

Convective and adsorptive removal of β_2 -microglobulin during predilutional and postdilutional hemofiltration

ROBERTO PADRINI, CRISTINA CANOVA, PIERO CONZ, ELENA MANCINI, EMANUELA RIZZIOLI, and ANTONIO SANTORO

Department of Pharmacology and Anesthesiology, University of Padova, Padova, Italy; Department of Nephrology and Dialysis, S. Orsola-Malpighi Hospital, Bologna, Italy; and Department of Nephrology and Dialysis, Monselice Hospital, Monselice (PD), Italy

Convective and adsorptive removal of β_2 -microglobulin during predilutional and postdilutional hemofiltration.

Background. Beta₂-microglobulin (β_2 -m) removal in patients with end-stage renal disease (ESRD) is maximal with convective techniques, such as hemofiltration (HF) or hemodiafiltration (HDF). Although the infusion mode of the replacement solution (predilution or postdilution) is expected to influence the efficiency of HF, experimental data in this respect are scanty. We therefore investigated the impact of the fluid re-infusion mode on the efficiency of HF in 11 ESRD patients who underwent both treatments.

Methods. The dialyzer (AK 200 ULTRA) was equipped with a 3-layer polyamide membrane (Poliflux 21 S, surface 2.1 m²) and blood flow was kept between 300 and 400 mL/min. β_2 -m concentrations were measured in plasma water and ultrafiltrate at appropriate times during a 240-minute treatment. The following dialytic parameters were calculated: total amount of β_2 -m removed (A_{tot}), β_2 -m removed by convection (A_{con}) and by adsorption (A_{ads}), percent reduction in β_2 -m plasma water concentration (% $C_{pw,in}$), total plasma water clearance ($CL_{pw,tot}$), convective plasma water clearance ($CL_{pw,con}$), adsorptive plasma water clearance ($CL_{pw,ads}$), and sieving coefficient (SC).

Results. $CL_{pw,tot}$, $CL_{pw,ads}$, and % $C_{pw,in}$ were similar in pre- and postdilutional conditions, whereas $CL_{pw,con}$ and SC were higher and $CL_{pw,ads}$ was lower in postdilution than in predilution HF. Since a significant inverse correlation was found between A_{ads} and SC, predilution probably determines greater protein fouling than postdilution.

Conclusion. The 2 techniques appear to be equivalent in terms of total β_2 -m removal, although this final result is obtained by different contributions of convective and adsorptive elimination.

Beta₂-microglobulin (β_2 -m) is a low-molecular-weight protein that is almost exclusively eliminated by glomerular

filtration and accumulates in the plasma of patients with end-stage renal disease (ESRD) [1]. The persistently high β_2 -m plasma levels during long-term dialytic therapy are thought to play a pathogenetic role in amyloid deposition in the osteo-articular system and related clinical signs (carpal tunnel syndrome, peripheral osteoarthropathy) [2,3]. The ability to remove β_2 -m varies considerably among the available dialysis techniques, being negligible with low-flux hemodialysis (HD), intermediate with high-flux HD, and greatest with techniques employing convective transport, such as hemodiafiltration (HDF) [4]. Although no renal replacement treatment proved to normalize β_2 -m plasma levels [4], it is noteworthy that ESRD patients undergoing convective procedures have a 42% reduction of risk for carpal tunnel syndrome surgery, compared with those submitted to diffusive procedures (HD) [5]. Accordingly, it seems logical to keep β_2 -m plasma concentrations as low as possible by means of the most suitable renal replacement therapy. By using convective transport techniques, the efficiency of solute removal chiefly depends on the membrane characteristics and the infusion site (before or after the filter). Membrane composition, structure, and thickness determine the relative amount of β_2 -m removed by convection or adsorption [6–8]; the 2 alternative infusion modes offer different potential advantages and drawbacks [9–12]. Investigations comparing β_2 -m removal in pre- and postdilution modes have mainly been carried out using the convective-diffusive HDF technique [13–15]. Little information exists on this issue concerning the pure convective HF technique [16]. The main aim of our study was to evaluate the β_2 -m kinetics in postdilution as well as in predilution hemofiltration. As a secondary aim, we analyzed the single separate effects of both convection and adsorption by the dialysis membrane on the β_2 -m overall removal. The study was planned as an in vivo study in order to take into account the effect of the plasma proteins and, consequently, the protein-cake on both the components, convective and adsorptive, of β_2 -m removal.

Key words: beta₂-microglobulin, hemofiltration, predilution, postdilution.

