

MINI REVIEW

published: 09 March 2016 doi: 10.3389/fonc.2016.00054



The Sweet Side of Immune Evasion: Role of Glycans in the Mechanisms of Cancer Progression

Ana Flávia Fernandes Ribas Nardy¹, Leonardo Freire-de-Lima², Célio Geraldo Freire-de-Lima²* and Alexandre Morrot¹*

¹ Instituto de Microbiologia, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil, ² Laboratório de Glicobiologia, Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

Glycans are part of the essential components of a cell. These compounds play a fundamental role in several physiopathological processes, including cell differentiation, adhesion, motility, signal transduction, host–pathogen interactions, tumor cell invasion, and metastasis development. Glycans are also able to exert control over the changes in tumor immunogenecity, interfering with tumor editing events and leading to immune-resistant cancer cells. The involvement of glycans in cancer progression is related to glycosylation alterations. Understanding such changes is, therefore, extremely useful to set the stage for their use as biomarkers, improving the diagnostics and therapeutic strategies. Herein, we discuss the basis of how modifications in glycosylation patterns may contribute to cancer genesis and progression as well as their importance in oncology field.

Keywords: glycosylation, immunesurveillance, cancer immunology, immune evasion, tumor virulence

OPEN ACCESS

Edited by:

Patrizia Agostinis, University of Leuven, Belgium

Reviewed by:

Martin Herrmann, Universitätsklinikum Erlangen, Germany Giovanna Schiavoni, Istituto Superiore di Sanita', Italy

*Correspondence:

Célio Geraldo Freire-de-Lima celio@biof.ufrj.br; Alexandre Morrot morrot@micro.ufrj.br

Specialty section:

This article was submitted to Molecular and Cellular Oncology, a section of the journal Frontiers in Oncology

Received: 20 November 2015 Accepted: 24 February 2016 Published: 09 March 2016

Citation:

Nardy AFFR, Freire-de-Lima L, Freire-de-Lima CG and Morrot A (2016) The Sweet Side of Immune Evasion: Role of Glycans in the Mechanisms of Cancer Progression. Front. Oncol. 6:54. doi: 10.3389/fonc.2016.00054

GLYCOSYLATION AS AN ESSENTIAL PROTEIN POST-TRANSLATIONAL MODIFICATION

1

Post-translational modifications (PTMs) exert an important role in controlling protein function in eukaryotes. PTMs comprise processes such as acetylation, carbonylation, methylation, hydroxylation, nitration, palmitoylation, phosphorylation, sulfation, ubiquitination, and glycosylation (1, 2). It is universally accepted that deregulation of such PTMs may lead to the development of a number of diseases. The glycosylation is the most common PTM and occurs in all domains of life. As a result, they impart an additional level of "information content" to underlying protein structures (3, 4). Two basic types of protein glycosylations are *N*- and *O*-glycosylations (**Figure 1**) with significant differences in terms of their biosynthesis and structures, as well as their location within the protein chain (5). In *N*-linked glycans, the nitrogen atom in the side chain of asparagine is attached to *N*-acetylglucosamine (GlcNAc). The sequence can be Asn–X–Ser or Asn–X–Thr, where X is any kind of amino acid except proline. In O-linked glycans, the oxygen atom in the side chain of serine or threonine is attached to *N*-acetylgalactosamine (GalNAc) (5). Furthermore, the glycopeptides Asn-GlcNAc or Ser/Thr-GalNAc may be extended by numerous and specific glycosyltransferase activities (6, 7) (**Figure 1**).

ALTERED TUMOR-CELL GLYCOSYLATION PROMOTES CANCER VIRULENCE

During the past few years, we have seen a breakthrough in understanding the molecular and cellular mechanisms of immune cell activation and homeostasis. Several studies have shown

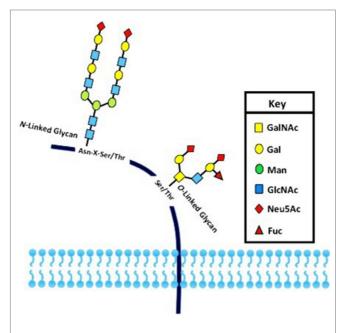


FIGURE 1 | The two major types of protein glycosylation. The attachment of sugar moieties to proteins is a post-translational modification that provides greater proteomic diversity to the proteins. N-linked glycosylation occurs through the asparagine residues of the protein, while O-linked glycosylation occurs through serine or threonine.

that glycosylation orchestrates important features in several pathological processes, mainly in cancer development and/or progression (8–11). Extensive progresses in defining the cellular and molecular networks that regulate the immune responses against different sort of tumors have renewed our enthusiasm to search for potential cancer immunotherapies. However, the successful translation of novel mechanistic insights into effective tumor immunotherapy is hindered by a number of obstacles, including the ability of tumor cells to generate a tolerant microenvironment (12-15). The immune response is crucial not only to elicit protection against pathogens but also to maintain immune surveillance against the development of malignant cells. From this perspective, the development of cancer can be seen as a failure of immune surveillance (16-18). The notion that the immune system can recognize and extinguish developing transformed cells was originally exemplified by the Burnet and Thomas' hypothesis concerning the mechanisms of cancer immune surveillance (19, 20). This hypothesis supports that the immunoediting of tumor antigens is responsible for sculpting the immunogenic phenotypes of transformed cells that eventually induce immunocompetent hosts. However, most tumor-associated carbohydrate antigens (TACAs) do not elicit strong humoral responses and, in fact, pieces of evidence have shown that the aberrant expression of glycan structures, as well as occurrence of truncated structures, precursors, or novel structures of glycan might prevent effective immune responses against tumor cells (21-23). Some TACAs with immunomodulatory or immunosuppressive properties are presented in Table 1.

