



Meristem identity and phyllotaxis in inflorescence development

Madelaine E. Bartlett^{1*} and Beth Thompson²

¹ Biology Department, University of Massachusetts Amherst, Amherst, MA, USA

² Biology Department, East Carolina University, Greenville, NC, USA

Inflorescence morphology is incredibly diverse. This diversity of form has been a fruitful source of inquiry for plant morphologists for more than a century. Work in the grasses (Poaceae), the tomato family (Solanaceae), and *Arabidopsis thaliana* (Brassicaceae) has led to a richer understanding of the molecular genetics underlying this diversity. The character of individual meristems, a combination of the number (determinacy) and nature (identity) of the products a meristem produces, is key in the development of plant form. A framework that describes inflorescence development in terms of shifting meristem identities has emerged and garnered empirical support in a number of model systems. We discuss this framework and highlight one important aspect of meristem identity that is often considered in isolation, phyllotaxis. Phyllotaxis refers to the arrangement of lateral organs around a central axis. The development and evolution of phyllotaxis in the inflorescence remains underexplored, but recent work analyzing early inflorescence development in the grasses identified an evolutionary shift in primary branch phyllotaxis in the Pooideae. We discuss the evidence for an intimate connection between meristem identity and phyllotaxis in both the inflorescence and vegetative shoot, and touch on what is known about the establishment of phyllotactic patterns in the meristem. Localized auxin maxima are instrumental in determining the position of lateral primordia. Upstream factors that regulate the position of these maxima remain unclear, and how phyllotactic patterns change over the course of a plant's lifetime and evolutionary time, is largely unknown. A more complete understanding of the molecular underpinnings of phyllotaxis and architectural diversity in inflorescences will require capitalizing on the extensive resources available in existing genetic systems, and developing new model systems that more fully represent the diversity of plant morphology.

Edited by:

Neelima Roy Sinha, University of California, Davis, USA

Reviewed by:

Naomi Nakayama, University of Edinburgh, UK
Toshi Marie Foster, Plant and Food Research, New Zealand
Andrea Gallavotti, Rutgers University, USA

*Correspondence:



Madelaine E. Bartlett is currently working as an assistant professor in the Biology department at the University of Massachusetts Amherst. The Bartlett lab is working on understanding the evolution and development of flowers and inflorescences, particularly in the monocots. We are focused on uncovering the molecular genetic underpinnings of floral development, and how these molecular networks have been altered over evolutionary time.
mbartlett@bio.umass.edu

Keywords: inflorescence, phyllotaxis, inflorescence morphology, auxin, meristem identity, inflorescence evolution

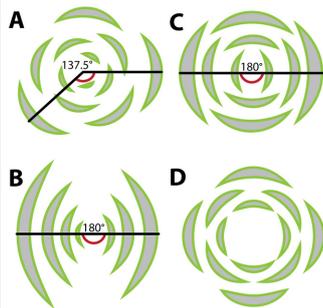
INTRODUCTION

Inflorescence morphology is an important determinant of yield in agricultural settings, and fitness in natural ones (Wyatt, 1982; Harder et al., 2004). Inflorescence form shows a startling degree of diversity. This diversity is evident at both broad scales, such as across angiosperms, and at finer scales, such as in the grass family (Weberling, 1989; Vegetti and Anton, 2000).

Although all grass inflorescences are termed “panicles,” this descriptor belies the immense diversity within the family (**Box 1**). A recent paper from Kellogg et al. highlights the evolution of one aspect of

Box 1 | A terminology primer.

Phyllotaxis and inflorescence morphology have both been the subject of much study and discussion, both recently and historically. Both aspects of plant architecture show considerable diversity across the plant kingdom. The combination of long study, naturally occurring diversity, and morphological complexity has led to a proliferation of terminology and, particularly in the case of inflorescence morphology, confusion. For the purposes of this review, we have defined only the most basic terms we feel are essential for understanding the work we discuss. For more in-depth discussion, we direct readers to Weberling, 1989, Prenner et al, 2009, and Endress, 2010, and references therein. For simplicity, not all leaves (e.g. bracts, prophylls) are shown in E - H.

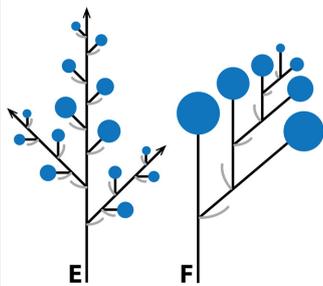
**Phyllotactic patterns discussed in this review**

A. Spiral. Primordia are initiated one at a time at a certain angle distant from the last primordium formed, often 137.5° .

B. Distichous. Successive primordia are initiated one at a time at an angle of 180° . Distichous phyllotaxis represents a subset of a more general pattern: two-ranked. In two-ranked phyllotaxis, two rows of leaves develop, but the two rows may not lie exactly 180° apart.

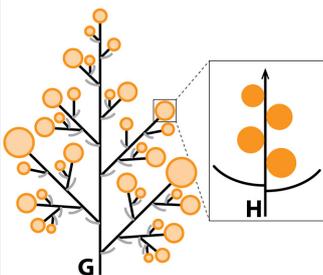
C. Opposite decussate. Primordia are initiated at an angle of 180° from one another, two at a time. Successive pairs of primordia are initiated at right angles to the previous pair.

D. Whorled. Characteristic of many flowers, in whorled phyllotaxis groups of three or more organs are initiated at once. In flowers, organs may all be initiated simultaneously, as is common for the petal whorls of many eudicots, or they may be initiated in very quick succession, often in a spiral (Endress, 1987).

**Inflorescence branching can be divided into two main classes**

E. Racemose branching. In racemes, whether simple or compound (shown here), flowers are formed in the axils of leaves (bracts, shown in grey, suppressed in some taxa) produced by the terminal inflorescence meristem. There is a continuous main axis (monopodial growth).

F. Cymose branching. In a cyme, the apical meristem forms a terminal flower, and either one or two axillary buds and their subtending leaves (prophylls). In an iterative pattern of development, the axillary buds themselves form terminal flowers and more axillary buds. There is no single main axis, but rather a sequence of axes terminating in flowers (sympodial growth). Cymes are named according to the number and arrangement of the axillary buds and their subtending leaves.

**Inflorescences in the grasses**

Grass inflorescences are panicles (**G**) with groups of flowers borne in spikelets as the terminal units (**H**) in all but the earliest-diverging lineages. The spikelet-enclosing bracts are termed glumes (shown in black in **H**). Branching in panicles is neither strictly racemose nor strictly cymose. By definition, the main and lateral axes of panicles all terminate in flowers, and the degree of branching increases towards the base of the inflorescence. In the grass family, branching occurs from the axils of suppressed bracts (shown in grey in **G**). Both panicles and spikelets can vary considerably from the generalized structures shown here. For example, grass panicles can vary in terms of degree of branching and/or in whether the inflorescence axes terminate in flowers, or as blind ends (Vegetti and Anton, 2000; Kellogg, 2013). Spikelets can vary in the number, arrangement, and/or sexuality of the flowers (florets) they contain. In some grasses (e.g. the Andropogoneae), the spikelets themselves are arranged into characteristic higher-order groupings.

inflorescence morphology in the grasses: the evolution of primary branch **phyllotaxis** (Kellogg et al., 2013). Through investigations of early inflorescence development in grass relatives, the authors establish that the ancestral primary branch phyllotaxis in the grass inflorescence was likely spiral. Most extant grasses still exhibit

spiral phyllotaxis of primary branches, but there has been an evolutionary shift in the Pooideae. The earliest-diverging member of the Pooideae, *Brachyelytrum*, shows spiral phyllotaxis of its primary inflorescence branches. Following the divergence of *Brachyelytrum*, there is a shift to two-ranked phyllotaxis, with

KEY CONCEPT 1 | Phyllotaxis

The geometric arrangement of organs around a central axis. Phyllotactic patterns are determined largely by auxin and cytokinin gradients in the meristem. Altered hormone activity likely underlies developmental shifts in phyllotaxis.

an angle of divergence less than 180°, and then a final evolutionary transition to true distichy (**Box 1B**) in the later diverging Pooids (including *Hordeum*, *Avena*, and *Brachypodium*). This paper highlights two phyllotactic shifts, one at the evolutionary level (spiral to distichous) and another at the developmental level (from vegetative to reproductive development).

The development of inflorescence morphology involves a complex interplay between a number of interacting processes: establishment of phyllotaxis and meristem identity, as well as variable patterns of cell division and expansion. As in all plant development, meristems and meristematic cells are of key importance in inflorescence development. Meristems are pools of undifferentiated cells that generate both more meristematic cells, as well as differentiated cells that form lateral organs. The particular character of a meristem can be described using the interrelated concepts of meristem identity and determinacy. **Meristem identity** refers to the type of primordia produced, whereas **meristem determinacy** refers to the number of primordia produced and whether or not the meristem is consumed in the production of primordia.

KEY CONCEPT 2 | Meristem identity and meristem determinacy

Meristem identity refers to the type of primordia produced by a meristem, whereas meristem determinacy refers to the number of primordia produced and whether or not the meristem is consumed in the production of primordia.

For example, a meristem has floral meristem identity if it initiates floral organs and floral meristems are determinate because they are consumed in the production of a limited number of organ primordia. Each one of these factors can vary across both developmental and evolutionary time, leading to adult **inflorescence architectures** that show vast morphological diversity.