Received for publication January 12, 2005
and in revised form March 24, 2005
Accepted for publication June 1, 2005

© 2005 by the International Society of Nephrology

Table 1. Patient demographic and clinical characteristics ($N = 11$)

Sex	4 F; 7 M
Age years	66.8 \pm 11.9 (46–86) ^a
Weight kg	64.1 \pm 9.4 (45–82) ^a
Time on dialysis months	80.9 \pm 66.9 (14–242) ^a
Renal disease	Glomerulonephritis ($N = 3$) Polycystic disease ($N = 1$) Nephroangiosclerosis ($N = 4$) Interstitial nephritis ($N = 2$) End-stage renal disease ($N = 1$)

^aRange.

METHODS

Patients and HF set-up

Eleven uremic patients (7 males, 4 females, mean age 67 \pm 12 years, weighing 64 \pm 9 kg) who had been on periodical renal replacement therapy for at least 3 months entered this study after providing their informed consent. The study was approved by the local Ethics Committee and was conducted in compliance with the principles of the Declaration of Helsinki. The inclusion criteria were age > 18 years and permanent blood access granting a flow rate of at least 300 mL/min. The exclusion criteria were the occurrence of severe vascular instability during treatment, active HBV or HCV hepatitis, HIV positivity, coagulation disorders, and neoplasms. Patients' demographic and clinical characteristics are listed in Table 1.

Each patient underwent 2 online hemofiltration treatments, 1 in predilution (pre-HF) and 1 in postdilution mode (post-HF). The dialyzer (AK 200 ULTRA; Gambro, Lund, Sweden) was equipped with a 3-layer polyamide membrane (Poliflux 21 S, surface 2.1 m²) and with a system that prepared sterile substitution solution online. A reverse-bi-osmosis system was used for the treatment of the main tap water. The incoming reverse-osmosis-treated water is filtered in a polyamide ultrafilter before entering the dialysis monitor. The infusion solution is then prepared on-line by the AK200 ULTRA, as a sterile, nonpyrogenic solution, by stepwise ultrafiltration of water and bicarbonate-containing dialysis fluid (BiCart), thanks to 2 polyamide ultrafilters sterilizing water, the first inserted into the machine, the second, disposable, inserted in the infusion line.

Blood flow was kept between 300 and 400 mL/min. The exchange volume was \approx 45% and \approx 100% of the body weight in post-HF and pre-HF, respectively. Previous and hitherto unpublished observations of ours have shown that these exchange volumes could guarantee an equivalence in the urea removal.

The main treatment parameters are shown in Table 2.

Hematocrit (Hct), total plasma protein, plasma urea, and creatinine concentrations were measured at the start and at end of each treatment; their values are listed in Table 3.

Table 2. Treatment parameters

	Predilution HF	Postdilution HF	<i>P</i>
Fluid replacement <i>L</i>	61.3 \pm 15.7	25.6 \pm 3.5	<0.001
Body weight decrease <i>kg</i>	2.9 \pm 0.6	3.0 \pm 0.6	NS
Blood flow (Qb) <i>mL/min</i>	317.3 \pm 18.49	338.2 \pm 26.4	<0.05
Ultrafiltration rate (Quf) <i>mL/min</i>	273.7 \pm 56.4	121.4 \pm 14.9	<0.001
Infusion rate (Qinf) <i>mL/min</i>	262.9 \pm 56.1	109.0 \pm 15.0	<0.001
Treatment duration <i>minutes</i>	240 \pm 8	240 \pm 3	NS

β 2-microglobulin concentration measurements

The β 2-microglobulin (β 2-m) plasma concentrations entering the hemofilter ($C_{p_{in}}$) have been determined immediately before starting HF (time 0) and after 30, 60, 120, 180, and 240 minutes. In pre-HF mode, blood samples were taken before dilution with replacement fluid. The sampling times for the β 2-m concentrations in plasma leaving the hemofilter ($C_{p_{out}}$) and in the ultrafiltrate (C_{uf}) were the same except for the first time, which was 5 minutes instead of 0 minutes (when $C_{p_{out}}$ and C_{uf} are presumed to be zero). In post-HF mode, $C_{p_{out}}$ was measured before dilution with replacement fluid.

β 2-m concentrations were assayed by means of a turbidimetric method using rabbit antihuman β 2-m for agglutination (Cobas Mira, Roche, Rotkreuz, Switzerland).

Pharmacokinetic analysis

The time courses of $C_{p_{in}}$, $C_{p_{out}}$, and C_{uf} were empirically described by cubic spline curves. The area under the curve (AUC) of β 2-m concentrations versus time (0–4 h) was calculated for $C_{p_{in}}$ ($AUC_{p_{in}}$), $C_{p_{out}}$ ($AUC_{p_{out}}$), and C_{uf} (AUC_{uf}) by means of the trapezoidal rule.