TUMOR-ASSOCIATED GLYCAN DETERMINANTS SUBVERT KEY IMMUNOLOGICAL DEFENSE MECHANISMS

The first demonstration that tumor cells express altered glycans came from studies showing that monoclonal antibodies may recognize such abnormal structures (38). Many of these alterations are accompanied by expression of oncofetal antigens on tumor glycoproteins as well as glycosphingolipids (38, 39). Furthermore, it is well known that modifications in glycan structures may contribute to early stages of invasion (40–42), although it is not clear if such alterations may also play a role in the genesis of neoplastic cells (43, 44).

Glycosphingolipids as Immunosuppressive Components in Tumor Cell Progression

Glycosphingolipids might also be involved in tumor cell progression by causing immune silencing. It has been also described that different types of cancer cells occasionally secrete gangliosides into the bloodstream (45, 46). In spite of their immunogenic properties, shaded membrane sialoglycolipids (gangliosides) may cause inhibition of co-stimulatory molecules synthesis, thus, promoting an impairment of dendritic cell (DC) maturation, leading to immune silencing by means of the inability of these cells to arm effective antitumor T cells responses (47, 48). A promising antitumor therapeutic approach using glycosphingolipids relies on therapeutic manipulation of these molecules to generate passive as well as active immunity (49–51).

Role of Endogenous Lectins Blocking the Adaptive Immunity Against Tumor Cells

Alterations in the glycosylation profile in tumor compared to health cells are mainly attributed to the gene expression deregulation of glycosyltransferases, enzymes responsible to transfer sugars from donor to acceptor substrate molecules, leading to the synthesis of immature core glycans (52, 53). Specific glycans may bind to cell surface lectins such as galectins, C-type lectins, and siglecs [sialic acid (Sia)-binding immunoglobulin-type lectins], resulting in the regulation of cancer immune responses due to interference with the tumor immunoediting, characterized by changes in the immunogenicity of target antigens that could favor the dissemination of cancer cells (54–56). Galectins are β -galactoside-binding proteins that share homology in the amino acid sequence of their carbohydrate-recognition domain. Their role in immune responses against tumor cells has been studied over the past years (57–59).

In cancer, galectins secreted by tumor cells exhibit tolerogenic effects over effector T cells, promoting a cytokine imbalance that can either result in T cell anergy or favoring T regulatory (T reg) cell activity (60, 61). This immune modulation leading to tolerogenic responses against tumor cells can be associated with galectin-1 expression (62). The galectin-3, another member of galectin family, is known to induce apoptosis of antitumor CD8+ T cells (CTLs) in murine model of colorectal cancer (63). In addition, galectin-3 was demonstrated to increase the distance

Glycan type Glycan structure Enzyme Immunobiological effect Reference N-linked glycan Tri-antennary Tetra-antennary ↑ β1,6 GlcNAc-T (MGATS) Helping the growth of cancer cells through (24)o.[□]•• inactivation of CD4+ T cells and macrophages TOTO TOTO O O-linked glycan β1,4 Gal-T Enhancing the production of anti-inflammatory (25, 26)Tn Sialyl-Tn cytokines; inducing a tolerogenic phenotype in **↓** COSMC **□**♦ innate and adaptive immune cells N-linked glycan Sialyl-Le^x n α 1,3 Fuc-T III Potentiating cancer metastasis; leading to (27 - 29)lung tumor formation or rejection by NK cells O-linked glycan Poly-N-Acetyllactosamine chain N-linked glycan N-glycan ↑ β1,6 GlclMAc-T Resistance against NK cell attack, promoting (30 - 33)tumor metastasis O-linked glycan O-glycan ↑ Core 2GnT R-0000 Neu5Gc-containing glycans N-linked alvcan Neu5Gc is not synthesized in humans, it Neu5Gc-containing glycans are recognized as (34 - 37)O-linked glycan is incorporated into human tissues from foreign antigens by the immune system and **○ □ ○ □ ○ □ ○ □ ○ □** dietary sources. Different sialyltransferases induce chronic inflammation ---can use Neu5Gc as substrate Kev GalNAc Gal GlcNAc Neu5Gc Man ▲ Fuc Neu5Ac

TABLE 1 | Glycan types and their main role in the subversion of antitumor immune responses.

between the TCR and CD8 molecule in human CTLs infiltrating the tumors, a matter that turn them to be anergic (64). Besides their role in attenuating adaptive immune responses, galectins are also described to be able to impair the antitumor functions of natural killer (NK) cells (54).

The C-type lectins constitute by far the largest family of animal lectins found as part of membrane proteins and in soluble forms, comprising L-selectin, P-selectin, and E-selectin glycoproteins (65-67). These molecules act promoting the adhesion of leukocytes to the vascular endothelium, recognizing sialyl Lewis X (SLeX), sialyl Lewis A (SLeA), and Sia found in O-linked glycans (68). Such Lewis carbohydrate antigens can be also found on tumor cells as part of mucin glycoproteins and selectins that play a major role in migration processes by binding to endothelium during tissue infiltration along tumor metastasis (69). In murin models, mice displaying deficiency in L- and P-selectins presented a reduction of the metastisation (70). In addition, specific intercellular adhesion molecule-3 grabbing non-integrin 1 expressed on DCs (DC-SIGN), another transmembrane protein belonging to the C-type lectin family, is expressed by DCs and binds to aberrant O-glycosylation structures on cancer cells (71, 72). These abnormal glycoconjugates expressed by tumor cells are also able to interact with other class of receptors on DCs as well as macrophages, known as macrophage galactose-type C-type lectin, which are able to induce cellular cytotoxicity (73).

