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# Protein structure and ionic selectivity in calcium channels: Selectivity filter size, not shape, matters

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## ABSTRACT

Calcium channels have highly charged selectivity filters ( $4 \text{ COO}^-$  groups) that attract cations in to balance this charge and minimize free energy, forcing the cations (Na<sup>+</sup> and Ca<sup>2+</sup>) to compete for space in the filter. A reduced model was developed to better understand the mechanism of ion selectivity in calcium channels. The charge/space competition (CSC) mechanism implies that Ca<sup>2+</sup> is more efficient in balancing the charge of the filter because it provides twice the charge as Na<sup>+</sup> while occupying the same space. The CSC mechanism further implies that the main determinant of Ca<sup>2+</sup> versus Na<sup>+</sup> selectivity is the density of charged particles in the selectivity filter, i.e., the volume of the filter (after fixing the number of charged groups in the filter). In this paper we test this hypothesis by changing filter length and/or radius (shape) of the cylindrical selectivity filter of our reduced model. We show that varying volume and shape together has substantially stronger effects than varying shape alone with volume fixed. Our simulations show the importance of depletion zones of ions in determining channel conductance calculated with the integrated Nernst–Planck equation. We show that confining the protein side chains with soft or hard walls does not influence selectivity.

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# 1. Introduction

Ion channels control many biological functions from signalling in the nervous system to initiating muscle contraction. These channel proteins form pores that conduct ions down their electrochemical potential gradient across otherwise impermeable membranes of both the cell and the organelles within it. Ion channels can select which ionic species they conduct, with different channel proteins conducting different ionic species. For example, potassium channels conduct K<sup>+</sup> and (statistically speaking) exclude Na<sup>+</sup>. The quite different sodium channel protein conducts Na<sup>+</sup> and excludes K<sup>+</sup> [1].

Another important class of ion channels preferentially conducts Ca<sup>2+</sup> compared to monovalent cations like Na<sup>+</sup> and K<sup>+</sup>. A biologically important subclass of calcium channels are highly Ca<sup>2+</sup> selective. For example, the L-type calcium channel has a Ca<sup>2+</sup> affinity of 1  $\mu$ M; in 30 mM Na<sup>+</sup> as little as 1  $\mu$ M Ca<sup>2+</sup> reduces the Na<sup>+</sup> current measured in 0 Ca<sup>2+</sup> by 50% [2,3]. While the main physiological function of the L-type calcium channel is to conduct Ca<sup>2+</sup> in the background of a much higher concentration of monovalent cations, it also discriminates between ions of the same charge (e.g., Ca<sup>2+</sup> vs. Ba<sup>2+</sup> or Na<sup>+</sup> vs. K<sup>+</sup> [4]).

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The physical mechanism of the selectivity of the L-type calcium channel has often been discussed because of the profound medical importance of the channel. In the first theories of permeation, ions were said to "hop" over energy barriers (reviewed by Sather and McCleskey [5], criticized by Eisenberg et al. [6–9]). This kind of helpful but phenomenological description has been replaced by modern theories and computer simulations. Corry et al. [10-12] have studied Ca<sup>2+</sup> selectivity, but their simulations, using canonical ensembles, necessarily used  $Ca^{2+}$  bath concentrations >18 mM - 10,000 times larger than the micromolar affinity of the pore. When grand canonical Monte Carlo (MC) simulations were performed using the model of Corry et al. [13] for the L-type calcium channel, they showed only 200  $\mu$ M Ca<sup>2+</sup> affinity, much less than the natural channel. Just as significantly, their model cannot distinguish between ions of the same valence, but different size [11,12] even though natural channels can do this.

So far, the theory that best describes experimental properties of calcium channels is the charge/space competition (CSC) mechanism. This theory not only produces micromolar  $Ca^{2+}$  affinity for the L-type calcium channel [14–16] but also describes how different calcium channel subclasses can have the wide range of  $Ca^{2+}$  affinities seen in experiments [14]. The CSC model correctly distinguishes between monovalent cations of different size [16–21]. The model uses only a few adjustable parameters to account for the main selectivity

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properties of the calcium channel and its cousin, the sodium channel [22,23].

The CSC mechanism describes ion selectivity in calcium and sodium channels as a balance of the ions' electrostatic attraction into the channel's highly charged selectivity filter and the ions' excluded volume in the small selectivity filter that is crowded with ions and protein side chains [14–31]. In this mechanism, selectivity of Ca<sup>2+</sup> over monovalent cations increases when the number of cations in the pore increases (with the shape and size of the pore unchanged). It also increases when the radius of the pore decreases. Thus, pore volume is a critical parameter for Ca<sup>2+</sup> versus monovalent cation selectivity [24]. Another important parameter is the dielectric coefficient of the protein that surrounds the permeation pathway; a low protein dielectric coefficient (relative to the pore) attracts more cations into the pore and thereby increases Ca<sup>2+</sup> affinity [14,19].

The role of the pore volume in selectivity was first suggested by Nonner et al. [24]. In fact, they suggested that pore volume is the fundamental determinant of  $Ca^{2+}$  versus monovalent cation selectivity. Both theoretical [14,17] and experimental [30,31] studies have shown that reducing the pore *radius* has large effects on this kind of selectivity. To test the CSC mechanism experimentally, Miedema et al. [30,31] chemically modified the pore of the otherwise non-selective OmpF channel. They showed that adding negative charge turned the non-selective porin into a calcium channel and reducing the pore volume improved  $Ca^{2+}$  selectivity.

In this paper, we test the "volume-is-fundamental hypothesis" directly by changing the shape of our model calcium channel. By "shape" we mean the pair of H and R parameters of our reduced model, namely, the length and radius of the cylindrical selectivity filter. We change these two parameters in two ways: (1) by simultaneously changing the volume varying R at fixed H (making the pore wider/narrower) or varying H at fixed R (making the pore longer/shorter) and (2) by keeping the volume fixed (changing the pore from a long and narrow to a short and wide cylinder).

Reducing the pore length (while increasing the pore radius) changes packing effects of the hard-sphere ions both axially and radially. In complex systems like ion binding proteins, details of structure are believed to be of the greatest importance in determining binding selectivity (i.e., how many ions accumulate in the pore) and so one would expect channels of different length to have different selectivity, even if they have the same volume. We find, however, that changes in pore length have relatively little effect on  $Ca^{2+}$ -binding affinity if volume is held constant. If the volume is changed simultaneously with the *H* and *R* parameters, however, selectivity can change substantially in a wide range.

Stronger packing in the axial dimension does have effects on ion currents. It produces deeper depletion zones (zones where the density of certain type of ions is significantly small). It was clear earlier [32] and it was shown explicitly recently [15,21] that these depletion zones determine the amount of current each ionic species conducts, similar to the role of depletion zones in transistors [7,33]. In this paper we analyze the effect of these depletion zones in terms of the integrated Nernst–Planck equation for various shapes of the channel. The effect of the nature of confinement of protein side chains is also examined and found to have small effects.