Mass transfer and clearance calculations

The total amount of β 2-m removed from plasma during the 4-hour HF period (A_{tot}) was calculated by the formula:

$$A_{tot} = (AUC_{b_{in}} - AUC_{b_{out}}) \times Q_b \quad (\text{Equation 1})$$

where $AUC_{b_{in}}$ is the AUC of β 2-m concentrations in the blood entering the hemofilter, $AUC_{b_{out}}$ is the AUC of β 2-m concentrations in the blood leaving it, and Q_b is blood flow. Since β 2-m is distributed outside red cells, β 2-m blood concentrations (C_b) can be calculated by using the β 2-m plasma concentration (C_p) and the hematocrit value expressed as a decimal fraction (Hct):

$$C_b = C_p \times (1 - \text{Hct}) \quad (\text{Equation 2})$$

The Hct of the blood entering the hemofilter (Hct_{in}) was directly measured, and that leaving it (Hct_{out}) were

Table 3. Hematochemical parameters at start and end of HF

	Start				End			
	Hct %	Proteins g/dL	Urea mg/dL	Creat mg/dL	Hct %	Proteins g/dL	Urea mg/dL	Creat mg/dL
Pre-HF	32.7 ± 4.3	6.40 ± 0.66	145.1 ± 30.0	8.91 ± 1.78	36.6 ^b ± 5.0	7.43 ^a ± 0.84	59.4 ^b ± 19.6	3.88 ^b ± 1.15
Post- HF	32.7 ± 3.5	6.57 ± 0.56	135.6 ± 49.0	8.30 ± 1.74	35.9 ^a ± 4.7	7.28 ^a ± 0.87	61.3 ^b ± 21.3	3.45 ^b ± 1.03

^a*P* < 0.001; ^b*P* < 0.0001.

calculated from Hct_{in}, Q_b, and the input/output fluid balance, according to the following formulas:

$$\text{for predilutional HF : Hct}_{\text{out}} = \frac{(Q_b \times \text{Hct}_{\text{in}})}{(Q_b - Q_{\text{uf}} + Q_{\text{inf}})} \quad \text{(Equation 3)}$$

$$\text{for postdilutional HF : Hct}_{\text{out}} = \frac{(Q_b \times \text{Hct}_{\text{in}})}{(Q_b - Q_{\text{uf}})} \quad \text{(Equation 4)}$$

where Q_{uf} is the ultrafiltration flow and Q_{inf} is the infusion rate. (See **Appendix** for calculations yielding Equations 3 and 4).

The amount of β 2-m removed by convection [i.e., recovered in the ultrafiltrate during the 4-hour HF period (A_{con})], was calculated by the formula:

$$A_{\text{con}} = \text{AUC}_{\text{uf}} \times Q_{\text{uf}} \quad \text{(Equation 5)}$$

The amount of β 2-m removed by membrane adsorption (A_{ads}) was calculated as the difference between A_{tot} and A_{con}.

The mean dialytic β 2-m plasma water clearances for total, convective, and adsorptive removal were calculated using A_{tot}, A_{con}, and A_{ads}, respectively:

$$\text{CLpw}_{\text{tot}} = A_{\text{tot}} / \text{AUCp}_{\text{w}_{\text{in}}} \quad \text{(Equation 6)}$$

$$\text{CLpw}_{\text{con}} = A_{\text{con}} / \text{AUCp}_{\text{w}_{\text{in}}} \quad \text{(Equation 7)}$$

$$\text{CLpw}_{\text{con}} = A_{\text{ads}} / \text{AUCp}_{\text{w}_{\text{in}}} \quad \text{(Equation 8)}$$

where AUCp_{w_{in}} is the AUC of β 2-m plasma water concentrations entering the hemofilter (Cp_{w_{in}}). Cp_{w_{in}} was obtained from Cp_{in} and the total plasma protein concentration (Pt) using Colton's formula [17]:

$$\text{Cp}_{\text{w}_{\text{in}}} = \text{Cp}_{\text{in}} / (1 - 0.0107 \times \text{Pt}) \quad \text{(Equation 9)}$$

where the Pt unit is g/dL.

To investigate possible time-dependent changes in β 2-m clearances, CLpw_{tot}, CLpw_{to}, CLpw_{tot} were also determined at each sampling interval (0–0.5, 0.5–1, 1–2, 2–3, 3–4 hours).

As an additional index of β 2-m removal, the percentage difference between Cp_{w_{in}} measured at the end and start of HF (% Cp_{w_{in}}) was calculated.