Role of Sialic Acid Domains in Avoiding Cell-Mediated Immunity Against Tumor Cells

Immune cells continually screen the glycan structures that are expressed on cell surface glycoproteins from pathogen and host cells. The Sias are part of these multiple cell surface carbohydrates

(74). The Sia motifs are differently expressed among the species, thus, allowing the immune system to distinguish *self* from *non-self*. In this sense, pathogens evolved to express Sia molecules mimicking the one present in host cells, therefore, subverting the host immunity (75, 76). The idea of Sia as self-associated molecular patterns (SAMPs) came from the fact that they may elicit inhibitory signals in order to prevent inadequate immune responses (77). Nonetheless, a growing body of evidence has been pointed out to an extensive contribution of sialoglycan motifs to tumor immune subversion (78, 79).

A multiplicity of ways whereby Sia molecules contribute to immune evasion mechanisms has been described. Complement system has evolved as a first line of defense against non-self or invading pathogens (80, 81). In neoplastic transformation, inhibition of complement activation allows the tumor cells to escape from immune responses (82). In fact, lung cancer cells and glioblastomas, for instance, are able to produce the inhibitory complement factor H, thus avoiding their elimination (83, 84). Tumor cells may evade the complement system activity through binding of Sia motifs present on their surface to polyanionic sites of the complement factor H component, thus activating a complement negative regulatory pathway (85).

Sialoglycans also play a role in tumor immunity mediated by NK cells, which are able to recognize transformed cells due to the lack of MCH class I molecules. However, tumor cells in turn may express inhibitory receptors impairing the cytotoxicity of NK cells (85). The presence of a dense layer of sialoglycans on tumor cell surfaces avoids the normal occurrence of immunological synapses between cancer and NK cells. Such reduced recognition is believed to be enhanced by hypersialylation of tumor ligands for the CD94/ NKG2 family of transmembrane C-type lectin-like receptors (NKG2D) expressed by not only NK cells but also NK1.1+ T cells, $\gamma\delta$ T cells, and activated CD8+ $\alpha\beta$ T cells and macrophages (86). The

NKG2D receptors specifically recognize self-proteins from MIC and RAET1/ULBP families induced on the surface of stressed, malignant transformed, and infected cells (87). The hypersialylation of tumor ligands is thought to repulse their interaction with NKG2D receptors via highly negative charge (88).

The tumor-derived sialoglycans can also affect antitumor functions of NK cells via Sia/siglecs binding (89, 90). In fact, it has been shown that the immunomodulatory effects of tumor cells in part have influence of interactions between sialoglycans derived from transformed cells and the immune inhibitory siglec receptors (69). In fact, studies have demonstrated that overexpression of siglec ligands in tumor cells leads to impairment of the protective immune responses elicited by NK cells and neutrophils (91, 92). In addition, blockade of siglec-9 improved antitumor neutrophil responses *in vitro* (92). Siglec receptor triggering with sialylated mucins derived from tumor cells is able to induce inhibitory signals to immune cells, a process that is thought to be associated with tumor progress (41).

Tumor-derived sialoglycans can target different aspects of the immune system to promote evasion responses. It has been shown that tumor-derived Sia inhibits the traffic and subsequent exocytosis of lytic granules from CTLs to the immunological synapse, disabling a key mechanism whereby these lymphocytes eradicate tumor cells (93). Moreover, Sias have also been described to take part in the hypersialylation process of Fas receptor (CD95) on tumor cells, damping its binding to the Fas-ligand (CD95L) expressed by CTLs (94). The blockage of CD95/CD95L interaction impairs the downstream activation of caspases and consequently disarming the apoptosis machinery that would lead to tumor cell elimination (69). In this context, hypersialylation of Fas receptor by upregulation of sialyltransferase ST6Gal-I in tumor cells has also been described (95).

Besides its effects on cytotoxic T cells, tumor-derived sialogly-cans are also able to dampen DC functions (96–100). Studies have shown that tumor-derived sialogangliosides inhibit the upregulation of co-stimulatory molecules (CD80/CD86) as well as IL-12 production by DCs, thus impairing T cell effector lymphocyte activation (101). This immunosuppressive effect is thought to be elicited by the interaction of highly sialylated tumor antigens