# 2. Model and methods

#### 2.1. Simple model of a calcium channel

The selectivity filter of the L-type calcium channel contains four glutamates (E) that determine the channel's selectivity properties [34,35] (the EEEE locus). Each glutamate has a terminal COO<sup>-</sup> group so the pore has a charge of -4e. Channel proteins have an atomic structure that in favorable cases can be determined from X-ray diffraction experiments, usually at 100 K [36], although the structures

of calcium channels have not yet been reported. This large-scale structure of alpha helices and the backbones provides a pore with a relatively rigid wall as implied by the stable currents flowing through these channels [37,38]. At the same time, channel proteins have a certain amount of flexibility because they are in a constant thermal motion at room temperature. The most flexible parts of the protein are the side chains. To take into account both aspects in our reduced model, we use a rigid confinement modeled by a pore with hard walls as shown in Fig. 1 and we place mobile protein side chains (the carboxyl groups) inside this confinement. The region -H/2 < x < H/2 defines the selectivity filter. The two important geometrical parameters of this model are the radius *R* and the length *H* of the filter. Fig. 2 shows snapshots of three different channel shapes determined by three different sets of the *R* and *H* values.

The CSC model states that the packing fraction (the volume occupied by the bodies of the side chains relative to the filter volume) and the charge of these side chains are the main determinants of selectivity. Therefore, we chose the simplest model that includes both the charge of the side chains and the volume that they occupy: each carboxyl group is represented as two half-charged, independent oxygen ions (two  $O^{1/2-}$ ) that are free to move within the selectivity filter but are prevented from leaving it (illustrated by red spheres in Fig. 2). The diameter of the oxygen ions is  $d_{\Omega} = 2.8$  Å.

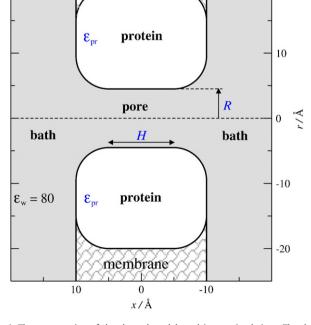
The oxygen ions are confined within the filter by a soft confining potential that is defined by the following Boltzmann factor

$$\exp(-U_{\rm conf}(x) / kT) = 0.5 + 0.5 \tanh[\pm a(x \pm H_{\rm c})], \tag{1}$$

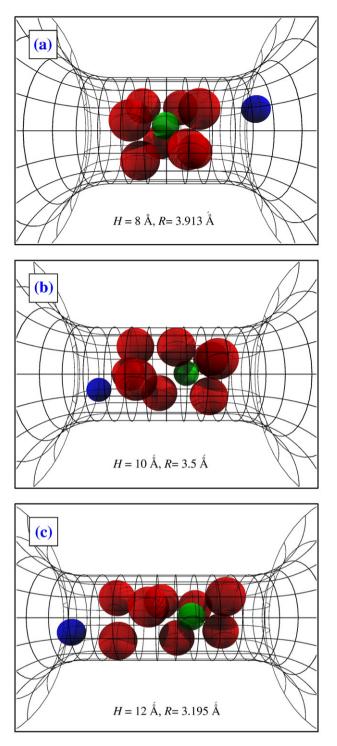
where  $H_c = H/2 - d_0/2$ , *x* is the longitudinal coordinate along the axis of the channel, and the signs are positive for x < 0 and negative for x > 0. Fig. 3 shows this function for three values of the *a* parameter that defines the "softness" of the wall. In the limit  $a \rightarrow \infty$  the usual hard wall confinement is recovered. The Boltzmann factor describes the

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**Fig. 1.** The cross-section of the channel model used in our simulations. The threedimensional model is obtained by rotating it around the *x*-axis. The two important parameters – changed in this paper – are the length H and radius R of the selectivity filter. The figure focuses on the channel and does not show the entire baths; the size of the simulation box is much larger than indicated in the figure. Distances are measured in Ångström throughout this paper.



**Fig. 2.** Panels A–C show snapshots of the packing of the 8  $O^{1/2-}$  ions (red spheres) in the pore axially. The three panels show snapshots of three different geometries used to study the effect of pore shape in Section 2. Green and blue spheres represent Ca<sup>2+</sup> and Na<sup>+</sup>, respectively.

probability that an oxygen ion can be found somewhere along the *x*-axis. Its integral  $V_{\text{eff}} = A \int \exp(-U_{\text{conf}}(x) / kT) dx$  can be identified as the effective volume that is available for the oxygen ion and is the same for different values of *a*:  $V_{\text{eff}} = 2AH_{c}$ .

The dielectric coefficient of the electrolyte solution is  $\varepsilon_w = 80$ , while the dielectric coefficient of the protein is  $\varepsilon_{pr} = 10$ . The low dielectric environment of the pore (1) creates a dielectric barrier for the passing ion (through the interaction with its own induced

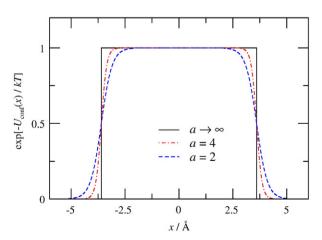
charge) and (2) enhances electrostatic attraction between oppositely charged ions (through the interaction with the induced charge of the counter-ions). These effects are thought to be present in real channels. The second term dominates in the highly charged selectivity filter of calcium channels and makes the filter more selective for Ca<sup>2+</sup> over Na<sup>+</sup> as  $\varepsilon_{\rm pr}$  is decreased [29]. The case when the pore has a dielectric coefficient that is lower than  $\varepsilon_{\rm w}$  (due to, for example, dielectric saturation [39]) is computationally problematic because of ions crossing sharp dielectric boundaries.

The Na<sup>+</sup>, Ca<sup>2+</sup> and Cl<sup>-</sup> ions are free to move everywhere in the solution (in the channel and in the baths). They are modeled as charged hard spheres with diameters 1.9, 1.98 and 3.62 Å, respectively. The simulation cell is a large finite cylinder that includes the baths, the membrane and the channel protein. The simulation cell is chosen to be large enough so that (1) bathing solutions have bulk properties independent of special effects at the walls of the simulation cell is increased in size.

This same model of the EEEE locus has been used to replicate experimental data of the L-type channel and other calcium channels with success [14,15,17–21,24–29]. A nearly identical model accounts for a wide range of data from the DEKA sodium channel and its DEEA mutant (which behaves like a calcium channel [40]) with the same two parameter values, despite the very different properties of the two channel types [23]. A closely related model accounts for a wide range of data for the RyR calcium channel and in fact provided detailed predictions of experimental results before they were measured [18–21].

## 2.2. Monte Carlo simulations

Details of our Metropolis MC simulations are given elsewhere [13,29,41,42]. The ionic concentrations in the bath have to be established accurately because the main output of calculations is the ionic content of the channel as a function of bath electrolyte composition. In order to efficiently simulate micromolar concentrations for  $Ca^{2+}$  in the bath, we used the grand canonical ensemble [43], where the simulation cell is connected to an external bath of fixed chemical potentials through insertion/deletion of neutral ionic groups (1 Na<sup>+</sup> + 1 Cl<sup>-</sup> or 1 Ca<sup>2+</sup> + 2 Cl<sup>-</sup>). The chemical potentials that correspond to prescribed concentrations of these salts were calculated with an iterative grand canonical MC method of Malasics et al. [44]. To accelerate convergence, special insertion/deletion steps were applied, where Na<sup>+</sup> or Ca<sup>2+</sup> were inserted (or deleted) into (or from) the channel itself instead of being inserted (or deleted) into (or from) the entire simulation cylinder [13].