The sieving coefficient was calculated for convective β 2-m removal as the ratio between measured CLpw_{con} and maximal theoretical plasma water clearance (CLpw_{max}, i.e., the CLpw expected if S = 1), which are:

$$\text{for postdilutional HF} \quad \text{CLpw}_{\text{max}} = Q_{\text{uf}}; \quad \text{(Equation 10)}$$

$$\text{for predilutional HF} \quad \text{CLpw}_{\text{max}} = Q_{\text{pw}} \times Q_{\text{uf}} / (Q_{\text{pw}} + Q_{\text{inf}}) \quad \text{(Equation 11)}$$

where Q_{pw} is mean the plasma water flow during the 4-hour HF period. Q_{pw} was calculated from the mean plasma flow [= Q_b × (1 – Hct_{mean})] corrected by the mean plasma protein concentration:

$$Q_{\text{pw}} = Q_b \times (1 - \text{Hct}_{\text{mean}}) \times (1 - 0.0107 \times \text{Pt}_{\text{mean}}) \quad \text{(Equation 12)}$$

Statistical analysis

Data are reported as mean ± SD. Comparisons between HF parameters measured in pre-HF and post-HF conditions were carried out by means of Student *t* test for paired data or the Wilcoxon paired rank test (for not normally distributed data). Time-related differences in fractional clearances and sieving coefficients were tested by means of a one-way analysis of variance (ANOVA) for repeated measures, followed by the Newman-Keuls post-hoc test. Correlations between parameters were assessed by linear regression analysis and the determination coefficient (r²). The level of significance was considered as *P* < 0.05.

RESULTS

The urea and creatinine reduction ratios were 59 ± 11% and 56 ± 10%, respectively, in pre-HF, and 54 ± 7% and 58 ± 10% in post-HF, with the differences between the 2 modes not being significant.

The time courses of the mean β 2-m concentrations in the plasma (Cp_{in}, Cp_{out}) and the ultrafiltrate (C_{uf}), after

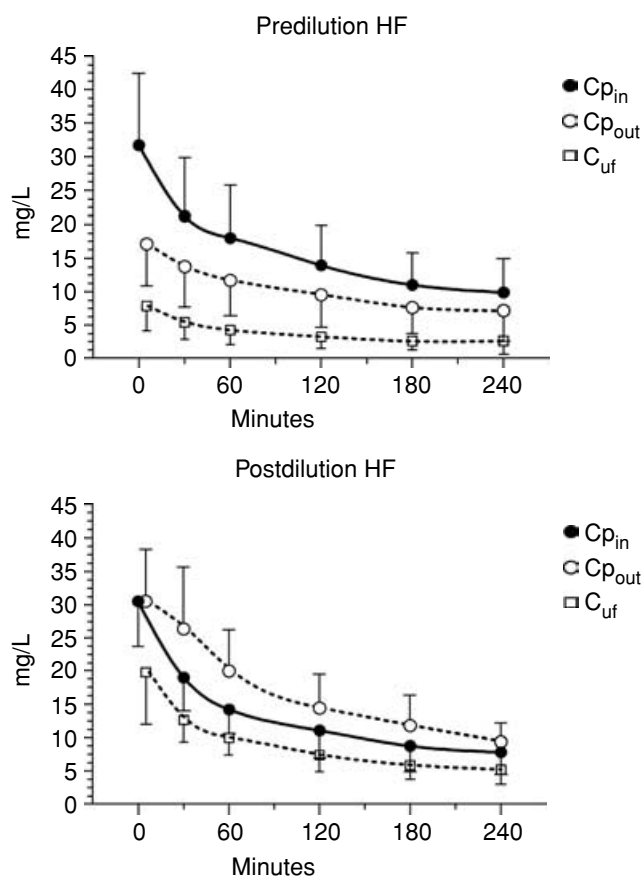


Fig. 1. Time courses of mean β 2-m concentrations in plasma ($C_{p_{in}}$, $C_{p_{out}}$) and ultrafiltrate (C_{uf}).

Table 4. Model-independent pharmacokinetic parameters of β 2-m based on $C_{p_{in}}$, $C_{p_{out}}$, and C_{uf}

	Predilution ($N = 11$)	Postdilution ($N = 11$)	P
C_{p_0} mg/L	31.7 ± 10.7	30.4 ± 6.8	NS
$C_{p_{4h}}$ mg/L	9.8 ± 5.1	7.7 ± 3.3	NS
$AUC_{p_{in}}$ mg/L \cdot min	3705 ± 1556	3074 ± 945	NS
$AUC_{p_{out}}$ mg/L \cdot min	2463 ± 1160	4191 ± 1174	<0.0005
AUC_{uf} mg/L \cdot min	907 ± 471	2090 ± 656	<0.0001

both pre- and postdilution modes are shown in Figure 1. No significant differences between pre- and postdilution were found for pharmacokinetic parameters C_{p_0} , $C_{p_{4h}}$, and $AUC_{p_{in}}$, but $AUC_{p_{out}}$ and AUC_{uf} were higher in post-HF than in pre-HF (Table 4). As expected, Hct and Pt increased, and urea and creatinine plasma concentrations decreased from the start to the end of each procedure (Table 3).

