buch, S. J. (2008). Cocaine-mediated alteration in tight junction protein expression and modulation of CCL2/CCR2 axis across the blood-brain barrier: implications for HIV-dementia. *J. Neuroimmune Pharmacol.* 3, 52–56.
- Diaz-Flores, L., Gutierrez, R., Varela, H., Rancel, N., and Valladares, F. (1991). Microvascular pericytes: a review of their morphological and functional characteristics. *Histol. Histopathol.* 6, 269–286.
- Dunn, M., Day, C., Bruno, R., Degenhardt, L., and Campbell, G. (2010). Sexual and injecting risk behaviours among regular ecstasy users. *Addict. Behav.* 35, 157–160.
- Farkas, E., and Luiten, P. G. (2001). Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog. Neurobiol.* 64, 575–611.
- Feldman, J. G., Minkoff, H., Schneider, M. F., Gange, S. J., Cohen, M., Watts, D. H., Gandhi, M., Mocharnuk, R. S., and Anastos, K. (2006). Association of cigarette smoking with HIV prognosis among women in the HAART era: a report from the women's inter-agency HIV study. *Am. J. Public Health* 96, 1060–1065.
- Fiala, M., Eshleman, A. J., Cashman, J., Lin, J., Lossinsky, A. S., Suarez, V., Yang, W., Zhang, J., Popik, W., Singer, E., Chiappelli, F., Carro, E., Weinand, M., Witte, M., and Arthos, J. (2005). Cocaine increases human immunodeficiency virus type 1 neuroinvasion through remodeling brain microvascular endothelial cells. *J. Neurovirol.* 11, 281–291.
- Fiala, M., Gan, X. H., Newton, T., Chiappelli, F., Shapshak, P., Kermani, V., Kung, M. A., Diagne, A., Martinez, O., Way, D., Weinand, M., Witte, M., and Graves, M. (1996). Divergent effects of cocaine on cytokine production by lymphocytes and monocyte/macrophages: HIV-1 enhancement by cocaine within the blood-brain barrier. *Adv. Exp. Med. Biol.* 402, 145–156.
- Fiala, M., Gan, X. H., Zhang, L., House, S. D., Newton, T., Graves, M. C., Shapshak, P., Stins, M., Kim, K. S., Witte, M., and Chang, S. L. (1998). Cocaine enhances monocyte migration across the blood-brain barrier. Cocaine's connection to AIDS dementia and vasculitis? *Adv. Exp. Med. Biol.* 437, 199–205.
- Fiala, M., Singer, E. J., Commins, D., Mirzapioazova, T., Verin, A., Espinosa, A., Ugen, K., Bernas, M., Witte, M., Weinand, M., and Lossinsky, A. S. (2008). HIV-1 Antigens in neurons of cocaine-abusing patients. *Open Virol. J.* 2, 24–31.
- Fletcher, N. F., Bexiga, M. G., Brayden, D. J., Brankin, B., Willett, B. J., Hosie, M. J., Jacque, J. M., and Callanan, J. J. (2009). Lymphocyte migration through the blood-brain barrier (BBB) in feline immunodeficiency virus infection is significantly influenced by the pre-existence of virus and tumour necrosis factor (TNF)-alpha within the central nervous system (CNS): studies using an in vitro feline BBB model. *Neuropathol. Appl. Neurobiol.* 35, 592–602.
- Gan, X., Zhang, L., Berger, O., Stins, M. F., Way, D., Taub, D. D., Chang, S. L., Kim, K. S., House, S. D., Weinand, M., Witte, M., Graves, M. C., and Fiala, M. (1999). Cocaine enhances brain endothelial adhesion molecules and leukocyte migration. *Clin. Immunol.* 91, 68–76.
- Gandhi, N., Saiyed, Z. M., Napuri, J., Samikkannu, T., Reddy, P. V., Agudelo, M., Khatavkar, P., Saxena, S. K., and Nair, M. P. (2010). Interactive role of human immunodeficiency virus type 1 (HIV-1) clade-specific Tat protein and cocaine in blood-brain barrier dysfunction: implications for HIV-1-associated neurocognitive disorder. *J. Neurovirol.* 16, 294–305.
- Garden, G. A. (2002). Microglia in human immunodeficiency virus-associated neurodegeneration. *Glia* 40, 240–251.
- Gerzanich, V., Ivanova, S., and Simard, J. M. (2003). Early pathophysiological changes in cerebral vessels predisposing to stroke. *Clin. Hemorheol. Microcirc.* 29, 291–294.