**Fig. 3.** The Boltzmann factor that confines oxygen ions in the selectivity filter for various values of parameter *a* that tunes the "softness" of the confining potential  $U_{\text{conf}}(x)$  (Eq. 1).

The heart of the MC simulation is the energy calculation. In the case of this model, the system contains dielectric boundaries where a polarization surface charge is induced by the ions. This induced charge is determined in every simulation step by the induced charge computation (ICC) method [45], a numerical method to calculate the discretized surface charge on the dielectric interfaces. The method was thoroughly tested and has been shown to be efficient and accurate [24,29,45,46].

The main results of the simulations are the axial density profiles  $c_i(x)$  of ionic species *i*. They are calculated as the average number of a given ionic species in predefined slabs along the *x*-axis divided by the volume of the slab that is accessible to the centers of the ions.

## 2.3. The integrated Nernst-Planck equation

The MC method provides equilibrium density profiles of the various ionic species. In recent papers [15,16], we have used an integrated form of the Nernst–Planck equation to relate equilibrium MC profiles to current. The Nernst–Planck equation (also known as the drift-diffusion or electrodiffusion equation) is

$$-J_i(\mathbf{r}) = \frac{1}{kT} D_i(\mathbf{r}) c_i(\mathbf{r}) \nabla \mu_i(\mathbf{r}), \qquad (2)$$

where  $J_i(\mathbf{r})$  is the particle flux density of species *i*, *k* is the Boltzmann's constant, *T* is the temperature,  $D_i(\mathbf{r})$  is the possibly position-dependent diffusion coefficient, and  $\mu_i(\mathbf{r})$  is the chemical potential. This chemical potential includes the ideal term  $(kT \log c_i(\mathbf{r}))$  and an excess term due to correlation between hard sphere ions and the external voltage across the membrane *V*. We assume that (1) the resistance of the selectivity filter limits the flux, (2) the chemical potential is uniform in the radial dimension, (3) the diffusion coefficients are constant in the selectivity filter, (4) the two baths are identical, but at different constant electric potentials, and that (5) the applied voltage *V* is small so that the current/voltage relation is linear. With these approximations, the conductance of the channel for ionic species *i* is

$$g_{i} = \frac{z_{i}^{2} e^{2} D_{i}}{kT} \frac{1}{\frac{1}{A} \int_{-H/2}^{H/2} \frac{dx}{c_{i}(x)}}.$$
(3)

and the total conductance is  $g = \sum_i g_i$ . Stevens [47] presents this equation, but it seems not to have been used in subsequent literature. Eisenberg [48] shows that the conductance is a rate constant equal exactly to the conditional probability of a well-defined stochastic process.

This equation shows why depletion (low concentration) zones can be so important. The conductance depends on the reciprocal of the integral of the reciprocal of the density profile. Neighboring slabs of the channel along the ionic pathway can be represented as resistors connected in series [15,16]. If the number of charge carriers (ions) in any of these slabs is depleted ( $c_i(x)$  is close to zero), then the integral for this slab (the resistance of the slab) becomes large making the whole resistance large. Its reciprocal (conductance) then becomes small. Thus, deep depletion zones for a given ionic species make the conductance of the channel for this ionic species small as discussed at length by Nonner et al. [32]. They were certainly not the first to understand the importance of depletion zones. Depletion zones determine most properties of semiconductor devices like transistors [33,49,50].

In current/voltage measurements for ion channels, currents (conductances) are commonly given as normalized quantities. In particular, for the classic  $Ca^{2+}$ -block experiment [2,3], where  $CaCl_2$  is added to a fixed NaCl background (30 mM, in this work), the current (conductance) at a given [CaCl<sub>2</sub>] is normalized by the Na<sup>+</sup> current

(conductance) in the absence of  $Ca^{2+}$ . Thus, the total normalized conductance of the channel is

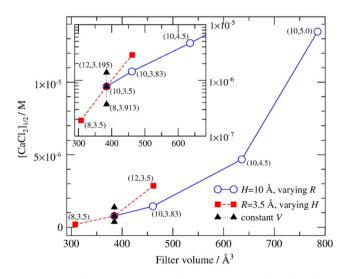
$$g^{*} = \frac{\int \frac{dx}{c_{Na}^{0}(x)}}{\int \frac{dx}{c_{Na}(x)}} + \frac{z_{Ca}^{2}}{z_{Na}^{2}} \frac{D_{Ca}}{D_{Na}} \frac{\int \frac{dx}{c_{Na}^{0}(x)}}{\int \frac{dx}{c_{Ca}(x)}},$$
(4)

where the superscript 0 refers to the density profile at zero [CaCl<sub>2</sub>], while  $c_{Na}(x)$ ,  $c_{Ca}(x)$  and  $g^*$  are taken at a given [CaCl<sub>2</sub>]. The first term of the sum is the normalized Na<sup>+</sup> conductance, while the second term is the normalized Ca<sup>2+</sup> conductance. The density profiles are obtained from simulations and the only adjustable parameter of this model is the ratio of the diffusion coefficients of Ca<sup>2+</sup> and Na<sup>+</sup> in the filter. For this, the value  $D_{Ca}/D_{Na} = 0.1$  was used (the mobility of Ca<sup>2+</sup> is much smaller in the channel, we suppose, because of the large electrostatic attraction imposed by the oxygen ions). The CaCl<sub>2</sub> concentration at which Ca<sup>2+</sup> block occurs is insensitive to this parameter [15].

## 3. Results and discussion

Our basic setup is that of the classical Almers et al. [2,3] experiment which added CaCl<sub>2</sub> to a background concentration of 30 mM NaCl using channel parameters in our model of R = 3.5 Å, H = 10 Å and  $\varepsilon_{\rm pr}$  = 10. These reproduce the micromolar Ca<sup>2+</sup> block. We change the filter radius R and filter length H in various ways to study the effect of these parameters on selectivity.

In the experimental literature, selectivity is commonly characterized by  $[CaCl_2]_{1/2}$ , the  $[CaCl_2]$  at which the current through the channel is half of that found when  $[Ca^{2+}] = 0$ , i.e.,  $g^* = 0.5$  (from Eq. 4). The smaller  $[CaCl_2]_{1/2}$ , the more selective the pore is for  $Ca^{2+}$ . To summarize the dependence of selectivity on various combinations of the parameters *H* and *R*, in Fig. 4 we plot  $[CaCl_2]_{1/2}$  as a function of the filter volume ( $\pi R^2 H$ ). How steeply  $[CaCl_2]_{1/2}$  changes with volume indicates how sensitive the  $Ca^{2+}$  affinity is to filter volume. On the other hand, for a fixed volume, the figure shows how sensitive the  $Ca^{2+}$  affinity is to filter shape (i.e., different combinations of *H* and *R* that keep the volume fixed). Visually, the horizontal change indicates volume sensitivity, while the vertical change indicates shape sensitivity. The inset shows the same data in a logarithmic scale. A detailed discussion of the results in the figure is given in the text below.