The parameters characterizing the ultrafiltration process with the 2 techniques are listed in Table 5. CL_{tot} was slightly but not significantly ($P = 0.064$) higher during post-HF. By contrast, CL_{con} and CL_{ads} significantly differed between the 2 techniques [i.e., CL_{con} was signif-

Table 5. Mean values \pm SD of the main parameters of HF efficiency

	Predilution ($N = 11$)	Postdilution ($N = 11$)	P
Plasma water clearance mL/min			
CL_{max}	114.5 ± 14.1	121.3 ± 14.9	NS
CL_{tot}	73.4 ± 12.4	82.8 ± 12.5	NS
CL_{con}	60.8 ± 13.1	$79.6^* \pm 11.8$	<0.005
CL_{ads}	12.6 ± 9.3	$3.2^* \pm 8.3$	<0.005
Sieving coefficient	0.53 ± 0.10	$0.65^* \pm 0.07$	<0.005
% $C_{p_{in}}$	-70.5 ± 7.4	-74.7 ± 6.6	NS
β 2-microglobulin removal mg			
A_{tot}	288.9 ± 113.8	273.2 ± 77.6	NS
A_{con}	236.6 ± 93.9	260.5 ± 65.1	NS
A_{ads}	52.3 ± 40.0	$12.7^* \pm 28.6$	<0.005

icantly higher (79.6 mL/min vs. 60.8 mL/min) and CL_{ads} was significantly lower (3.2 mL/min vs. 12.6 mL/min) in post-HF than in pre-HF]. The same trend was observed for the amounts of β 2-m removed (A_{tot} , A_{con} , A_{ads}), although only A_{ads} was significantly lower in post-HF than in pre-HF (52.3 mg vs. 12.7 mg). Using pooled pre- and post-HF data, a significant inverse correlation ($r^2 = 0.43$; $P = 0.0009$) was found between the amount of β 2-m adsorbed by the membrane (A_{ads}) and the sieving coefficient.

The percentage decrease in $C_{p_{in}}$ from the start to the end of the treatment was similar (-70.5% in pre-HF vs. -74.6% in post-HF). The sieving coefficient was significantly higher in post- than in predilution (0.65 vs. 0.55).

The changes in $CL_{pw_{tot}}$, $CL_{pw_{con}}$, $CL_{pw_{ads}}$, and sieving coefficient throughout the 4-hour HF period are shown in Figures 2 and 3. The one-way ANOVA revealed that $CL_{pw_{tot}}$, $CL_{pw_{con}}$ underwent a significant time-dependent decrease in both HF modes, whereas $CL_{pw_{ads}}$ decreased only with post-HF and the sieving coefficient decreased only with pre-HF (Table 6).

DISCUSSION

Different strategies have been adopted in the course of time in order to prevent the accumulation of β 2-m. The routine use of polyacrylonitrile or, more in general, the switch from conventional to high-flux dialysis membranes have proven to be useful to reduce the risk of carpal tunnel syndrome [18]. The bacteriologic control of all the dialysis fluids, avoiding the endotoxin contamination, may also play a preventative role. In actual fact, some studies have demonstrated that the use of high-quality dialysis fluids alone may reduce the incidence of carpal tunnel syndrome [19, 20]. The use of convective dialysis therapies, in addition to the control of the biocompatibility aspects of dialysis, may add the strength of an enhanced removal, to the reduced production of β 2-m, thus resulting to be the best strategy to prevent β 2-m-related pathologies.