KEY CONCEPT 3 | Inflorescence architecture

The three dimensional arrangement of branches, leaves, flowers, and floral buds in an inflorescence (**Box 1**). Inflorescence architecture is determined in large part by shifting meristem identities over developmental time.

As highlighted by the Kellogg et al. (2013) paper, there is a need for understanding inflorescence morphology in terms of development, evolution, and at the intersection of these fields. There are a few examples where the adaptive value of individual aspects of inflorescence morphology have been investigated, but a very small number of systematic investigations into evolutionary patterns. This work was reviewed recently (Harder and Prusinkiewicz, 2013), and we will not revisit that topic here. Computational modeling and molecular genetic studies of inflorescence development in the grasses, petunia, tomato, and *Arabidopsis*, has led to greater understanding of the regulatory logic that underlies inflorescence development (Rijkema et al., 2006; Prusinkiewicz et al., 2007; Castel et al., 2010; Pautler et al., 2013; Tanaka et al., 2013; Zhang and Yuan, 2014). Similarly, strides have been made into understanding the molecular underpinnings of phyllotaxis,

particularly in *Arabidopsis* (Kuhlemeier, 2007; Sassi and Vernoux, 2013; Traas, 2013). What has been missing is integration between these two closely allied topics.

Here, we attempt to seat what we know about the molecular and genetic factors that control phyllotaxis within the reigning paradigm of how we understand inflorescence development. We discuss recent investigations of inflorescence development in terms of shifting meristem identities, establishment of phyllotaxis and shifting phyllotactic patterns over the lifetime of a plant, and the close link between meristem identity and phyllotaxis.

SHIFTING MERISTEM IDENTITIES DURING INFLORESCENCE DEVELOPMENT SHAPE INFLORESCENCE ARCHITECTURE

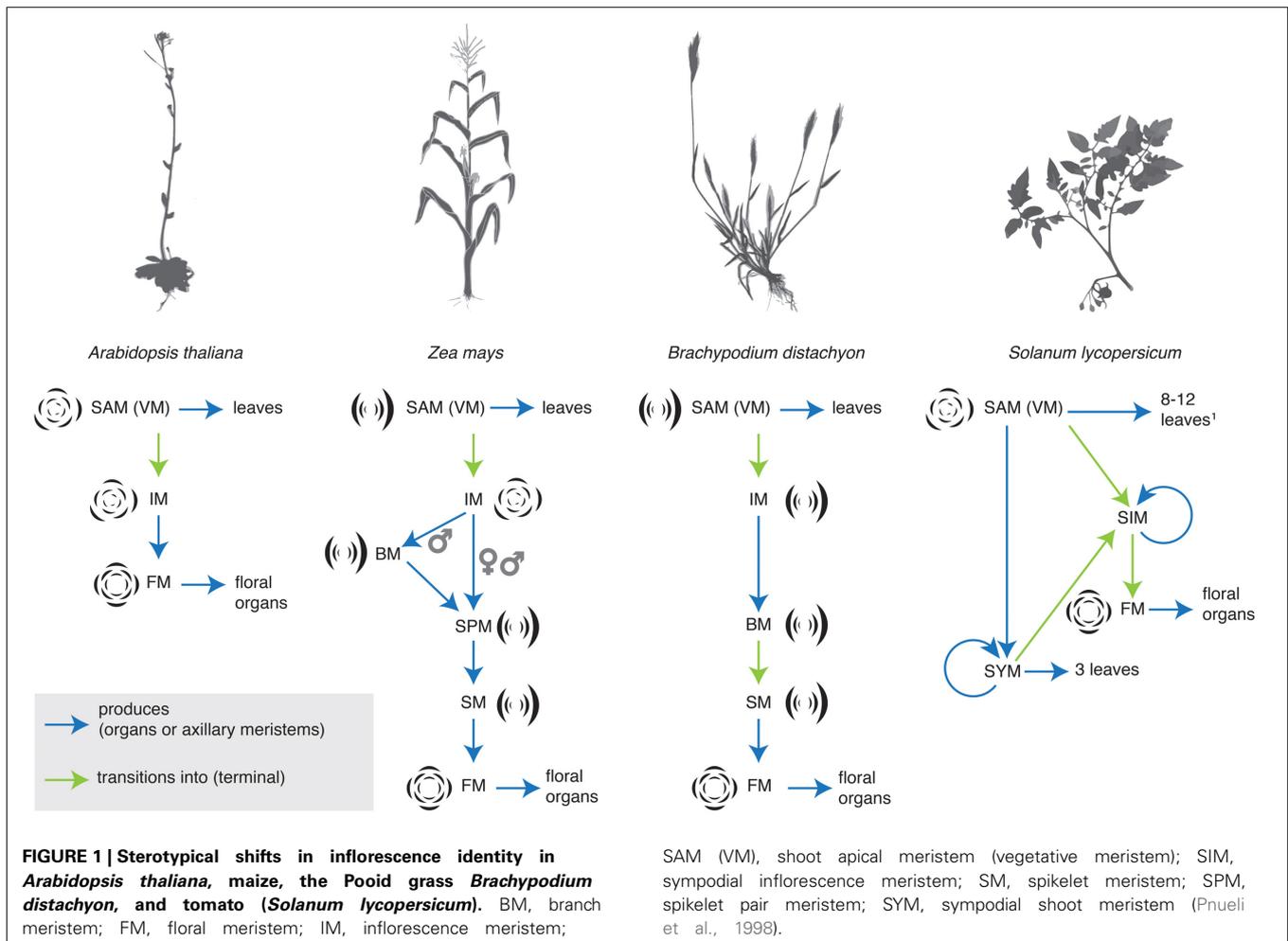
Meristem identity changes during plant development. One key change in meristem identity is at the transition to flowering, where the vegetative meristem (VM) or shoot apical meristem (SAM) transitions to an inflorescence meristem (IM). A conceptual framework is emerging for understanding inflorescence development after this point in terms of further transitions between meristem identities. Computational modeling and molecular analyses of inflorescence development in the grasses, tomato (*Solanum lycopersicum*), petunia (*Petunia hybrida*), *Arabidopsis thaliana*, and *Pisum sativum* (Fabaceae) have revealed a framework of progressive changes in meristem identity underlying morphology.

An early model (Kellogg, 2000) portrayed inflorescence development as an iterative series of developmental switches that controlled the transition between meristem identities. In later computational modeling experiments, Prusinkiewicz et al. (2007) proposed a model wherein competency to flower is specified by a particular meristem's degree of *vegetativeness* (*veg*). For a meristem to transition to floral meristem fate, *veg* must drop below a threshold. *veg* in individual meristems could be specified by two major regulators of inflorescence development in *Arabidopsis*: *LEAFY* (*LFY*) and *TERMINAL FLOWER1* (*TFL1*). *LFY* and *TFL1* are transcription factors with roughly opposite roles. *LFY* promotes the transition from inflorescence to determinate floral meristem fate, while *TFL1* acts to repress floral meristem fate, effectively maintaining indeterminacy of the IM (Weigel et al., 1992; Bradley et al., 1997; Ratcliffe et al., 1999; Moyroud et al., 2010). In their model, *LFY* decreases *veg*, while *TFL* activity increases *veg*. Thus, variation in *veg*, a measure of meristem identity, acts over developmental time and morphological space to produce final inflorescence architecture (Prusinkiewicz et al., 2007). Common to both models (Kellogg, 2000; Prusinkiewicz et al., 2007) is the concept that variable meristem identity across the inflorescence is key in generating final structure. This concept continues to garner empirical support from work conducted in the grasses, the tomato family, and *Pisum*.

In the grasses maize and rice, studies of molecular genetics have revealed a regimented and precise transition between meristem identities during inflorescence development (Tanaka et al., 2013; Thompson, 2014). The grasses bear their flowers in spikelets (**Box 1**). Grass inflorescences undergo a series of meristem transitions, generalized as branch (BM), to spikelet (SM), to floral meristem (FM) identity, although there are variations between taxa and male and female inflorescences. Maize, and

its allied grasses in the Andropogoneae, bear their spikelets in pairs, and thus possess a separate spikelet pair meristem (SPM) identity (Figure 1). Over the years, many key developmental regulators that control these transitions have been identified. The developmental regulators have been described and cataloged in a number of recent reviews, and so we do not discuss them here at length (reviewed in Pautler et al., 2013; Tanaka et al., 2013; Teo et al., 2014; Thompson, 2014). Meristem identity in the inflorescence is controlled by a sequentially active series of genes. Some genes and gene functions are conserved between maize and rice, while others appear unique to a subset of grasses. For example, both *BRANCHED SILKLESS1* (*BD1*) and its rice ortholog, *FRIZZY PANICLE1* (*FZP1*) regulate SM identity and determinacy. In *bd1* and *fzp1* mutants, SMs take on a more BM-like identity (Chuck et al., 2002; Komatsu et al., 2003). In contrast, the *RAMOSA* (*RA*) genes are key regulators of SPM (*RA1*, *RA2*, *RA3*) and SM (*RA3*) determinacy (Vollbrecht et al., 2005; Bortiri et al., 2006; Satoh-Nagasawa et al., 2006). Rice has no ortholog of *RA1*, indeed *RA1* appears to be unique to the Andropogoneae (Vollbrecht et al., 2005). The homologs of *RA2* and *RA3* have not been characterized in rice, but the barley *RA2* ortholog, *Vrs4*, appears to play an analogous role in regulating meristem determinacy in the inflorescence (Koppolu et al., 2013).