- Gavrilin, M. A., Mathes, L. E., and Podell, M. (2002). Methamphetamine enhances cell-associated feline immunodeficiency virus replication in astrocytes. *J. Neurovirol.* 8, 240–249.
- Goncalves, J., Baptista, S., Martins, T., Milhazes, N., Borges, F., Ribeiro, C. F., Malva, J. O., and Silva, A. P. (2010). Methamphetamine-induced neuroinflammation and neuronal dysfunction in the mice hippocampus: preventive effect of indomethacin. *Eur. J. Neurosci.* 31, 315–326.
- Goncalves, J., Martins, T., Ferreira, R., Milhazes, N., Borges, F., Ribeiro, C. F., Malva, J. O., Macedo, T. R., and Silva, A. P. (2008). Methamphetamine-induced early increase of IL-6 and TNF-alpha mRNA expression in the mouse brain. *Ann. N. Y. Acad. Sci.* 1139, 103–111.
- Gritz, E. R., Vidrine, D. J., Lazev, A. B., Amick, B. C. III, and Arduino, R. C. (2004). Smoking behavior in a low-income multiethnic HIV/AIDS population. *Nicotine Tob. Res.* 6, 71–77.
- Gu, Y., Zheng, G., Xu, M., Li, Y., Chen, X., Zhu, W., Tong, Y., Chung, S. K., Liu, K. J., and Shen, J. (2012). Caveolin-1 regulates nitric oxide-mediated matrix metalloproteinases activity and blood-brain barrier permeability in focal cerebral ischemia and reperfusion injury. *J. Neurochem.* 120, 147–156.
- Guilarte, T. R., Nihei, M. K., McGlothlan, J. L., and Howard, A. S. (2003). Methamphetamine-induced deficits of brain monoaminergic neuronal markers: distal axotomy or neuronal plasticity. *Neuroscience* 122, 499–513.
- Hawkins, B. T., Abbruscato, T. J., Egleton, R. D., Brown, R. C., Huber, J. D., Campos, C. R., and Davis, T. P. (2004). Nicotine increases in vivo blood-brain barrier permeability and alters cerebral microvascular tight junction protein distribution. *Brain Res.* 1027, 48–58.
- Hawkins, B. T., and Davis, T. P. (2005). The blood-brain barrier/neurovascular unit in health and disease. *Pharmacol. Rev.* 57, 173–185.

- Hawkins, B. T., Egleton, R. D., and Davis, T. P. (2005). Modulation of cerebral microvascular permeability by endothelial nicotinic acetylcholine receptors. *Am. J. Physiol. Heart Circ. Physiol.* 289, H212–H219.
- Hirschtick, R. E., Glassroth, J., Jordan, M. C., Wilcosky, T. C., Wallace, J. M., Kvale, P. A., Markowitz, N., Rosen, M. J., Mangura, B. T., and Hopewell, P. C. (1995). Bacterial pneumonia in persons infected with the human immunodeficiency virus. Pulmonary Complications of HIV Infection Study Group. *N. Engl. J. Med.* 333, 845–851.
- Humfleet, G. L., Delucchi, K., Kelley, K., Hall, S. M., Dilley, J., and Harrison, G. (2009). Characteristics of HIV-positive cigarette smokers: a sample of smokers facing multiple challenges. *AIDS Educ. Prev.* 21, 54–64.
- Hutamekalin, P., Farkas, A. E., Orbok, A., Wilhelm, I., Nagyoszi, P., Veszelka, S., Deli, M. A., Buzas, K., Hunyadi-Gulyas, E., Medzihradzky, K. F., Meksuriyen, D., and Krizbai, I. A. (2008). Effect of nicotine and polyaromatic hydrocarbons on cerebral endothelial cells. *Cell Biol. Int.* 32, 198–209.
- Iadecola, C. (2004). Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat. Rev. Neurosci.* 5, 347–360.
- Janzer, R. C., and Raff, M. C. (1987). Astrocytes induce blood-brain barrier properties in endothelial cells. *Nature* 325, 253–257.
- Kiyatkin, E. A., Brown, P. L., and Sharma, H. S. (2007). Brain edema and breakdown of the blood-brain barrier during methamphetamine intoxication: critical role of brain hyperthermia. *Eur. J. Neurosci.* 26, 1242–1253.
- Kiyatkin, E. A., and Sharma, H. S. (2009). Acute methamphetamine intoxication: brain hyperthermia, blood-brain barrier, brain edema, and morphological cell abnormalities. *Int. Rev. Neurobiol.* 88, 65–100.
