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# Review Endothelial dysfunction — A major mediator of diabetic vascular disease



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# ABSTRACT

The vascular endothelium is a multifunctional organ and is critically involved in modulating vascular tone and structure. Endothelial cells produce a wide range of factors that also regulate cellular adhesion, thromboresistance, smooth muscle cell proliferation, and vessel wall inflammation. Thus, endothelial function is important for the homeostasis of the body and its dysfunction is associated with several pathophysiological conditions, including atherosclerosis, hypertension and diabetes. Patients with diabetes invariably show an impairment of endothelium-dependent vasodilation. Therefore, understanding and treating endothelial dysfunction is a major focus in the prevention of vascular complications associated with all forms of diabetes mellitus. The mechanisms of endothelial dysfunction in diabetes may point to new management strategies for the prevention of cardiovascular disease in diabetes. This review will focus on the mechanisms and therapeutics that specifically target endothelial dysfunction in the context of a diabetic setting. Mechanisms including altered glucose metabolism, impaired insulin signaling, low-grade inflammatory state, and increased reactive oxygen species generation will be discussed. The importance of developing new pharmacological approaches that upregulate endothelium-derived nitric oxide synthesis and target key vascular ROS-producing enzymes will be highlighted and new strategies that might prove clinically relevant in preventing the development and/or retarding the progression of diabetes associated vascular complications.

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# 1. Introduction

# 1.1. Vascular function and endothelium

The endothelium is a monolayer of cells covering the vascular lumen. For many years this cell layer was thought to be relatively inert, a mere physical barrier between circulating blood and the underlying tissues. It is now recognized, however, that endothelial cells are metabolically active with important paracrine, endocrine and autocrine functions, indispensable for the maintenance of vascular homeostasis under physiological conditions [1,2]. The multiple functions of vascular endothelium are summarized in Fig. 1 and include regulation of vessel integrity, vascular growth and remodeling, tissue growth and metabolism, immune responses, cell adhesion, angiogenesis, hemostasis and vascular permeability. The endothelium plays a pivotal role in the regulation of vascular tone, controlling tissue blood flow and inflammatory responses and maintaining blood fluidity [3–5].

Endothelium-derived factors with vasodilatory and antiproliferative effects include endothelium-derived hyperpolarization factor (EDHF) [], nitric oxide (NO) [8,9] and prostacyclin (PGI<sub>2</sub>) [10], while endothelin-1

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(ET-1) [11], angiotensin II and reactive oxygen species (ROS) are among the mediators that exert vasoconstrictor effects [12,13]. Endothelial cells also produce antithrombotic (NO and PGI<sub>2</sub> both inhibit platelet aggregation) and prothrombotic molecules [von Willebrand factor, which promotes platelet aggregation, and plasminogen activator inhibitor-1 (PAI-1), which inhibits fibrinolysis] [5].

As a major regulator of vascular homeostasis, the endothelium maintains the balance between vasodilation and vasoconstriction, inhibition and promotion of the migration and proliferation of smooth muscle cells, fibrinolysis and thrombogenesis as well as prevention and stimulation of the adhesion and aggregation of platelets (Fig. 2) [5]. Disturbing this tightly regulated equilibrium leads to endothelial dysfunction.

#### 1.2. Nitric oxide

NO is a crucial player in vascular homeostasis. NO is synthesized within endothelial cells during conversion of L-arginine to L-citrulline by endothelial nitric oxide synthase (eNOS) [15]. It is released from endothelial cells mainly in response to shear stress elicited by the circulating blood or receptor-operated substances such as acetylcholine, bradykinin, or serotonin [16]. NO diffuses to vascular smooth muscle cells (VSMC) and activates soluble guanylate cyclase (sGC), yielding increased levels of cyclic guanosine-3,5-monophosphate (cGMP) and relaxation of VSMC [1,17]. Additionally, NO also prevents leukocyte adhesion and migration, smooth muscle cell proliferation, platelet adhesion and aggregation, and opposes

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Fig. 1. Multiple functions of endothelium

apoptosis and inflammation having an overall antiatherogenic effect (Fig. 3) [18].

The half-life of NO is very short (less than 4 s). It is rapidly metabolized to nitrite and then to nitrate before being excreted in the urine [4]. Alternatively, NO can also be an endocrine vasoregulator, modulating blood flow in the microcirculation [19]. Importantly, reduced eNOS expression and/or NO bioavailability is associated with endothelial dysfunction [20,21].

# 1.2.1. Decreased formation of NO

eNOS is a dimeric enzyme depending on multiple cofactors for its physiological activity and optimal function. eNOS resides in the caveolae and is bound to the caveolar protein, caveolin-1 that inhibits its activity. Elevations in cytoplasmic Ca<sup>2+</sup> promote binding of calmodulin to eNOS that subsequently displaces caveolin and activates eNOS [22,23].

eNOS utilizes L-arginine as the substrate, and molecular oxygen and reduced nicotinamide adenine dinucleotide phosphate (NADPH) as co-substrates. Flavin adenine dinucleotide, flavin mononucleotide, tetrahydrobiopterin (BH<sub>4</sub>) and calmodulin are the cofactors [4]. A reduced expression and/or activity of eNOS could be responsible for a decrease in NO production. Oxidative stress leads to eNOS uncoupling, a process where eNOS is converted from an NO-producing enzyme to an enzyme that generates superoxide anion ( $O_2^{*-}$ ). Mechanisms implicated in eNOS uncoupling include oxidation of BH<sub>4</sub> (a critical eNOS cofactor; [24]), depletion of the enzyme substrate L-arginine, and accumulation of eNOS has also been proposed as a mechanism that leads to eNOS uncoupling and decreased NO bioavailability [26]. Additionally, increased expression of caveolin-1 in the endothelium (as described in diabetes and obesity [190]) leads to impaired activation of eNOS.

eNOS activity within the endothelial cell is also modulated by circulating factors like insulin. Insulin is an essential hormone in metabolic homeostasis with a vasodilator action exerted through the phosphatidylinositol-3 kinase (PI-3K)/AKT pathway-dependent eNOS activation [27]. Insulin can modulate eNOS activity by increasing BH<sub>4</sub> synthesis [28]. Insulin-stimulated endothelial dependent vasodilatation is impaired in insulin resistance [29,18]. Conversely, eNOS plays a major role in the regulation of insulin sensitivity due to the functions of NO in peripheral tissues [30]. Previous studies have shown that mice lacking eNOS are more likely to develop insulin resistance [227]. Apparently, modulation of eNOS phosphorylation in mice is sufficient to affect systemic insulin sensitivity indicating that eNOS phosphorylation may be a novel target for the treatment of insulin resistance [228].



**Fig. 2.** Endothelial cells are responsible for a number of physiological functions, including: 1) regulation of vascular tone through balanced production of vasodilators and vasoconstrictors; 2) control of blood fluidity and coagulation through production of factors that regulate platelet activity, the clotting cascade, and the fibrinolytic system; and 3) regulation of inflammatory processes through expression of cytokines and adhesion molecules. ACh, acetylcholine; ATR, angiotensin-II receptor; BK, bradykinin; EDHF, endothelium-derived hyperpolarization factor; NO, nitric oxide; PAI-1, plasminogen activator inhibitor-1; PGH<sub>2</sub>, prostaglandin H<sub>2</sub>; PGI<sub>2</sub>, prostacyclin; O<sub>2</sub>•<sup>-</sup>, superoxide anion; t-PA, tissue plasminogen activator; TM, thrombomodulin; TxA<sub>2</sub>, thromboxane A<sub>2</sub>; vWF, von Willebrand factor.



Fig. 3. Atheroprotective properties of nitric oxide generated by endothelial nitric oxide synthase.

eNOS may be inhibited by endogenous products of arginine metabolism such as asymmetric dimethyl-L-arginine (ADMA) [1]. Following oxidative stress or angiotensin II administration, the observed elevation in ADMA levels reduces NO formation and leads to endothelial dysfunction. Indeed, in several prospective studies, ADMA has been noted to be an independent predictor of cardiovascular events [33–35].

Another factor that regulates eNOS activity in the setting of metabolic disease is adropin, which was recently recognized to be an important regulator of energy homeostasis and insulin sensitivity. Lovren and colleagues [36] demonstrated that adropin is expressed in endothelial cells and improves angiogenesis-related responses via activation of Akt, eNOS, and extracellular signal regulated kinase 1/2. Like adiponectin and leptin, adropin may be an endocrine factor that influences both insulin resistance and endothelial functions such as vasodilation and angiogenesis [36].

#### 1.2.2. Accelerated breakdown of NO

Accelerated degradation of NO by ROS is probably the major mechanism impairing NO bioavailability in states of cardiovascular disease [37,38]. In a diabetic milieu, an increment in  $O_2^{\bullet^-}$  levels is observed in the vasculature (Fig. 4).  $O_2$ •<sup>-</sup> readily reacts with NO to form peroxynitrite (ONOO<sup>-</sup>), reducing NO bioavailability and contributing to impaired vasorelaxation [39]. Fig. 5 shows an increase in nitrotyrosine staining in the aorta of a type 2 diabetic animal model, indicative of peroxynitrite formation. Additionally, lipid peroxyl radicals react with NO at almost diffusionlimited rates and may be a source of NO inactivation [40]. Also, oxidized low-density lipoprotein (LDL) cholesterol may react with endothelial NO before it reaches the vascular smooth muscle cell and therefore reduce total NO-mediated vasodilation [41].

# 1.3. Prostacyclin

PGI<sub>2</sub> is the major metabolite of arachidonic acid produced by cyclooxygenase in the endothelium. PGI<sub>2</sub> activates adenylate cyclase, leading to increased production of cyclic AMP and VSMC vasodilation. Additionally, PGI<sub>2</sub> is a potent antiproliferative agent in vascular smooth muscle cells, and it reduces oxidative stress and prevents cellular adhesion to the vascular wall [42]. PGI<sub>2</sub> also inhibits platelet aggregation. Clinical and experimental models of diabetes are associated with decreased secretion of PGI<sub>2</sub> [3,27].



Fig. 4. In situ detection of superoxide in arterial vessels of normal Wistar (left panels) and diabetic Goto–Kakizaki rats (GK, right panels). Superoxide production was detected as red fluorescence after incubation with dihydroethidium. Representative fluorescent staining of superoxide with dihydroethidium in the abdominal aorta (upper panels) and kidney arterial vessels (lower panels). O<sub>2</sub>\*<sup>-</sup> formation significantly increased in diabetic animals when compared to age-matched controls.



Fig. 5. Nitrotyrosine (3-NT) immunoreactivity increases in arterial vessels from diabetic Goto–Kakizaki (GK) rats. Immunofluorescence staining for 3-NT (green) in aortic (upper panels) and kidney arterial sections (lower panels) isolated from Wistar (left panels) and diabetic GK (right panels) rats. Nuclei were counter-stained with DAPI (blue). The extent and intensity of immunofluorescence for nitrotyrosine was much greater in arterial rings from diabetic rat animals when compared to age-matched controls. Measuring NT levels is thought to be a reliable index to analyze peroxynitrite formation.

# 1.4. Endothelium derived hyperpolarizing factor

There are smaller arteries in which endothelium-mediated vasodilation is predominately affected by endothelium-dependent hyperpolarization of vascular smooth muscle cells. The mechanism partially responsible for the endothelium-dependent vasodilation of these arteries, which persists in the presence of inhibitors of eNOS and prostacyclin, was first attributed to a non-characterized endothelial factor termed EDHF [43-45]. The relative importance of the EDHF mediated mechanisms to NO mediated mechanisms alters with vessel size [46]. NO is an important endothelium-dependent mediator of vascular tone in relatively large arteries and larger arterioles. At the level of the aorta, reduced NO bioavailability is proposed to be the main marker for endothelial dysfunction. In resistance arteries, NO, prostacyclins and EDHFs are thought to be involved in mediating endothelial function [47]. Alterations in EDHF-mediated responses have been reported in diabetes [48]. Interestingly, EDHF synthase/cytochrome P450 epoxygenase is also a source of superoxide anion [52].

# 2. Endothelial dysfunction

In the earliest stages, the principal endothelial alteration is merely functional. Functional impairment of the vascular endothelium is found in all forms of cardiovascular disease [3,12] and also in people with insulin resistance, obesity and type 2 diabetes [18]. The hallmark of endothelial dysfunction is the impaired NO bioavailability. Additionally, endothelial dysfunction is characterized by one or more of the following features: reduced endothelium-mediated vasorelaxation, hemodynamic deregulation, impaired fibrinolytic ability, enhanced turnover, overproduction of growth factors, increased expression of adhesion molecules and inflammatory genes, excessive generation of ROS, increased oxidative stress, and enhanced permeability of the cell layer [53–57].

Numerous risk factors directly contribute to endothelial dysfunction. Some of the more important are: elevated LDL cholesterol and oxidized LDL; low high-density lipoprotein (HDL) cholesterol; elevated triglycerides; hypertension; elevated C-reactive protein (CRP) and circulating lipoprotein-associated phospholipase A2 (Lp-PLA 2 – a specific marker of vascular inflammation); hyperglycemia; elevated omega-6:omega-3 ratio [58]; hyperinsulinemia; elevated homocysteine levels; increased fibrinogen and PAI-1; smoking; insufficient vitamin D; among others [59–61].

The presence of endothelial dysfunction has been implicated in the pathogenesis of atherosclerosis and thrombosis, both for the loss of its protective capability and for the induction of proatherothrombotic mechanisms. The major features associated with endothelial dysfunction are summarized in Table 1.

### 2.1. The impact of diabetes on the vasculature

Diabetes is not only a metabolic disease but also considered as a vascular disease because of its effect on macro and microcirculation of many vascular beds. The link between diabetes and an increased incidence of cardiovascular disease is well established [62,63]. Recent evidence shows that etiopathogenesis of endothelial dysfunction differs in types 1 and 2 diabetes [64]; it is present at the earliest stages of metabolic syndrome and insulin resistance, and may precede the clinical diagnosis of type 2 diabetes by several years [65].