In the Solanaceae (tomato, peppers, and petunia), several genetic mutants that disrupt inflorescence architecture have been characterized and cloned (reviewed most recently in Cohen et al., 2012; Park et al., 2014). Transcriptional evidence also supports the concept of shifting meristem identity in tomato inflorescences (Park et al., 2012). The branching pattern in tomato is cymose (but see Prenner et al., 2009) and the progression from inflorescence to floral meristem occurs iteratively across the inflorescence through a stereotypical progression of meristem identities (Figure 1). Transcriptional dynamics across this progression reveal that these meristems are distinct both in their products and in their molecular profiles. Variation in meristem identity across the inflorescence, expressed as differing maturation states, may be key in generating final inflorescence architecture, both in *Solanum* and in the broader Solanaceae (Park et al., 2012, 2014). One important point arising from this work (Park et al., 2012), and transcriptional profiling of *ra* mutants in maize (Eveland et al., 2014) is that the molecular sequence of meristem identities is much more fine-grained than the morphologically identifiable sequence of identities. Morphologically distinct meristem identities serve as useful markers, but they overlie a much larger degree of change that occurs at the molecular level.



The first IM identity gene was recently cloned in the legume *P. sativum*. *Pisum*, and indeed most legumes, bears its flowers in compound racemes (**Box 1E**). The IM produces secondary inflorescence meristems that ultimately produce flowers. *vegetative1* (*veg1*) plays a role in specifying secondary inflorescence meristem identity. In *veg1* mutants, VMs form in the position of secondary inflorescence meristems, which never produce flowers. *VEG1* encodes an AGL79-like MADS-box gene that appears to be eudicot-specific (Berbel et al., 2012).

Apart from meristem identity, positional effects within an inflorescence have long been known to play a role in determining architecture. By studying inflorescence morphology in thousands of species, Troll inferred a generalized inflorescence architecture composed of four zones (the innovation, inhibition, enrichment, and terminal zones), occupying distinct regions of the inflorescence (Troll, 1969; Weberling, 1989). The concept of distinct inflorescence zones is supported by studies of both genetic mutants, and natural diversity. For example, in *ral* mutants, later meristems don't have the capacity to respond to the lack of *ral* in the same way as earlier ones do, leading to a grade of branching in the inflorescence (Vollbrecht et al., 2005). In the Solanaceae, inflorescence position also plays a key role in determining floral meristem fate and floral sexuality (Diggle and Miller, 2004). The molecular nature of these positional signals remains unknown (transcriptional profiling experiments are giving hints, see Eveland et al., 2014). What seems clear, however, is that positional information along the proximo-distal inflorescence axis interacts with meristem identity networks to generate inflorescence architecture.

Modulation of meristem identity allows for morphological differentiation in the inflorescence (Kellogg, 2007). A striking example of this can be seen in plants that produce vegetative plantlets termed "bulbils" in place of flowers in their inflorescences (Elmqvist and Cox, 1996). In *Titanotrichum oldhamii* (Gesneriaceae), for example, later "floral" meristems are replaced with bulbils under short day conditions. A *LFY* homolog is down-regulated in these bulbils, as expected for a transition to VM identity (Wang et al., 2004). Taxa where bulbils form often live in alpine, arctic, or arid habitats. Thus, this innovation in inflorescence architecture might represent an adaptation to a short growing season, or to the heterogeneity characteristic of these environments (Elmqvist and Cox, 1996). A more familiar example of the reversion of a reproductive meristem to a vegetative meristematic state can be found in pineapples (*Ananas comosus*, Bromeliaceae), where vegetative growth continues after flowering (Bell and Bryan, 1991).

These examples illustrate the lability of meristem identity under certain conditions. Altering and repurposing existing meristem identity networks is one pathway evolution can take within a clade (e.g., bulbils, inflorescence diversity in the Solanaceae) (Park et al., 2014). Between clades, unique meristem identities seem to be specified by a mix of both unique and conserved genes. Some of the same genes were identified in transcriptional profiling of both *ra* mutants in maize, and in tomato inflorescences (e.g., TCP transcription factors and *ga-2 oxidase*) (Park et al., 2012; Eveland et al., 2014). However, a number of the upstream regulators seem unique to specific clades

(e.g., *ral* and the SPM in maize, *veg1* and the secondary inflorescence meristem in *Pisum*). As researchers sample further there may be more examples of genetic conservation, but we need more genetic models, throughout the angiosperms, to find the genes. For example, thyrses (cymose branching on a racemose main axis, Endress, 2010) are fairly common, yet remain unmentioned in the developmental genetic literature.

PHYLLOTAXIS IS ONE ASPECT OF MERISTEM IDENTITY

Meristem identity and phyllotaxis are intimately linked. This link has been revealed by studies of both natural diversity and genetic mutants. In *Arabidopsis*, the grasses, and *Solanum*, the meristem identity transitions that determine inflorescence architecture are often accompanied by changes in phyllotaxis (**Figure 1**). In the grasses, for example, the VM initiates leaves in a distichous phyllotaxis, IMs produce branches spirally (except in the Pooideae, see above Kellogg et al., 2013), branch meristems produce either branches or spikelets in two ranks, and floral organs are patterned in a whorled phyllotaxis (although they may be initiated spirally, see **Box 1**). The main axis eventually switches to producing spikelets in two ranks (**Box 1**), except in the maize tassel, where the main axis continues to produce spikelet pairs spirally (Kellogg et al., 2013).

One of the phyllotactic shifts observed in grasses is common to many flowering plants. The transition from IM identity (indeterminate, initiates floral meristems) to floral meristem identity (determinate, initiates floral organs) is often accompanied by a shift to whorled phyllotaxis (**Figure 1**) (Bell and Bryan, 1991; but see Endress, 1987; Specht and Bartlett, 2009). Earlier in development, dicotyledonous plants often undergo a shift in phyllotaxis from opposite decussate to spiral phyllotaxis because cotyledons and the first leaves are initiated as opposite pairs (Bell and Bryan, 1991). Furthermore, mutants or double mutants that significantly perturb meristem fate often cause a dramatic shift in phyllotaxis. For example, in *Arabidopsis lfy* mutants and in *Antirrhinum floricaula* (*flo*) and *squamosa* (*squ*) mutants, floral meristems are converted to IMs, and whorled floral phyllotaxis is replaced by spiral phyllotaxis (Coen et al., 1990; Huijser et al., 1992; Weigel et al., 1992; Carpenter et al., 1995). Similarly, the maize *bde zag1* and *ifa1 ids* double mutants cause a determinate meristem (FM or SM) to revert to an indeterminate branch-like meristem, and have BM-like phyllotaxis, rather than FM-like whorled phyllotaxis (Laudencia-Chingcuanco and Hake, 2002; Thompson et al., 2009).

These meristem identity and phyllotactic shifts must be tied into the regulatory pathways that regulate the transition from juvenile to adult growth, and in turn from vegetative to reproductive growth. The juvenile to adult transition in plants is largely controlled by the activity of two opposing microRNAs (miRNAs) (Wu and Poethig, 2006; Chuck et al., 2007; Wang et al., 2011; Zhang et al., 2011; Fu et al., 2012; Shikata et al., 2012; Xie et al., 2012; Poethig, 2013). MiR156 promotes juvenile development, while miR172 promotes reproductive development. Mutants that perturb this network also perturb phyllotaxis. For example, the dominant maize mutant *Corngrass1* (*Cg1*) overexpresses miR156 and extends the juvenile phase. *Cg1* inflorescences, while still spiral, have abnormal phyllotaxis and do not produce ordered rows

of meristems (Chuck et al., 2007). These phyllotactic transitions governed by phase transitions can also be observed by studying natural diversity. In *Eucalyptus globulus*, for example, the transition from juvenile to adult growth is accompanied by a shift from opposite decussate to spiral phyllotaxis (Zotz et al., 2011).

Meristem determinacy mutants also affect phyllotaxis. Mutants that increase meristem indeterminacy due to increased stem cell activity often have abnormal phyllotaxis (e.g., *unusual floral organs (ufo)* in *Arabidopsis*, *bearded-ear1 (bde)* in maize, and *OsMads3/53* in rice), albeit these mutants usually do not exhibit a discrete shift from one phyllotactic pattern to another (Levin and Meyerowitz, 1995; Thompson et al., 2009; Dreni et al., 2011). Meristem determinacy is impacted not only by stem cell activity, but also by cell partitioning to the three components of the phytomer (leaf/bract/prophyll, axillary meristem, and stem). The links between cell partitioning, determinacy and phyllotaxis are illustrated by analysis of the maize *tasselsheath4 (tsh4)* mutant. *tsh4* suppresses bract outgrowth in the inflorescence and in *tsh4* mutants, cells that would normally be partitioned to the axillary meristem are partitioned to the bract. This reallocation of cells results in increased meristem determinacy (fewer lateral primordia are produced), as well as disruption of phyllotaxis in the ear (Chuck et al., 2010). Intriguingly, *tsh4* is targeted by miR156, which as mentioned above plays a fundamental role in the juvenile to adult transition. *tsh4* also regulates the meristem determinacy gene *ra2*. In *tsh4* mutants, expression of RA2 protein is expanded outside of its normal domain in the SPM, into the de-repressed bract (Chuck and Bortiri, 2010).