- Kiyatkin, E. A., and Sharma, H. S. (2011). Expression of heat shock protein (HSP 72 kDa) during acute methamphetamine intoxication depends on brain hyperthermia: neurotoxicity or neuroprotection? *J. Neural. Transm.* 118, 47–60.
- Klitzman, R. L., Greenberg, J. D., Pollock, L. M., and Dolezal, C. (2002). MDMA ("ecstasy") use, and its association with high risk behaviors, mental health, and other factors among gay/bisexual men in New York City. *Drug Alcohol Depend.* 66, 115–125.
- Koehler, R. C., Roman, R. J., and Harder, D. R. (2009). Astrocytes and the regulation of cerebral blood flow. *Trends Neurosci.* 32, 160–169.
- Kousik, S. M., Graves, S. M., Napier, T. C., Zhao, C., and Carvey, P. M. (2011). Methamphetamine-induced vascular changes lead to striatal hypoxia and dopamine reduction. *Neuroreport* 22, 923–928.
- Koutsilieris, E., Gotz, M. E., Sopper, S., Sauer, U., Demuth, M., ter Meulen, V., and Riederer, P. (1997). Regulation of glutathione and cell toxicity following exposure to neurotropic substances and human immunodeficiency virus-1 in vitro. *J. Neuroviral.* 3, 342–349.
- Krasnova, I. N., and Cadet, J. L. (2009). Methamphetamine toxicity and messengers of death. *Brain Res.* 60, 379–407.
- Krueger, M., and Bechmann, I. (2010). CNS pericytes: concepts, misconceptions, and a way out. *Glia* 58, 1–10.
- Kuhn, D. M., Francescutti-Verbeem, D. M., and Thomas, D. M. (2006). Dopamine quinones activate microglia and induce a neurotoxic gene expression profile: relationship to methamphetamine-induced nerve ending damage. *Ann. N. Y. Acad. Sci.* 1074, 31–41.
- Kuroda, K. O., Ornthanalai, V. G., Kato, T., and Murphy, N. P. (2010). FosB null mutant mice show enhanced methamphetamine neurotoxicity: potential involvement of FosB in intracellular feedback signaling and astroglial function. *Neuropsychopharmacology* 35, 641–655.
- Lai, Y. T., Tsai, Y. P., Cherng, C. G., Ke, J. J., Ho, M. C., Tsai, C. W., and Yu, L. (2009). Lipopolysaccharide mitigates methamphetamine-induced striatal dopamine depletion via modulating local TNF- $\alpha$  and dopamine transporter expression. *J. Neural. Transm.* 116, 405–415.
- Le, H. J. (2003). Role of nicotine pharmacokinetics in nicotine addiction and nicotine replacement therapy: a review. *Int. J. Tuberc. Lung Dis.* 7, 811–819.
- Lecrux, C., and Hamel, E. (2011). The neurovascular unit in brain function and disease. *Acta Physiol. (Oxf.)* 203, 47–59.
- Lee, Y. W., Hennig, B., Fiala, M., Kim, K. S., and Toborek, M. (2001). Cocaine activates redox-regulated transcription factors and induces TNF- $\alpha$  expression in human brain endothelial cells. *Brain Res.* 920, 125–133.
- Letendre, S., Paulino, A. D., Rockenstein, E., Adame, A., Crews, L., Cherner, M., Heaton, R., Ellis, R., Everall, I. P., Grant, I., and Masliah, E. (2007). Pathogenesis of hepatitis C virus coinfection in the brains of patients infected with HIV. *J. Infect. Dis.* 196, 361–370.
- Letendre, S. L., Cherner, M., Ellis, R. J., Marquie-Beck, J., Gragg, B., Marcotte, T., Heaton, R. K., McCutchan, J. A., and Grant, I. (2005). The effects of hepatitis C, HIV, and methamphetamine dependence on neuropsychological performance: biological correlates of disease. *AIDS* 19(Suppl. 3), S72–S78.
- Liang, H., Wang, X., Chen, H., Song, L., Ye, L., Wang, S. H., Wang, Y. J., Zhou, L., and Ho, W. Z. (2008). Methamphetamine enhances HIV infection of macrophages. *Am. J. Pathol.* 172, 1617–1624.
- Lin, S. J., Hong, C. Y., Chang, M. S., Chiang, B. N., and Chien, S. (1992). Long-term nicotine exposure increases aortic endothelial cell death and enhances transendothelial macromolecular transport in rats. *Arterioscler. Thromb.* 12, 1305–1312.
- Liou, H. H., Hsu, H. J., Tsai, Y. F., Shih, C. Y., Chang, Y. C., and Lin, C. J. (2007). Interaction between nicotine and MPTP/MPP+ in rat brain endothelial cells. *Life Sci.* 81, 664–672.