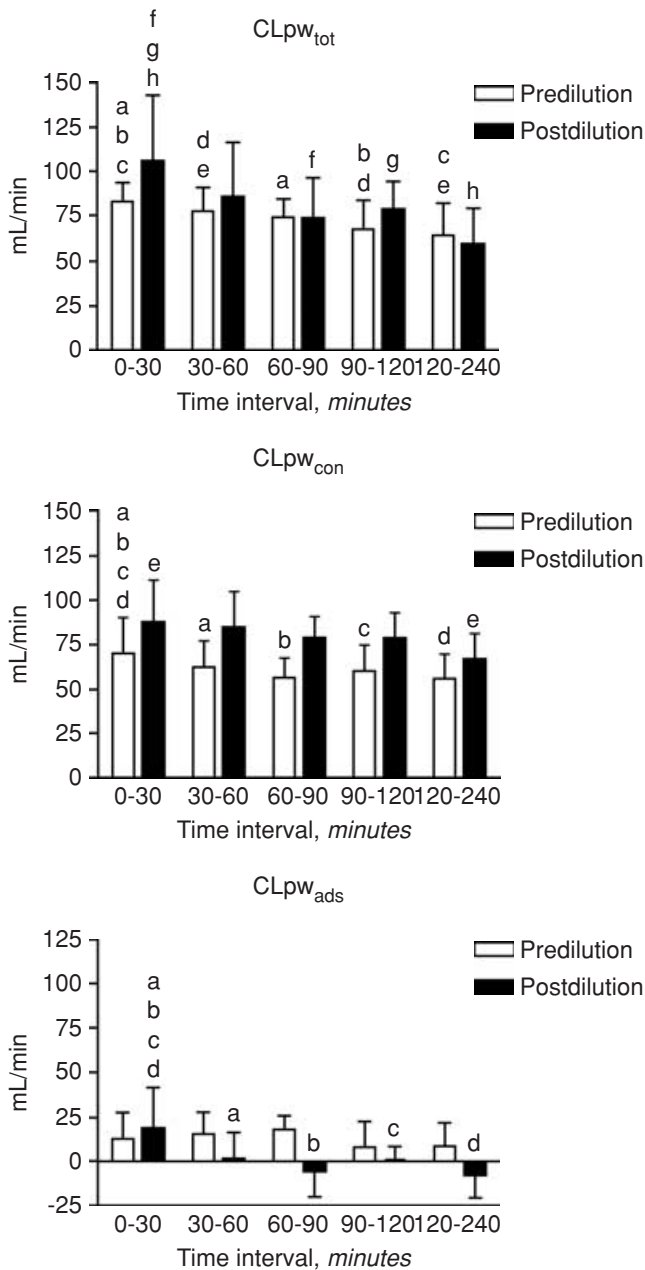


Fig. 2. Changes in fractional CLpw_{tot}, CLpw_{con} and CLpw_{ads} during HF procedure. Columns labeled with same letter differ significantly (post-hoc test).

The main result of our study is that the overall HF efficiency in removing β 2-m from plasma, evaluated as CLpw_{tot}, A_{tot}, and % Cpw_{in}, is similar in both pre- and postdilutional conditions. These findings are in contrast with what is generally stated in the literature (i.e., that the removal of middle molecules is higher in predilution modes) [21–24]. However, another recent study [15] reported no substantial differences between pre- and post-dilution HDF in regards to the total amount of β 2-m removed from plasma.

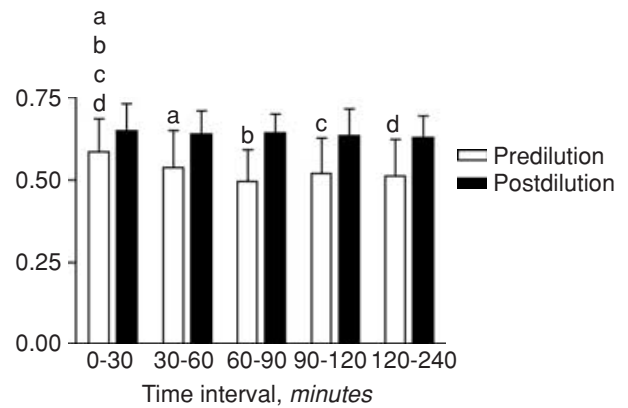


Fig. 3. Changes in sieving coefficient during HF procedure. Columns labeled with same letter differ significantly (post-hoc test).

Table 6. Influence of time on HF parameters

	Predilution HF	Postdilution HF
CLpw _{tot}	<i>P</i> < 0.0001	<i>P</i> < 0.005
CLpw _{con}	<i>P</i> < 0.005	<i>P</i> < 0.05
CLpw _{ads}	NS	<i>P</i> < 0.005
Sieving coefficient	<i>P</i> < 0.005	NS

One-way ANOVA.