Together, these analyses highlight the **integrated nature of plant development**. Developmental programs that regulate meristem determinacy and phyllotaxis cannot be considered independently of meristem identity. Determinacy and phyllotactic pattern are not imposed on a particular meristem, but are rather intrinsic to a particular meristem. In addition, these examples illustrate that the programs that regulate life span progression must also be tied into phyllotactic programs. Recently, great progress has been made in understanding the molecular underpinnings of these phyllotactic patterns.

KEY CONCEPT 4 | The integrated nature of meristem development

Meristem identity, meristem determinacy, and phyllotaxis are intimately linked. Regulation of meristem fate and phyllotaxis appear to be linked into the developmental programs that control life progression.

AUXIN AND MOLECULAR MODELS OF PHYLLOTAXIS

The robust and reproducible nature of phyllotaxis has intrigued and fascinated biologists, mathematicians, and physicists for centuries (Adler et al., 1997). Numerous models have been proposed to describe phyllotaxis. Central to these models is the idea that emerging primordia create an inhibitory field that suppresses the formation of new primordia (Hofmeister, 1868; Snow and Snow, 1962; Douady and Couder, 1996; Adler et al., 1997). The position of the next primordium is therefore determined by where the inhibitory field is lowest. This concept has been confirmed by microsurgery and laser ablation experiments that disrupt primordia and also phyllotaxis (Snow and Snow, 1932; Reinhardt et al., 2005).

Work conducted in the past decade has greatly increased our understanding of the molecular mechanisms that regulate primordium initiation and phyllotaxis. The phytohormone auxin has emerged as a central player in the regulation of both processes (for detailed reviews see Kuhlemeier, 2007; Sassi and Vernoux, 2013; Traas, 2013). Auxin is required to initiate all lateral primordia. Mutants that disrupt auxin synthesis or transport have reduced numbers or completely lack lateral primordia in *Arabidopsis*, petunia, and maize (Okada et al., 1991; Galweiler et al., 1998; Tobeña-Santamaria et al., 2002; Friml et al., 2004; Cheng et al., 2006; McSteen et al., 2007; Gallavotti et al., 2008a; Phillips et al., 2011). At least in some contexts, primordium initiation defects can be rescued by application of exogenous auxin, indicating that auxin plays an instructive role in the initiation of lateral primordia (Reinhardt et al., 2000). Indeed, lateral primordia initiate at regions of high auxin, and high auxin activity (as measured by the synthetic auxin reporter, *DR5*) coincides with primordium initiation (Heisler et al., 2005; Gallavotti et al., 2008b; O'Connor et al., 2014).

The mechanisms by which auxin maxima are formed are complex and include both auxin synthesis and auxin transport. Central players in auxin localization are the PIN-FORMED (PIN) auxin efflux carriers, of which PIN1 appears to be most critical for initiation of lateral primordia during reproductive development (Okada et al., 1991; Galweiler et al., 1998; Reinhardt et al., 2000; Vernoux et al., 2000). PIN1 is asymmetrically localized in the cell in response to auxin. During the initiation of lateral primordia, PIN1 forms convergence points that are instrumental in creating auxin maxima. In this developmental context, PIN1 is asymmetrically localized on cell membranes, and is proposed to pump auxin up a concentration gradient to ultimately form auxin maxima (Heisler et al., 2005; O'Connor et al., 2014). In addition to determining the position of primordia, these auxin maxima function as auxin sinks and deplete the surrounding cells of auxin. Polarized auxin transport thus provides mechanistic support for the “inhibitory field” hypothesis, and explains why two primordia cannot form adjacent to one another (Reinhardt et al., 2003; Reinhardt, 2005; De Reuille et al., 2006).

Auxin transport and regulation of phyllotaxis is undoubtedly more complex, as evidenced by the existence of multiple PIN1-like proteins in multiple contexts (Bennett et al., 2014a). In *Arabidopsis*, PIN1 is the main PIN that functions in the SAM (Benková et al., 2003; Reinhardt et al., 2003; Scarpella et al., 2006; Guenot et al., 2012). Outside of the Brassicaceae, Sister of PIN1 (SoPIN1, absent from the Brassicaceae) seems to have a more important role in auxin distribution on the meristem surface (O'Connor et al., 2014). In the grasses, PIN1 has been duplicated, resulting in three PIN proteins that regulate auxin distribution in developing inflorescences. In *Brachypodium* and maize, SoPIN1 is localized to the surface, while PIN1a and PIN1b are more prominent in internal tissues and likely function in vasculature formation. A model that successfully recapitulates experimental evidence varies how each PIN transporter responds to auxin (O'Connor et al., 2014). For example, SoPIN1 moves auxin up the gradient and PIN1a and PIN1b canalize auxin flow by transporting auxin in the direction of greatest auxin flux. Thus, the number, localization, and activity of PIN

proteins are likely important drivers in the positioning of auxin maxima, and perhaps in the evolution of different phyllotactic patterns.

Changes in PIN localization and auxin signaling pathways also have the potential to alter auxin activity and phyllotaxis during development. Very little is known about the molecular mechanisms by which PIN proteins are localized in response to auxin (Michniewicz et al., 2007; Robert et al., 2010), which is a major limitation in generating biologically meaningful auxin transport models (Bennett et al., 2014b). Furthermore, not all cells are competent to respond to auxin, and localization of auxin signaling molecules might also play a role in phyllotaxis. For example, cells within the central zone have high auxin levels, but do not respond to auxin and activate auxin reporter genes (De Reuille et al., 2006; Brunoud et al., 2012).

Auxin synthesis also plays an important role in the initiation of lateral primordia. In maize, *vanishing tassels2* and *sparse inflorescence1* encode auxin biosynthesis enzymes and both *vt2* and *spi1* mutant inflorescences have dramatically reduced numbers of lateral primordia (Gallavotti et al., 2008a; Phillips et al., 2011). Interestingly, although both *vt2* and *spi1* function in auxin biosynthesis, they are not co-expressed in the same domain. *vt2* is expressed in the epidermal layer of initiating primordia, prior to any morphological outgrowth, whereas *spi1* is expressed throughout incipient SPMs, and has restricted expression in the SM and FM. Thus, *vt2* and *spi1* appear to make unique contributions to auxin levels and primordium initiation in developing inflorescences.

Players upstream and downstream of auxin are starting to be identified. The PLETHORA (PLT) transcription factors are required for spiral phyllotaxis in the *Arabidopsis* shoot and are required to upregulate two auxin biosynthesis genes (*YUC1* and *YUC4*) in the central zone of the SAM (Pinon et al., 2013). As mentioned above, *Arabidopsis* goes through a phyllotactic shift common to many dicotyledonous plants. The cotyledons and first two leaves of *Arabidopsis* are initiated in an opposite decussate phyllotaxis and all subsequent leaves are initiated spirally. *plt3plt5plt7* triple mutants delay this phyllotactic shift (Prasad et al., 2011). Most models of phyllotaxis only take into account auxin transport (De Reuille et al., 2006; Jönsson et al., 2006; Smith et al., 2006), but analysis of auxin biosynthesis mutants such as *plt* and *yuc*, indicate that auxin biosynthesis likely plays a significant role (Cheng et al., 2006; Pinon et al., 2013). Along this line, the INDETERMINATE DOMAIN (IDD) transcription factors IDD14, IDD15, and IDD16 have recently been implicated as directly regulating both *PIN1* and *YUC5* in the combinatorial control of auxin signaling and organ growth in *Arabidopsis* (Cui et al., 2013).

Auxin promotes organogenesis at least in part by affecting cell wall rigidity. Demethylesterification of pectin, which increases elasticity in the meristem, is necessary and sufficient for primordium initiation. Overexpression of pectin methylesterase 5 results in increased numbers of lateral primordia, with aberrant phyllotaxis (Peaucelle et al., 2008, 2011). Auxin promotes organ outgrowth at least in part by promoting pectin demethylesterification, since PME13 overexpression lines (which block demethylesterification) have pin-like inflorescences that

fail to form primordia in response to exogenous auxin (Braybrook and Peaucelle, 2013). *PIN1* also seems to respond to mechanical cues in the meristem. *PIN1* localization is correlated with the direction of microtubule arrays, and genetic or chemical disruption of the cell wall disrupts *PIN1* localization (Heisler et al., 2010; Braybrook and Peaucelle, 2013). Thus, differences in cell wall composition and mechanical forces within the meristem may also play an instructive role in creating auxin maxima and determining phyllotaxis. Mechanical forces have a demonstrated effect on the appearance of phyllotactic pattern post-initiation. Lateral primordia may be initiated in one phyllotactic pattern, but twisting of the stem after initiation can lead to an altered final leaf position (Bell and Bryan, 1991; Landrein et al., 2013).

In addition to auxin, the phytohormone cytokinin plays a critical role in the regulation of meristem size and phyllotaxis. The maize *aberrant phyllotaxy1* (*abph1*) and the rice *decussate* (*dec*) mutants stably change leaf phyllotaxis from distichous (alternate) to opposite decussate (Jackson and Hake, 1999; Itoh et al., 2012). Both *abph1* and *dec* mutants are defective in cytokinin signaling. *abph1* encodes a cytokinin-inducible type A response regulator (ARR) that negatively regulates cytokinin signaling (Giulini et al., 2004). *dec* encodes a plant-specific protein that functions in cytokinin signaling, although the exact role of the DEC protein is unknown (Itoh et al., 2012). In addition to phyllotactic defects, both *abph1* and *dec* mutants have enlarged SAMs, lending support to the idea that meristem size impacts phyllotaxis (Snow and Snow, 1932). Indeed, *Arabidopsis* mutants with enlarged meristems often have perturbed or shifted phyllotaxes (Leyser and Furner, 1992; Goldshmidt et al., 2008; Mandel et al., 2014). Cytokinin also plays a role in meristem size and phyllotaxis in *Arabidopsis*. Cytokinin signaling has been directly linked the transcriptional regulatory network that controls stem cell number. The transcription factor WUSCHEL (WUL) controls the size of the stem cell niche by negatively regulating stem cell number and directly represses *ARR* genes (Leibfried et al., 2005). However, mutants that disrupt cytokinin signaling in *Arabidopsis* are not as severe as their grass counterparts and do not show a discrete phyllotactic shift (Leibfried et al., 2005; Zhao et al., 2010).