- Liuzzi, G. M., Mastroianni, C. M., Latronico, T., Mengoni, F., Fasano, A., Lichtner, M., Vullo, V., and Riccio, P. (2004). Anti-HIV drugs decrease the expression of matrix metalloproteinases in astrocytes and microglia. *Brain* 127, 398–407.
- Lockman, P. R., Van der Schyf, C. J., Abbruscato, T. J., and Allen, D. D. (2005). Chronic nicotine exposure alters blood-brain barrier permeability and diminishes brain uptake of methyllycaconitine. *J. Neurochem.* 94, 37–44.
- Madsen, S. J., and Hirschberg, H. (2010). Site-specific opening of the blood-brain barrier. *J. Biophotonics* 3, 356–367.
- Mahajan, S. D., Aalinkeel, R., Sykes, D. E., Reynolds, J. L., Bindukumar, B., Adal, A., Qi, M., Toh, J., Xu, G., Prasad, P. N., and Schwartz, S. A. (2008). Methamphetamine alters blood brain barrier permeability via the modulation of tight junction expression: Implication for HIV-1 neuropathogenesis in the context of drug abuse. *Brain Res.* 1203, 133–148.
- Manda, V. K., Mittapalli, R. K., Bohn, K. A., Adkins, C. E., and Lockman, P. R. (2010a). Nicotine and cotinine increases the brain penetration of saquinavir in rat. *J. Neurochem.* 115, 1495–1507.
- Manda, V. K., Mittapalli, R. K., Geldenhuys, W. J., and Lockman, P. R. (2010b). Chronic exposure to nicotine and saquinavir decreases endothelial Notch-4 expression and disrupts blood-brain barrier integrity. *J. Neurochem.* 115, 515–525.
- Maragos, W. F., Young, K. L., Turchan, J. T., Guseva, M., Pauly, J. R., Nath, A., and Cass, W. A. (2002). Human immunodeficiency virus-1 Tat protein and methamphetamine interact synergistically to impair striatal dopaminergic function. *J. Neurochem.* 83, 955–963.
- Mark, K. S., and Davis, T. P. (2002). Cerebral microvascular changes in permeability and tight junctions induced by hypoxia-reoxygenation. *Am. J. Physiol. Heart Circ. Physiol.* 282, H1485–H1494.
- Martins, T., Baptista, S., Goncalves, J., Leal, E., Milhazes, N., Borges, F., Ribeiro, C. F., Quintela, O., Lendoiro, E., Lopez-Rivadulla, M., Ambrosio, A. F., and Silva, A. P. (2011). Methamphetamine transiently increases the blood-brain barrier permeability in the hippocampus: role of tight junction proteins and matrix metalloproteinase-9. *Brain Res.* 1411, 28–40.
- McCann, U. D., Kuwabara, H., Kumar, A., Palermo, M., Abbey, R., and Brasic, J. (2008). Persistent cognitive and dopamine transporter deficits in abstinent methamphetamine users. *Synapse* 62, 91–100.
- Mitcheson, L., McCambridge, J., Byrne, A., Hunt, N., and Winstock, A. (2008). Sexual health risk among dance drug users: cross-sectional comparisons with nationally representative data. *Int. J. Drug Policy* 19, 304–310.
- Monks, T. J., Jones, D. C., Bai, F., and Lau, S. S. (2004). The role of metabolism in 3,4-(+)-methylenedioxyamphetamine and 3,4-(+)-methylenedioxymethamphetamine (ecstasy) toxicity. *Ther. Drug. Monit.* 26, 132–136.
- Morgan, M. E., and Gibb, J. W. (1980). Short-term and long-term effects of methamphetamine on biogenic amine metabolism in extra-striatal dopaminergic nuclei. *Neuropharmacology* 19, 989–995.
- Morganti-Kossmann, M. C., Lenzlinger, P. M., Hans, V., Stahel, P., Csuka, E., Ammann, E., Stocker, R., Trentz, O., and Kossmann, T. (1997). Production of cytokines following brain injury: beneficial and deleterious for

- the damaged tissue. *Mol. Psychiatry* 2, 133–136.
- Nair, M. P., Saiyed, Z. M., Nair, N., Gandhi, N. H., Rodriguez, J. W., Boukhi, N., Provencio-Vasquez, E., Malow, R. M., and Miguez-Burbano, M. J. (2009). Methamphetamine enhances HIV-1 infectivity in monocyte derived dendritic cells. *J. Neuroimmune Pharmacol* 4, 129–139.