The metabolic milieu in diabetes (i.e. hyperglycemia, excess free fatty acid release and insulin resistance) induces a vicious circle of events in the vascular wall, involving increased endothelial dysfunction, oxidative stress, low-grade inflammation and platelet hyperactivity, in the early stages of diabetic disease. Thereby, activation of these systems impairs endothelial function, augments vasoconstriction, increases inflammation, and promotes thrombosis [62,63]. In Fig. 6 multiple mechanisms that promote atherogenesis are summarized.

#### Table 1

Differences between a healthy and a dysfunctional endothelium. Besides impaired vasodilation ( $\downarrow$ NO, PGI<sub>2</sub>), endothelial dysfunction is characterized by increase oxidative stress († nitrotyrosine and uric acid), pro-coagulant (†PAI-1, vWF, P-selectin), and pro-inflammatory biomarkers (†slCAM, sVCAM, E-selectin, CRP, TNF-alpha, IL-6, MCP-1); decrement in endothelial progenitor cells and increased molecular markers of damage (circulating endothelial cells, microparticles, MPs).

Healthy Endothelium	Dysfunctional Endothelium
<ul> <li>Vasodilatory (↑ NO, PGI<sub>2</sub>)</li> <li>↓ Oxidative stress , low uric acid</li> <li>Anti-coagulant (↓ PAI-1, vWF, P-selectin)</li> <li>Anti-inflammatory (↓ sICAM, sVCAM, E-selectin, CRP, TNF-α, IL-6, MCP-1)</li> <li>↑ Repair (EPCs), ↓ Damage (CECs, MPs)</li> </ul>	<ul> <li>Impaired vasodilation (↓ NO, PGI<sub>2</sub>)</li> <li>↑ Oxidative stress, uric acid</li> <li>Pro-coagulant (↑ PAI-1, vWF, P-selectin)</li> <li>Pro-inflammatory (↑ sICAM, sVCAM, E-selectin, CRP, TNF-α, IL-6, MCP-1)</li> <li>↓ Repair (EPCs), ↑ Damage (CECs, MPs)</li> </ul>

CECs, circulating endothelial cells; CRP, C-reactive protein; EMPs, endothelial microparticles; EPCs, endothelial progenitor cells; IL-6, interleukin-6; MPs, microparticles; NO, nitric oxide; PAI-1, plasminogen activator inhibitor 1; PGI<sub>2</sub>, prostacyclin; ROS, reactive oxygen species; sICAM, soluble intercellular adhesion molecule; sVCAM, soluble vascular cell adhesion molecule; TNF- $\alpha$ , tumor necrosis factor alpha; vWF, von Willebrand factor.

# 2.1.1. Hyperglycemia

Prolonged hyperglycemia and also transient, acute hyperglycemia have been proven to impair endothelial function in both macro- and microvascular beds in animal studies and in human subjects [67–69]. Although the effect of intensive glycemic control on the prevention of macrovascular disease is less profound than on the reduction of microvascular complications [70].

Hyperglycemia causes vascular damage in different cells of the vascular wall (Table 2). The mechanisms are diverse and include: 1) increased flux of glucose and other sugars through the polyol pathway; 2) augmented intracellular formation of advanced glycation end products (AGEs); 3) increment in the expression of the receptor for AGEs (RAGE) and its activating ligands; 4) activation of protein kinase protein kinase C (PKC) isoforms; and 5) overactivation of the hexosamine pathway [74]. The common pathway is oxidative stress. ROS decreases the metabolism of glucose through glycolysis, and the flux through the alternative polyol and hexosamine pathways is increased. Hyperglycemia induced oxidative stress [71] leads to DNA damage and activation of nuclear poly(ADP-ribose) polymerase (PARP) that in turn increases production of polymers of ADP-ribose reducing glyceraldehyde 3-phosphate dehydrogenase activity. Ultimately the levels of all upstream glycolytic intermediates increase. The accumulation of glycolytic intermediates activates damaging mechanisms: PKC pathway, hexosamine and polyol pathways and AGE formation. The overall effects of these mechanisms are increased oxidative stress, apoptosis and vascular permeability [74].

Additionally, glucotoxicity induces a low-grade proinflammatory condition, due to the activation of transcription factors such as nuclear



**Fig. 6.** Several mechanisms that foster endothelial dysfunction and vascular damage in type 2 diabetes. Various risk factors converge on the artery (center) to promote atherogenesis under diabetic conditions. These factors include: hypertension, genetic predisposition, hyperglycemia, hyperinsulinemia, oxidative stress, advanced glycation end products (AGEs), insulin resistance and increased free fatty acids (FFAs) in circulation, lipemia, increased obesity as related to some factors which characterize life-style (sedentary, drinking, smoking and eating habits), enhanced proinflammatory and prothrombotic cytokines. Peripheral tissues are resistant to insulin action, which promotes hyperglycemia and increased levels of FFAs. In insulin resistance states, the pancreas initially tries to compensate by producing more insulin, resulting in hyperinsulinemia, itself a risk factor for angiopathy. High levels of abdominal fat present the liver with elevated levels of FFAs through the portal circulation. This excess of FFAs will lead to excess production of triglyceride (TG)-rich lipoprotein particles. Hypertriglyceridemia is accompanied by a concomitant decrease in HDL. The adipocyte can also release proinflammatory cytokines such as TNF-  $\alpha$ , which not only have direct effects on vascular wall promoting atherogenesis, but also can elicit the production of acute phase reactants by the liver, including CRP, increased fibrinogen and PAI-1. Finally, the formation of advanced glycation end products (AGEs) from glycated macromolecules can damage vasculature through different mechanisms. VLDL, very low-density lipoprotein; TNF-  $\alpha$ , tumor necrosis factor- $\alpha$ ; CRP, C-reactive protein; and PAI-1, plasminogen activator inhibitor-1.

#### Table 2

Examples of mechanisms implicated in diabetic macrovascular disease.

Cellular players	Mechanisms
Endothelial cells	Increased reactive oxygen species Decreased NO bioavailability Increased harmful metabolites (peroxynitrite, nitrotyrosine) NF-xB activation Increased lipid peroxidation products Increased glycation (AGEs) Impaired endothelial-dependent relaxation
Monocyte-derived macrophages Vascular smooth muscle cells	Increased IL1β, IL6, CD36, MCP-1 Activation of protein kinase C Increased proliferation Increased migration into intima Increased matrix degradation Altered matrix components (chondroitin, dermatan sulphate proteoglycans) Increased nonenzymatic collagen glycation Increased reactive oxygen species

AGEs — advanced glycation end products; IL — interleukin; MCP-1 — monocyte chemoattractant protein-1; NF- $\kappa$ B — nuclear factor-kappa B; NO — nitric oxide.

factor- $\kappa$ B (NF- $\kappa$ B) [74–76]. NF- $\kappa$ B is a key mediator that regulates multiple proinflammatory and proatherosclerotic target genes in endothelial cells, VSMC, and macrophages. Activation of NF- $\kappa$ B leads to an increased production of adhesion molecules, leukocyte-attracting chemokines and cytokines activating inflammatory cells in the vascular wall. A prothrombotic state is generated by the increased production of lesion-based coagulants, such as tissue factor, and the inhibitors of fibrinolysis, such as PAI-1 (Table 1).

Vascular tone and remodeling are enhanced through reduced NO and an increased activity and production of vasoconstrictors (ET-1, angiotensin II, and prostanoids [74–76]) due to postprandial increases in glucose, LDL cholesterol, and hyperinsulinemia (Fig. 7). Glucose may also activate matrix-degrading metalloproteinases, enzymes implicated in plaque rupture and arterial remodeling, inducing similar responses in VSMC. Glucose may also stimulate VSMC proliferation, migration, and altered reactivity, for example, through renin–angiotensin activation.

Elevated glucose can foster glycation of proteins, promoting formation of AGEs (Fig. 8), protein cross-linking, and ROS formation. Accumulation of AGEs alters the functional property of matrix components and mediates sustained cellular changes. Glycation modifies the structure of the molecules and disturbs their function and receptor recognition properties. In turn, binding of AGEs to their RAGE receptor increases intracellular enzymatic superoxide production [79,80] and promotes macrophage-mediated inflammation in the vessel wall [81]. AGEs also decrease NO bioavailability and eNOS expression [82,83] and increase expression of ET-1 in endothelial cells [84]; therefore altering the balance between NO and ET-1 to favor vasoconstriction and endothelial dysfunction.

Thus, accelerated formation of multiple biochemical species under hyperglycemic conditions such as nonenzymatic reactive Amadori products, 3-deoxyglucosone, diacylglycerol, methylglyoxal [85], AGEs, ROS, and nitrosylated species, greatly contributes to endothelial dysfunction in diabetes. The increased oxidative stress seems to be the common alteration, triggered by a type 2 diabetes milieu, in which hyperglycemia is adjoined by insulin resistance, hyperinsulinemia, and dyslipidemia [86].

#### 2.1.2. Insulin resistance

Insulin resistance refers to a decreased ability of insulin to promote glucose uptake in skeletal muscle and adipose tissue and to suppress hepatic glucose output [87]. Insulin signaling is transduced via two major pathways: metabolic and hemodynamic effects are mediated by PI-3K and the Ras-mitogen-activated protein kinase (MAPK) pathway is mainly involved in gene expression regulation, cell growth and differentiation [88]. Normally, insulin stimulates NO production in endothelial



**Fig. 7.** Endothelial dysfunction in diabetes. Prolonged exposure to hyperglycemia is the major culprit in the pathogenesis of diabetic complications, involving increased ROS and RNS production. Oxidative stress leads to an imbalance in the vascular homeostasis due to increased vasoconstriction and impaired vasorelaxation that ultimately fosters diabetic endothelial dysfunction. AGEs, advanced glycation end products; EDCF, endothelium-derived contracting factors; eNOS, endothelial nitric oxide synthase; FFAs, free fatty acids; PKC, protein kinase C; PGIS, prostacyclin synthase; NF+xB, nuclear factor-kappa B; NO, nitric oxide; RNS, reactive nitrogen species; ROS, reactive oxygen species. [167].

cells by activating NO synthase via the PI-3K pathway. In insulin resistance (IR) this pathway is impaired, and the production of NO is diminished [89]. Instead, insulin resistance activates MAPK leading to endothelial dysfunction. Insulin stimulates production of the vasoconstrictor ET-1, and increases PAI-1 and cellular adhesion molecule expression [90]. In addition to the direct effects of IR on the endothelium, it also stimulates VSMC proliferation and migration and in adipose tissue is associated with excessive release of free fatty acids (FFAs), which evokes pathogenic gene expression through PKC activation and increased oxidative stress [93]. IR- induced excess of FFAs is essential also in the development of dyslipidemia, which further promotes the development of a proatherogenic lipid profile.

Ultimately, insulin resistance and type 2 diabetes are associated with low-grade inflammation being reflected in increased serum levels of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-6, PAI-1, ET-1 and high-sensitive C-reactive protein, also related to endothelial dysfunction [98].

#### 2.1.3. FFAs

Excessive release from adipose tissue and diminished uptake by skeletal muscles increase circulating levels of FFAs in diabetes [99,100]. Acute infusion of FFAs reduces endothelium-dependent vasodilation in animal models and in humans in vivo [75,101].

Lipotoxicity by FFAs may impair endothelial function by a number of related mechanisms, including increased production of ROS, increase AGE formation and activate PKC, the hexosamine pathway, and proinflammatory signaling to the same extent as diabetic levels of hyperglycemia. FFAs have been shown to induce ROS production in the vasculature via mitochondrial uncoupling and by increasing the expression and protein content of NADPH oxidases [74,93]. FFAinduced overproduction of superoxide inactivates two important antiatherogenic enzymes: prostacyclin synthase and eNOS. ROS also decreases the concentration of intracellular glutathione and makes vasculature more prone to oxidative damage.



Fig. 8. The formation of advanced glycation end products (AGEs) can involve early glucose metabolites such as glyoxal and methylglyoxal, highly reactive dicarbonyls and key precursors of AGEs.

FFA-induced ROS also activate NF-KB, which further stimulates production of other proinflammatory cytokines [103–105]. By activating IKK $\alpha$ , FFA treatment impairs insulin stimulated activation of eNOS and NO production in endothelial cells [106]. Activation of PKC by FFAs also results in increased serine phosphorylation of IRS-1 that leads to reduced insulin-stimulated activation of PI-3 kinase, phosphoinositide-dependent kinase-1, Akt, and eNOS, and culminates with impaired NO production in endothelium [102,107]. Ultimately, FFAs stimulate endothelial apoptosis, augment vascular oxidative stress, reduce NO bioavailability, enhance endothelial and monocyte activation and increase inflammation [32].

The activation of metabolite sensitive pathways of vascular damage by increased FFA flux from insulin resistant visceral adipocytes to arterial endothelial cells may be the metabolic link between insulin resistance and macrovascular disease [74,94]. Increased oxidation of fatty acids, derived in part from insulin resistance leads to oxidative stress in diabetic macrovasculature, while in diabetic microvascular disease, ROS are derived mainly from intracellular hyperglycemia [91,92]. In both cases, under diabetic conditions oxidative stress seems to be the common mechanism that triggers vascular dysfunction.

# 2.2. Oxidative stress

Oxidative stress describes the condition wherein an excessive production of ROS overwhelms endogenous antioxidant defense mechanisms. The resultant elevation in ROS levels has a detrimental effect on cellular function, a consequence of ROS-induced damage to lipid membranes, enzymes and nucleic acids [108].

Risk factors for cardiovascular disease (CVD), including type 2 diabetes, are characterized by excess vascular production of ROS [108,109]. One of the earliest consequences of oxidative stress in human subjects is impaired endothelium-dependent vasodilation [108]. Thus, accessing oxidative stress in the vasculature could evaluate the risk for development of vascular disease (Table 3).