The cytokinin network intersects with the auxin network and this interplay seems to be particularly important in determining the timing between primordium initiation events, or the plastochron. Since phyllotaxis is determined by a combination of spatial determinates (where primordia arise on the meristem periphery) and timing determinates (time between subsequent primordium initiation events), this interplay is a key determinate of phyllotaxis. In maize *abph1* mutants, both *PIN1* and auxin levels are decreased, suggesting that cytokinin negatively regulates *PIN1* (Lee et al., 2009). This observation led the authors to hypothesize that the phyllotactic shift in *abph1* mutants may be due to a delay in organ initiation, not just due to an increase in meristem size. Similarly, reduced plastochron duration in the *corkscrew1* (*cks1*) mutant of maize occasionally leads to a shift to opposite decussate phyllotaxis (Alexander et al., 2005). In *Arabidopsis*, cytokinin does not appear to affect *PIN1* localization or levels, but inhibitory cytokinin signaling fields are required downstream of auxin to determine the timing of primordium initiation (Besnard et al., 2014a) and auxin and cytokinin appear

to function synergistically to initiate primordia (Besnard et al., 2014b). Direct links between auxin and cytokinin have been established in *Arabidopsis* shoots, in which the auxin response factor (ARF), ARF5/MP, directly modulates the expression of two *ARR* genes, *ARR7* and *ARR15* and the cytokinin inhibitor, *AHP6* (Zhao et al., 2010; Besnard et al., 2014a). The mechanism of crosstalk between auxin and cytokinin pathways has not yet been determined in the grasses.

Molecular analyses clearly indicate that the position of auxin maxima determines the position of lateral primordia and thereby phyllotactic pattern. Cytokinin also appears to play a key role in generating robust phyllotactic patterns. How then do phyllotactic patterns change, both over the course of development, and across evolutionary time? The positioning of auxin maxima, modulated by cytokinin signaling, appears to be key. This suggests that regulation of auxin synthesis and transport is different in meristems with different phyllotactic patterns. Differences in auxin transport and regulation are likely due to differential expression or regulation of PIN proteins or auxin biosynthesis genes (e.g., through the *PLT* or *IDD* genes), which can differ depending on meristem identity. Thus, phyllotaxis is intimately linked to and one aspect of meristem identity.

MOVING FORWARD

Inflorescence architecture is a major determinant of reproductive success and fitness (Wyatt, 1982; Harder et al., 2004) that cannot be understood by looking only to *Arabidopsis*, or indeed only to the grasses. The development of new model systems has opened up possibilities for investigating meristem dynamics and inflorescence development in a range of taxa with divergent morphologies. Exploring this diversity will allow for a more nuanced understanding of the underlying principles governing shifting meristem identity in particular, and plant development in general.

One important aspect of meristem identity and plant architecture, both in the inflorescence and in the vegetative shoot, is phyllotaxis. Developmental and molecular biology have converged with mathematical modeling to generate a framework to describe the role of auxin in initiating primordia and the establishment of phyllotaxis. Very little is known, however, about how phyllotactic shifts occur either within species (over developmental time) or between species (over evolutionary time). One of the first steps to understanding evolutionary shifts in phyllotaxis is to continue to trace phyllotactic patterns across the tree (Kellogg et al., 2013), at both broad and fine scales.

How do phyllotactic shifts occur? We propose that phyllotaxis is intimately connected to meristem identity. Since phyllotaxis is ultimately determined by auxin (and perhaps cytokinin) signaling within the meristem, meristems with different identities (and phyllotaxes) must also differentially regulate auxin accumulation and signaling. Indeed, in ChIP-Seq and RNA-Seq experiments, transcriptional regulators of meristem identity (e.g., *SEP4*, *RA1*) directly regulate genes in the auxin regulatory network (Kaufmann et al., 2009; Eveland et al., 2014). *KNOTTED1* (*KN1*), a canonical regulator of meristematic activity (Vollbrecht et al., 1991, 2000), binds and modulates many genes in hormonal pathways, particularly genes in the auxin pathway (Bolduc et al.,

2012). In *ra* mutants, auxin signaling is perturbed before any morphological phenotype can be observed (Eveland et al., 2014). Only a small fraction of hormone-associated genes were regulated by *RA1*, but a significant fraction of the auxin-related genes were bound and co-regulated by *RA1* and *KN1*, suggesting a link between auxin signaling and meristem determinacy (Bolduc et al., 2012; Eveland et al., 2014). Three *IDD* genes (mentioned above) are direct targets of *RA1* (Eveland et al., 2014). Thus, the remodeling of auxin signaling networks (and therefore phyllotaxis) might be one aspect of how meristem identity is realized. Similarly, differences in auxin biosynthesis, transport, and signaling likely account for phyllotactic differences between species. To address these questions, we must integrate molecular and genomic approaches with a deep understanding of plant morphology and diversity, and expand analyses beyond a few model species. These endeavors will undoubtedly shed light onto how plants generate their amazing diversity of form.

ACKNOWLEDGMENTS

We would like to thank the reviewers for their helpful comments and suggestions for improving the manuscript. We also apologize to those authors whose work we have inadvertently omitted, or could not review at length due to space limitations. Research in the Thompson lab is supported by NSF award 1148971.

REFERENCES

- Adler, I., Barabe, D., and Jean, R. V. (1997). A history of the study of phyllotaxis. *Ann. Bot.* 80, 231–244. doi: 10.1006/anbo.1997.0422
- Alexander, D. L., Mellor, E. A., and Langdale, J. A. (2005). CORKSCREW1 defines a novel mechanism of domain specification in the maize shoot. *Plant Physiol.* 138, 1396–1408. doi: 10.1104/pp.105.063909
- Bell, A. D., and Bryan, A. L. (1991). *Plant Form: An Illustrated Guide to Flowering Plant Morphology*. Oxford; New York: Oxford University Press.
- Benková, E., Michniewicz, M., Sauer, M., Teichmann, T., Seifertová, D., Jürgens, G., et al. (2003). Local, efflux-dependent auxin gradients as a common module for plant organ formation. *Cell* 115, 591–602. doi: 10.1016/S0092-8674(03)00924-3
- Bennett, T., Brockington, S. F., Rothfels, C., Graham, S. W., Stevenson, D., Kutchan, T., et al. (2014a). Paralogous radiations of PIN proteins with multiple origins of noncanonical PIN structure. *Mol. Biol. Evol.* 31, 2042–2060. doi: 10.1093/molbev/msu147
- Bennett, T., Hines, G., and Leyser, O. (2014b). Canalization: what the flux? *Trends Genet.* 30, 41–48. doi: 10.1016/j.tig.2013.11.001
- Berbel, A., Ferrandiz, C., Hecht, V., Dalmis, M., Lund, O. S., Sussmilch, F. C., et al. (2012). *VEGETATIVE1* is essential for development of the compound inflorescence in pea. *Nat. Commun.* 3, 797. doi: 10.1038/ncomms1801
- Besnard, F., Refahi, Y., Morin, V., Marteaux, B., Brunoud, G., Chambrier, P., et al. (2014a). Cytokinin signalling inhibitory fields provide robustness to phyllotaxis. *Nature* 505, 417–421. doi: 10.1038/nature12791
- Besnard, F., Rozier, F., and Vernoux, T. (2014b). The AHP6 cytokinin signaling inhibitor mediates an auxin-cytokinin crosstalk that regulates the timing of organ initiation at the shoot apical meristem. *Plant Signal. Behav.* 9:e28788. doi: 10.4161/psb.28788
- Bolduc, N., Yilmaz, A., Mejia-Guerra, M. K., Morohashi, K., O'Connor, D., Grotewold, E., et al. (2012). Unraveling the *KNOTTED1* regulatory network in maize meristems. *Genes Dev.* 26, 1685–1690. doi: 10.1101/gad.193433.112
- Bortiri, E., Chuck, G., Vollbrecht, E., Rocheford, T., Martienssen, R., and Hake, S. (2006). *ramosa2* encodes a LATERAL ORGAN BOUNDARY domain protein that determines the fate of stem cells in branch meristems of maize. *Plant Cell* 18, 574–585. doi: 10.1105/tpc.105.039032
- Bradley, D., Ratcliffe, O., Vincent, C., Carpenter, R., and Coen, E. (1997). Inflorescence commitment and architecture in *Arabidopsis*. *Science* 275, 80–83. doi: 10.1126/science.275.5296.80