- Nair, M. P., Schwartz, S. A., Mahajan, S. D., Tsiao, C., Chawda, R. P., Whitney, R., Don Sykes, B. B., and Hewitt, R. (2004). Drug abuse and neuropathogenesis of HIV infection: role of DC-SIGN and IDO. *J. Neuroimmunol.* 157, 56–60.
- Napier, T. C., Zhang, J., and Hu, X. T. (2010). *HIV-1 Tat Dysregulates Activity of Rat mPFC Pyramidal Neurons via Up-Regulating L-Type Calcium Channels. Program No. 764.2. 2010 Neuroscience Meeting Planner. San Diego, CA: Society for Neuroscience.*
- Nash, J. F., and Yamamoto, B. K. (1992). Methamphetamine neurotoxicity and striatal glutamate release: comparison to 3,4-methylenedioxymethamphetamine. *Brain Res.* 581, 237–243.
- Nath, A. (2010). Human immunodeficiency virus-associated neurocognitive disorder: pathophysiology in relation to drug addiction. *Ann. N. Y. Acad. Sci.* 1187, 122–128.
- Nath, A., Haughey, N. J., Jones, M., Anderson, C., Bell, J. E., and Geiger, J. D. (2000). Synergistic neurotoxicity by human immunodeficiency virus proteins Tat and gp120: protection by memantine. *Ann. Neurol.* 47, 186–194.
- Nath, A., Hauser, K. F., Wojna, V., Booze, R. M., Maragos, W., Prendergast, M., Cass, W., and Turchan, J. T. (2002). Molecular basis for interactions of HIV and drugs of abuse. *J. Acquir. Immune Defic. Syndr.* 31(Suppl. 2), S62–S69.
- Nath, A., Maragos, W. F., Avison, M. J., Schmitt, F. A., and Berger, J. R. (2001). Acceleration of HIV dementia with methamphetamine and cocaine. *J. Neurovirol.* 7, 66–71.
- Ohtsuki, S. (2004). Physiological function of blood-brain barrier transporters as the CNS supporting and protecting system. *Yakugaku Zasshi* 124, 791–802.
- Orio, L., O'Shea, E., Sanchez, V., Pradillo, J. M., Escobedo, I., Camarero, J., Moro, M. A., Green, A. R., and Colado, M. I. (2004). 3,4-Methylenedioxymethamphetamine increases interleukin-1beta levels and activates microglia in rat brain: studies on the relationship with acute hyperthermia and 5-HT depletion. *J. Neurochem.* 89, 1445–1453.
- O'Shea, E., Sanchez, V., Orio, L., Escobedo, I., Green, A. R., and Colado, M. I. (2005). 3,4-Methylenedioxymethamphetamine increases pro-interleukin-1beta production and caspase-1 protease activity in frontal cortex, but not in hypothalamus, of dark agouti rats: role of interleukin-1beta in neurotoxicity. *Neuroscience* 135, 1095–1105.
- Paulson, J. R., Roder, K. E., McAfee, G., Allen, D. D., Van der Schyf, C. J., and Abbruscato, T. J. (2006). Tobacco smoke chemicals attenuate brain-to-blood potassium transport mediated by the Na,K,2Cl-cotransporter during hypoxia-reoxygenation. *J. Pharmacol. Exp. Ther.* 316, 248–254.
- Paulson, J. R., Yang, T., Selvaraj, P. K., Mdzinarishvili, A., Van der Schyf, C. J., Klein, J., Bickel, U., and Abbruscato, T. J. (2010). Nicotine exacerbates brain edema during in vitro and in vivo focal ischemic conditions. *J. Pharmacol. Exp. Ther.* 332, 371–379.
- Peppiatt, C. M., Howarth, C., Mobbs, P., and Attwell, D. (2006). Bidirectional control of CNS capillary diameter by pericytes. *Nature* 443, 700–704.
- Perdisky, Y., Ho, W., Ramirez, S. H., Potula, R., Abood, M. E., Unterwald, E., and Tuma, R. (2011). HIV-1 infection and alcohol abuse: neurocognitive impairment, mechanisms of neurodegeneration and therapeutic interventions. *Brain Behav. Immun.* 25(Suppl. 1), S61–S70.
- Peterson, P. K., Gekker, G., Chao, C. C., Schut, R., Molitor, T. W., and Balfour, H. H. Jr. (1991). Cocaine potentiates HIV-1 replication in human peripheral blood mononuclear cell cocultures. Involvement of transforming growth factor-beta. *J. Immunol.* 146, 81–84.