#### 2.2.1. Reactive oxygen species: major sources in the vasculature

All layers of the vascular wall have enzymatic systems capable of producing ROS. ROS include the superoxide anion, the hydroxyl radical, NO, lipid radicals, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hypochlorous acid and peroxynitrite [108].

The most important sources of ROS generation in the vasculature include the mitochondrial electron transport chain [114–116], NADPH oxidases [117–119] and xanthine oxidase [108,120]. In addition,

uncoupled eNOS and enzymes, such as lipoxygenase and cyclooxygenase, cytochrome P450s, peroxidases and other hemoproteins [108] are sources of ROS.

2.2.1.1. NADPH oxidases. NADPH oxidases are multicomponent enzymes functional in membranes of various cell types including endothelial cells and smooth muscle cells. NADPH family is the predominant source of  $O_2^{\bullet^-}$  in the human vasculature [117,119]. Of the seven Nox isoforms discovered, only Nox1, Nox2, Nox4 and Nox5 are expressed in blood vessels, with different cell-specific expressions, mode of activations and functions (for a review see [110,112]).

Activation of NADPH oxidases in the vasculature occurs in response to angiotensin II, other vasoactive hormones (e.g., ET-1), growth factors (e.g., transforming growth factor- $\beta$ ), cytokines, and mechanical stimuli (shear stress and stretch), among others [121,110]. Evidences from the literature clearly point to a role of Nox isoforms in vascular disease although their relative contribution remains unclear [110]. Nox1 and

# Table 3

Approaches to access oxidative stress in biological systems.

Approach	Examples
Markers of increased pro-oxidant activity	Increase in oxidant-generating systems (NADPH oxidases, xanthine oxidase, mitochondrial ROS, NOS) Direct measurements of ROS/RNS generation (reduction of NBT, oxidant-sensitive dyes, direct radicals measurement by ESR)
Markers of	Low-molecular-weight antioxidants
decrease in	(vitamins C and E, GSH)
antioxidant	Antioxidant enzymes
activity	(SODs, GPx, GR, catalase, thioredoxin system, paraoxonase)
	Total antioxidant capacity
	Resistant to an external oxidant
Altered cellular	Overall reducing activity (cyclic voltammetry)
redox state	GSH/GSSG ratio
Oxidative damage	Lipid oxidation (MDA, isoprostanes, 4-HNE)
parameters	Protein oxidation
	(protein carbonylation, S-nitrosylation, nitrotyrosine, glutathionylation) DNA oxidation
	(8-hydroxydeoxyguanosine, dihydropropidium iodide)

ROS, reactive oxygen species; RNS, reactive nitrogen species; NOS, nitric oxide synthase; NADPH oxidases, nicotinamide adenine dinucleotide phosphate oxidases; NBT, nitroblue tetrazolium; ESR, electron spin resonance; SOD, superoxide dismutase; GR, glutathione reductase; GSH/GSSG, reduced glutathione/oxidized glutathione ratio; GPx, glutathione peroxidase; 4-HNE, 4-hydroxynonenal; MDA, malondialdehyde. Nox2 have distinct roles in atherogenesis promoting vascular damage [112]. Recent data suggest an important role for Nox1 in diabetesassociated atherosclerosis [111]. Sukumar and co-workers showed that endothelial cell-specific insulin resistance increases Nox2 expression and leads to  $O_2^{\bullet-}$  generation in endothelial cells sufficient to foster arterial dysfunction [122,225]. Contrary to Nox1 and Nox2, expression of Nox4 was recently suggested to be vasculoprotective [112,113]. Apparently, under ischemia, hypertension or inflammatory stress Nox4-derived  $H_2O_2$  was suggested to have a protective role [112,113]. Finally, Nox5 (an isoform expressed in humans but not in rodents) is also able to generate ROS in blood vessels and seems to have a role in endothelial and VSMC growth [112].

# 2.2.2. Endothelial nitric oxide synthase

Nitric oxide generation is dependent on eNOS homodimerization in the presence of  $BH_4$ . However,  $BH_4$  is highly susceptible to oxidative degradation by ONOO<sup>-</sup> and in the absence of its cofactor, eNOS fails to dimerize fully, resulting in uncoupling of the enzyme and amplification of oxidative stress and generation of  $O_2^{\bullet-}$  rather than NO [73]. Uncoupled eNOS has been shown to contribute to increased superoxide production and endothelial dysfunction in a number of CVD, including coronary artery disease [129] and type 2 diabetes [130].

#### 2.2.3. Mitochondria

Enzymes of the inner mitochondrial membrane transfer electrons along the electron transport chain which generates a proton gradient, enabling ATP synthase to generate ATP. Under physiological conditions, this process produces ROS as byproducts [116,131,132]. Several mitochondrial antioxidant systems are in place to protect against ROS-induced damage to mitochondrial proteins, lipids and nucleic acids. However, under conditions of oxidative stress, these antioxidant systems are overwhelmed, allowing ROS to exert their deleterious effects and ultimately change mitochondrial function [116,131,134].

# 2.3. Mechanisms of defense against oxidative stress

The vasculature is endowed with protective antioxidant defense mechanisms, both enzymatic and nonenzymatic, to counteract the detrimental effects of ROS. Non-enzymatic antioxidant molecules include ascorbic acid (vitamin C),  $\alpha$ -tocopherol (vitamin E) and glutathione, while superoxide dismutases (SODs), catalase, glutathione peroxidases (GPxs) and thioredoxins represent important antioxidant enzymes which act to directly scavenge ROS, converting them to less reactive species [138–140].

#### 2.3.1. Superoxide dismutases

The SODs represent the first and most important line of enzymatic antioxidant defense against ROS. A ubiquitous family of enzymes, SODs catalyzes the conversion of  $O_2^{\bullet-}$  to  $H_2O_2$  and  $O_2$  [138,139,141]. Three distinct isoforms of SOD have been identified in vascular tissue: Cu/Zn SOD (encoded by *SOD1* gene) is located in the cytoplasm, MnSOD (encoded by *SOD2* gene) in the mitochondria and extracellular SOD (encoded by *SOD3* gene).

The importance of SODs as an antioxidant defense mechanism has been highlighted by gene transfer studies wherein SOD overexpression improved endothelial function [142,143]. Overexpression of SOD2 has also been shown to prevent hyperglycemia-associated production of  $O_2^{\bullet-}$ , activation of PKC and AGE formation [144], supporting a role for mitochondrial ROS production in diabetic macrovascular disease.

# 2.3.2. Catalase

Catalase is a highly catalytically efficient enzyme, primarily located in peroxisomes but also functions in the cytosol and catalyzes the conversion of  $H_2O_2$  to water following dismutation of  $O_2^{\bullet^-}$  by SOD [138,139,145]. Inherited catalase deficiency has been linked to elevated cardiovascular risk and increased incidence of diabetes mellitus [139]. However, experimental investigation has provided evidence that catalase provides only moderate protection against oxidative stress [146].

#### 2.3.3. Glutathione peroxidases

GPxs are a family of enzymes with an important role in antioxidant defense. Like catalase, GPxs reduce  $H_2O_2$  to water and lipid hydroperoxides to their corresponding alcohols. Detoxification of secondary oxidation products is vital and GPxs play an important role, reducing lipid peroxides [138,139,147].

There are several isozymes, GPx1 is the most abundant form in mammalian tissues. Mice with a disrupted GPx1 gene exhibit increased susceptibility to oxidative stress-inducing agents [148], while induction of this isozyme has been shown to provide protection against oxidative damage in endothelial cells [149]. In apoE-deficient mice, the deficiency of GPx1 accelerates and modifies atherosclerotic lesion progression [31,150]. Furthermore, transgenic GPx1 expression was observed to impair endothelial dysfunction [151]. Similarly, deficiency of GPx3 has been associated with decreased NO bioavailability and increased platelet-dependent thrombosis [139]. GPx4 knockout mice are not viable; they die during early embryonic development.

Glutathione is the principal low molecular weight, non-protein thiol in the cell [138]. Mainly found in the reduced state, glutathione has numerous functions in metabolism, signal transduction and gene expression [152]. GSH acts as an electron donor and can directly scavenge ROS but also acts as a cofactor in the conversion of H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O by GPxs [138,139].

Additional selenoproteins with similar antioxidant activities to GPxs include the thioredoxins [139], while the glutathione-S-transferases (GSTs) are examples of nonselenocysteine containing enzymes of significant importance in secondary oxidative stress defense, acting to detoxify reactive electrophiles [139,147].

#### 2.3.4. Thioredoxin

Thioredoxin (Trx) seems to exert most of its ROS-scavenging properties through Trx peroxidase (peroxiredoxin), which uses endogenous SH groups as reducing equivalents. Thioredoxin is present in endothelial- and vascular smooth muscle cells. Trx scavenges ROS and ONOO<sup>-</sup> and also reduce disulfides in proteins, peptides, and oxidized glutathione (GSSG) [140,153].

#### 2.3.5. Heme oxygenase

Heme oxygenase (HO) has indirect antioxidant effects through breakdown of free heme and the production of carbon monoxide, as well as biliverdin and bilirubin, which themselves have antioxidant properties [154]. There are two isoforms of this enzyme, a constitutive heme oxygenase, HO2, which is ubiquitously expressed in endothelial cells, and HO1, which is induced in response to oxidative stress, probably as an adaptive response. There is extensive evidence that HO1 can protect against vascular damage and atherogenesis [14,124,155]. The carbon monoxide has antiproliferative and anti-inflammatory as well as vasodilatory properties [156]. Genetic models of HO1 deficiency or overexpression of HO1 suggest that the actions of HO1 are important in modulating the severity of atherosclerosis [157].

#### 2.3.6. Paraoxonase

The paraoxonase (PON) family of enzymes acts as vascular antioxidant defense and protects against vascular disease [158]. The PON1 and PON3 enzymes are synthesized in the liver and circulate in plasma associated with HDL. The capacity of HDL in decreasing HDL and LDL lipid peroxidation largely depends on its PON1 content [158].

PON1 knockout mice are more prone to atherosclerosis [159] and low PON1 activity predicts acute cardiovascular events in human prospective studies [160]. Deletion of PON1 gene increases oxidative stress in mouse macrophages [161]. PON2 is expressed in many cell types. The enzyme has been shown to reduce ROS in human endothelial cells and vascular smooth muscle cells [162]. PON2-deficient mice with an  $apoE^{-/-}$  background developed more atherosclerotic lesions, whereas PON2-overexpressing mice were protected against those lesions [163].

Diabetes is characterized by increased oxidative stress and by decreased PON1 activity [164]. The ability of PON1 to protect against oxidative stress involved in major diseases such atherosclerosis and diabetes underlines the notion that strategies aimed at increasing PON1 activity and/or expression would have several benefits.

## 3. Potential therapeutic targets

In type 2 diabetes, glucotoxicity, lipotoxicity, insulin resistance and a mutual interaction between these factors occur to foster the development and progression of endothelial dysfunction. Conventional therapies to reduce hyperglycemia, dyslipidemia and insulin resistance represent important clinical options to improve endothelial function and delay the progression of vascular complications [165]. These conventional therapies and their effect on vascular function have been evaluated and reviewed elsewhere [166–169]. Noteworthy, most of these therapies are not completely effective in slowing vascular disease and would benefit from adjuvant cardiovascular protective therapies.

### 3.1. Cardiovascular therapies targeting the endothelium

The endothelium is a highly important target for therapy in cardiovascular disease [170]. It is rapidly and preferentially exposed to systemically administered agents and establishes a link with the underlying tissue, providing the researcher with a useful therapeutic target.

# 3.1.1. Potential therapeutic options for treating endothelial dysfunction by modulating eNOS

The vascular tone of arteries is primarily controlled by the bioavailability of NO, a key factor in vascular protection by preserving vessel reactivity. Thus, multiple potential therapeutic targets have been identified along the L-arginine–NOS pathway that could increase NO bioavailability. In Fig. 9 these sites are identified and include: at the level of the substrate, L-arginine; at the level of the NO-generating enzyme, eNOS; at the level of the soluble guanylyl cyclase, its main target; and at the level of the main effector of NO action, cGMP [5].

Administration of NO donors such as pentaerythritol tetranitrate (PETN) reduces oxidative stress (probably by inducing HO1) and improves endothelial dysfunction [135]. Thus, diabetic patients would benefit greatly from organic nitrate treatment devoid of classical adverse effects, such as nitrate-induced vascular oxidative stress, nitrate tolerance, and endothelial dysfunction (cross-tolerance) [125].

NO availability can be increased augmenting NO production by eNOS. The simplest way to modulate eNOS is administration of its substrate L-arginine [126] or its essential cofactor BH<sub>4</sub> or BH<sub>4</sub> analogs (Figs. 9, 10). Folic acid and its active form 5-methyltetrahydrofolate can modulate eNOS by improving BH<sub>4</sub> bioavailability in the vasculature by preventing its oxidation [218]. Midostaurin, betulinic acid and ursolic acid upregulate eNOS and simultaneously decrease NADPH oxidase expression. Novel small molecules AVE9488 and AVE3085 are two eNOS transcription enhancers that reverse eNOS uncoupling and preserve eNOS functionality and consequently increase NO bioavailability [136,171]. There is evidence that a cell-permeable peptide antagonizes the inhibitory actions of caveolin-1 on eNOS leading to increase in NO production [217]. Statins, angiotensin II type 1 receptor blockers, estrogens and erythropoietin enhance BH<sub>4</sub> synthesis by stimulating GTP cyclohydrolase I (GCH1) expression or activity. In vivo activation of AMP-activated protein kinase (AMPK) normalizes endothelial function due to an inhibition of GCH1 degradation associated with diabetes [137]. Statins, angiotensin II receptor blockers, ACE inhibitors, the aldosterone antagonist eplerenone and the renin inhibitor aliskiren prevent BH<sub>4</sub> oxidation by decreasing the expression and/or activity of NADPH oxidase (Figs. 9, 10). Statins can also directly activate eNOS via posttranslational mechanisms involving activation of the phosphatidylinositol 3-kinase/protein kinase Akt pathway [172].