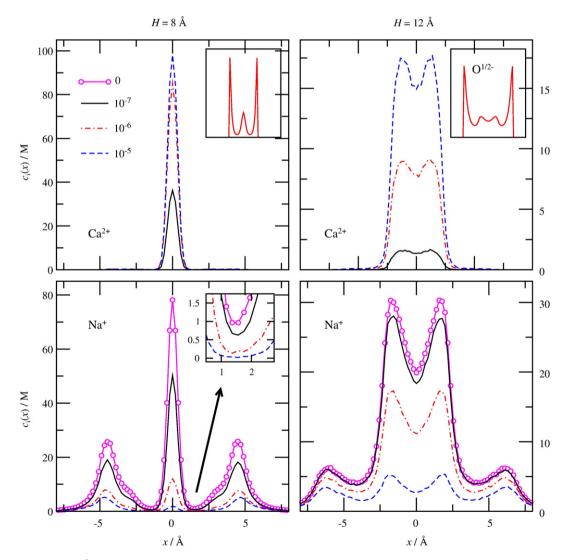
**Fig. 4.** Summary of the simulation results of this work in terms of the  $[CaCl_2]_{1/2}$ , the concentration of  $CaCl_2$  at which channel conductance drops to half its value in the absence of  $CaCl_2$ . This value  $[CaCl_2]_{1/2}$  is plotted against filter volume for various experiments when filter length, filter radius or filter volume are kept constant. The inset plots a part of the main graph for small volumes with a logarithmic scale of the ordinate. The numbers near symbols indicate the values of *H* and *R*.

3.1. Changing pore shape and volume simultaneously has strong effect on selectivity

The effect of pore radius was studied in our previous work [14] by examining the occupancy of the pore by various ions for [NaCl] = 100 mM at fixed channel length H. The concentration of CaCl<sub>2</sub> at which the average number of Na<sup>+</sup> drops to half its value in the absence of Ca<sup>2+</sup> depends sensitively on pore radius. (As Ca<sup>2+</sup> is added to the bath, Na<sup>+</sup> in the filter is gradually replaced by Ca<sup>2+</sup>.) Smaller pore volume makes the selectivity filter even more crowded, which makes it even harder for cations to find space in the high-density filter. This competition favors Ca<sup>2+</sup> because Ca<sup>2+</sup> provides twice the charge to balance the negative charge of the oxygen in the filter than Na<sup>+</sup> while occupying about the same space. This competition can be described in terms of a balance of energy and entropy. By absorbing  $Ca^{2+}$ instead of Na<sup>+</sup>, the filter reduces free energy. It decreases electrostatic energy without decreasing entropy too much. (Note that entropy can also be changed by tethering oxygen ions together as in real COO<sup>-</sup> groups. Such details will be built into the model in future studies.)

Because we first used the integrated Nernst–Planck equation [15] after the publication of the above results [14], we repeated the simulations for [NaCl]=30 mM and analyze the results in terms of conductance. Keeping H=10 Å fixed, we change pore radius and compute the normalized conductance of the channel using Eq. 4 with  $D_{ca}/D_{Na}=0.1$ . The channel shape is changed in a way that the pore is made wider keeping its length fixed (open circles with solid line in Fig. 4). The conclusions that we can draw are the same that were drawn in Ref. [14]: Ca<sup>2+</sup> versus Na<sup>+</sup> selectivity improves considerably with decreasing filter volume. Density profiles (not shown) have more structure in narrower filters, those with smaller R.

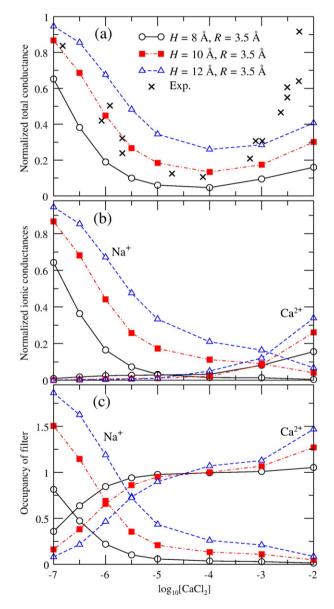
Then, we changed pore length keeping the radius R = 3.5 Å fixed. Fig. 5 shows the density profiles for length H = 8 Å (left panels) and H = 12 Å (right panels) for different Ca<sup>2+</sup> concentrations. Profiles in the short channel have more structure; the depletion zones are more pronounced, and Ca<sup>2+</sup> is more efficient in squeezing Na<sup>+</sup> out from the filter compared to the long channel. In the short channel, the Na<sup>+</sup> profiles decrease almost to half of the zero-Ca<sup>2+</sup> profiles (magenta open circles) at [CaCl<sub>2</sub>]=10<sup>-7</sup> M, while this happens only at [CaCl<sub>2</sub>]=10<sup>-6</sup> M in the case of the long channel. Furthermore, as Ca<sup>2+</sup> gradually replaces Na<sup>+</sup> in the selectivity filter, the depletion



**Fig. 5.** Axial distribution profiles for  $Ca^{2+}$  (top panels) and  $Na^+$  (bottom panels) for a short (H=8 Å, left panels) and a long (H=12 Å, right panels) selectivity filter at different  $Ca^{2+}$  concentrations (R=3.5 Å and  $\varepsilon_{pr}=10$ ). The insets of the top panels show the oxygen profiles for [ $CaCl_2$ ] = 0 M. The profiles are symmetric about x=0 because the simulation system is symmetric. Any deviation from symmetry is a result of statistical uncertainty of the simulation. The inset of the bottom-left panel magnifies the depletion zone region as indicated by the arrow. Concentrations are given in M.

zones of Na<sup>+</sup> become more pronounced in the shorter channel (see inset of Fig. 5, bottom-left).

Fig. 6a shows that as  $Ca^{2+}$  is added to the bath, the current drops at lower  $Ca^{2+}$  concentration in the short channel than in the long channel. The shift of the curves along the abscissa shows the change in  $Ca^{2+}$  selectivity as *H* (and so the volume) is varied. Fig. 4 shows this shift in term of  $[CaCl_2]_{1/2}$  (filled red squares with dashed line). We can change the filter volume in a narrower regime by changing *H* at fixed *R* (as opposed to changing *R* at fixed *H*). The variation of  $[CaCl_2]_{1/2}$  as a function of filter volume is still large as seen in the inset where the results for small volumes are shown on a logarithmic scale. This inset shows that  $log_{10}[CaCl_2]_{1/2}$  changes linearly with pore volume. The inset also shows that  $Ca^{2+}$  affinity is more sensitive to changes in pore length than in pore radius.