- Petty, M. A., and Lo, E. H. (2002). Junctional complexes of the blood-brain barrier: permeability changes in neuroinflammation. *Prog. Neurobiol.* 68, 311–323.
- Price, T. O., Ercal, N., Nakaoka, R., and Banks, W. A. (2005). HIV-1 viral proteins gp120 and Tat induce oxidative stress in brain endothelial cells. *Brain Res.* 1045, 57–63.
- Quinton, M. S., and Yamamoto, B. K. (2006). Causes and consequences of methamphetamine and MDMA toxicity. *AAPS J.* 8, E337–E347.
- Ramirez, S. H., Potula, R., Fan, S., Eidem, T., Papugani, A., Reichenbach, N., Dykstra, H., Weksler, B. B., Romero, I. A., Couraud, P. O., and Persidsky, Y. (2009). Methamphetamine disrupts blood-brain barrier function by induction of oxidative stress in brain endothelial cells. *J. Cereb. Blood Flow Metab.* 29, 1933–1945.
- Ramsauer, M., Krause, D., Dermietzel, R. (2002). Angiogenesis of the blood-brain barrier in vitro and the function of cerebral pericytes. *FASEB J.* 16, 1274–1276.
- Ransohoff, R. M., Kivisakk, P., and Kidd, G. (2003). Three or more routes for leukocyte migration into the central nervous system. *Nat. Rev. Immunol.* 3, 569–581.
- Reynolds, J. L., Mahajan, S. D., Bindukumar, B., Sykes, D., Schwartz, S. A., and Nair, M. P. (2006). Proteomic analysis of the effects of cocaine on the enhancement of HIV-1 replication in normal human astrocytes (NHA). *Brain Res.* 1123, 226–236.
- Ricaurte, G., Bryan, G., Strauss, L., Seiden, L., and Schuster, C. (1985). Hallucinogenic amphetamine selectively destroys brain serotonin nerve terminals. *Science* 229, 986–988.
- Ricaurte, G. A., Forno, L. S., Wilson, M. A., DeLanney, L. E., Irwin, I., Molliver, M. E., and Langston, J. W. (1988). (±)3,4-Methylenedioxymethamphetamine selectively damages central serotonergic neurons in nonhuman primates. *JAMA* 260, 51–55.
- Ricaurte, G. A., Schuster, C. R., and Seiden, L. S. (1980). Long-term effects of repeated methylamphetamine administration on dopamine and serotonin neurons in the rat brain: a regional study. *Brain Res.* 193, 153–163.
- Risdahl, J. M., Khanna, K. V., Peterson, P. K., and Molitor, T. W. (1998). Opiates and infection. *J. Neuroimmunol.* 83, 4–18.
- Rock, R. B., Gekker, G., Aravalli, R. N., Hu, S., Sheng, W. S., and Peterson, P. K. (2008). Potentiation of HIV-1 expression in microglial cells by nicotine: involvement of transforming growth factor-beta 1. *J. Neuroimmune Pharmacol.* 3, 143–149.
- Rodriguez-Gaztelumendi, A., Alvehus, M., Andersson, T., and Jacobsson, S. O. (2011). Comparison of the effects of nicotine upon the transcellular electrical resistance and sucrose permeability of human ECV304/rat C6 co-cultures and human CaCo cells. *Toxicol. Lett.* 207, 1–6.
- Romanelli, R. M., Pinto, J. A., Melo, L. J., Vasconcelos, M. A., and Pereira, R. M. (2006). Effectiveness of dual and triple antiretroviral therapy in the treatment of HIV-infected children. *J. Pediatr. (Rio J)* 82, 260–265.
- Roth, M. D., Tashkin, D. P., Choi, R., Jamieson, B. D., Zack, J. A., and Baldwin, G. C. (2002). Cocaine enhances human immunodeficiency virus replication in a model of severe combined immunodeficient mice implanted with human peripheral blood leukocytes. *J. Infect. Dis.* 185, 701–705.
- Roth, M. D., Whittaker, K. M., Choi, R., Tashkin, D. P., and Baldwin, G. C. (2005). Cocaine and sigma-1 receptors modulate HIV infection, chemokine receptors, and the HPA axis in the huPBL-SCID model. *J. Leukoc. Biol.* 78, 1198–1203.
- Rothman, R. B., and Baumann, M. H. (2003). Monoamine transporters and psychostimulant drugs. *Eur. J. Pharmacol.* 479, 23–40.
- Rothman, R. B., Baumann, M. H., Dersch, C. M., Romero, D. V., Rice, K. C., Carroll, F. I., and Partilla, J. S. (2001). Amphetamine-type central nervous system stimulants release norepinephrine more potently than they release dopamine and serotonin. *Synapse* 39, 32–41.