In addition, cGMP levels may also be increased by inhibiting its metabolism by the phosphodiesterase-5 (PDE5) enzyme. The strategy of increasing the downstream mediator cGMP without affecting NO levels may be preferred due to the mixed outcomes in stroke reported in animal models following alterations in NO levels [173].

sGC stimulators and activators can treat the 2 forms of sGC insufficiency (i.e., diminished NO bioavailability and reduction of the catalytic capacity of sGC). Preliminary studies with both PDE5 inhibitors and sGC-targeted drugs have shown promising results [174–176].

# 3.1.2. Therapeutic approaches to reduce oxidative stress and/or increase antioxidant defense systems

Given the crucial role of ROS in endothelial function, considerable efforts have been made to discover therapies to reduce ROS in the vasculature. Despite promising initial observations, clinical trials with antioxidant vitamins C and E failed to show an improved cardiovascular outcome. Eventually new antioxidant molecules, targeted to the precise locations where ROS concentrations are elevated may, at an early stage, inhibit the mechanisms leading to diabetic complications [177].

The ability of PON1 to protect against oxidative stress and hydrolyze homocysteine thiolactone, a metabolite of homocysteine that can impair protein function promoting endothelial dysfunction, underlines the notion that strategies aimed at increasing PON1 activity and/or expression can be beneficial. Certain drugs (e.g. hypolipemic and antidiabetic compounds), dietary and life-style factors (e.g. antioxidants, polyphenols, moderate wine consumption) appear to increase PON1 activity [180,195]. Promoting an increment in PON1 activity may prove beneficial to prevent diabetes development [229] and slow down its cardiovascular complications [180,181,230].

Substances able to inhibit NADPH oxidases and prevent superoxide production may be useful for treatment of endothelial dysfunction [224]. Several inhibitors of the NADPH oxidase have been developed to specifically target NADPH oxidases with potential benefits [182-184, reviewed in 110 and 112]. Many cardiovascular drugs interfere with NADPH oxidases although most likely by indirect mechanisms. Additionally, flavonoids exhibit an inhibitory effect on NADPH oxidase combined with O<sub>2</sub>•<sup>-</sup> scavenging [184]. Nox-signaling pathways in the vasculature are likely to offer novel therapies. Discovering gene therapy targets towards enzymes involved in the homeostasis of vascular redox state is essential. Recently, the design and application of nanocarriers for delivery of antioxidants to the endothelium was performed with favorable outcomes [178]. Additionally, it has been described that delivery of genes encoding antioxidant defense enzymes (e.g. superoxide dismutase, catalase, glutathione peroxidase, PON1 or HO1) or eNOS suppress atherogenesis in animal models [49-51,66,97,123]. Similarly, delivery of genes encoding regulators of redox sensitive transcriptional factors (e.g. NFkappa B, AP-1, and NF-E2-related factor-2) or reactive oxygen species scavengers has been successfully used in experimental studies [72,179].

Induction of endogenous antioxidant enzymes by activators of the NF-E2-related factor-2/antioxidant response element pathway may be an interesting approach to obtain sufficient levels of antioxidants and reduce oxidative stress [72,77]. Additionally, SIRT1-mediated inhibition of p66Shc (a key effector driving vascular memory in diabetes) may also contribute to the prevention of oxidative stress-induced endothelial dysfunction in vascular diseases [78]. Despite the promising results from basic science, the clinical applicability of these strategies has proven to be difficult and challenging.

#### 3.2. Other therapeutic approaches

Novel therapeutic approaches designed to inhibit AGE formation and signaling (Fig. 11) [185,186], specifically directed to reduce inflammation [187,226] and restore the ox/redox balance in the endothelium may represent promising strategies to ameliorate vascular function in



**Fig. 9.** Potential sites of therapeutic intervention in the L-arginine–NO-synthase–soluble guanylyl cyclase pathway. They are indicated by numbers (from 1 to 14). The schematic diagram shows an endothelial cell in yellow and a vascular smooth muscle cell in light pink. The numbers indicate: (1) L-arginine supplementation. (2) Inhibition of protein arginine N-methyltransferase type I (PRMT-I) to prevent the formation of asymmetric dimethyl-L-arginine (ADMA). (3) Increasing the expression and/or the activity of dimethylarginine dimethylaminohydrolase (DDAH) to increase ADMA degradation. (4) Inhibition of arginase-2 to preclude L-arginine metabolism. (5) Increasing the expression and/or activity of endothelial ritric oxide synthase (eNOS). (6) Stimulation of endothelium-derived nitric oxide release. (7) Enhancing the expression and/or activity of guanosine triphosphate cyclohydrolase (GCH1), to increase tetrahydrobiopterin synthesis (BH4), or direct supplementation with BH4, or with its precursors. (8) Enhancing the expression and/or activity of dinydrofolate reductase (DHFR), to increase BH4 regeneration. (9) Scavengers of reactive oxygen species (ROS). (10) Inhibition of the activity and/or expression and/or activity of enzymes that generate ROS such as NADPH oxidases (NOX), cyclooxygenases (COX), lipoxygenases (LOX) or cytochrome P450 monoxygenases (P450). (11) Enhancing the expression and/or activity of enzymes that metabolized ROS such as superoxide dismutase (SOD) or gutathione peroxidase. (12) Activators of soluble guanylyl cyclase (sGC). (13) Activators of sGC. (14) Inhibitors of phosphodiesterase-5 (PDE-5). BH<sub>2</sub>, dihydrobiopterin; CAT-1, cationic amino acid transporters; CaV, voltage-activated calcium channel; cGMP, cyclic guanosine monophosphate; FAD, flavin adenine dinucleotide; FMN, flavin mononucleotide; O<sub>2</sub>-<sup>-</sup>, superoxide anion; ONOO<sup>-</sup>, peroxynitrite; PKG, protein kinase G.

diabetic state. Potential therapies also include: AMPK activators, PKC inhibitors, PARP inhibitors and rho-associated coiled-coil protein kinase (ROCK) inhibitors, among others.

AMPK is recognized as a key regulator of cellular energy status that has favorable effects on eNOS activity, insulin sensitivity, and mitochondrial function in a variety of cell types, including vascular cells. Therefore, pharmacological therapeutics that activate AMPK can be an important target in treating vascular complications in diabetes [137,219].

Inhibition of protein kinase C is another therapeutic approach. LY333531 (ruboxistaurin mesylate) has been shown to reduce oxidative stress and inflammation by blocking PKC-β isoform activation [188]. This inhibitory approach may also decrease vascular insulin resistance [133].

Furthermore, chronic treatment with the PARP inhibitors in rodent models has been demonstrated to improve endothelial dysfunction associated with aging [189,191]. In addition, pharmacological inhibition of PARP with PJ-34 restored endothelium-dependent vasodilation and reduced the levels of cytokines and inflammatory response [192,193]. Additionally, PARP-1 knockout protects against dyslipidemia-induced autonomic and vascular dysfunction in  $ApoE^{-/-}$  mice [194]. PARP inhibitors are potential therapies for diabetic vasculopathy. Pharmacological catalytic decomposition of peroxynitrite with FP15 has been demonstrated to effectively eliminate peroxynitrite and prevent PARP activation both in vitro and in vivo [220,221], thereby improving cardiovascular function in various disease models.

Rho-associated coiled-coil protein kinases are potential targets for treatment in vascular disease as suggested by the use of specific inhibitors as fasudil. Treatment with fasudil was protective against vascular-injury-induced leukocyte recruitment in wild type but not eNOS KO mice [222]. In diabetic animal models, studies have demonstrated a significant correlation between increased RhoA activity and impaired vascular function [223]. Thus, testing of fasudil and newer more specific second generation ROCK inhibitors in a diabetic setting would be of great interest in an effort to limit vascular complications [95].

Overall it is important to identify new targets for therapy and develop new agents for clinical use.

#### 3.3. Targeting vascular disease risk factors with nutritional therapeutics

Several nutritional agents such as lipoic acid, polyphenols, resveratrol, pomegranate, omega-3 fatty acids and bioavailable SOD have been shown to effectively improve and/or protect against endothelial dysfunction. Indeed, a comprehensive nutritional regimen can be adjoined with pharmacological approaches in order to target all of the risk factors that contribute to atherosclerosis.

Lipoic acid (LA) is a naturally occurring antioxidant that serves as a coenzyme in energy metabolism of fats, carbohydrates, and proteins. It can regenerate thioredoxin, vitamin C, and glutathione, which in turn can recycle vitamin E. LA reduces serum glucose levels in diabetic



**Fig. 10.** Focus on the potential eVOS-based therapeutic approaches for endothenal dystituction. The essential NOS collator feel any distribution (BH<sub>4</sub>) is synthesized from guardistic S – triphosphate (GTP) via a de novo pathway by the rate-limiting enzyme GTP cyclohydrolase I (GCH1). Alternatively, the synthesis of BH<sub>4</sub> can occur via other pathways including the salvage pathway, from dihydrobiopterin (BH<sub>2</sub>) back to BH<sub>4</sub>. As a substrate, L-arginine stimulates NO release from eNOS. Folic acid may improve eNOS functionality by stabilizing BH<sub>4</sub> and stimulating the endogenous regeneration of BH<sub>2</sub> back to BH<sub>4</sub>. Midostaurin, betulinic acid and ursolic acid upregulate eNOS and simultaneously decrease NADPH oxidase expression. AVE9488 and AVE3085 are two eNOS transcription enhancers that reverse eNOS uncoupling and improve eNOS functionality. Statins, angiotensin II type 1 receptor blockers (ARBs), estrogens and erythropoietin (EPO) enhance BH<sub>4</sub> synthesis by stimulating GCH1 expression or activity. Statins, ARBs, angiotensin-converting enzyme (ACE) inhibitors, the aldosterone antagonist eplerenone and the renin inhibitor aliskiren prevent BH<sub>4</sub> oxidation by decreasing the expression and/or activity of NADPH oxidase.

patients [195] and improves endothelial function in subjects with metabolic syndrome [196]. In type 2 diabetic animal models, we have previously shown a reduction of endothelial dysfunction after treatment with LA [21].

Different natural polyphenols have been shown to preserve endothelial function and prevent cardiovascular disease. Epidemiological evidence suggests a negative correlation between the consumption of polyphenol-rich foods (fruits, vegetables, and cocoa contained in chocolate) or beverages (wine, especially red wine, grape juice, green tea, among others.) and the incidence of cardiovascular disease [197–199]. Most polyphenols are mild antioxidants, some can reduce the activity of prooxidative NADPH oxidases, and others can stimulate antioxidative enzymes and eNOS [200–203]. The beneficial effects of silibinin on ADMA levels and endothelial dysfunction in db/db mice were recently described. The endothelium-dependent vasodilatation to ACh was impaired in db/db mice and was restored in the silibinin group, accompanied with a reduction of plasma and vascular levels of ADMA [128].

Several molecules with antioxidant properties (such as resveratrol, piceatannol, probucol, taurine) enhance dimethyl arginine dimethyl amino hydrolase activity, increasing ADMA catabolism [127].

The benefits of resveratrol include improvements in endothelial function [204–206]. Resveratrol seems to increase the number and activity of endothelial progenitor cells [205]. Resveratrol benefits the circulatory system by eliciting a decrease in the oxidation of LDL; by fostering decreases in platelet aggregation; and by promoting relaxation of arterioles [207]. Thus, resveratrol improves cardiovascular system by decreasing factors that contribute to the development of atherosclerosis and atherothrombosis [96,208].

Previous studies indicate that pomegranate and its extracts reduce oxidation and inflammation mainly through their effect on PON-1 activity, intervening at each step in the development of atherosclerosis [209–211].

Intake of omega-3 fatty acids might reduce Lp-PLA 2 levels and reduce the risk of vascular disease [212,213]. Omega-3 fatty acids serve as substrates for the conversion to a novel series of lipid mediators designated resolvins and protectins, with potent anti-inflammatory properties [187]. Studies have found that when omega-3 fatty acids were combined with rosuvastatin or other conventional therapies, the combination improved endothelial function [214,215].

Diminished levels of the antioxidant enzyme SOD have been linked with cardiovascular disease. Supplementation with GliSODin, a vegetal



Fig. 11. Potential sites of therapeutic intervention in order to reduce hyperglycemia and its downstream effects. On the left there are the potential target sites: glycemic control; glycosylation inhibition; crosslink breakers; RAGE blockers; blocking of PKC signaling pathway; and blocking of apoptosis. AGEs, advanced glycation end products; MAPK, mitogenic activated protein kinase; PKC, protein kinase C; RAGE, receptor for advanced glycation end products.

SOD associated with gliadin, was effective in controlling the thickness of the carotid artery intima and media layers as measured by ultrasonography-B [216]. Previous studies have demonstrated the preventive efficacy of GliSODin at a preclinical stage in subjects with risk factors of cardiovascular disease.

#### 4. Conclusions

Endothelial function is important for the homeostasis of the body and its dysfunction is associated with several pathophysiological conditions, including atherosclerosis, hypertension and diabetes. Understanding and treating endothelial dysfunction is a major issue in the prevention of vascular complications associated with all forms of diabetes mellitus.

Controlling a variety of risk factors causing inflammation and oxidative stress with combination therapy targeting intracellular mechanisms underlying metabolic alterations (such as inhibiting AGE formation and signaling and suppressing PKC activation) may simultaneously address multiple mechanisms underlying the pathogenesis of atherosclerosis. Since therapy addressing a single metabolic abnormality has not been effective, to reduce cardiovascular complications in type 2 diabetes may require simultaneous interventions within multiple metabolic and signaling pathways. Concurrent reduction of hyperglycemia, oxidative stress, inflammation and insulin resistance may be necessary to ameliorate the adverse effects that progress to diabetic vasculopathy in patients with cardiovascular risk factors.