**Fig. 6.** The classical anomalous mole fraction experiment: (a) normalized conductances as functions of  $\log_{10}[CaCl_2]$  in M added to a 30-mM NaCl background for channels with varying filter lengths at fixed radius (R = 3.5 Å and  $\varepsilon_{pr} = 10$ ). The × symbols show the experimental results of Almers et al. [2,3]. Their results are different from ours at high [CaCl\_2] because in the experiments Ca<sup>2+</sup> was added only to one side of the pore (increasing Ca<sup>2+</sup> driving force at high [CaCl\_2] [15]), while in our equilibrium simulations Ca<sup>2+</sup> was added symmetrically. (b) Normalized ionic conductances of the pore for Na<sup>+</sup> and Ca<sup>2+</sup> as given by Eq. 4. (c) The average number of Na<sup>2+</sup> and Ca<sup>2+</sup>, while decreasing curves refer to Na<sup>+</sup>.

Eq. 4 makes it possible to separate the currents carried by  $Ca^{2+}$ and Na<sup>+</sup>. Fig. 6b shows these currents with  $D_{Ca}/D_{Na} = 0.1$ . When  $Ca^{2+}$  concentration of the bath is less than  $10^{-4}$  M, only a very small current is carried by  $Ca^{2+}$  (Fig. 6b). Na<sup>+</sup> then carries almost all of the total current (Fig. 6a,b). This Na<sup>+</sup> current, however, is reduced by low micromolar concentrations of Ca<sup>2+</sup>, to the extent that  $Na^+$  in the pore is displaced by  $Ca^{2+}$  (Fig. 6c). It is a rare event that a Ca<sup>2+</sup> ion enters the selectivity filter through the depletion zones, but once it is there, it stays there for a long time. Therefore, this first Ca<sup>2+</sup> ion tends to dwell in the pore but does not contribute significant current. Na<sup>+</sup> ions, on the contrary, can enter and leave the filter and carry current but only in the time when the  $Ca^{2+}$  ion is not there. A substantial Ca<sup>2+</sup> current starts to flow at millimolar  $Ca^{2+}$  concentrations, at which a second  $Ca^{2+}$  tends to occupy the pore (Fig. 6c). Our simulations measure the long-time average of this process, showing a reduced Na<sup>+</sup> occupancy and current.

Therefore, for small bath  $Ca^{2+}$  concentrations (where  $Ca^{2+}$  current is vanishingly small), the total conductance curves (Fig. 6a) reflect only the Na<sup>+</sup> conductances (Fig. 6b), which, in turn, reflect the Na<sup>+</sup> occupancies (Fig. 6c). This implies that, in this regime, the selective binding of  $Ca^{2+}$  to the pore is the first-order determinant of the variation of conductance with [CaCl<sub>2</sub>]. This selective binding is sensitive to the volume of the pore. Depletion zones of Na<sup>+</sup> have little role in this because the Na<sup>+</sup> concentration changes in the depletion zones just as quickly (as a function of [CaCl<sub>2</sub>]) as it changes in the binding site (compare the bottom-left panel of Fig. 5 and its inset).

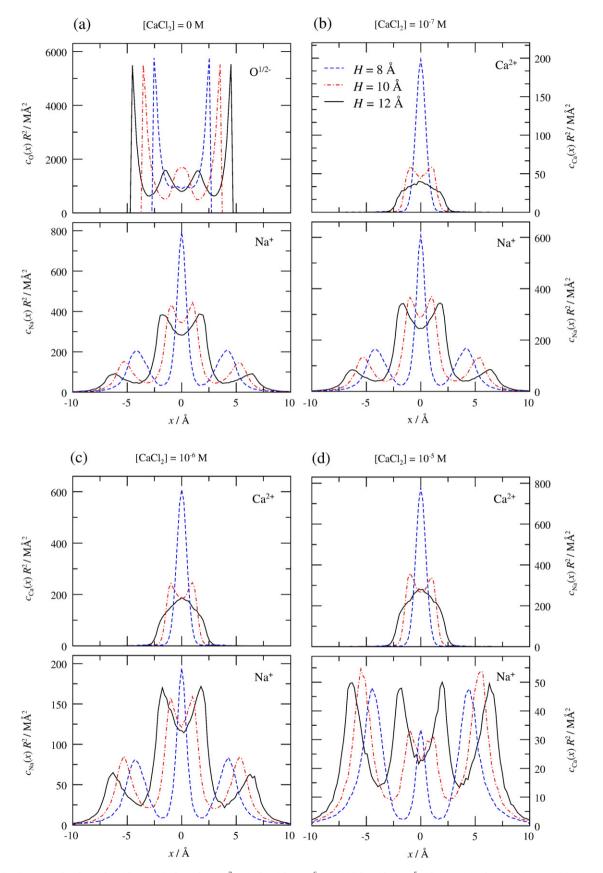
The qualitative findings of Fig. 6a agree with experiments. In a highly selective calcium channel with a small selectivity filter  $Ca^{2+}$  block of current happens at smaller bath [CaCl<sub>2</sub>] and current has a deeper minimum as in the case of the L-type calcium channel [2,3]. In a less  $Ca^{2+}$ -selective calcium channel with a larger selectivity filter,  $Ca^{2+}$  block of current occurs at larger bath [CaCl<sub>2</sub>] with a less pronounced minimum as in the case of the RyR calcium channel [21].

3.2. Changing pore shape under conserved volume has small effect on selectivity

Fig. 4 shows that the shape of the pore keeping the volume fixed has a real, but small effect on selectivity. Consider the results for volume 460.6 Å for which there are two points: one with H = 10 Å and R = 3.83 Å (open circle) and another with H = 12 Å and R = 3.5 Å (filled square). The difference in  $[CaCl_2]_{1/2}$  between these two points implies that shorter pore with larger radius has better Ca<sup>2+</sup>-selectivity properties than a longer pore with smaller radius at the same volume.

To explore this effect, we performed additional simulations on channels with different shapes, but the same volume as the previously studied geometry H = 10 Å and R = 3.5 Å. Fig. 2 shows these channels with dimensions H = 8 Å and R = 3.913 Å, H = 10 Å and R = 3.5 Å and H = 12 Å and R = 3.195 Å. All have the same volume 384.65 Å<sup>3</sup>.

Fig. 7 shows the density profiles of different species for different  $Ca^{2+}$  concentrations and channel lengths. Each panel shows results for a different bath  $Ca^{2+}$  concentration: in Fig. 7a,  $[Ca^{2+}]=0$  M; in Fig. 7b,  $[Ca^{2+}]=10^{-7}$  M; in Fig. 7c,  $[Ca^{2+}]=10^{-6}$  M; in Fig. 7d,  $[Ca^{2+}]=10^{-5}$  M. Each curve in a given panel is computed for a different pore length. The densities are multiplied by  $R^2$ , so the height of curves are proportional to the average number of ions in the cross-section of the pore instead of the local density (compare to Eq. 3 for the conductance).  $Ca^{2+}$  profiles are absent in the case of  $[Ca^{2+}]=0$  M, so we show the curves for the oxygen ions in the top panel of Fig. 7a. They are quite independent of bath  $Ca^{2+}$  concentration. In the longer pores, additional peaks in  $O^{1/2-}$  concentration are found because there is space for additional layers of oxygen ions in the axial dimension. The distributions for Na<sup>+</sup> and  $Ca^{2+}$  are quite different for various pore lengths due to different