REFERENCES

- Karve TM, Cheema AK. Small changes huge impact: the role of protein post-translational modifications in cellular homeostasis and disease. *J Amino Acids* (2011) 2011:207691. doi:10.4061/2011/207691
- Seo J, Lee KJ. Post-translational modifications and their biological functions: proteomic analysis andsystematic approaches. J Biochem Mol Biol (2004) 37(1):35–44. doi:10.5483/BMBRep.2004.37.1.035
- 3. Fuster MM, Esko JD. The sweet and sour of cancer: glycans as novel therapeutic targets. *Nat Rev Cancer* (2005) 5(7):526–42. doi:10.1038/nrc1649
- Moremen KW, Tiemeyer M, Nairn AV. Vertebrate protein glycosylation: diversity, synthesis and function. Nat Rev Mol Cell Biol (2012) 13(7):448–62. doi:10.1038/nrm3383
- Spiro RG. Protein glycosylation: nature, distribution, enzymatic formation, and disease implications of glycopeptide bonds. *Glycobiology* (2002) 12(4):43R–56R. doi:10.1093/glycob/12.4.43R
- Schwarz F, Aebi M. Mechanisms and principles of N-linked protein glycosylation. Curr Opin Struct Biol (2011) 21(5):576–82. doi:10.1016/j. sbi.2011.08.005

with the siglec receptors expressed by DCs (102, 103). Moreover, the interaction of tumor-derived sialylated antigens with siglec receptors expressed by macrophages has also been described as a mechanism influencing tumor progression (104). In these studies, siglec-9 expressed by macrophages is shown to induce high levels of the immunosuppressive IL-10 cytokine together with reduction of the pro-inflammatory TNF- α cytokine associated with antitumoral responses (105).

CONCLUDING REMARKS

Cell surface glycosylation patterns may suffer important changes during pathological conditions, especially in tumor invasion processes. Such alterations are the result of genetics as well as epigenetics changes, conferring to the tumor cells the ability of dissemination by escaping the immunesurveillance mechanisms (10, 106). This immune evasion phenomenon is being clarified, pointing out an important role for the shield created by altered sialylated glycans on the surface of cancer cells on the subversion of the immune system. This evolutionary conserved strategy can also be observed in some pathogens such as trypanosomatids, bacteria, and fungi (57). Understanding how abnormal glycosylation patterns effectively contribute to tumor-induced immune deviation would lead to early detection of potential tissue alterations and ultimately the development of therapeutic approaches against cancer.

AUTHOR CONTRIBUTIONS

AN, LF-L, CGF-L, and AM wrote the paper. All authors read and approved the final version of the manuscript.

FUNDING

This work was supported by grants from Conselho Nacional de Desenvolvimento Científico e Tecnológico do Brasil (CNPq), Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ). AM and CGF-L are recipients of fellowships from CNPq.

- Lehle L, Strahl S, Tanner W. Protein glycosylation, conserved from yeast to man: a model organism helps elucidate congenital human diseases. *Angew Chem Int Ed Engl* (2006) 45(41):6802–18. doi:10.1002/anie.200601645
- Nicholas NS, Apollonio B, Ramsay AG. Tumor microenvironment (TME)driven immune suppression in B cell malignancy. *Biochim Biophys Act* (2016) 1863(3):471–82. doi:10.1016/j.bbamcr.2015.11.003
- Kölbl AC, Andergassen U, Jeschke U. The role of glycosylation in breast cancer metastasis and cancer control. Front Oncol (2015) 5:219. doi:10.3389/ fonc.2015.00219
- Ohtsubo K, Marth JD. Glycosylation in cellular mechanisms of health and disease. Cell (2006) 126(5):855–67. doi:10.1016/j.cell.2006.08.019
- Hakomori S. Glycosylation defining cancer malignancy: new wine in an old bottle. Proc Natl Acad Sci U S A (2002) 99(16):10231–3. doi:10.1073/ pnas.172380699
- Eyre TA, Collins GP. Immune checkpoint inhibition in lymphoid disease. Br J Haematol (2015) 170(3):291–304. doi:10.1111/bjh.13397
- Guo Y, Wang AY. Novel immune check-point regulators in tolerance maintenance. Front Immunol (2015) 18(6):421. doi:10.3389/ fimmu.2015.00421

- Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, et al. Immune evasion in cancer: mechanistic basis and therapeutic strategies. *Semin Cancer Biol* (2015) 35:S185–98. doi:10.1016/j.semcancer.2015.03.004
- 15. Liu Y, Cao X. Immunosuppressive cells in tumor immune escape and metastasis. *J Mol Med* (2015) **93**:1–14. doi:10.1007/s00109-015-1376-x
- Prestwich RJ, Errington F, Hatfield P, Merrick AE, Ilett EJ, Selby PJ, et al. The immune system – is it relevant to cancer development, progression and treatment? Clin Oncol (2008) 20(2):101–12. doi:10.1016/j.clon.2007.10.011
- Kim R, Emi M, Tanabe K. Cancer immunoediting from immune surveillance to immune escape. *Immunology* (2007) 121(1):1–14. doi:10.1111/j.1365-2567.2007.02587.x
- 18. Morvan MG, Lanier LL. NK cells and cancer: you can teach innate cells new tricks. *Nat Rev Cancer* (2015) **16**(1):7–19. doi:10.1038/nrc.2015.5
- Burnet M. Cancer; a biological approach. I. The processes of control. *Br Med J* (1957) 1(5022):779–86. doi:10.1136/bmj.1.5022.779
- Thomas L. On immunosurveillance in human cancer. Yale J Biol Med (1982) 55(3-4):329-33.
- Monzavi-Karbassi B, Pashov A, Kieber-Emmons T. Tumor-associated glycans and immune surveillance. *Vaccines (Basel)* (2013) 1(2):174–203. doi:10.3390/ vaccines1020174
- Pashov A, Monzavi-Karbassi B, Kieber-Emmons T. Immune surveillance and immunotherapy: lessons from carbohydrate mimotopes. *Vaccine* (2009) 27(25–26):3405–15. doi:10.1016/j.vaccine.2009.01.074
- Pashov A, Monzavi-Karbassi B, Raghava GP, Kieber-Emmons T. Bridging innate and adaptive antitumor immunity targeting glycans. *J Biomed Biotechnol* (2010) 2010:354068. doi:10.1155/2010/354068
- Li D, Li Y, Wu X, Li Q, Yu J, Gen J, et al. Knockdown of Mgat5 inhibits breast cancer cell growth with activation of CD4+ T cells and macrophages. J Immunol (2008) 180(5):3158–65. doi:10.4049/jimmunol.180.5.3158
- Takamiya R, Ohtsubo K, Takamatsu S, Taniguchi N, Angata T. The interaction between Siglec-15 and tumor-associated sialyl-Tn antigen enhances TGF-beta secretion from monocytes/macrophages through the DAP12-Syk pathway. Glycobiology (2013) 23(2):178–87. doi:10.1093/glycob/cws139
- Carrascal MA, Severino PF, Guadalupe Cabral M, Silva M, Ferreira JA, Calais
 F, et al. Sialyl Tn-expressing bladder cancer cells induce a tolerogenic phenotype in innate and adaptive immune cells. *Mol Oncol* (2014) 8(3):753–65. doi:10.1016/j.molonc.2014.02.0088
- Kannagi R, Izawa M, Koike T, Miyazaki K, Kimura N. Carbohydrate-mediated cell adhesion in cancer metastasis and angiogenesis. *Cancer Sci* (2004) 95(5):377–84. doi:10.1111/j.1349-7006.2004.tb03219.x
- Schultz MJ, Swindall AF, Bellis SL. Regulation of the metastatic cell phenotype by sialylated glycans. *Cancer Metastasis Rev* (2012) 31(3–4):501–18. doi:10.1007/s10555-012-9359-7
- Ohyama C, Tsuboi S, Fukuda M. Dual roles of sialyl Lewis X oligosaccharides in tumor metastasis and rejection by natural killer cells. *EMBO J* (1999) 18(6):1516–25. doi:10.1093/emboj/18.6.1516
- Tsuboi S, Hatakeyama S, Ohyama C, Fukuda M. Two opposing roles of O-glycans in tumor metastasis. *Trends Mol Med* (2012) 18(4):224–32. doi:10.1016/j.molmed.2012.02.001
- Suzuki Y, Sutoh M, Hatakeyama S, Mori K, Yamamoto H, Koie T, et al. MUC1 carrying core 2 O-glycans functions as a molecular shield against NK cell attack, promoting bladder tumor metastasis. *Int J Oncol* (2012) 40(6):1831–8. doi:10.3892/ijo.2012.1411
- 32. Tsuboi S. Tumor defense systems using O-glycans. *Biol Pharm Bull* (2012) 35(10):1633–6. doi:10.1248/bpb.b12-00367
- Okamoto T, Yoneyama MS, Hatakeyama S, Mori K, Yamamoto H, Koie T, et al. Core2 O-glycan-expressing prostate cancer cells are resistant to NK cell immunity. Mol Med Rep (2013) 7(2):359–64. doi:10.3892/mmr.2012.1189
- Samraj AN, Pearce OM, Läubli H, Crittenden AN, Bergfeld AK, Banda K, et al. A red meat-derived glycan promotes inflammation and cancer progression. Proc Natl Acad Sci USA (2015) 112(2):542–7. doi:10.1073/pnas.1417508112
- Pearce OM, Läubli H, Verhagen A, Secrest P, Zhang J, Varki NM, et al. Inverse hormesis of cancer growth mediated by narrow ranges of tumor-directed antibodies. *Proc Natl Acad Sci U S A* (2014) 111(16):5998–6003. doi:10.1073/ pnas.1209067111
- Pearce OM, Läubli H. Sialic acids in cancer biology and immunity. Glycobiology (2016) 26(2):111–28. doi:10.1093/glycob/cwv097