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#### References

- P. Vallance, Importance of asymmetrical dimethylarginine in cardiovascular risk, Lancet 358 (2001) 2096–2097.
- [2] P.O. Bonetti, L.O. Lerman, A. Lerman, Endothelial dysfunction: a marker of atherosclerotic risk, Arterioscler. Thromb. Vasc. Biol. 23 (2003) 168–175.
- [3] M. Félétou, P.M. Vanhoutte, Endothelial dysfunction: a multifaceted disorder (the Wiggers award lecture), Am. J. Physiol. Heart Circ. Physiol. 291 (2006) H985-H1002.
- [4] S. Moncada, E.A. Higgs, Nitric oxide and the vascular endothelium, Handb. Exp. Pharmacol. 176 (2006) 213–254.
- [5] M. Félétou, The Endothelium: Part 1: Multiple Functions of the Endothelial Cells— Focus on Endothelium-derived Vasoactive Mediators, Morgan & Claypool Life Sciences, San Rafael (CA), 2011.
- [6] J.L. Beny, P. Brunet, H. Huggel, Interaction of bradykinin and des-Arg9-bradykinin with isolated pig coronary arteries: mechanical and electrophysiological events, Regul. Pept. 17 (1987) 181–190.
- [7] G. Chen, H. Suzuki, A.H. Weston, Acetylcholine releases endothelium-derived hyperpolarizing factor and EDRF from rat blood vessels, Br. J. Pharmacol. 95 (1988) 1165–1174.
- [8] R.M. Palmer, A.G. Ferrige, S. Moncada, Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor, Nature 327 (1987) 524–526.
- [9] L.J. Ignarro, G.M. Buga, K.S. Wood, R.E. Byrns, G. Chaudhuri, Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide, Proc. Natl. Acad. Sci. U. S. A. 84 (1987) 9265–9269.

- [10] S. Moncada, R. Gryglewski, S. Bunting, J.R. Vane, An enzyme isolated from arteries transforms prostaglandin endoperoxides to an unstable substance that inhibits platelet aggregation, Nature 263 (1976) 663–665.
- [11] M. Yanagisawa, H. Kurihara, S. Kimura, Y. Tomobe, M. Kobayashi, Y. Mitsui, Y. Yazaki, K. Goto, T. Masaki, A novel potent vasoconstrictor peptide produced by vascular endothelial cells, Nature 332 (1988) 411–415.
- [12] D.H. Endemann, E.L. Schiffrin, Endothelial dysfunction, J. Am. Soc. Nephrol. 15 (2004) 1983–1992.
- [13] A. Just, C.L. Whitten, W.J. Arendshorst, Reactive oxygen species participate in acute renal vasoconstrictor responses induced by ET<sub>A</sub> and ET<sub>B</sub> receptors, Am. J. Physiol. Renal Physiol. 294 (2008) F719–F728.
- [14] H.J. Duckers, M. Boehm, A.L. True, S.F. Yet, H. San, J.L. Park, R. Clinton Webb, M.E. Lee, G.J. Nabel, E.G. Nabel, Heme oxygenase-1 protects against vascular constriction and proliferation, Nat. Med. 7 (2001) 693–698.
- [15] P. Vallance, N. Chan, Endothelial function and nitric oxide: clinical relevance, Heart 85 (2001) 342–350.
- [16] C. Boulanger, T.F. Luscher, Release of endothelin from the porcine aorta: inhibition by endothelium-derived nitric oxide, J. Clin. Invest. 85 (1990) 587–590.
- [17] R. Joannides, W.E. Haefeli, L. Linder, V. Richard, E.H. Bakkali, C. Thuillez, T.F. Luscher, Nitric oxide is responsible for flow-dependent dilatation of human peripheral conduit arteries in vivo, Circulation 91 (1995) 1314–1319.
- [18] S.B. Wheatcroft, I.L. Williams, A.M. Shah, M.T. Kearney, Pathophysiological implications of insulin resistance on vascular endothelial function, Diabet. Med. 20 (2003) 255–268.
- [19] B. Datta, T. Tufnell-Barrett, R.A. Bleasdale, C.J. Jones, I. Beeton, V. Paul, M. Frenneaux, P. James, Red blood cell nitric oxide as an endocrine vasoregulator: a potential role in congestive heart failure, Circulation 109 (2004) 1339–1342.
- [20] B.S. Oemar, M.R. Tschudi, N. Godoy, V. Brovkovich, T. Malinski, T.F. Luscher, Reduced endothelial nitric oxide synthase expression and production in human atherosclerosis, Circulation 97 (1998) 2494–2498.
- [21] C.M. Sena, E. Nunes, T. Louro, T. Proença, R. Fernandes, M.R. Boarder, R.M. Seiça, Br. J. Pharmacol. 153 (2008) 894–906.
- [22] C. Dessy, O. Feron, J.L. Balligand, The regulation of endothelial nitric oxide synthase by caveolin: a paradigm validated in vivo and shared by the 'endothelium-derived hyperpolarizing factor', Pflugers Arch. 459 (2010) 817–827.
- [23] W.N. Duran, J.W. Breslin, F.A. Sanchez, The NO cascade, eNOS location, and microvascular permeability, Cardiovasc. Res. 87 (2010) 254–261.
- [24] J. Vásquez-Vivar, B. Kalyanaraman, P. Martásek, N. Hogg, B.S. Masters, H. Karoui, P. Tordo, K.A. Pritchard Jr., Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors, Proc. Natl. Acad. Sci. U. S. A. 95 (1998) 9220–9225.
- [25] C. Antoniades, C. Shirodaria, P. Leeson, A. Antonopoulos, N. Warrick, T. Van-Assche, C. Cunnington, D. Tousoulis, R. Pillai, C. Ratnatunga, C. Stefanadis, K.M. Channon, Association of plasma asymmetrical dimethylarginine (ADMA) with elevated vascular superoxide production and endothelial nitric oxide synthase uncoupling: implications for endothelial function in human atherosclerosis, Eur. Heart J. 30 (2009) 1142–1150.
- [26] C.A. Chen, T.Y. Wang, S. Varadharaj, L.A. Reyes, C. Hemann, M.A. Talukder, Y.R. Chen, LJ. Druhan, J.L. Zweier, S-glutathionylation uncouples eNOS and regulates its cellular and vascular function, Nature 468 (2010) 1115–1118.
- [27] R. Muniyappa, M. Montagnani, K.K. Kwang Kon Koh, M.J. Quon, Cardiovascular actions of insulin, Endocrine 28 (2007) 463–491.
- [28] K. Shinozaki, A. Kashiwagi, Y. Nishio, T. Okamura, Y. Yoshida, M. Masada, N. Toda, R. Kikkawa, Abnormal biopterin metabolism is a major cause of impaired endothelium-dependent relaxation through nitric oxide/O<sub>2</sub><sup>-</sup> imbalance in insulin-resistant rat aorta, Diabetes 48 (1999) 2437–2445.
- [29] S.J. Cleland, J.R. Petrie, M. Small, H.L. Elliott, J.M. Connell, Insulin action is associated with endothelial function in hypertension and type 2 diabetes, Hypertension 35 (2000) 507–511.
- [30] M.A. Vincent, E.J. Barrett, J.R. Lindner, M.G. Clark, S. Rattigan, Inhibiting NOS blocks microvascular recruitment and blunts muscle glucose uptake in response to insulin, Am. J. Physiol. Endocrinol. Metab. 285 (2003) E123–E129.
- [31] P. Lewis, N. Stefanovic, J. Pete, A.C. Calkin, S. Giunti, V. Thallas-Bonke, K.A. Jandeleit-Dahm, T.J. Allen, I. Kola, M.E. Cooper, J.B. de Haan, Lack of the antioxidant enzyme glutathione peroxidase-1 accelerates atherosclerosis in diabetic apolipoprotein E-deficient mice, Circulation 115 (2007) 2178–2187.
- [32] H. Zhang, K.C. Dellsperger, C. Zhang, The link between metabolic abnormalities and endothelial dysfunction in type 2 diabetes: an update, Basic Res. Cardiol. 107 (2012) 237–248.
- [33] V.P. Valkonen, H. Päivä, J.T. Salonen, T.A. Lakka, T. Lehtimäki, J. Laakso, R. Laaksonen, Risk of acute coronary events and serum concentration of asymmetrical dimethylarginine, Lancet 358 (2001) 2127–2128.
- [34] F. Mittermayer, K. Krzyzanowska, M. Exner, W. Mlekusch, J. Amighi, S. Sabeti, E. Minar, M. Muller, M. Wolzt, M. Schillinger, Asymmetric dimethylarginine predicts major adverse cardiovascular events in patients with advanced peripheral artery disease, Arterioscler. Thromb. Vasc. Biol. 26 (2006) 2536–2540.
- [35] K. Krzyzanowska, F. Mittermayer, M. Wolzt, G. Schernthaner, Asymmetric dimethylarginine predicts cardiovascular events in patients with type 2 diabetes, Diabetes Care 30 (2007) 1834–1839.
- [36] F. Lovren, Y. Pan, A. Quan, K.K. Singh, P.C. Shukla, M. Gupta, M. Al Omran, H. Teoh, S. Verma, Adropin is a novel regulator of endothelial function, Circulation 122 (2010) S185–S192.
- [37] D.G. Harrison, Endothelial function and oxidant stress, Clin. Cardiol. 20 (1997) 11–17.
   [38] D. Behrendt, P. Ganz, Endothelial function. From vascular biology to clinical appli-
- [56] D. Semener, T. Ganz, Encontential function. From Vascular biology to chilled applications, Am. J. Cardiol. 21 (2002) 40L–48L.
- [39] M.S. Wolin, S.A. Gupte, B.H. Neo, Q. Gao, M. Ahmad, Oxidant-redox regulation of pulmonary vascular responses to hypoxia and nitric oxide-cGMP signaling, Cardiol. Rev. 18 (2010) 89–93.

- [40] V.B. O'Donnell, B.A. Freeman, Interactions between nitric oxide and lipid oxidation pathways: implications for vascular disease, Circ. Res. 88 (2001) 12–21.
- [41] G. Kojda, D. Harrison, Interactions between NO and reactive oxygen species: pathophysiological importance in atherosclerosis, hypertension, diabetes and heart failure, Cardiovasc. Res. 43 (1999) 562–571.
- [42] K. Egan, G.A. FitzGerald, Eicosanoids and the vascular endothelium, Handb. Exp. Pharmacol. 176 (2006) 189–211.
- [43] M. Félétou, P.M. Vanhoutte, Endothelium-dependent hyperpolarization of canine coronary smooth muscle, Br. J. Pharmacol. 93 (1988) 515–524.
- [44] C.J. Garland, C.R. Hiley, K.A. Dora, EDHF: spreading the influence of the endothelium, Br. J. Pharmacol. 164 (2011) 839–852.
- [45] M. Félétou, P.M. Vanhoutte, Endothelium-derived hyperpolarizing factor: where are we now? Arterioscler. Thromb. Vasc. Biol. 26 (2006) 1215–1225.
- [46] H. Shimokawa, H. Yasutake, K. Fujii, M.K. Owada, R. Nakaike, Y. Fukumoto, T. Takayanagi, T. Nagao, K. Egashira, M. Fujishima, A. Takeshita, The importance of the hyperpolarizing mechanism increases as the vessel size decreases in endothelium-dependent relaxations in rat mesenteric circulation, J. Cardiovasc. Pharmacol. 28 (1996) 703–711.
- [47] K.T. Kang, J.C. Sullivan, J.M. Sasser, J.D. Imig, J.S. Pollock, Novel nitric oxide synthase – dependent mechanism of vasorelaxation in small arteries from hypertensive rats, Hypertension 49 (2007) 893–901.
- [48] I. Grgic, B.P. Kaistha, J. Hoyer, R. Köhler, Endothelial Ca2+-activated K+ channels in normal and impaired EDHF-dilator responses-relevance to cardiovascular pathologies and drug discovery, Br. J. Pharmacol. 157 (2009) 509–526.
- [49] J.H. Bräsen, O. Leppänen, M. Inkala, T. Heikura, M. Levin, F. Ahrens, J. Rutanen, H. Pietsch, D. Bergqvist, A.L. Levonen, S. Basu, T. Zeller, G. Klöppel, M.O. Laukkanen, S. Ylä-Herttuala, Extracellular superoxide dismutase accelerates endothelial recovery and inhibits in-stent restenosis in stented atherosclerotic Watanabe heritable hyperlipidemic rabbit aorta, J. Am. Coll. Cardiol. 50 (2007) 2249–2253.
- [50] H. Yang, L.J. Roberts, M.J. Shi, L.C. Zhou, B.R. Ballard, A. Richardson, Z.M. Guo, Retardation of atherosclerosis by overexpression of catalase or both Cu/Zn-superoxide dismutase and catalase in mice lacking apolipoprotein E, Circ. Res. 95 (2004) 1075–1081.
- [51] P.J. Guns, T. Van Assche, W. Verreth, P. Fransen, B. Mackness, M. Mackness, P. Holvoet, H. Bult, Paraoxonase 1 gene transfer lowers vascular oxidative stress and improves vasomotor function in apolipoprotein E-deficient mice with pre-existing atherosclerosis, Br. J. Pharmacol. 153 (2008) 508–516.
- [52] I. Fleming, U.R. Michaelis, D. Bredenkötter, B. Fisslthaler, F. Dehghani, R.P. Brandes, R. Busse, Endothelium-derived hyperpolarizing factor synthase (cytochrome P450 2C9) is a functionally significant source of reactive oxygen species in coronary arteries, Circ. Res. 88 (2001) 44–51.
- [53] S. Taddei, L. Ghiadoni, A. Virdis, D. Versari, A. Salvetti, Mechanisms of endothelial dysfunction: clinical significance and preventive non-pharmacological therapeutic strategies, Curr. Pharm. Des. 9 (2003) 2385–2402.
- [54] M.H. Laughlin, S.C. Newcomer, S.B. Bender, Importance of hemodynamic forces as signals for exercise-induced changes in endothelial cell phenotype, J. Appl. Physiol. 104 (2008) 588–600.
- [55] W.T. Cade, Diabetes-related microvascular and macrovascular diseases in the physical therapy setting, Phys. Ther. 88 (2008) 322–335.
- [56] F. Addabbo, M. Montagnani, M.S. Goligorsky, Mitochondria and reactive oxygen species, Hypertension 53 (2009) 885–892.
- [57] A. Hirose, T. Tanikawa, H. Mori, Y. Okada, Y. Tanaka, Advanced glycation endproducts increase endothelial permeability through the RAGE/Rho signaling pathway, FEBS Lett. 584 (2010) 61–66.
- [58] J.B. Wan, L.L. Huang, R. Rong, R. Tan, J. Wang, J.X. Kang, Endogenously decreasing tissue n – 6/n – 3 fatty acid ratio reduces atherosclerotic lesions in apolipoprotein E-deficient mice by inhibiting systemic and vascular inflammation, Arterioscler. Thromb. Vasc. Biol. 30 (2010) 2487–2494.
- [59] D. Versari, E. Daghini, A. Virdis, L. Ghiadoni, S. Taddei, Endothelial dysfunction as a target for prevention of cardiovascular disease, Diabetes Care 32 (2009) S314–S321.
- [60] F. Grover-Páez, A.B. Zavalza-Gómez, Endothelial dysfunction and cardiovascular risk factors, Diabetes Res. Clin. Pract. 84 (2009) 1–10.
- [61] S. Bhatti, A. Hakeem, M. Cilingiroglu, Lp-PLA(2) as a marker of cardiovascular diseases, Curr. Atheroscler. Rep. 12 (2010) 140–144.
- [62] J.A. Beckman, M.A. Creager, P. Libby, Diabetes and atherosclerosis: epidemiology, pathophysiology, and management, JAMA 15 (2002) 2570–2581.
- [63] Nesto, Correlation between cardiovascular disease and diabetes mellitus: current concepts, Am. J. Med. 116 (2004) 11S–22S.
- [64] D.K. Singh, P. Winocour, K. Farrington, Endothelial cell dysfunction, medial arterial calcification and osteoprotegerin in diabetes, Br. J. Diabetes Vasc. Dis. 10 (2010) 71–77.
- [65] P. Highlander, G.P. Shaw, Current pharmacotherapeutic concepts for the treatment of cardiovascular disease in diabetics, Therap. Adv. Cardiovasc. Dis. 4 (2010) 43–54.
- [66] S.H. Juan, T.S. Lee, K.W. Tseng, J.Y. Liou, S.K. Shyue, K.K. Wu, L.Y. Chau, Adenovirus-mediated heme oxygenase-1 gene transfer inhibits the development of atherosclerosis in apolipoprotein E-deficient mice, Circulation 104 (2001) 1519–1525.
- [67] C. Riccardo, B. Stella, J.A. Terri, Linking diabetes and atherosclerosis, Expert. Rev. Endocrinol. Metab. 4 (2009) 603–624.
- [68] A. Ceriello, Point: postprandial glucose levels are a clinically important treatment target, Diabetes Care 33 (2010) 1905–1907.
- [69] D. Grassi, G. Desideri, S. Necozione, F. Ruggieri, J.B. Blumberg, M. Stornello, C. Ferri, Protective effects of flavanol-rich dark chocolate on endothelial function and wave reflection during acute hyperglycemia, Hypertension 60 (2012) 827–832.