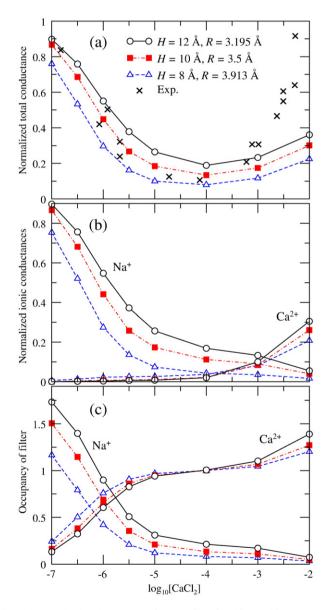


**Fig. 7.** Axial distribution profiles for (a)  $[CaCl_2] = 0 \text{ M}$ , (b)  $[CaCl_2] = 10^{-7} \text{ M}$ , (c)  $[CaCl_2] = 10^{-6} \text{ M}$  and (d)  $[CaCl_2] = 10^{-5} \text{ M}$  for channels of various shapes. Solid, dotted and dashed lines refer to selectivity filters of lengths H = 12, 10 and 8 Å, respectively. The top and bottom panels show  $Ca^{2+}$  and  $Na^+$  profiles, respectively. The top panel of panel a shows oxygen profiles (they are practically the same for every  $Ca^{2+}$  concentration). The profiles are multiplied by  $R^2$  thus obtaining a quantity that is related to the average number of ions along the x-axis and to the conductance defined by the one-dimensional Eq. 3. This profile provides a more straightforward comparison between pores of different cross-sections.

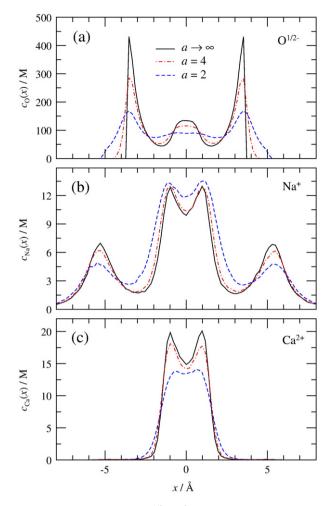
packing, but they change together as  $Ca^{2+}$  concentration changes, which implies that selectivity should be similar in the three cases.

Stronger packing results in considerable depletion zones for Na<sup>+</sup> in the short pore at the locations of the peaks in  $O^{1/2-}$  profiles. These depletion zones decrease the absolute value of the conductance of the channel. The normalized conductance, however, is less influenced because we normalize by the conductance at  $[CaCl_2] = 0$  M, where these depletion zones are also present. As pore length is increased, these depletion zones become less pronounced.

Fig. 8 is analogous to Fig. 6. It shows the same quantities in panels a–c that Fig. 6 does as functions of  $log_{10}[CaCl_2]$ , but here the pore radius is not constant. The curves in Fig. 8a, computed for selectivity filters with the same volume, are closer together than those in Fig. 6a, which were computed for selectivity filters with different volumes, showing that changing pore shape has a larger effect on selectivity if we also change the pore volume. Fig. 4 shows this result in terms of  $[CaCl_2]_{1/2}$  (filled triangles with dotted line).



**Fig. 8.** (a) Normalized conductances as a function of  $\log_{10}[CaCl_2]$  in M added to a 30-mM NaCl background for channels with varying filter lengths and radii while keeping filter volume constant ( $\epsilon_{pr} = 10$ ). The times symbols show the experimental results of Almers et al. [2,3]. (b) Normalized ionic conductances of the pore for Na<sup>+</sup> and Ca<sup>2+</sup> as given by Eq. 4. (c) The average number of Na<sup>2+</sup> and Ca<sup>2+</sup> ions in the selectivity filter. In panels b and c, increasing curves refer to Ca<sup>2+</sup>, while decreasing curves refer to Na<sup>+</sup>.



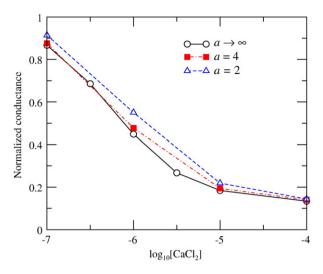
**Fig. 9.** Axial distribution profiles for  $O^{1/2-}$ ,  $Ca^{2+}$  and  $Na^+$  (from top to bottom) for various values of the "softness" parameter *a* for  $[CaCl_2] = 10^{-6}$  M (channel parameters are R = 3.5 Å, H = 10 Å and  $\varepsilon_{pr} = 10$ ).

Similar conclusions can be drawn from Fig. 8 as from Fig. 6: The occupancy curves for Na<sup>+</sup> (Fig. 8c) behave similarly to Na<sup>+</sup>-conductance curves (Fig. 8b) in the regime of Ca<sup>2+</sup> block ([CaCl]<sub>2</sub> <10<sup>-5</sup> M), which implies that the block is a result of selective binding of Ca<sup>2+</sup> over Na<sup>+</sup> in the selectivity filter; that is, as the number of Na<sup>+</sup> ions decreases in the filter, the current carried by them decreases simultaneously.

The main difference between Fig. 8 and Fig. 6 is that the curves for different combinations of H and R are farther apart when filter volume is changed (Fig. 6) than in the case when it is unchanged (Fig. 8). We conclude that changing the pore shape has less effect if the pore volume is conserved. Thus, pore shape (at constant volume) provides an additional engineering variable with which we can fine-tune the selectivity properties of the model. Pore radius can be especially important in the competition between ions of different size, in particular for the sodium channel, where Na<sup>+</sup> versus K<sup>+</sup> selectivity seems to arise from a deeper depletion zone for K<sup>+</sup> than for Na<sup>+</sup> [23].

#### 3.3. Softness of oxygen confinement does not influence selectivity

Our results so far imply that particle density in the filter and packing effects resulting in depletion zones are important determinants of selectivity. Packing of oxygen ions is strongly influenced by the way we confine them. In our earlier studies (and in the simulations shown so far in this work), the oxygen was confined by hard walls at  $\pm H/2$  (corresponding to  $a \rightarrow \infty$  in Eq. 1). This hard wall confinement results in an artificially high peak of oxygen at these



**Fig. 10.** Normalized conductances as a function of  $\log_{10}[\text{CaCl}_2]$  in M added to a 30-mM NaCl background for values of the "softness" parameter *a* (channel parameters are R = 3.5 Å, H = 10 Å and  $\varepsilon_{\text{pr}} = 10$ ). The × symbols show the experimental results of Almers et al. [2,3].

walls which is a well-known hard sphere packing effect (see, for example, insets of Fig. 5). In real selectivity filters, made by proteins, confinement is unlikely to be so hard. Therefore, we examine the role of the hard wall. Does the hard wall produce artifactual depletion zones of  $Ca^{2+}$  or  $Na^+$ ? Does it have any artificial effect that would seriously influence our conclusions drawn from this channel model?