- Braybrook, S. A., and Peaucelle, A. (2013). Mechano-chemical aspects of organ formation in *Arabidopsis thaliana*: the relationship between auxin and pectin. *PLoS ONE* 8:e57813. doi: 10.1371/journal.pone.0057813
- Brunoud, G., Wells, D. M., Oliva, M., Larriau, A., Mirabet, V., Burrow, A. H., et al. (2012). A novel sensor to map auxin response and distribution at high spatio-temporal resolution. *Nature* 482, 103–106. doi: 10.1038/nature10791
- Carpenter, R., Copeley, L., Vincent, C., Doyle, S., Magrath, R., and Coen, E. (1995). Control of flower development and phyllotaxy by meristem identity genes in *Antirrhinum*. *Plant Cell* 7, 2001–2011. doi: 10.1105/tpc.7.12.2001
- Castel, R., Kusters, E., and Koes, R. (2010). Inflorescence development in petunia: through the maze of botanical terminology. *J. Exp. Bot.* 61, 2235–2246. doi: 10.1093/jxb/erq061
- Cheng, Y., Dai, X., and Zhao, Y. (2006). Auxin biosynthesis by the YUCCA flavin monooxygenases controls the formation of floral organs and vascular tissues in *Arabidopsis*. *Genes Dev.* 20, 1790–1799. doi: 10.1101/gad.1415106
- Chuck, G., and Bortiri, E. (2010). The unique relationship between *tsh4* and *ra2* in patterning floral phytomers. *Plant Signal. Behav.* 5, 979–981. doi: 10.4161/psb.5.8.12220
- Chuck, G., Cigan, A. M., Saetern, K., and Hake, S. (2007). The heterochronic maize mutant *Corngrass1* results from overexpression of a tandem microRNA. *Nat. Genet.* 39, 544–549. doi: 10.1038/ng2001
- Chuck, G., Muszynski, M., Kellogg, E., Hake, S., and Schmidt, R. J. (2002). The control of spikelet meristem identity by the *branched silkleless1* gene in maize. *Science* 298, 1238–1241. doi: 10.1126/science.1076920
- Chuck, G., Whipple, C., Jackson, D., and Hake, S. (2010). The maize SBP-box transcription factor encoded by *tassel sheath4* regulates bract development and the establishment of meristem boundaries. *Development* 137, 1243–1250. doi: 10.1242/dev.048348
- Coen, E. S., Romero, J. M., Doyle, S., Elliott, R., Murphy, G., and Carpenter, R. (1990). *floricaula*: a homeotic gene required for flower development in *Antirrhinum majus*. *Cell* 63, 1311–1322. doi: 10.1016/0092-8674(90)90426-F
- Cohen, O., Borovsky, Y., David-Schwartz, R., and Paran, I. (2012). *CAJOINTLESS* is a MADS-box gene involved in suppression of vegetative growth in all shoot meristems in pepper. *J. Exp. Bot.* 63, 4947–4957. doi: 10.1093/jxb/ers172
- Cui, D., Zhao, J., Jing, Y., Fan, M., Liu, J., Wang, Z., et al. (2013). The *Arabidopsis* IDD14, IDD15, and IDD16 cooperatively regulate lateral organ morphogenesis and gravitropism by promoting auxin biosynthesis and transport. *PLoS Genet.* 9:e1003759. doi: 10.1371/journal.pgen.1003759
- De Reuille, P. B., Bohn-Courseau, I., Ljung, K., Morin, H., Carraro, N., Godin, C., et al. (2006). Computer simulations reveal properties of the cell-cell signaling network at the shoot apex in *Arabidopsis*. *Proc. Natl. Acad. Sci. U.S.A.* 103, 1627–1632. doi: 10.1073/pnas.0510130103
- Diggle, P. K., and Miller, J. S. (2004). Architectural effects mimic floral sexual dimorphism in *Solanum* (Solanaceae). *Am. J. Bot.* 91, 2030–2040. doi: 10.3732/ajb.91.12.2030
- Douady, S., and Couder, Y. (1996). Phyllotaxis as a dynamical self organizing process.3. The simulation of the transient regimes of ontogeny. *J. Theor. Biol.* 178, 295. doi: 10.1006/jtbi.1996.0026
- Dreni, L., Pilatone, A., Yun, D., Erreni, S., Pajoro, A., Caporali, E., et al. (2011). Functional analysis of all AGAMOUS subfamily members in rice reveals their roles in reproductive organ identity determination and meristem determinacy. *Plant Cell* 23, 2850–2863. doi: 10.1105/tpc.111.087007
- Elmqvist, T., and Cox, P. A. (1996). The evolution of vivipary in flowering plants. *Oikos* 77, 3–9. doi: 10.2307/3545579
- Endress, P. K. (1987). Floral phyllotaxis and floral evolution. *Bot. Jahrb. Syst.* 108, 417–438.
- Endress, P. K. (2010). Disentangling confusions in inflorescence morphology: patterns and diversity of reproductive shoot ramification in angiosperms. *J. Syst. Evol.* 48, 225–239. doi: 10.1111/j.1759-6831.2010.00087.x
- Eveland, A. L., Goldshmidt, A., Pautler, M., Morohashi, K., Liseron-Monfils, C., Lewis, M. W., et al. (2014). Regulatory modules controlling maize inflorescence architecture. *Genome Res.* 24, 431–443. doi: 10.1101/gr.166397.113
- Friml, J., Yang, X., Michniewicz, M., Weijers, D., Quint, A., Tietz, O., et al. (2004). A PINOID-dependent binary switch in apical-basal PIN polar targeting directs auxin efflux. *Science* 306, 862–865. doi: 10.1126/science.1100618
- Fu, C., Sunkar, R., Zhou, C., Shen, H., Zhang, J. Y., Matts, J., et al. (2012). Overexpression of miR156 in switchgrass (*Panicum virgatum* L.) results in various morphological alterations and leads to improved biomass production. *Plant Biotechnol. J.* 10, 443–452. doi: 10.1111/j.1467-7652.2011.00677.x
- Gallavotti, A., Barazesh, S., Malcomber, S., Hall, D., Jackson, D., Schmidt, R. J., et al. (2008a). *sparse inflorescence1* encodes a monocot-specific YUCCA-like gene required for vegetative and reproductive development in maize. *Proc. Natl. Acad. Sci. U.S.A.* 105, 15196–15201. doi: 10.1073/pnas.0805596105
- Gallavotti, A., Yang, Y., Schmidt, R. J., and Jackson, D. (2008b). The relationship between auxin transport and maize branching. *Plant Physiol.* 147, 1913–1923. doi: 10.1104/pp.108.121541
- Galweiler, L., Guan, C., Muller, A., Wisman, E., Mendgen, K., Yephremov, A., et al. (1998). Regulation of polar auxin transport by AtPIN1 in *Arabidopsis* vascular tissue. *Science* 282, 2226–2230. doi: 10.1126/science.282.5397.2226
- Giulini, A., Wang, J., and Jackson, D. (2004). Control of phyllotaxy by the cytokinin-inducible response regulator homologue ABPHYLL1. *Nature* 430, 1031–1034. doi: 10.1038/nature02778
- Goldshmidt, A., Alvarez, J. P., Bowman, J. L., and Eshed, Y. (2008). Signals derived from YABBY gene activities in organ primordia regulate growth and partitioning of *Arabidopsis* shoot apical meristems. *Plant Cell* 20, 1217–1230. doi: 10.1105/tpc.107.057877
- Guenot, B., Bayer, E., Kierzkowski, D., Smith, R. S., Mandel, T., Žádníková, P., et al. (2012). PIN1-independent leaf initiation in *Arabidopsis*. *Plant Physiol.* 159, 1501–1510. doi: 10.1104/pp.112.200402
- Harder, L. D., Jordan, C. Y., Gross, W., and Routley, M. B. (2004). Beyond floricentrism: the pollination function of inflorescences. *Plant Species Biol.* 19, 137–148. doi: 10.1111/j.1442-1984.2004.00110.x
- Harder, L. D., and Prusinkiewicz, P. (2013). The interplay between inflorescence development and function as the crucible of architectural diversity. *Ann. Bot.* 112, 1477–1493. doi: 10.1093/aob/mcs252
- Heisler, M. G., Hamant, O., Krupinski, P., Uyttewaal, M., Ohno, C., Jonsson, H., et al. (2010). Alignment between PIN1 polarity and microtubule orientation in the shoot apical meristem reveals a tight coupling between morphogenesis and auxin transport. *PLoS Biol.* 8:e1000516. doi: 10.1371/journal.pbio.1000516
- Heisler, M. G., Ohno, C., Das, P., Sieber, P., Reddy, G. V., Long, J. A., et al. (2005). Patterns of auxin transport and gene expression during primordium development revealed by live imaging of the *Arabidopsis* inflorescence meristem. *Curr. Biol.* 15, 1899–1911. doi: 10.1016/j.cub.2005.09.052
- Hofmeister, W. F. B. (1868). *Allgemeine Morphologie der Gewächse von Wilh: Hofmeister, Mit 134 Holzschnitten*. Leipzig: W. Engelmann.
- Huijser, P., Klein, J., Lonnig, W. E., Meijer, H., Saedler, H., and Sommer, H. (1992). Bracteomania, an inflorescence anomaly, is caused by the loss of function of the MADS-box gene *squamosa* in *Antirrhinum majus*. *EMBO J.* 11, 1239–1249.
- Itoh, J. I., Hibara, K. I., Kojima, M., Sakakibara, H., and Nagato, Y. (2012). Rice *DECUSSATE* controls phyllotaxy by affecting the cytokinin signaling pathway. *Plant J.* 72, 869–881. doi: 10.1111/j.1365-313X.2012.05123.x
- Jackson, D., and Hake, S. (1999). Control of phyllotaxy in maize by the *abphyll1* gene. *Development* 126, 315–323.
- Jönsson, H., Heisler, M. G., Shapiro, B. E., Meyerowitz, E. M., and Mjolsness, E. (2006). An auxin-driven polarized transport model for phyllotaxis. *Proc. Natl. Acad. Sci. U.S.A.* 103, 1633–1638. doi: 10.1073/pnas.0509839103
- Kaufmann, K., Muino, J. M., Jauregui, R., Airolidi, C. A., Smaczniak, C., Krajewski, P. I., et al. (2009). Target genes of the MADS transcription factor SEPALLATA3: integration of developmental and hormonal pathways in the *Arabidopsis* flower. *PLoS Biol.* 7:e1000090. doi:10.1371/journal.pbio.1000090
- Kellogg, E. A. (2000). “A model of inflorescence development,” in *Monocots: Systematics and Evolution*, eds K. L. Wilson and D. A. Morrison (Melbourne: CSIRO), 84–88.
- Kellogg, E. A. (2007). Floral displays: genetic control of grass inflorescences. *Curr. Opin. Plant Biol.* 10, 26–31. doi: 10.1016/j.pbi.2006.11.009
- Kellogg, E. A., Camara, P. E., Rudall, P. J., Ladd, P., Malcomber, S. T., Whipple, C. J., et al. (2013). Early inflorescence development in the grasses (Poaceae). *Front. Plant Sci.* 4:250. doi: 10.3389/fpls.2013.00250
- Komatsu, K., Chujo, A., Nagato, Y., Shimamoto, K., and Kyoizuka, J. (2003). *FRIZZY PANICLE* is required to prevent the formation of axillary meristems and to establish floral meristem identity in rice spikelets. *Development* 130, 3841–3850. doi: 10.1242/dev.00564
- Koppolu, R., Anwar, N., Sakuma, S., Tagiri, A., Lundqvist, U., Pourkheirandish, M., et al. (2013). *Six-rowed spike4 (Vrs4)* controls spikelet determinacy and row-type in barley. *Proc. Natl. Acad. Sci. U.S.A.* 110, 13198–13203. doi: 10.1073/pnas.1221950110