- [70] J.S. Skyler, R. Bergenstal, R.O. Bonow, J. Buse, P. Deedwania, E.A. Gale, B.V. Howard, M.S. Kirkman, M. Kosiborod, P. Reaven, R.S. Sherwin, Intensive glycemic control and the prevention of cardiovascular events: implications of the ACCORD, ADVANCE, and VA diabetes trials: a position statement of the American Diabetes Association and a scientific statement of the American College of Cardiology Foundation and the American Heart Association, Circulation 119 (2009) 351–357.
- [71] M. Brownlee, Biochemistry and molecular cell biology of diabetic complications, Nature 13 (2001) 813–820.
- [72] A.L. Levonen, M. Inkala, T. Heikura, S. Jauhiainen, H.K. Jyrkkänen, E. Kansanen, K. Määttä, E. Romppanen, P. Turunen, J. Rutanen, S. Ylä-Herttuala, Nrf2 gene transfer induces antioxidant enzymes and suppresses smooth muscle cell growth in vitro and reduces oxidative stress in rabbit aorta in vivo, Arterioscler. Thromb. Vasc. Biol. 27 (2007) 741–747.
- [73] S. Milstien, Z. Katusic, Oxidation of tetrahydrobiopterin by peroxynitrite: implications for vascular endothelial function, Biochem. Biophys. Res. Commun. 263 (1999) 681–684.
- [74] M. Brownlee, The pathobiology of diabetes complications: a unifying mechanism, Diabetes 54 (2005) 1615–1625.
- [75] M.A. Creager, T.F. Luscher, F. Cosentino, J.A. Beckman, Diabetes and vascular disease: pathophysiology, Clin. Consequences Med. Ther. I Circ. 108 (2003) 1527–1532.
- [76] A. D'Souza, M. Hussain, F.C. Howarth, N.M. Woods, K. Bidasee, J. Singh, Pathogenesis and pathophysiology of accelerated atherosclerosis in the diabetic heart, Mol. Cell. Biochem, 331 (2009) 89–116.
- [77] G.V. Velmurugan, N.R. Sundaresan, M.P. Gupta, C. White, Defective Nrf2-dependent redox signaling contributes to microvascular dysfunction in type 2 diabetes, Cardiovasc. Res. (2013), http://dx.doi.org/10.1093/cvr/cvt125 (in press).
- [78] S. Zhou, H.Z. Chen, Y.Z. Wan, Q.J. Zhang, Y.S. Wei, S. Huang, J.J. Liu, Y.B. Lu, Z.Q. Zhang, R.F. Yang, R. Zhang, H. Cai, D.P. Liu, C.C. Liang, Repression of P66Shc expression by SIRT1 contributes to the prevention of hyperglycemia-induced endothelial dysfunction, Circ. Res. 109 (2011) 639–648.
- [79] A.M. Schmidt, S.D. Yan, J.L. Wautier, D. Stern, Activation of receptor for advanced glycation end products: a mechanism for chronic vascular dysfunction in diabetic vasculopathy and atherosclerosis, Circ. Res. 84 (1999) 489–497.
- [80] M.P. Wautier, O. Chappey, S. Corda, D.M. Stern, A.M. Schmidt, J.L. Wautier, Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE, Am. J. Physiol. Endocrinol. Metab. 280 (2001) E685–E694.
- [81] T. Chavakis, A. Bierhaus, P.P. Nawroth, RAGE (receptor for advanced glycation end products): a central player in the inflammatory response, Microbes Infect. 6 (2004) 1219–1225.
- [82] U. Chakravarthy, R.G. Hayes, A.W. Stitt, E. McAuley, D.B. Archer, Constitutive nitric oxide synthase expression in retinal vascular endothelial cells is suppressed by high glucose and advanced glycation end products, Diabetes 47 (1998) 945–952.
- [83] B. Xu, R. Chibber, D. Ruggiero, E. Kohner, J. Ritter, A. Ferro, Impairment of vascular endothelial nitric oxide synthase activity by advanced glycation end products, FASEB J. 17 (2003) 1289–1291.
- [84] P. Quehenberger, A. Bierhaus, P. Fasching, C. Muellner, M. Klevesath, M. Hong, G. Stier, M. Sattler, E. Schleicher, W. Speiser, P.P. Nawroth, Endothelin 1 transcription is controlled by nuclear factor-kappaB in AGE-stimulated cultured endothelial cells, Diabetes 49 (2000) 1561–1570.
- [85] C.M. Sena, P. Matafome, J. Crisóstomo, L. Rodrigues, R. Fernandes, P. Pereira, R.M. Seiça, Methylglyoxal promotes oxidative stress and endothelial dysfunction, Pharmacol. Res. 65 (2012) 497–506.
- [86] M.A. Potenza, S. Gagliardi, C. Nacci, M.R. Carratu, M. Montagnani, Endothelial dysfunction in diabetes: from mechanisms to therapeutic targets, Curr. Med. Chem. 16 (2009) 94–112.
- [87] G.M. Reaven, Banting lecture 1988. Role of insulin resistance in human disease, Diabetes 37 (1988) 1595–1607.
- [88] J. Avruch, Insulin signal transduction through protein kinase cascades, Mol. Cell. Biochem. 182 (1998) 31–48.
- [89] J. Kim, M. Montagnani, K.K. Koh, M.J. Quon, Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms, Circulation 113 (2006) 1888–1904.
- [90] R. Muniyappa, J.R. Sowers, Roles of insulin resistance in endothelial dysfunction, Rev. Endocr. Metab. Disord. 14 (2013) 5–12.
- [91] X. Du, D. Edelstein, S. Obici, N. Higham, M.H. Zou, M. Brownlee, Insulin resistance reduces arterial prostacylcin synthase and eNOS activities by increasing endothelial fatty acid oxidation, J. Clin. Invest. 116 (2006) 1071–1080.
- [92] F. Giacco, M. Brownlee, Oxidative stress and diabetic complications, Circ. Res. 107 (2010) 1058-1070.
- [93] T. Inoguchi, P. Li, F. Umeda, H.Y. Yu, M. Kakimoto, M. Imamura, T. Aoki, T. Etoh, T. Hashimoto, M. Naruse, H. Sano, H. Utsumi, H. Nawata, High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells, Diabetes 49 (2000) 1939–1945.
- [94] C. Capurso, A. Capurso, From excess adiposity to insulin resistance: the role of free fatty acids, Vascul. Pharmacol. 57 (2012) 91–97.
- [95] A. Sharma, P.N. Bernatchez, J.B. de Haan, Targeting endothelial dysfunction in vascular complications associated with diabetes, Int. J. Vasc. Med. 2012 (2012) 750126, http://dx.doi.org/10.1155/2012/750126.
- [96] H. Li, U. Förstermann, Red wine and cardiovascular health, Circ. Res. 111 (2012) 959–961.
- [97] M.O. Laukkanen, A. Kivela, T. Rissanen, J. Rutanen, M.K. Karkkainen, O. Leppanen, J.H. Brasen, S. Yla-Herttuala, Adenovirus-mediated extracellular superoxide dismutase gene therapy reduces neointima formation in balloon-denuded rabbit aorta, Circulation 106 (2002) 1999–2003.