- Samii, A., Nutt, J. G., and Ransom, B. R. (2004). Parkinson's disease. *Lancet* 363, 1783–1793.
- Sandoval, K. E., and Witt, K. A. (2008). Blood-brain barrier tight junction permeability and ischemic stroke. *Neurobiol. Dis.* 32, 200–219.
- Schep, L. J., Slaughter, R. J., and Beasley, D. M. (2010). The clinical toxicology of metamfetamine. *Clin. Toxicol. (Phila.)* 48, 675–694.
- Schilling, L., Bultmann, A., and Wahl, M. (1992). Lack of effect of topically applied nicotine on pial arteriole diameter and blood-brain barrier integrity in the cat. *Clin. Investig.* 70, 210–217.
- Semple, S. J., Patterson, T. L., and Grant, I. (2004). The context of sexual risk behavior among heterosexual methamphetamine users. *Addict. Behav.* 29, 807–810.
- Sharma, H. S., and Ali, S. F. (2006). Alterations in blood-brain barrier function by morphine and methamphetamine. *Ann. N. Y. Acad. Sci.* 1074, 198–224.
- Sharma, H. S., and Ali, S. F. (2008). Acute administration of 3,4-methylenedioxymethamphetamine induces profound hyperthermia, blood-brain barrier disruption, brain edema formation, and cell injury. *Ann. N. Y. Acad. Sci.* 1139, 242–258.
- Sharma, H. S., and Kiyatkin, E. A. (2009). Rapid morphological brain abnormalities during

- acute methamphetamine intoxication in the rat: an experimental study using light and electron microscopy. *J. Chem. Neuroanat.* 37, 18–32.
- Sharma, H. S., Muresanu, D., Sharma, A., and Patnaik, R. (2009). Cocaine-induced breakdown of the blood-brain barrier and neurotoxicity. *Int. Rev. Neurobiol.* 88, 297–334.
- Sopori, M. L., and Kozak, W. (1998). Immunomodulatory effects of cigarette smoke. *J. Neuroimmunol.* 83, 148–156.
- Speer, P., Zhang, Y., Gu, Y., Lucas, M. J., and Wang, Y. (2002). Effects of nicotine on intercellular adhesion molecule expression in endothelial cells and integrin expression in neutrophils in vitro. *Am. J. Obstet. Gynecol.* 186, 551–556.
- Stewart, P. A., Hayakawa, K., Akers, M. A., and Vinters, H. V. (1992). A morphometric study of the blood-brain barrier in Alzheimer's disease. *Lab. Invest.* 67, 734–742.
- Stolp, H. B., and Dziegielewska, K. M. (2009). Review: Role of developmental inflammation and blood-brain barrier dysfunction in neurodevelopmental and neurodegenerative diseases. *Neuropathol. Appl. Neurobiol.* 35, 132–146.
- Strazza, M., Pirrone, V., Wigdahl, B., and Nonnemacher, M. R. (2011). Breaking down the barrier: the effects of HIV-1 on the blood-brain barrier. *Brain Res.* 1399, 96–115.
- Sulzer, D., Sonders, M. S., Poulsen, N. W., and Galli, A. (2005). Mechanisms of neurotransmitter release by amphetamines: a review. *Prog. Neurobiol.* 75, 406–433.
- Theodore, S., Cass, W. A., and Maragos, W. F. (2006). Methamphetamine and human immunodeficiency virus protein Tat synergize to destroy dopaminergic terminals in the rat striatum. *Neuroscience* 137, 925–935.
- Torres, E., Gutierrez-Lopez, M. D., Mayado, A., Rubio, A., O'Shea, E., and Colado, M. I. (2011). Changes in interleukin-1 signal modulators induced by 3,4-methylenedioxymethamphetamine (MDMA): regulation by CB2 receptors and implications for neurotoxicity. *J. Neuroinflammation* 8, 53.
- Uzum, G., Diler, A. S., and Ziyilan, Y. Z. (1999). Chronic nicotine pre-treatment protects the blood-brain barrier against nicotine-induced seizures in the rat. *Pharmacol. Res.* 40, 263–269.
- Venisnik, K. M. (2000). Nicotine and cotinine modulate in vitro and in situ blood-brain barrier permeability. *Proc. Western Pharm. Soc.* 43, 128.
- Verma, S., and Szmítko, P. E. (2006). The vascular biology of peroxisome proliferator-activated receptors: modulation of atherosclerosis. *Can. J. Cardiol.* 22(Suppl. B), 12B–17B.