After a more in-depth analysis of removal mechanisms, it appears that the equivalence of CLpw_{tot} arises from balanced, opposite changes in convective and adsorptive clearances with the 2 techniques, as testified by the higher CLpw_{con} and lower CLpw_{ads} values found in post-HF in comparison with pre-HF. The former was almost entirely attributable to a higher sieving coefficient (0.65 vs. 0.55), as CLpw_{max} was virtually the same in both infusion modes (114 mL/min vs. 121 mL/min). In line with this finding, we have recently observed [25] that, at equivalent transmembrane pressure, the loss of albumin through the dialysis membrane is greater in post-HF than in pre-HF. In this respect, it is noteworthy that the sieving coefficient was similar at the beginning of the 2 procedures and then significantly decreased only during pre-HF treatment (Fig. 3). Therefore, the sieving coefficient may change during HF depending on the technique used. Interestingly, using pooled data, a significant inverse correlation was found between the amount of β 2-m adsorbed to the membrane (A_{ads}) and the sieving coefficient. If we accept that β 2-m adsorption is a marker of the more general phenomenon of protein deposition on the filter surface, we could conclude that the reduced convective β 2-m removal in predilution is due to greater protein fouling. This finding contrasts with the intuitive notion that plasma and blood concentrations occurring during post-HF should reduce HF efficiency by increasing blood viscosity and clogging the membrane pores. However, ultrafiltration is a process ruled by several factors

[10, 12], whose analysis may provide explanations. To justify the greater A_{ads} found with pre-HF than with post-HF, it may be assumed that the true adsorbing surface is different with the 2 techniques, even though the same hemofilter is used. As membrane pores are structured like water channels rather than simple holes, they probably oppose differing degrees of hydraulic resistance. If so, the higher ultrafiltration flow used in pre-HF may open a greater number of channels, thus increasing the actual membrane surface and the amount of β 2-m adsorbed.

Regarding the differences in sieving efficiency, it is known that the sieving coefficient is not constant for any given solute-membrane combination, but depends on the blood and the ultrafiltration flows. The high blood flows used in predilution (true $Q_b + Q_{\text{inf}}$) are expected to increase the wall shear rate and decrease the solute polarization (i.e., the rise in β 2-m concentration at the membrane surface that improves the sieving coefficient) [10, 21]. Solute polarization is considered to be particularly important for macromolecules, like β 2-m, which are confined to plasma water and accumulate on the membrane surface more than small molecules. Indeed, experimental data indicate that, if Q_b is gradually increased in post-HF while Q_{uf} is kept constant, the sieving coefficient for β 2-m first increases (for Q_b up to 450–500 mL/min) and then decreases [10]. Furthermore, it has been shown that, if Q_{uf} is increased in constant Q_b conditions, the β 2-m sieving coefficient uniformly decreases [10]. Therefore, on the whole, the high blood flow and ultrafiltrate rate typical of pre-HF tend to reduce the sieving coefficient for β 2-m.

CONCLUSION

Our data show a substantial equivalence between pre-HF and post-HF in removing β 2-m from plasma, although the contribution of convective and adsorptive mechanisms is different in the 2 techniques. In the long term, predilution HF could prove more useful in patients with high hematocrit levels. This is a frequent condition today in view of the routine use of erythropoietin to maintain the hemoglobin levels within an ideal range (11–12 g/dL), and may favor the clotting of the hemofilter in the post-dilution modality.

Since the efficiency of urea and creatinine removal, measured as percent decreases in baseline plasma concentrations, is also comparable, the choice between the 2 HF modes should be solely based on feasibility and economic assessment.

APPENDIX

Hct_{out} may be calculated using the formula:

$$Hct_{\text{out}} = Q_e/Q_{b_{\text{out}}} \quad (\text{Equation a})$$

where Q_e is erythrocyte flow through the hemofilter and $Q_{b_{\text{out}}}$ is blood flow leaving the hemofilter. Q_e is equal to

$$Q_{b_{\text{in}}} \times Hct_{\text{in}} \quad (\text{Equation b})$$

$Q_{b_{\text{out}}}$ depends on the amount of fluid infused and/or lost and on the HF technique:

$$Q_{b_{\text{out}}} = Q_{b_{\text{in}}} - Q_{\text{uf}} + Q_{\text{inf}}, \text{ for predilutional HF} \quad (\text{Equation c})$$

$$Q_{b_{\text{out}}} = Q_{b_{\text{in}}} - Q_{\text{uf}}, \text{ for postdilutional HF} \quad (\text{Equation d})$$

Substituting Q_e and $Q_{b_{\text{out}}}$ in Eq. a for Eqs. b, c, and d yields Eqs. 3 and 4 (see **Methods**).

Reprint requests to Roberto Padrini, Department of Pharmacology and Anesthesiology, University of Padova, Largo Meneghetti 2, 35131 Padova, Italy.