- [98] A. Natali, E. Toschi, S. Baldeweg, D. Ciociaro, S. Favilla, L. Saccà, E. Ferrannini, Clustering of insulin resistance with vascular dysfunction and low-grade inflammation in type 2 diabetes, Diabetes 55 (2006) 1133–1140.
- [99] S., Kawashima, M. Yokoyama, Dysfunction of endothelial nitric oxide synthase and atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 24 (2004) 998–1005.
- [100] A. Dresner, D. Laurent, M. Marcucci, M.E. Griffin, S. Dufour, G.W. Cline, L.A. Slezak, D.K. Andersen, R.S. Hundal, D.L. Rothman, K.F. Petersen, G.I. Shulman, Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity, J. Clin. Invest. 103 (1999) 253–259.
   [101] H.O. Steinberg, M. Tarshoby, R. Monestel, G. Hook, J. Cronin, A. Johnson, B.
- [101] H.O. Steinberg, M. Tarshoby, R. Monestel, G. Hook, J. Cronin, A. Johnson, B. Bayazeed, A.D. Baron, Elevated circulating free fatty acid levels impair endothelium-dependent vasodilation, J. Clin. Invest. 100 (1997) 1230–1239.
- [102] X.L. Wang, L. Zhang, K. Youker, M.X. Zhang, J. Wang, S.A. Le-Maire, J.S. Coselli, Y.H. Shen, Free fatty acids inhibit insulin signaling-stimulated endothelial nitric oxide synthase activation through upregulating PTEN or inhibiting Akt kinase, Diabetes 55 (2006) 2301–2310.
- [103] G. Boden, P. She, M. Mozzoli, P. Cheung, K. Gumireddy, P. Reddy, X. Xiang, Z. Luo, N. Ruderman, Free fatty acids produce insulin resistance and activate the proinflammatory nuclear factor-kappaB pathway in rat liver, Diabetes 54 (2005) 3458–3465.
- [104] Z. Gao, X. Zhang, A. Zuberi, D. Hwang, M.J. Quon, M. Lefevre, J. Ye, Inhibition of insulin sensitivity by free fatty acids requires activation of multiple serine kinases in 3T3-L1 adipocytes, Mol. Endocrinol. 18 (2004) 2024–2034.
- [105] M. Jove, A. Planavila, R.M. Sanchez, M. Merlos, J.C. Laguna, M. Vazquez-Carrera, Palmitate induces tumor necrosis factor-alpha expression in C2C12 skeletal muscle cells by a mechanism involving protein kinase C and nuclear factor-kappaB activation, Endocrinology 147 (2006) 552–561.
- [106] F. Kim, K.A. Tysseling, J. Rice, M. Pham, L. Haji, B.M. Gallis, A.S. Baas, P. Paramsothy, C.M. Giachelli, M.A. Corson, E.W. Raines, Free fatty acid impairment of nitric oxide production in endothelial cells is mediated by IKKbeta, Arterioscler. Thromb. Vasc. Biol. 25 (2005) 989–994.
- [107] K. Naruse, C. Rask-Madsen, N. Takahara, S.W. Ha, K. Suzuma, K.J. Way, J.R. Jacobs, A.C. Clermont, K. Ueki, Y. Ohshiro, J. Zhang, A.B. Goldfine, G.L. King, Activation of vascular protein kinase C-beta inhibits Akt-dependent endothelial nitric oxide synthase function in obesity-associated insulin resistance, Diabetes 55 (2006) 691–698.
- [108] H. Cai, D.G. Harrison, Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress, Circ. Res. 87 (2000) 840–844.
- [109] D. Harrison, K.K. Griendling, U. Landmesser, B. Hornig, H. Drexler, Role of oxidative stress in atherosclerosis, Am. J. Cardiol. 91 (2003) 7A–11A.
- [110] A. Schramm, P. Matusik, G. Osmenda, T.J. Gusik, Targeting NADPH oxidases in vascular pharmacology, Vasc. Pharmacol. 56 (2012) 216–231.
- [111] S.P. Gray, E. Di Marco, J. Okabe, C. Szyndralewiez, F. Heitz, A.C. Montezano, J.B. de Haan, C. Koulis, A. El-Osta, K.L. Andrews, J.P. Chin-Dusting, R.M. Touyz, K. Wingler, M.E. Cooper, H.H. Schmidt, K.A. Jandeleit-Dahm, NADPH oxidase 1 plays a key role in diabetes mellitus-accelerated atherosclerosis, Circulation 127 (2013) 1888–1902.
- [112] B. Lassegue, K.K. Griendling, NADPH oxidases: functions and pathologies in the vasculature, Arterioscler. Thromb. Vasc. Biol. 30 (2010) 653–661.
- [113] K. Schroder, M. Zhang, S. Benkhoff, A. Mieth, R. Pliquett, J. Kosowski, C. Kruse, P. Luedike, U.R. Michaelis, N. Weissmann, S. Dimmeler, A.M. Shah, R.P. Brandes, Nox4 is a protective reactive oxygen species generating vascular NADPH oxidase, Circ. Res. 110 (2012) 1217–1225.
- [114] B. Chance, H. Sies, A. Boveris, Hydroperoxide metabolism in mammalian organs, Physiol. Rev. 59 (1979) 527–605.
- [115] P. Jezek, L. Hlavata, Mitochondria in homeostasis of reactive oxygen species in cell, tissues, and organism, Int. J. Biochem. Cell Biol. 37 (2005) 2478–2503.
- [116] J.F. Turrens, Mitochondrial formation of reactive oxygen species, J. Physiol. 552 (2003) 335–344.
- [117] K.K. Griendling, M. Ushio-Fukai, B. Lassegue, R.W. Alexander, Angiotensin II signaling in vascular smooth muscle. New concepts, Hypertension 29 (1997) 366–373.
- [118] R. Ray, A.M. Shah, NADPH oxidase and endothelial cell function, Clin. Sci. (Lond.) 109 (2005) 217–226.
- [119] M. Soccio, E. Toniato, V. Evangelista, M. Carluccio, R. De Caterina, Oxidative stress and cardiovascular risk: the role of vascular NAD(P)H oxidase and its genetic variants, Eur. J. Clin. Invest. 35 (2005) 305–314.
- [120] H. Suzuki, A. Swei, B.W. Zweifach, G.W. Schmid-Schonbein, In vivo evidence for microvascular oxidative stress in spontaneously hypertensive rats. Hydroethidine microfluorography, Hypertension 25 (1995) 1083–1089.
- [121] G.W. De Keulenaer, D.C. Chappell, N. Ishizaka, R.M. Nerem, R.W. Alexander, K.K. Griendling, Oscillatory and steady laminar shear stress differentially affect human endothelial redox state: role of a superoxide-producing NADH oxidase, Circ. Res. 82 (1998) 1094–1101.
- [122] P. Sukumar, H. Viswambharan, H. Imrie, R.M. Cubbon, N. Yuldasheva, M. Gage, S. Galloway, A. Skromna, P. Kandavelu, C.X. Santos, V.K. Gatenby, J. Smith, D.J. Beech, S.B. Wheatcroft, K.M. Channon, A.M. Shah, M.T. Kearney, Nox2 NADPH oxidase has a critical role in insulin resistance-related endothelial cell dysfunction, Diabetes 62 (2013) 2130–2134.
- [123] T. Van Assche, V. Huygelen, M.J. Crabtree, Targeting vascular redox biology through antioxidant gene delivery: a historical view and current perspectives, Recent Pat. Cardiovasc. Drug Discov. 6 (2011) 89–102.
- [124] E. Marcantoni, L. Di Francesco, M. Dovizio, A. Bruno, P. Patrignani, Novel insights into the vasoprotective role of heme oxygenase-1, Int. J. Hypertens. (2012), http://dx.doi.org/10.1155/2012/127910.
- [125] J.C. Irvine, B.K. Kemp-Harper, R.E. Widdop, Chronic administration of the HNO donor Angeli's salt does not lead to tolerance, cross-tolerance, or endothelial

dysfunction: comparison with GTN and DEA/NO, Antioxid. Redox Signal. 14 (2011) 1615–1624.

- [126] T. Ramprasath, P.H. Kumar, S.S. Puhari, P.S. Murugan, V. Vasudevan, G.S. Selvam, L-Arginine ameliorates cardiac left ventricular oxidative stress by upregulating eNOS and Nrf2 target genes in alloxan-induced hyperglycemic rats, Biochem. Biophys. Res. Commun. 428 (2012) 389–394.
- [127] M. Frombaum, S. Le Clanche, D. Bonnefont-Rousselot, D. Borderie, Antioxidant effects of resveratrol and other stilbene derivatives on oxidative stress and NO bioavailability: potential benefits to cardiovascular diseases, Biochimie 94 (2012) 269–276.
- [128] G.L. Volti, S. Salomone, V. Sorrenti, A. Mangiameli, V. Urso, I. Siarkos, F. Galvano, F. Salamone, Effect of silibinin on endothelial dysfunction and ADMA levels in obese diabetic mice, Cardiovasc. Diabetol. (2011) 10–62.
- [129] D.E. Kelley, J.A. Simoneau, Impaired free fatty acid utilization by skeletal muscle in non-insulin dependent diabetes mellitus, J. Clin. Invest. 94 (1994) 2349–2356.
- [130] T.J. Guzik, S. Mussa, D. Gastaldi, J. Sadowski, C. Ratnatunga, R. Pillai, K.M. Channon, Mechanisms of increased vascular superoxide production in human diabetes mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase, Circulation 105 (2002) 1656–1662.
- [131] S.W. Ballinger, Mitochondrial dysfunction in cardiovascular disease, Free Radic. Biol. Med. 38 (2005) 1278–1295.
- [132] M.P. Murphy, How mitochondria produce reactive oxygen species, Biochem. J. 417 (2009) 1–13.
- [133] Q. Li, K. Park, C. Li, C. Rask-Madsen, A. Mima, W. Qi, K. Mizutani, P.L. Huang, G.L. King, Induction of vascular insulin resistance, endothelin-1 expression, and acceleration of atherosclerosis by the overexpression of protein kinase C  $\beta$  isoform in the endothelium, Circ. Res. 113 (2013) 418–427.
- [134] B.B. Lowell, G.I. Shulman, Mitochondrial dysfunction and type 2 diabetes, Science 307 (2005) 384–387.
- [135] S. Schuhmacher, M. Oelze, F. Bollmann, H. Kleinert, C. Otto, T. Heeren, S. Steven, M. Hausding, M. Knorr, A. Pautz, K. Reifenberg, E. Schulz, T. Gori, P. Wenzel, T. Münzel, A. Daiber, Vascular dysfunction in experimental diabetes is improved by pentaerithrityltetranitrate but not isosorbide-5-mononitrate therapy, Diabetes 60 (2011) 2608–2616.
- [136] W.S. Cheang, W.T. Wong, X.Y. Tian, Q. Yang, H.K. Lee, G.W. He, X. Yao, Y. Huang, Endothelial nitric oxide synthase enhancer reduces oxidative stress and restores endothelial function in db/db mice, Cardiovasc. Res. 92 (2011) 267–275.
- [137] S. Wang, J. Xu, P. Song, B. Viollet, M.H. Zou, In vivo activation of AMP-activated protein kinase attenuates diabetes-enhanced degradation of GTP cyclohydrolase I, Diabetes 58 (2009) 1893–1901.
- [138] J.M. Li, A.M. Shah, Endothelial cell superoxide generation: regulation and relevance for cardiovascular pathophysiology, Am. J. Physiol. Regul. Integr. Comp. Physiol. 287 (2004) R1014–R1030.
- [139] J.A. Leopold, J. Loscalzo, Oxidative enzymopathies and vascular disease, Arterioscler. Thromb. Vasc. Biol. 25 (2005) 1332–1340.
- [140] N. Maulik, D.K. Das, Emerging potential of thioredoxin and thioredoxin interacting proteins in various disease conditions, Biochim. Biophys. Acta 1780 (2008) 1368–1382.
- [141] I.N. Zelko, T.J. Mariani, R.J. Folz, Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression, Free Radic. Biol. Med. 33 (2002) 337–349.
- [142] J.P. Fennell, M.J. Brosnan, A.J. Frater, C.A. Hamilton, M.Y. Alexander, S.A. Nicklin, D.D. Heistad, A.H. Baker, A.F. Dominiczak, Adenovirus-mediated overexpression of extracellular superoxide dismutase improves endothelial dysfunction in a rat model of hypertension, Gene Ther. 9 (2002) 110–117.
- [143] M. Zanetti, J. Sato, Z.S. Katusic, T. O'Brien, Gene transfer of superoxide dismutase isoforms reverses endothelial dysfunction in diabetic rabbit aorta, Am. J. Physiol. Heart Circ, Physiol. 280 (2001) H2516–H2523.
- [144] T. Nishikawa, D. Edelstein, X.L. Du, S. Yamagishi, T. Matsumura, Y. Kaneda, M.A. Yorek, D. Beebe, P.J. Oates, H.P. Hammes, I. Giardino, M. Brownlee, Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage, Nature 404 (2000) 787–790.
- [145] I. Fridovich, Oxygen toxicity: a radical explanation, J. Exp. Biol. 201 (1998) 1203–1209.
- [146] V.R. Muzykantov, Targeting of superoxide dismutase and catalase to vascular endothelium, J. Control. Release 71 (2001) 1–21.
- [147] J.D. Hayes, J.U. Flanagan, I.R. Jowsey, Glutathione transferases, Annu. Rev. Pharmacol. Toxicol. 45 (2005) 51–88.
- [148] J.B. de Haan, C. Bladier, P. Griffiths, M. Kelner, R.D. O'Shea, N.S. Cheung, R.T. Bronson, M.J. Silvestro, S. Wild, S.S. Zheng, P.M. Beart, P.J. Hertzog, I. Kola, Mice with a homozygous null mutation for the most abundant glutathione peroxidase, Gpx1, show increased susceptibility to the oxidative stressinducing agents paraquat and hydrogen peroxide, J. Biol. Chem. 273 (1998) 22528–22536.
- [149] Y. Zhang, D.E. Handy, J. Loscalzo, Adenosine-dependent induction of glutathione peroxidase 1 in human primary endothelial cells and protection against oxidative stress, Circ. Res. 96 (2005) 831–837.
- [150] M. Torzewski, V. Ochsenhirt, A.L. Kleschyov, M. Oelze, A. Daiber, H. Li, H. Rossmann, S. Tsimikas, K. Reifenberg, F. Cheng, H.A. Lehr, S. Blankenberg, U. Forstermann, T. Munzel, K.J. Lackner, Deficiency of glutathione peroxidase-1 accelerates the progression of atherosclerosis in apolipoprotein E-deficient mice, Arterioscler. Thromb. Vasc. Biol. 27 (2007) 850–857.
- [151] N. Weiss, Y.Y. Zhang, S. Heydrick, C. Bierl, J. Loscalzo, Overexpression of cellular glutathione peroxidase rescues homocyst(e)ine-induced endothelial dysfunction, Proc. Natl. Acad. Sci. U. S. A. 98 (2001) 12503–12508.