- Samraj AN, Läubli H, Varki N, Varki A. Involvement of a non-human sialic acid in human cancer. Front Oncol (2014) 19(4):33. doi:10.3389/ fonc.2014.00033
- 38. Feizi T. Demonstration by monoclonal antibodies that carbohydrate structures of glycoproteins and glycolipids are onco-developmental antigens. *Nature* (1985) **314**(6006):53–7. doi:10.1038/314053a0
- Holmes EH, Ostrander GK, Clausen H, Graem N. Oncofetal expression of Lex carbohydrate antigens in human colonic adenocarcinomas. Regulation through type 2 core chain synthesis rather than fucosylation. *J Biol Chem* (1987) 262(23):11331–8.
- Reticker-Flynn NE, Bhatia SN. Aberrant glycosylation promotes lung cancer metastasis through adhesion to galectins in the metastatic niche. Cancer Discov (2015) 5(2):168–81. doi:10.1158/2159-8290.CD-13-0760
- Boligan KF, Mesa C, Fernandez LE, von Gunten S. Cancer intelligence acquired (CIA): tumor glycosylation and sialylation codes dismantling antitumor defense. Cell Mol Life Sci (2015) 72(7):1231–48. doi:10.1007/ s00018-014-1799-5
- Freire-de-Lima L. Sweet and sour: the impact of differential glycosylation in cancer cells undergoing epithelial-mesenchymal transition. Front Oncol (2014) 25(4):59. doi:10.3389/fonc.2014.00059
- Kakugawa Y, Wada T, Yamaguchi K, Yamanami H, Ouchi K, Sato I, et al. Up-regulation of plasma membrane-associated ganglioside sialidase (Neu3) in human colon cancer and its involvement in apoptosis suppression. *Proc Natl Acad Sci U S A* (2002) 99(16):10718–23. doi:10.1073/ pnas.152597199
- Handa K, Hakomori SI. Carbohydrate to carbohydrate interaction in development process and cancer progression. *Glycoconj J* (2012) 29(8–9):627–37. doi:10.1007/s10719-012-9380-7
- Varki A, Cummings RD, Esko JD, Freeze H, Stanley P, Bertozzi C, et al. *Essentials of Glycobiology*. New York: Cold Spring Harbor Laboratory Press (2009).
- 46. Hakomori S. Tumor malignancy defined by aberrant glycosylation and sphingo(glyco)lipid metabolism. *Cancer Res* (1996) **56**(23):5309–18.
- Wolfl M, Batten WY, Posovszky C, Bernhard H, Berthold F. Gangliosides inhibit the development from monocytes to dendritic cells. *Clin Exp Immunol* (2002) 130(3):441–8. doi:10.1046/j.1365-2249.2002.02006.x
- Caldwell S, Heitger A, Shen W, Liu Y, Taylor B, Ladisch S. Mechanisms of ganglioside inhibition of APC function. *J Immunol* (2003) 171(4):1676–83. doi:10.4049/jimmunol.171.4.1676
- Bestagno M, Occhino M, Corrias MV, Burrone O, Pistoia V. Recombinant antibodies in the immunotherapy of neuroblastoma: perspectives of new developments. *Cancer Lett* (2003) 197(1–2):193–8. doi:10.1016/ S0304-3835(03)00109-5
- Carr A, Rodríguez E, Arango Mdel C, Camacho R, Osorio M, Gabri M, et al. Immunotherapy of advanced breast cancerwith a heterophilic ganglioside (NeuGcGM3) cancervaccine. *J Clin Oncol* (2003) 21(6):1015–21. doi:10.1200/JCO.2003.02.124
- 51. Ragupathi G, Livingston PO, Hood C, Gathuru J, Krown SE, Chapman PB, et al. Consistent antibody response against ganglioside GD2 induced in patients with melanoma by a GD2 lactone-keyhole limpet hemocyanin conjugate vaccine plus immunological adjuvant QS-21. *Clin Cancer Res* (2003) 9(14):5214–20.
- Brooks SA, Carter TM, Royle L, Harvey DJ, Fry SA, Kinch C, et al. Altered glycosylation of proteins in cancer: what is the potential for new anti-tumour strategies. *Anticancer Agents Med Chem* (2008) 8(1):2–21. doi:10.2174/187152008783330860
- Amon R, Reuven EM, Leviatan Ben-Arye S, Padler-Karavani V. Glycans in immune recognition and response. *Carbohydr Res* (2014) 7(389):115–22. doi:10.1016/j.carres.2014.02.004
- Rabinovich GA, Toscano MA. Turning 'sweet' on immunity: galectin-glycan interactions in immune tolerance and inflammation. *Nat Rev Immunol* (2009) 9(5):338–52. doi:10.1038/nri2536
- Macauley MS, Crocker PR, Paulson JC. Siglec-mediated regulation of immune cell function in disease. *Nat Rev Immunol* (2014) 14(10):653–66. doi:10.1038/nri3737
- Pinho SS, Reis CA. Glycosylation in cancer: mechanisms and clinical implications. Nat Rev Cancer (2015) 15(9):540–55. doi:10.1038/nrc3982