- Kuhlemeier, C. (2007). Phyllotaxis. *Trends Plant Sci.* 12, 143–150. doi: 10.1016/j.tplants.2007.03.004
- Landrein, B., Lathe, R., Bringmann, M., Vouillot, C., Ivakov, A., Boudaoud, A., et al. (2013). Impaired cellulose synthase guidance leads to stem torsion and twists phyllotactic patterns in *Arabidopsis*. *Curr. Biol.* 23, 895–900. doi: 10.1016/j.cub.2013.04.013
- Laudencia-Chinguanco, D., and Hake, S. (2002). The *indeterminate floral apex1* gene regulates meristem determinacy and identity in the maize inflorescence. *Development* 129, 2629–2638.
- Lee, B. H., Johnston, R., Yang, Y., Gallavotti, A., Kojima, M., Travencolo, B. A., et al. (2009). Studies of *aberrant phyllotaxy1* mutants of maize indicate complex interactions between auxin and cytokinin signaling in the shoot apical meristem. *Plant Physiol.* 150, 205–216. doi: 10.1104/pp.109.137034
- Leibfried, A., To, J. P., Busch, W., Stehling, S., Kehle, A., Demar, M., et al. (2005). WUSCHEL controls meristem function by direct regulation of cytokinin-inducible response regulators. *Nature* 438, 1172–1175. doi: 10.1038/nature04270
- Levin, J. Z., and Meyerowitz, E. M. (1995). *UFO*: an *Arabidopsis* gene involved in both floral meristem and floral organ development. *Plant Cell* 7, 529–548. doi: 10.1105/tpc.7.5.529
- Leyser, H. O., and Furner, I. (1992). Characterisation of three shoot apical meristem mutants of *Arabidopsis thaliana*. *Development* 116, 397–403.
- Mandel, T., Moreau, F., Kutsher, Y., Fletcher, J. C., Carles, C. C., and Eshed Williams, L. (2014). The *ERECTA* receptor kinase regulates *Arabidopsis* shoot apical meristem size, phyllotaxis and floral meristem identity. *Development* 141, 830–841. doi: 10.1242/dev.104687
- McSteen, P., Malcomber, S., Skirpan, A., Lunde, C., Wu, X., Kellogg, E., et al. (2007). *barren inflorescence2* Encodes a co-ortholog of the *PINOID* serine/threonine kinase and is required for organogenesis during inflorescence and vegetative development in maize. *Plant Physiol.* 144, 1000–1011. doi: 10.1104/pp.107.098558
- Michniewicz, M., Zago, M. K., Abas, L., Weijers, D., Schweighofer, A., Meskiene, I., et al. (2007). Antagonistic regulation of PIN phosphorylation by PP2A and PINOID directs auxin flux. *Cell* 130, 1044–1056. doi: 10.1016/j.cell.2007.07.033
- Moyroud, E., Kusters, E., Monniaux, M., Koes, R., and Parcy, F. (2010). *LEAFY* blossoms. *Trends Plant Sci.* 15, 346–352. doi: 10.1016/j.tplants.2010.03.007
- O'Connor, D. L., Runions, A., Sluis, A., Bragg, J., Vogel, J. P., Prusinkiewicz, P., et al. (2014). A division in PIN-mediated auxin patterning during organ initiation in grasses. *PLoS Comput. Biol.* 10:e1003447. doi: 10.1371/journal.pcbi.1003447
- Okada, K., Ueda, J., Komaki, M. K., Bell, C. J., and Shimura, Y. (1991). Requirement of the auxin polar transport system in early stages of *Arabidopsis* floral bud formation. *Plant Cell* 3, 677–684. doi: 10.1105/tpc.3.7.677
- Park, S. J., Eshed, Y., and Lippman, Z. B. (2014). Meristem maturation and inflorescence architecture—lessons from the Solanaceae. *Curr. Opin. Plant Biol.* 17, 70–77. doi: 10.1016/j.pbi.2013.11.006
- Park, S. J., Jiang, K., Schatz, M. C., and Lippman, Z. B. (2012). Rate of meristem maturation determines inflorescence architecture in tomato. *Proc. Natl. Acad. Sci. U.S.A.* 109, 639–644. doi: 10.1073/pnas.1114963109
- Pautler, M., Tanaka, W., Hirano, H. Y., and Jackson, D. (2013). Grass meristems I: shoot apical meristem maintenance, axillary meristem determinacy and the floral transition. *Plant Cell Physiol.* 54, 302–312. doi: 10.1093/pcp/pct025
- Peaucelle, A., Braybrook, S. A., Le Guillou, L., Bron, E., Kuhlemeier, C., and Höfte, H. (2011). Pectin-induced changes in cell wall mechanics underlie organ initiation in *Arabidopsis*. *Curr. Biol.* 21, 1720–1726. doi: 10.1016/j.cub.2011.08.057
- Peaucelle, A., Louvet, R., Johansen, J. N., Höfte, H., Laufs, P., Pelloux, J., et al. (2008). *Arabidopsis* phyllotaxis is controlled by the methyl-esterification status of cell-wall pectins. *Curr. Biol.* 18, 1943–1948. doi: 10.1016/j.cub.2008.10.065
- Phillips, K. A., Skirpan, A. L., Liu, X., Christensen, A., Slewinski, T. L., Hudson, C., et al. (2011). *vanishing tassell2* encodes a grass-specific tryptophan aminotransferase required for vegetative and reproductive development in maize. *Plant Cell* 23, 550–566. doi: 10.1105/tpc.110.075267
- Pinon, V., Prasad, K., Grigg, S. P., Sanchez-Perez, G. F., and Scheres, B. (2013). Local auxin biosynthesis regulation by *PLETHORA* transcription factors controls phyllotaxis in *Arabidopsis*. *Proc. Natl. Acad. Sci. U.S.A.* 110, 1107–1112. doi: 10.1073/pnas.1213497110
- Pnueli, L., Carmel-Goren, L., Hareven, D., Gutfinger, T., Alvarez, J., Ganai, M., et al. (1998). The *SELF-PRUNING* gene of tomato regulates vegetative to reproductive switching of sympodial meristems and is the ortholog of *CEN* and *TFL1*. *Development* 125, 1979–1989.
- Poethig, R. S. (2013). Vegetative phase change and shoot maturation in plants. *Curr. Top. Dev. Biol.* 105, 125–152. doi: 10.1016/B978-0-12-396968-2.00005-1
- Prasad, K., Grigg, S. P., Barkoulas, M., Yadav, R. K., Sanchez-Perez, G. F., Pinon, V., et al. (2011). *Arabidopsis* *PLETHORA* transcription factors control phyllotaxis. *Curr. Biol.* 21, 1123–1128. doi: 10.1016/j.cub.2011.05.009
- Prenner, G., Vergara-Silva, F., and Rudall, P. J. (2009). The key role of morphology in modelling inflorescence architecture. *Trends Plant Sci.* 14, 302–309. doi: 10.1016/j.tplants.2009.03.004
- Prusinkiewicz, P., Erasmus, Y., Lane, B., Harder, L. D., and Coen, E. (2007). Evolution and development of inflorescence architectures. *Science* 316, 1452–1456. doi: 10.1126/science.1140429
- Ratcliffe, O. J., Bradley, D. J., and Coen, E. S. (1999). Separation of shoot and floral identity in *Arabidopsis*. *Development* 126, 1109–1120.
- Reinhardt, D. (2005). Regulation of phyllotaxis. *Int. J. Dev. Biol.* 49, 539–546. doi: 10.1387/ijdb.041922dr
- Reinhardt, D., Frenz, M., Mandel, T., and Kuhlemeier, C. (2005). Microsurgical and laser ablation analysis of leaf positioning and dorsoventral patterning in tomato. *Development* 132, 15–26. doi: 10.1242/dev.01544
- Reinhardt, D., Mandel, T., and Kuhlemeier, C. (2000). Auxin regulates the initiation and radial position of plant lateral organs. *Plant Cell* 12, 507–518. doi: 10.1105/tpc.12.4.507
- Reinhardt, D., Pesce, E. R., Stieger, P., Mandel, T., Baltensperger, K., Bennett, M., et al. (2003). Regulation of phyllotaxis by polar auxin transport. *Nature* 426, 255–260. doi: 10.1038/nature02081
- Rijkema, A., Gerats, T., and Vandenbussche, M. (2006). Genetics of floral development in *Petunia*. *Adv. Bot. Res.* 44, 237–278. doi: 10.1016/S0065-2296(06)44006-4
- Robert, S., Kleine-Vehn, J., Barbez, E., Sauer, M., Paciorek, T., Baster, P., et al. (2010). ABP1 mediates auxin inhibition of clathrin-dependent endocytosis in *Arabidopsis*. *Cell* 143, 111–121. doi: 10.1016/j.cell.2010.09.027
- Sassi, M., and Vernoux, T. (2013). Auxin and self-organization at the shoot apical meristem. *J. Exp. Bot.* 64, 2579–2592. doi: 10.1093/jxb/ert101
- Sato-Nagasawa, N., Nagasawa, N., Malcomber, S., Sakai, H., and Jackson, D. (2006). A trehalose metabolic enzyme controls inflorescence architecture in maize. *Nature* 441, 227–230. doi: 10.1038/nature04725
- Scarpella, E., Marcos, D., Friml, J., and Berleth, T. (2006). Control of leaf vascular patterning by polar auxin transport. *Genes Dev.* 20, 1015–1027. doi: 10.1101/gad.1402406
- Shikata, M., Yamaguchi, H., Sasaki, K., and Ohtsubo, N. (2012). Overexpression of *Arabidopsis* miR157b induces bushy architecture and delayed phase transition in *Torenia fournieri*. *Planta* 236, 1027–1035. doi: 10.1007/s00425-012-1649-3
- Smith, R. S., Guyomarc'h, S., Mandel, T., Reinhardt, D., Kuhlemeier, C., and Prusinkiewicz, P. (2006). A plausible model of phyllotaxis. *Proc. Natl. Acad. Sci. U.S.A.* 103, 1301–1306. doi: 10.1073/pnas.0510457103
- Snow, M., and Snow, R. (1932). Experiments on phyllotaxis. I. The effect of isolating a primordium. *Philos. Trans. R. Soc. Lond. B Contain. Papers Biol. Character.* 221, 1–43. doi: 10.1098/rstb.1932.0001
- Snow, M., and Snow, R. (1962). A theory of the regulation of phyllotaxis based on *Lupinus albus*. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 244, 483–513. doi: 10.1098/rstb.1962.0003
- Specht, C. D., and Bartlett, M. E. (2009). Flower evolution: the origin and subsequent diversification of the angiosperm flower. *Annu. Rev. Ecol. Syst.* 40, 217–243. doi: 10.1146/annurev.ecolsys.110308.120203
- Tanaka, W., Pautler, M., Jackson, D., and Hirano, H. Y. (2013). Grass meristems II: inflorescence architecture, flower development and meristem fate. *Plant Cell Physiol.* 54, 313–324. doi: 10.1093/pcp/pct016
- Teo, Z. W., Song, S., Wang, Y. Q., Liu, J., and Yu, H. (2014). New insights into the regulation of inflorescence architecture. *Trends Plant Sci.* 19, 158–165. doi: 10.1016/j.tplants.2013.11.001
- Thompson, B. (2014). “Genetic and hormonal regulation of maize inflorescence development,” in *Advances in Botanical Research*, eds P. Gadal and J.-P. Jacquot (London: Academic Press, Elsevier Ltd.), 263–296.
- Thompson, B. E., Bartling, L., Whipple, C., Hall, D. H., Sakai, H., Schmidt, R., et al. (2009). *bearded-ear* encodes a MADS box transcription factor critical for maize floral development. *Plant Cell* 21, 2578–2590. doi: 10.1105/tpc.109.067751
- Tobena-Santamaria, R., Blied, M., Ljung, K., Sandberg, G., Mol, J. N., Souer, E., et al. (2002). *FLOOZY* of petunia is a flavin mono-oxygenase-like protein