We performed simulations with different values of the *a* parameter of the soft confining potential given in Eq. 1 for the geometry H = 10 Å and R = 3.5 Å. As expected, the distribution of  $O^{1/2-}$  ions is significantly influenced by the way they are confined as shown in the top panel of Fig. 9 for [CaCl<sub>2</sub>] =  $10^{-6}$  M. The oxygen ions distribute over the filter more evenly when a softer confinement allows them more flexibility at the filter entrances; the high peaks are suppressed. Remarkably, the profiles of Na<sup>+</sup> and Ca<sup>2+</sup> are much less influenced. The conductance of the channel is the same whether the channel is constructed with hard or soft walls (Fig. 10) even though the resulting distribution of the side chains is quite different.

This is a satisfying result because it supports our hypothesis: it is the confinement provided by the protein and the density of the charges in the selectivity filter that really matter, and as long as those coarse features are maintained, then smaller details of protein structure have minor effects on selectivity.

#### 4. Summary

Our reduced model of the selectivity filter is not intended to be a realistic structural representation of the calcium channel. Instead, it is used to study the role of fundamental classical physical interactions (electrostatics and hard sphere exclusion) and how these terms create selectivity to first order. The success of the model and the CSC mechanism implies that these terms should be present in more detailed models of the calcium channel. Molecular dynamics simulations of atomistic models of channels have been chiefly concentrated on potassium channels whose structure is known [51-55]. Simulations of assumed atomic structures of calcium channels using that of the potassium channel as a template have also been published [56-58]. Neither of these simulations, nevertheless, relate their results to experimental conductivities as functions of varying bath compositions as we do. Atomic structure, nevertheless, has a close relation to the parameters of our reduced model. The polarization properties of the protein surrounding the pore ( $\varepsilon_{pr}$  in our model), as well as the length H and radius R of the selectivity filter are all structural parameters that are essentially determined by protein structure. In place of highresolution structure (which is not available for  $Ca^{2+}$  channels), we model the bulk of the protein in an abstract, low-resolution structure. This structure is fully specified by a small number of parameters whose physical meaning is self-evident.

Our simulations showed that ionic density is an important determinant of  $Ca^{2+}$  versus  $Na^+$  selectivity in calcium channels. If the number of structural ions in the filter (8  $O^{1/2-}$  ions in this study) and polarization properties (dielectric coefficient of the protein) are fixed, it is the volume that determines the density. Simulations with different pore shapes with and without conserved volume showed that variation in selectivity is larger if the volume is changed. Channel shape at fixed volume is also important, but its influence is smaller.

Strong variations of particle packing have surprisingly small significance for selective (relative) conduction, but they do have interesting consequences regarding the absolute value of conductance (for example, the size of the Na<sup>+</sup> leak at physiological [Ca<sup>2+</sup>]). Finally, we have shown that confining the oxygen ions by soft walls instead of hard walls has little effect on selectivity.

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#### References

- B. Hille, Ion Channels of Excitable Membranes, Sinauer Associates Inc., Sunderland, 2001.
- [2] W. Almers, E.W. McCleskey, P.T. Palade, Non-selective cation conductance in frog muscle membrane blocked by micromolar external calcium ions, J. Physiol. 353 (1984) 565–583.
- [3] W. Almers, E. McCleskey, Non-selective conductance in calcium channels of frog muscle: calcium selectivity in a single-file pore, J. Physiol. 353 (1984) 585–608.
- [4] P. Hess, J.B. Lansman, R.W. Tsien, Calcium channel selectivity for divalent and monovalent cations. Voltage and concentration dependence of single channel current in ventricular heart cells, J. Gen. Physiol. 88 (1986) 293–319.
- [5] W.A. Sather, E.W. McCleskey, Permeation and selectivity in calcium channels, Annu. Rev. Physiol. 65 (2003) 133–159.
- [6] K.E. Cooper, P.Y. Gates, R.S. Eisenberg, Diffusion theory and discrete rate constants in ion permeation, J. Membr. Biol. 109 (1988) 95–105.
- [7] R.S. Eisenberg. New developments and theoretical studies of proteins, chap. Atomic biology, electrostatics, and ionic channels. World Scientific, Philadelphia (1996), 269-357. URL http://arxiv.org/abs/0807.0715v1.
- [8] D. Chen, L. Xu, A. Tripathy, G. Meissner, B. Eisenberg, Permeation through the calcium release channel of cardiac muscle, Biophys. J. 73 (1997) 1337–1354.
- [9] R.S. Eisenberg, From structure to function in open ionic channels, J. Membr. Biol. 171 (1999) 1–24.
- [10] B. Corry, T. Allen, S. Kuyucak, S.-H. Chung, Biophys. J. 80 (2001) 195-214.
- [11] B. Corry, T. Vora, S.-H. Chung, Electrostatic basis of valence selectivity in cationic channels, Biochim. Biophys. Acta 1711 (2005) 72–86.
- [12] S.H. Chung, B. Corry, Three computational methods for studying permeation, selectivity and dynamics in biological ion channels, Soft Matter 1 (2005) 417–427.
- [13] D. Boda, W. Nonner, D. Henderson, B. Eisenberg, D. Gillespie, Volume exclusion in calcium selective channels, Biophys. J. 94 (2008) 3486–3496.
- [14] D. Boda, M. Valiskó, B. Eisenberg, W. Nonner, D. Henderson, D. Gillespie, Combined effect of pore radius and protein dielectric coefficient on the selectivity of a calcium channel, Phys. Rev. Lett. 98 (2007) 168102.
- [15] D. Gillespie, D. Boda, The anomalous mole fraction effect in calcium channels: a measure of preferential selectivity, Biophys. J. 95 (2008) 2658–2672.
- [16] D. Boda, M. Valiskó, D. Henderson, B. Eisenberg, D. Gillespie, W. Nonner, Ion selectivity in L-type calcium channels by electrostatics and hard-core repulsion, J. Gen. Physiol. 133 (2009) 497–509.
- [17] D. Boda, D. Henderson, D.D. Busath, Monte Carlo study of the effect of ion and channel size on the selectivity of a model calcium channel, J. Phys. Chem. B 105 (2001) 11574–11577.
- [18] D. Gillespie, L. Xu, Y. Wang, G. Meissner, (De)constructing the ryanodine receptor: modeling ion permeation and selectivity of the calcium release channel, J. Phys. Chem. B 109 (2005) 15598–15610.
- [19] D. Gillespie, Energetics of divalent selectivity in a calcium channel: the Ryanodine Receptor Case Study, Biophys. J. 94 (2008) 1169–1184.
- [20] D. Gillespie, M. Fill, Intracellular calcium release channels mediate their own countercurrent: the ryanodine receptor case study, Biophys. J. 95 (2008) 3706–3714.
- [21] D. Gillespie, J. Giri, M. Fill, Reinterpreting the anomalous mole fraction effect: the Ryanodine Receptor Case Study, Biophys. J. 97 (2009) 2212–2221.