- Rambaruth ND, Dwek MV. Cell surface glycan-lectin interactions in tumor metastasis. Acta Histochem (2011) 113(6):591–600. doi:10.1016/j. acthis.2011.03.001
- Perone MJ, Bertera S, Shufesky WJ, Divito SJ, Montecalvo A, Mathers AR, et al. Suppression of autoimmune diabetes by soluble galectin-1. *J Immunol* (2009) 182(5):2641–53. doi:10.4049/jimmunol.0800839
- Rabinovich GA, Liu FT, Hirashima M, Anderson A. An emerging role for galectins in tuning the immune response: lessons from experimental models of inflammatory disease, autoimmunity and cancer. *Scand J Immunol* (2007) 66(2–3):143–58. doi:10.1111/j.1365-3083.2007.01986.x
- Rabinovich GA, Gabrilovich D, Sotomayor EM. Immunossupressive strategies that are mediated by tumor cells. *Annu Rev Immunol* (2007) 25:267–96. doi:10.116/annurev.immunol.25.022106.141609
- 61. Liu FT, Rabinovich GA. Galectins as modulators of tumour progression. *Nat Rev Cancer* (2005) 5(1):29–41. doi:10.1038/nrc1527
- Le QT, Shi G, Cao H, Nelson DW, Wang Y, Chen EY, et al. Galectin-1: a link between tumor hypoxia and tumor immune privilege. *J Clin Oncol* (2005) 23(35):8932–41. doi:10.1200/JCO.2005.02.0206
- Peng W, Wang HY, Miyahara Y, Peng G, Wang RF. Tumor-associated galectin-3 modulates the function of tumor-reactive T cells. *Cancer Res* (2008) 68(17):7228–36. doi:10.1158/0008-5472
- Demotte N, Stroobant V, Courtoy PJ, Van Der Smissen P, Colau D, Luescher IF, et al. Restoring the association of the T cell receptor with CD8 reverses anergy in human tumor-infiltrating lymphocytes. *Immunity* (2008) 28(3):414–24. doi:10.1016/j.immuni.2008.01.011
- Glavey SV, Huynh D, Reagan MR, Manier S, Moschetta M, Kawano Y, et al. The cancer glycome: carbohydrates as mediators of metastasis. *Blood Rev* (2015) 29(4):269–79. doi:10.1016/j.blre.2015.01.003
- Nakahara S, Raz A. Biological modulation by lectins and their ligands in tumor progression and metastasis. *Anticancer Agents Med Chem* (2008) 8(1):22–36. doi:10.2174/187152008783330833
- 67. Fukuda M, Hiraoka N, Yeh JC. C-type lectins and sialyl Lewis X oligosaccharides. Versatile roles in cell-cell interaction. *J Cell Biol* (1999) **147**(3):467–70. doi:10.1083/jcb.147.3.467
- Varki A. Selectin ligands. Proc Natl Acad Sci U S A (1994) 91(16):7390-7. doi:10.1073/pnas.91.16.7390
- Häuselmann I, Borsig L. Altered tumor-cell glycosylation promotes metastasis. Front Oncol (2014) 4:28. doi:10.3389/fonc.2014.00028
- Kohler S, Ullrich S, Richter U, Schumacher U. E-/P-selectins and colon carcinoma metastasis: first in vivo evidence for their crucial role in a clinically relevant model of spontaneous metastasis formation in the lung. Br J Cancer (2010) 102(3):602–9. doi:10.1038/sj.bjc.6605492
- Guo Y, Feinberg H, Conroy E, Mitchell DA, Alvarez R, Blixt O, et al. Structural basis for distinct ligand-binding and targeting properties of the receptors DC-SIGN and DC-SIGNR. Nat Struct Mol Biol (2004) 11(7):591–8. doi:10.1038/nsmb784
- Samsen A, Bogoevska V, Klampe B, Bamberger AM, Lucka L, Horst AK, et al. DC–SIGN and SRCL bind glycans of carcinoembryonic antigen (CEA) and CEA-related cell adhesion molecule 1 (CEACAM1): recombinant human glycan-binding receptors as analytical tools. *Eur J Cell Biol* (2010) 89(1):87–94. doi:10.1016/j.ejcb.2009.11.018
- Lavrsen K, Madsen CB, Rasch MG, Woetmann A, Ødum N, Mandel U, et al. Aberrantly glycosylated MUC1 is expressed on the surface of breast cancer cells and a target for antibody-dependent cell-mediated cytotoxicity. Glycoconj J (2013) 30(3):227–36. doi:10.1007/s10719-012-9437-7
- Varki A, Gagneux P. Multifarious roles of sialic acids in immunity. Ann N Y Acad Sci (2012) 1253:16–36. doi:10.1111/j.1749-6632.2012.06517.x
- Khatua B, Ghoshal A, Bhattacharya K, Mandal C, Saha B, Crocker PR, et al. Sialic acids acquired by *Pseudomonas aeruginosa* are involved in reduced complement deposition and siglec mediated host-cell recognition. *FEBS Lett* (2010) 584(3):555–61. doi:10.1016/j.febslet.2009.11.087
- Carlin AF, Uchiyama S, Chang YC, Lewis AL, Nizet V, Varki A. Molecular mimicry of host sialylated glycans allows a bacterial pathogen to engage neutrophil Siglec-9 and dampen the innate immune resposses. *Blood* (2009) 113(14):3333–6. doi:10.1182/blood-2008-11-187302
- Varki A. Since there are PAMPs and DAMPs, there must be SAMPs? Glycan "self-associatedmolecular patterns" dampen innate immunity, but pathogens can mimic them. *Glycobiology* (2011) 21(9):1121–4. doi:10.1093/glycob/cwr087