- required for the specification of leaf and flower architecture. *Genes Dev.* 16, 753–763. doi: 10.1101/gad.219502
- Traas, J. (2013). Phyllotaxis. *Development* 140, 249–253. doi: 10.1242/dev.074740
- Troll, W. (1969). *Die Infloreszenzen. Typologie und Stellung im Aufbau des Vegetationskörpers*. Vol. II/1. Jena: VEB Gustav Fischer Verlag.
- Vegetti, A., and Anton, A. (2000). “The grass inflorescence,” in *Grasses: Systematics and Evolution*, eds S. W. L. Jacobs and J. Everett (Collingwood, VIC: CSIRO), 29–31.
- Vernoux, T., Kronenberger, J., Grandjean, O., Laufs, P., and Traas, J. (2000). PIN-FORMED 1 regulates cell fate at the periphery of the shoot apical meristem. *Development* 127, 5157–5165.
- Vollbrecht, E., Reiser, L., and Hake, S. (2000). Shoot meristem size is dependent on inbred background and presence of the maize homeobox gene, *knotted1*. *Development* 127, 3161–3172.
- Vollbrecht, E., Springer, P. S., Goh, L., Buckler, E. S. T., and Martienssen, R. (2005). Architecture of floral branch systems in maize and related grasses. *Nature* 436, 1119–1126. doi: 10.1038/nature03892
- Vollbrecht, E., Veit, B., Sinha, N., and Hake, S. (1991). The developmental gene *Knotted-1* is a member of a maize homeobox gene family. *Nature* 350, 241–243. doi: 10.1038/350241a0
- Wang, C. N., Moller, M., and Cronk, Q. C. (2004). Altered expression of *GFL0*, the Gesneriaceae homologue of *FLORICAULA/LEAFY*, is associated with the transition to bulbil formation in *Titanotrichum oldhamii*. *Dev. Genes Evol.* 214, 122–127. doi: 10.1007/s00427-004-0388-2
- Wang, J.-W., Park, M. Y., Wang, L.-J., Koo, Y., Chen, X.-Y., Weigel, D., et al. (2011). miRNA control of vegetative phase change in trees. *PLoS Genet.* 7:e1002012. doi: 10.1371/journal.pgen.1002012
- Weberling, F. (1989). *Morphology of Flowers and Inflorescences*. Cambridge; New York: Cambridge University Press.
- Weigel, D., Alvarez, J., Smyth, D. R., Yanofsky, M. F., and Meyerowitz, E. M. (1992). *LEAFY* controls floral meristem identity in *Arabidopsis*. *Cell* 69, 843–859. doi: 10.1016/0092-8674(92)90295-N
- Wu, G., and Poethig, R. S. (2006). Temporal regulation of shoot development in *Arabidopsis thaliana* by miR156 and its target *SPL3*. *Development* 133, 3539–3547. doi: 10.1242/dev.02521
- Wyatt, R. (1982). Inflorescence architecture: how flower number, arrangement, and phenology affect pollination and fruit-set. *Am. J. Bot.* 69, 585–594. doi: 10.2307/2443068
- Xie, K., Shen, J., Hou, X., Yao, J., Li, X., Xiao, J., et al. (2012). Gradual increase of miR156 regulates temporal expression changes of numerous genes during leaf development in rice. *Plant Physiol.* 158, 1382–1394. doi: 10.1104/pp.111.190488
- Zhang, D., and Yuan, Z. (2014). Molecular control of grass inflorescence development. *Annu. Rev. Plant Biol.* 65, 553–578. doi: 10.1146/annurev-arplant-050213-040104
- Zhang, X., Zou, Z., Zhang, J., Zhang, Y., Han, Q., Hu, T., et al. (2011). Overexpression of sly-miR156a in tomato results in multiple vegetative and reproductive trait alterations and partial phenocopy of the *sft* mutant. *FEBS Lett.* 585, 435–439. doi: 10.1016/j.febslet.2010.12.036
- Zhao, Z., Andersen, S. U., Ljung, K., Dolezal, K., Miotk, A., Schultheiss, S. J., et al. (2010). Hormonal control of the shoot stem-cell niche. *Nature* 465, 1089–1092. doi: 10.1038/nature09126
- Zotz, G., Wilhelm, K., and Becker, A. (2011). Heteroblasty—a review. *Bot. Rev.* 77, 109–151. doi: 10.1007/s12229-010-9062-8

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 03 July 2014; accepted: 10 September 2014; published online: 14 October 2014.

Citation: Bartlett ME and Thompson B (2014) Meristem identity and phyllotaxis in inflorescence development. *Front. Plant Sci.* 5:508. doi: 10.3389/fpls.2014.00508
This article was submitted to the journal *Frontiers in Plant Science*.

Copyright © 2014 Bartlett and Thompson. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.