- [22] D. Boda, D.D. Busath, B. Eisenberg, D. Henderson, W. Nonner, Monte Carlo simulations of ion selectivity in a biological Na channel: charge-space competition, Phys. Chem. Chem. Phys. 4 (2002) 5154–5160.
- [23] D. Boda, W. Nonner, M. Valiskó, D. Henderson, B. Eisenberg, D. Gillespie, Steric selectivity in Na channels arising from protein polarization and mobile side chains, Biophys. J. 93 (2007) 1960–1980.
- [24] W. Nonner, L. Catacuzzeno, B. Eisenberg, Binding and selectivity in L-type calcium channels: a mean spherical approximation, Biophys. J. 79 (2000) 1976–1992.
- [25] W. Nonner, D. Gillespie, D. Henderson, B. Eisenberg, Ion accumulation in a biological calcium channel: effects of solvent and confining pressure, J. Phys. Chem. B 105 (2001) 6427–6436.
- [26] D. Boda, D.D. Busath, D. Henderson, S. Soko lowski, Monte Carlo simulations of the mechanism for channel selectivity: the competition between volume exclusion and charge neutrality, J. Phys. Chem. B 104 (2000) 8903–8910.
- [27] D. Boda, D. Henderson, D.D. Busath, Monte Carlo study of the selectivity of calcium channels: improved geometrical model, Mol. Phys. 100 (2002) 2361–2368.
- [28] D. Boda, D. Henderson, Computer simulation of the selectivity of a model calcium channel, J. Phys.: Cond. Matt. 14 (2002) 9485–9488.
- [29] D. Boda, M. Valiskó, B. Eisenberg, W. Nonner, D. Henderson, D. Gillespie, The effect of protein dielectric coefficient on the ionic selectivity of a calcium channel, J. Chem. Phys. 125 (2006) 034901.
- [30] M. Vrouenraets, J. Wierenga, W. Meijberg, H. Miedema, Chemical modification of the bacterial Porin OmpF: gain of selectivity by volume reduction, Biophys. J. 90 (2006) 1202–1211.
- [31] H. Miedema, M. Vrouenraets, J. Wierenga, D. Gillespie, B. Eisenberg, W. Meijberg, W. Nonner, Ca<sup>2+</sup> selectivity of a chemically modified OmpF with reduced pore volume, Biophys. J. 91 (2006) 4392–4400.
- [32] W. Nonner, D.P. Chen, B. Eisenberg, Anomalous mole fraction effect, electrostatics, and binding in ionic channels, Biophys. J. 74 (1998) 2327–2334.
- [33] R.S. Eisenberg. Living transistors: a physicist's view of ion channels. URL http:// arxiv.org/abs/q-bio/0506016v2. Version 2.
- [34] J. Yang, P.T. Ellinor, W.A. Sather, J.F. Zhang, R.W. Tsien, Molecular determinants of Ca<sup>2+</sup>
- selectivity and ion permeation in L-type Ca<sup>2+</sup> channels, Nature 366 (1993) 158–161. [35] P.T. Ellinor, J. Yang, W.A. Sather, J.F. Zhang, R.W. Tsien, Ca<sup>2+</sup> interactions, Neuron
- [35] P.T. Ellinor, J. Yang, W.A. Sather, J.F. Zhang, R.W. Tsien, Ca<sup>2+</sup> interactions, Neuron 15 (1995) 1121–1132.
- [36] D.A. Doyle, J.M. Cabral, R.A. Pfuetzner, A.L. Kuo, J.M. Gulbis, S.L. Cohen, B.T. Chait, R. MacKinnon, The structure of the potassium channel: molecular basis of K<sup>+</sup> conduction and selectivity, Science 280 (1998) 69–77.
- [37] F.J. Sigworth, Covariance of nonstationary sodium current fluctuations at the node of Ranvier, Biophys. J. 34 (1981) 111–133.
- [38] F.J. Sigworth, Voltage gating of ion channels, Q. Rev. Biophys. 27 (1994) 1-40.
- [39] M. Aguilella-Arzo, A. Andrio, V.M. Aguilella, A. Alcaraz, Dielectric saturation of water in a membrane protein channel, Phys. Chem. Chem. Phys. 11 (2009) 358–365.

- [40] S.H. Heinemann, H. Teriau, W. Stuhmer, K. Imoto, S. Numa, Calcium-channel characteristics conferred on the sodium-channel by single mutations, Nature 356 (1992) 441–443.
- [41] M.P. Allen, D.J. Tildesley, Computer Simulation of Liquids, Oxford, New York, 1987.
- [42] D. Frenkel, B. Smit, Understanding Molecular Simulations, Academic Press, San Diego, 1996.
- [43] J.P. Valleau, L.K. Cohen, Primitive model electrolytes. 1. grand canonical Monte-Carlo computations, J. Chem. Phys. 72 (1980) 5935–5941.
- [44] A. Malasics, D. Gillespie, D. Boda, Simulating prescribed particle densities in the grand canonical ensemble using iterative algorithms, J. Chem. Phys. 128 (2008) 124102.
- [45] D. Boda, D. Gillespie, W. Nonner, D. Henderson, B. Eisenberg, Computing induced charges in inhomogeneous dielectric media: application in a Monte Carlo simulation of complex ionic systems, Phys. Rev. E 69 (2004) 046702.
- [46] D. Boda, D. Gillespie, B. Eisenberg, W. Nonner, D. Henderson, Ionic soft matter: Novel trends in theory and applications, vol. 206 of NATO Science Series: II: Mathematics, Physics and Chemistry, chap. The Induced Charge Computation method and its application in Monte Carlo simulations of inhomogeneous dielectric systems, Springer, 2005.
- [47] C.F. Stevens, Neurophysiology: A Primer, John Wiley, New York, 1966.
- [48] B. Eisenberg. Biophysics textbook on line "Channels, receptors, and transporters", chap. Permeation as a diffusion process. URL http://arxiv.org/abs/0807.0721.
- [49] S.W., Electrons and Holes in Semiconductors to Applications in Transistor Electronics, Van Nostrand, New York, 1950.
- [50] S.M., Physics of Semiconductor Devices, Prentice Hall, New York, 1990.
- [51] N.S. Y. B. Roux, Importance of hydration and dynamics on the selectivity of the KcsA and NaK channels, J. Gen. Physiol. 129 (2007) 135–143.
- [52] S. Varma, S.B. Rempe, Tuning ion coordination architectures to enable selective partitioning, Biophys. J. 93 (2007) 1093–1099.
- [53] G.V. Miloshevsky, P.C. Jordan, Conformational changes in the selectivity filter of the open state KcsA channel: an energy minimization study, Biophys. J. 95 (2008) 3239–3251.
- [54] F.P.W., K. Tai, S.M.S., The selectivity of K<sup>+</sup> ion channels: testing the hypotheses, Biophys. J. 95 (2008) 5062–5072.
- [55] D.L. Bostick, A.K., B.C.L., K<sup>+</sup> /Na<sup>+</sup> selectivity in toy cation binding site models is determined by the 'host', Biophys. J. 96 (2009) 3887–3896.
- [56] G.M. Lipkind, H.A. Fozzard, Modeling of the outer vestibule and selectivity filter of the L-type Ca<sup>2+</sup> channel, Biochemistry 40 (2001) 6786–6794.
- 57] G. Barreiro, C.R.W. Guimaraes, R.B. de Alencastro, A molecular dynamics study of an L-type calcium channel model, Protein Eng. 15 (2002) 109–122.
- [58] G. Barreiro, C.R.W. Guimaraes, R.B. de Alencastro, Potential of mean force calculations on an L-type calcium channel model, Protein Eng. 16 (2003) 209–215.