- Sanford BH. An alteration in tumor histocompatibility induced by neuraminidase. *Transplantation* (1967) 5(5):1273–9. doi:10.1097/00007890-196709000-00005
- Bagshawe KD, Currie GA. Immunogenicity of L 1210 murine leukaemia cells after treatment with neuraminidase. *Nature* (1968) 218(5148):1254–5. doi:10.1038/2181254a0
- 80. Walport MJ. Complement. First of two parts. N Engl J Med (2001) $\bf 344(14):1058-66.$ doi:10.1056/NEJM200104053441406
- Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. *Nat Immunol* (2010) 11(9):785–97. doi:10.1038/ni.1923
- 82. Pio R, Corrales L, Lambris JD. The role of complement in tumor growth. *Adv Exp Med Biol* (2014) **772**:229–62. doi:10.1007/978-1-4614-5915-6_11
- Junnikkala S, Jokiranta TS, Friese MA, Jarva H, Zipfel PF, Meri S. Exceptional resistance of human H2 glioblastoma cells to complement-mediated killing by expression and utilization of factor H and factor H-like protein 1. *J Immunol* (2000) 164(11):6075–81. doi:10.4049/jimmunol.164.11.6075
- Ajona D, Hsu YF, Corrales L, Montuenga LM, Pio R. Down-regulation of human complement factor H sensitizes non-small cell lung cancer cells to complement attack and reduces in vivo tumor growth. *J Immunol* (2007) 178(9):5991–8. doi:10.4049/jimmunol.178.9.5991
- Büll C, den Brok MH, Adema GJ. Sweet escape: sialic acids in tumor immune evasion. *Biochim Biophys Acta* (2014) 1846(1):238–46. doi:10.1016/j. bbcan.2014.07.005
- Lanier LL. NKG2D receptor and its ligands in host defense. Cancer Immunol Res (2015) 3(6):575–82. doi:10.1158/2326-6066
- Cheng M, Chen Y, Xiao W, Sun R, Tian Z. NK cell-based immunotherapy for malignant diseases. *Cell Mol Immunol* (2013) 10(3):230–52. doi:10.1038/ cmi.2013.10
- Cohen M, Elkabets M, Perlmutter M, Porgador A, Voronov E, Apte RN, et al. Lichtenstein, sialylation of 3-methylcholanthrene-induced fibrosarcoma determinesantitumor immune responses during immunoediting. *J Immunol* (2010) 185(10):5869–78. doi:10.4049/jimmunol.1001635
- Park JE, Wu DY, Prendes M, Lu SX, Ragupathi G, Schrantz N, et al. Fine specificity of natural killer T cells against GD3 ganglioside and identification of GM3 as an inhibitory natural killer T-cell ligand. *Immunology* (2008) 123(1):145–55. doi:10.1111/j.1365-2567.2007.02760.x
- Webb TJ, Li X, Giuntoli RL II, Lopez PH, Heuser C, Schnaar RL, et al. Molecular identification of GD3 as a suppressor of the innate immune response in ovarian cancer. Cancer Res (2012) 72(15):3744–52. doi:10.1158/0008-5472.CAN-11-2695
- 91. Jandus C, Boligan KF, Chijioke O, Liu H, Dahlhaus M, Demoulins T, et al. Interactions between Siglec-7/9 receptors and ligands influence NK cell-dependent tumor immunosurveillance. *J Clin Invest* (2014) **124**(4):1810–20. doi:10.1172/JCI65899
- 92. Läubli H, Pearce OM, Schwarz F, Siddiqui SS, Deng L, Stanczak MA, et al. Engagement of myelomonocytic Siglecs by tumor-associated ligands modulates the innate immune response to cancer. *Proc Natl Acad Sci U S A* (2014) 111(39):14211–6. doi:10.1073/pnas.1409580111
- Lee HC, Wondimu A, Liu Y, Ma JS, Radoja S, Ladisch S. Ganglioside inhibition of CD8+T cell cytotoxicity: interference with lytic granule trafficking and exocytosis. *J Immunol* (2012) 189(7):3521-7. doi:10.4049/jimmunol.1201256
- Lee KH, Feig C, Tchikov V, Schickel R, Hallas C, Schütze S, et al. The role of receptor internalization in CD95 signaling. EMBO J (2006) 25(5):1009–23. doi:10.1038/sj.emboj.7601016
- Swindall AF, Bellis SL. Sialylation of the Fas death receptor by ST6Gal-I provides protection against Fas-mediated apoptosis in colon carcinoma cells. *J Biol Chem* (2011) 286(26):22982–90. doi:10.1074/jbc.M110.211375
- Steinman RM, Adams JC, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. IV. Identification and distribution in mouse spleen. J Exp Med (1975) 141:804–20.
- 97. Steinman RM, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. I. Morphology, quantitation, tissue distribution. *J Exp Med* (1973) 137:1142–62. doi:10.1084/jem.137.5.1142
- Steinman RM, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. II. Functional properties in vitro. *J Exp Med* (1974) 139:380–97. doi:10.1084/jem.139.2.380
- Steinman RM, Lustig DS, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. III. Functional properties in vivo. J Exp Med (1974) 139:1431–45. doi:10.1084/jem.139.6.1431

- 100. Steinman RM, Witmer MD. Lymphoid dendritic cells are potent stimulators of the primary mixed leukocyte reaction in mice. *Proc Natl Acad Sci U S A* (1978) 75:5132–6. doi:10.1073/pnas.75.10.5132
- 101. Jales A, Falahati R, Mari E, Stemmy EJ, Shen W, Southammakosane C, et al. Ganglioside-exposed dendritic cells inhibit T-cell effectorfunction by promoting regulatory cell activity. *Immunology* (2011) 132(1):134–43. doi:10.1111/j.1365-2567.2010.03348.x
- 102. Péguet-Navarro J, Sportouch M, Popa I, Berthier O, Schmitt D, Portoukalian J. Gangliosides from human melanoma tumors impair dendritic cell differentiation from monocytes and induce their apoptosis. *J Immunol* (2003) 170(7):3488–94. doi:10.4049/jimmunol.170.7.3488
- 103. Shurin GV, Shurin MR, Bykovskaia S, Shogan J, Lotze MT, Barksdale EM Jr. Neuroblastoma-derived gangliosides inhibit dendritic cell generation and function. *Cancer Res* (2001) 61(1):363–9.
- 104. Pillai S, Netravali IA, Cariappa A, Mattoo H. Siglecs and immune regulation. Annu Rev Immunol (2012) 30:357–92. doi:10.1146/ annurev-immunol-020711-075018

- Ando M, Tu W, Nishijima K, Iijima S. Siglec-9 enhances IL-10 production in macrophages via tyrosine-based motifs. *Biochem Biophys Res Commun* (2008) 369(3):878–83. doi:10.1016/j.bbrc.2008.02.11a
- 106. Kannagi R, Sakuma K, Miyazaki K, Lim KT, Yusa A, Yin J, et al. Altered expression of glycan genes in cancers induced by epigenetic silencing and tumor hypoxia: clues in the ongoing search for new tumor markers. Cancer Sci (2010) 101(3):586–93. doi:10.1111/j.1349-7006.2009.01455.x

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Nardy, Freire-de-Lima, Freire-de-Lima and Morrot. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.