- Wang, L., Kittaka, M., Sun, N., Schreiber, S. S., and Zlokovic, B. V. (1997). Chronic nicotine treatment enhances focal ischemic brain injury and depletes free pool of brain microvascular tissue plasminogen activator in rats. *J. Cereb. Blood Flow Metab.* 17, 136–146.
- Wang, L., McComb, J. G., Weiss, M. H., McDonough, A. A., and Zlokovic, B. V. (1994). Nicotine downregulates alpha 2 isoform of Na,K-ATPase at the blood-brain barrier and brain in rats. *Biochem. Biophys. Res. Commun.* 199, 1422–1427.
- Webber, M. P., Schoenbaum, E. E., Gourevitch, M. N., Buono, D., and Klein, R. S. (1999). A prospective study of HIV disease progression in female and male drug users. *AIDS* 13, 257–262.
- Wisor, J. P., Schmidt, M. A., and Clegern, W. C. (2011). Cerebral microglia mediate sleep/wake and neuroinflammatory effects of methamphetamine. *Brain Behav. Immun.* 25, 767–776.
- Yamamoto, B. K., and Bankson, M. G. (2005). Amphetamine neurotoxicity: cause and consequence of oxidative stress. *Crit. Rev. Neurobiol.* 17, 87–117.
- Yamamoto, B. K., and Raudensky, J. (2008). The role of oxidative stress, metabolic compromise, and inflammation in neuronal injury produced by amphetamine-related drugs of abuse. *J. Neuroimmune Pharmacol.* 3, 203–217.
- Yamamoto, Y., Yoshikawa, H., Nagano, S., Kondoh, G., Sadahiro, S., Gotow, T., Yanagihara, T., and Sakoda, S. (1999). Myelin-associated oligodendrocytic basic protein is essential for normal arrangement of the radial component in central nervous system myelin. *Eur. J. Neurosci.* 11, 847–855.
- Yang, T., Roder, K. E., Bhat, G. J., Thekkumkara, T. J., and Abbruscato, T. J. (2006). Protein kinase C family members as a target for regulation of blood-brain barrier Na,K,2Cl-cotransporter during in vitro stroke conditions and nicotine exposure. *Pharm. Res.* 23, 291–302.
- Yang, Y., Yao, H., Lu, Y., Wang, C., and Buch, S. (2010). Cocaine potentiates astrocyte toxicity mediated by human immunodeficiency virus (HIV-1) protein gp120. *PLoS ONE* 5, e13427. doi:10.1371/journal.pone.0013427
- Yao, H., Kim, K., Duan, M., Hayashi, T., Guo, M., Morgello, S., Prat, A., Wang, J., Su, T. P., and Buch, S. (2011). Cocaine hijacks sigma1 receptor to initiate induction of activated leukocyte cell adhesion molecule: implication for increased monocyte adhesion and migration in the CNS. *J. Neurosci.* 31, 5942–5955.
- Yenari, M. A., and Han, H. S. (2012). Neuroprotective mechanisms of hypothermia in brain ischaemia. *Nat. Rev. Neurosci.* 13, 267–278.
- Yong, T., Zheng, M. Q., and Linthicum, D. S. (1997). Nicotine induces leukocyte rolling and adhesion in the cerebral microcirculation of the mouse. *J. Neuroimmunol.* 80, 158–164.
- Zhang, L., Looney, D., Taub, D., Chang, S. L., Way, D., Witte, M. H., Graves, M. C., and Fiala, M. (1998). Cocaine opens the blood-brain barrier to HIV-1 invasion. *J. Neurovirol.* 4, 619–626.
- Zhang, X., Banerjee, A., Banks, W. A., and Ercal, N. (2009). N-Acetylcysteine amide protects against methamphetamine-induced oxidative stress and neurotoxicity in immortalized human brain endothelial cells. *Brain Res.* 1275, 87–95.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 16 April 2012; accepted: 06 June 2012; published online: 29 June 2012.

Citation: Kousik SM, Napier TC and Carvey PM (2012) The effects of psychostimulant drugs on blood brain barrier function and neuroinflammation. *Front. Pharmacol.* 3:121. doi: 10.3389/fphar.2012.00121

This article was submitted to *Frontiers in Neuropharmacology*, a specialty of *Frontiers in Pharmacology*.

Copyright © 2012 Kousik, Napier and Carvey. This is an open-access article distributed under the terms of the Creative Commons Attribution Non Commercial License, which permits non-commercial use, distribution, and reproduction in other forums, provided the original authors and source are credited.