E-mail: roberto.padrini@unipd.it

REFERENCES

1. BIANCHI C, DONADIO C, TRAMONTI G, et al: Reappraisal of serum beta2-microglobulin as marker of GFR. *Renal Failure* 23:419–429, 2001
2. KOCH KM: Dialysis-associated amyloidosis. *Kidney Int* 41:1416–1429, 1992
3. FLOEGE J, EHLERDING G: Beta2-microglobulin-associated amyloidosis. *Nephron* 72:9–26, 1996
4. FLOEGE J, KETTELER M: Beta2-microglobulin-derived amyloidosis: An update. *Kidney Int* 59(Suppl 78):S164–S171, 2001
5. LOCATELLI F, MARCELLI D, CONTE F, et al: Comparison of mortality in ESRD patients on convective and diffusive extracorporeal treatments. *Kidney Int* 55:286–293, 1999
6. DRUEKE T, URENA P, MAN NK, ZINGRAFF J: Membrane transfer, membrane adsorption and possible membrane-induced generation of beta-2-microglobulin. *Contrib Nephrol* 74:113–119, 1989
7. DAVID S, CANINO F, FERRARI ME, CAMBI V: The role of adsorption in beta 2-microglobulin removal. *Nephrol Dial Transplant* 6(Suppl 2):64–68, 1991
8. RONCO C, HEIFETZ A, FOX K, et al: Beta 2-microglobulin removal by synthetic dialysis membranes. Mechanisms and kinetics of the molecule. *Int J Artif Organs* 20:136–143, 1997
9. DAVID S, TAGLIAVINI D, CAMBI V: Pre-postdilution haemofiltration. *Nephrol Dial Transplant* 4:37–40, 1989
10. DAVID S, CAMBI V: Hemofiltration: Predilution versus postdilution. *Contrib Nephrol* 96:77–85, 1992
11. LEDEBO I: Predilution hemofiltration: A new technology applied to an old therapy. *Int J Artif Organs* 18:735–742, 1995
12. LEDEBO I: Principles and practice of hemofiltration and hemodiafiltration. *Artif Organs* 22:20–25, 1998
13. AHRENHOLZ P, WINKLER RE, RAMLOW W, et al: On-line hemodiafiltration with pre- and post-dilution: A comparison of efficacy. *Int J Artif Organs* 20:81–90, 1997
14. LORNOY W, BECAUS I, BILLIOUW JM, et al: Remarkable removal of beta-2-microglobulin by on-line hemodiafiltration. *Am J Nephrol* 18:105–108, 1998
15. PEDRINI LA, DE CRISTOFARO V, PAGLIARI B, SAMA F: Mixed predilution and postdilution online hemodiafiltration compared with the traditional infusion modes. *Kidney Int* 58:2155–2165, 2000
16. DAVID S, LOCATELLI F, REDAELLI F, et al: Predilution hemofiltration (pre-HF): Higher removal by convection of both small and large solutes, a multicenter study. *Blood Purif* 8:112–113, 1990
17. COLTON CK, SMITH KA, MERRIL EW, FRIEDMAN S: Diffusion of urea in flowing blood. *Ann Inst Chem Eng J* 17:800–801, 1971
18. KODA Y, NISHI S, MIYAZAKI S, et al: Switch from conventional to high-flux membrane reduces the risk of carpal tunnel syndrome and mortality of hemodialysis patients. *Kidney Int* 52:1096–1101, 1997

19. BERLAND Y, BRUNET P, RAGON A, REYNIER JP: Dialysis fluid and water: Their role in biocompatibility. *Nephrol Dial Transplant* 10(Suppl 10):45-47, 1995
20. KLEOPHAS W, HAASSTERT B, BACKUS G, et al: Long-term experience with an ultrapure individual dialysis fluid with a batch type machine. *Nephrol Dial Transplant* 13:3118-3125, 1998
21. PILLARELLA MR, ZYDNEY AL: Analysis of solute clearance and flux in pre- and post-dilution hemofiltration. *ASAIO Trans* 34:415-419, 1988
22. LOCATELLI F, DI FILIPPO S, MANZONI C: Removal of small and middle molecules by convective techniques. *Nephrol Dial Transplant* 15(Suppl 2):37-44, 2000
23. LEDEBO I: Online hemofiltration. *Contrib Nephrol* 137:221-226, 2002
24. BEERENHOUT C, DEJAGERE T, VAN DER SANDE F, et al: Haemodynamics and electrolyte balance: A comparison between online pre-dilution haemofiltration and hemodialysis. *Nephrol Dial Transplant* 19:2354-2359, 2000
25. SANTORO A, CANOVA C, MANCINI E, et al: Protein loss in on-line hemofiltration. *Blood Purif* 22:261-268, 2004