- [152] G. Wu, Y.Z. Fang, S. Yang, J.R. Lupton, N.D. Turner, Glutathione metabolism and its implications for health, J. Nutr. 134 (2004) 489–492.
- [153] H. Yamawaki, J. Haendeler, B.C. Berk, Thioredoxin: a key regulator of cardiovascular homeostasis, Circ. Res. 93 (2003) 1029–1033.
- [154] M.A. Perrella, S.F. Yet, Role of heme oxygenase-1 in cardiovascular function, Curr. Pharm. Des. 9 (2003) 2479–2487.
- [155] R. Stocker, M.A. Perrella, Heme oxygenase-1: a novel drug target for atherosclerotic diseases? Circulation 114 (2006) 2178–2189.
- [156] T. Morita, Heme oxygenase and atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 25 (2005) 1786–1795.
- [157] K.A. Hoekstra, D.V. Godin, K.M. Cheng, Protective role of heme oxygenase in the blood vessel wall during atherogenesis, Biochem. Cell Biol. 82 (2004) 351–359.
- [158] M. Aviram, M. Rosenblat, C.L. Bisgaier, R.S. Newton, S.L. Primo-Parmo, B.N. La Du, Paraoxonase inhibits high-density lipoprotein oxidation and preserves its functions. A possible peroxidative role for paraoxonase, J. Clin. Invest. 101 (1998) 1581–1590.
- [159] D.M. Shih, L. Gu, Y.R. Xia, M. Navab, W.F. Li, S. Hama, L.W. Castellani, C.E. Furlong, L.G. Costa, A.M. Fogelman, A.J. Lusis, Mice lacking serum paraoxonase are susceptible to organophosphate toxicity and atherosclerosis, Nature 394 (1998) 284–287.
- [160] B. Mackness, P. Durrington, P. McElduff, J. Yarnell, N. Azam, M. Watt, M. Mackness, Low paraoxonase activity predicts coronary events in the Caerphilly Prospective Study, Circulation 107 (2003) 2775–2779.
- [161] O. Rozenberg, D.M. Shih, M. Aviram, Paraoxonase 1 (PON1) attenuates macrophage oxidative status: studies in PON1 transfected cells and in PON1 transgenic mice, Atherosclerosis 181 (2005) 9–18.
- [162] S. Horke, I. Witte, P. Wilgenbus, M. Kruger, D. Strand, U. Forstermann, Paraoxonase-2 reduces oxidative stress in vascular cells and decreases endoplasmic reticulum stress-induced caspase activation, Circulation 115 (2007) 2055–2064.
- [163] C.J. Ng, N. Bourquard, V. Grijalva, S. Hama, D.M. Shih, M. Navab, A.M. Fogelman, A.J. Lusis, S. Young, S.T. Reddy, Paraoxonase-2 deficiency aggravates atherosclerosis in mice despite lower apolipoprotein-B-containing lipoproteins: antiatherogenic role for paraoxonase-2, J. Biol. Chem. 281 (2006) 29491–29500.
- [164] Y. Ikeda, T. Suehiro, M. Inoue, Y. Nakauchi, T. Morita, K. Arii, H. Ito, Y. Kumon, K. Hashimoto, Serum paraoxonase activity and its relationship to diabetic complications in patients with noninsulin-dependent diabetes mellitus, Metabolism 47 (1998) 598–602.
- [165] C.M. Sena, P. Matafome, T. Louro, E. Nunes, R. Fernandes, R.M. Seiça, Metformin restores endothelial function in aorta of diabetic rats Br, J. Pharmacol. 163 (2011) 424–437.
- [166] D. Nathanson, T. Nyström, Hypoglycemic pharmacological treatment of type 2 diabetes: targeting the endothelium, Mol. Cell. Endocrinol. 297 (2009) 112–126.
- [167] J. Xu, M.H. Zou, Molecular insights and therapeutic targets for diabetic endothelial dysfunction, Circulation 120 (2009) 1266–1286.
- [168] C.M. Sena, R.M. Seiça, Oxidative stress and endothelial dysfunction: novel therapeutic interventions, in: M. Stefek (Ed.), Advances in Molecular Mechanisms and Pharmacology of Diabetic Complications, Transworld Research Network, India, 2010, pp. 153–174.
- [169] U. Campia, M. Tesauro, C. Cardillo, Human obesity and endothelium-dependent responsiveness, Br. J. Pharmacol. 165 (2012) 561–573.
- [170] G. Mancia, G. De Backer, A. Dominiczak, R. Cifkova, R. Fagard, G. Germano, G. Grassi, A.M. Heagerty, S.E. Kjeldsen, S. Laurent, K. Narkiewicz, L. Ruilope, A. Rynkiewicz, R.E. Schmieder, H.A. Boudier, A. Zanchetti, A. Vahanian, J. Camm, R. De Caterina, V. Dean, K. Dickstein, G. Filippatos, C. Funck-Brentano, I. Hellemans, S.D. Kristensen, K. McGregor, U. Sechtem, S. Silber, M. Tendera, P. Widimsky, J.L. Zamorano, S. Erdine, W. Kiowski, E. Agabiti-Rosei, E. Ambrosioni, L.H. Lindholm, M. Viigimaa, S. Adamopoulos, E. Agabiti-Rosei, E. Ambrosioni, V. Bertomeu, D. Clement, S. Erdine, C. Farsang, D. Gaita, G. Lip, J.M. Mallion, A.J. Manolis, P.M. Nilsson, E. O'Brien, P. Ponikowski, J. Redon, F. Ruschitzka, J. Tamargo, P. van Zwieten, B. Waeber, B. Williams, Guidelines for the management of arterial hypertension: the task force for the management of arterial hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC), J. Hypertens. 25 (2007) 1105–1187.
- [171] R. Kietadisorn, R.P. Juni, A.L. Moens, Therapeutic possibilities uncoupling: new insights into its pathogenesis and tackling endothelial dysfunction by modulating NOS uncoupling: new insights into its pathogenesis and therapeutic possibilities, Am. J. Physiol. Endocrinol. Metab. 302 (2012) E481–E495.
- [172] Y. Kureishi, Z. Luo, I. Shiojima, A. Bialik, D. Fulton, D.J. Lefer, W.C. Sessa, K. Walsh, The HMG-CoA reductase inhibitor simvastatin activates the protein kinase Akt and promotes angiogenesis in normocholesterolemic animals, Nat. Med. 6 (2000) 1004–1010.
- [173] U. Dirnagl, C. ladecola, M.A. Moskowitz, Pathobiology of ischaemic stroke: an integrated view, Trends Neurosci. 22 (1999) 391–397.
- [174] J.P. Stasch, K. Dembowsky, E. Perzborn, E. Stahl, M. Schramm, Cardiovascular actions of a novel NO-independent guanylyl cyclase stimulator, BAY 41-8543: in vivo studies, Br. J. Pharmacol. 135 (2002) 344–355.
- [175] M. Hoenicka, C. Schmid, Cardiovascular effects of modulators of soluble guanylylcyclase activity, Cardiovasc. Hematol. Agents Med. Chem. 6 (2008) 287–301.
- [176] P.R. Evora, P.M. Evora, A.C. Celotto, A.J. Rodrigues, E.E. Joviliano, Cardiovascular therapeutics targets on the NO-sGC-cGMP signaling pathway: a critical overview, Curr. Drug Targets 13 (2012) 1207–1214.
- [177] J.B. De Haan, M.E. Cooper, Targeted antioxidant therapies in hyperglycemia-mediated endothelial dysfunction, Front. Biosci. 3 (2011) 709–729.
- [178] E. Hood, E. Simone, P. Wattamwar, T. Dziubla, V. Muzykantov, Nanocarriers for vascular delivery of antioxidants, Nanomedicine (Lond.) 6 (2011) 1257–1272.

- [179] T. Van-Assche, V. Huygelen, M.J. Crabtree, C. Antoniades, Gene therapy targeting inflammation in atherosclerosis, Curr. Pharm. Des. 17 (2011) 4210–4223.
- [180] M. Rosenblat, T. Hayek, M. Aviram, Anti-oxidative effects of pomegranate juice (PJ) consumption by diabetic patients on serum and on macrophages, Atherosclerosis 187 (2006) 363–371.
- [181] M. Aviram, B. Fuhrman, Wine flavonoids protect against LDL oxidation and atherosclerosis, Ann. N. Y. Acad. Sci. 957 (2002) 146–161.
- [182] A.J. Cayatte, A. Rupin, J. Oliver-Krasinski, K. Maitland, P. Sansilvestri-Morel, M.F. Boussard, M. Wierzbicki, T.J. Verbeuren, R.A. Cohen, S17834, a new inhibitor of cell adhesion and atherosclerosis that targets NADPH oxidase, Arterioscler. Thromb. Vasc. Biol. 21 (2001) 1577–1584.
- [183] S. Wind, K. Beuerlein, T. Eucker, H. Müller, P. Scheurer, M.E. Armitage, H. Ho, H.H. Schmidt, K. Wingler, Comparative pharmacology of chemically distinct NADPH oxidase inhibitors, Br. J. Pharmacol. 161 (2010) 885–898.
- [184] A.R. Weseler, A. Bast, Oxidative stress and vascular function: implications for pharmacologic treatments, Curr. Hypertens. Rep. 12 (2010) 154–161.
- [185] A. Goldin, J.A. Beckman, A.M. Schmidt, M.A. Creager, Vascular injury advanced glycation end products: sparking the development of diabetic vascular injury, Circulation 114 (2006) 597–605.
- [186] S.Y. Goh, M.E. Cooper, The role of advanced glycation end products in progression and complications of diabetes, J. Clin. Endocrinol. Metab. 93 (2008) 1143–1152.
- [187] H.N. Lee, Y.J. Surh, Therapeutic potential of resolvins in the prevention and treatment of inflammatory disorders, Biochem. Pharmacol. 84 (2012) 1340–1350.
- [188] Y. Xu, S. Wang, L. Feng, Q. Zhu, P. Xiang, B. He, Blockade of PKC-beta protects HUVEC from advanced glycation end products induced inflammation, Int. Immunopharmacol. 10 (2010) 1552–1559.
- [189] P. Pacher, C. Szabo, Role of poly(ADP-ribose) polymerase 1 (PARP-1) in cardiovascular diseases: the therapeutic potential of PARP inhibitors, Cardiovasc. Drug Rev. 25 (2007) 235–260.
- [190] M. Bucci, F. Roviezzo, V. Brancaleone, M.I. Lin, A. Di Lorenzo, C. Cicala, A. Pinto, W.C. Sessa, S. Farneti, S. Fiorucci, G. Cirino, Diabetic mouse angiopathy is linked to progressive sympathetic receptor deletion coupled to an enhanced caveolin-1 expression, Arterioscler. Thromb. Vasc. Biol. 24 (2004) 721–726.
- [191] F. Garcia Soriano, L. Virag, P. Jagtap, E. Szabo, J.G. Mabley, L. Liaudet, A. Marton, D.G. Hoyt, K.G. Murthy, A.L. Salzman, G.J. Southan, C. Szabo, Diabetic endothelial dysfunction: the role of poly(ADP-ribose) polymerase activation, Nat. Med. 7 (2001) 108–113.
- [192] L. Zheng, C. Szabo, T.S. Kern, Poly(ADP-ribose) polymerase is involved in the development of diabetic retinopathy via regulation of nuclear factor-kappaB, Diabetes 53 (2004) 2960–2967.
- [193] F.A. English, F.P. McCarthy, I.J. Andersson, J.L. Stanley, S.T. Davidge, P.N. Baker, S.K. Walsh, L.C. Kenny, Administration of the PARP inhibitor Pj34 ameliorates the impaired vascular function associated with eNOS(-/-) mice, Reprod. Sci. 19 (2012) 806–813.
- [194] C.P. Hans, Y. Feng, A.S. Naura, M. Zerfaoui, B.M. Rezk, H. Xia, A.D. Kaye, K. Matrougui, E. Lazartigues, A.H. Boulares, Protective effects of PARP-1 knockout on dyslipidemia-induced autonomic and vascular dysfunction in ApoE<sup>-/-</sup> mice: effects on eNOS and oxidative stress, PLoS One 4 (2009) e7430.
- [195] L. Packer, K. Kraemer, G. Rimbach, Molecular aspects of lipoic acid in the prevention of diabetes complications, Nutrition 17 (2001) 888–895.
- [196] S. Sola, M.Q. Mir, F.A. Cheema, N. Khan-Merchant, R.G. Menon, S. Parthasarathy, B.V. Khan, Irbesartan and lipoic acid improve endothelial function and reduce markers of inflammation in the metabolic syndrome: results of the Irbesartan and Lipoic Acid in Endothelial Dysfunction (ISLAND) study, Circulation 111 (2005) 343–348.
- [197] I.C. Arts, P.C. Hollman, Polyphenols and disease risk in epidemiologic studies, Am. J. Clin. Nutr. 81 (2005) 317S–325S.
- [198] J.A. Vita, Polyphenols and cardiovascular disease: effects on endothelial and platelet function, Am. J. Clin. Nutr. 81 (2005) 292S–297S.
- [199] V. Habauzit, C. Morand, Evidence for a protective effect of polyphenols-containing foods on cardiovascular health: an update for clinicians, Ther. Adv. Chronic Dis. 3 (2012) 87–106.
- [200] H. Li, U. Forstermann, Resveratrol: a multifunctional compound improving endothelial function. Editorial to: "resveratrol supplementation gender independently improves endothelial reactivity and suppresses superoxide production in healthy rats", by S. Soylemez et al. Cardiovasc. Drugs Ther. 23 (2009) 425–429.
- [201] K. Steinkamp-Fenske, L. Bollinger, N. Voller, H. Xu, Y. Yao, R. Bauer, U. Forstermann, H. Li, Ursolic acid from the Chinese herb danshen (*Salvia miltiorrhiza* L.) upregulates eNOS and downregulates Nox4 expression in human endothelial cells, Atherosclerosis 195 (2007) e104–e111.
- [202] K. Steinkamp-Fenske, L. Bollinger, H. Xu, Y. Yao, S. Horke, U. Forstermann, H. Li, Reciprocal regulation of endothelial nitric-oxide synthase and NADPH oxidase by betulinic acid in human endothelial cells, J. Pharmacol. Exp. Ther. 322 (2007) 836–842.
- [203] T. Wallerath, D. Poleo, H. Li, U. Forstermann, Red wine increases the expression of human endothelial nitric oxide synthase: a mechanism that may contribute to its beneficial cardiovascular effects, J. Am. Coll. Cardiol. 41 (2003) 471–478.
- [204] Z. Ungvari, Z. Orosz, A. Rivera, N. Labinskyy, Z. Xiangmin, S. Olson, A. Podlutsky, A. Csiszar, Resveratrol increases vascular oxidative stress resistance, Am. J. Physiol. Heart Circ. Physiol. 292 (2007) H2417–H2424.
- [205] X.B. Wang, J. Huang, J.G. Zou, É.B. Su, Q.J. Shan, Z.J. Yang, K.J. Cao, Effects of resveratrol on number and activity of endothelial progenitor cells from human peripheral blood, Clin. Exp. Pharmacol. Physiol. 34 (2007) 1109–1115.
- [206] R.H. Wong, P.R. Howe, J.D. Buckley, A.M. Coates, I. Kunz, N.M. Berry, Acute resveratrol supplementation improves flow-mediated dilatation in overweight/obese

individuals with mildly elevated blood pressure, Nutr. Metab. Cardiovasc. Dis. 21 (2011) 851–856.

- [207] X. Han, T. Shen, H. Hongxiang Lou, Dietary polyphenols and their biological significance, Int. J. Mol. Sci. 8 (2007) 950–988.
- [208] W.P. Chen, M.J. Su, L.M. Hung, In vitro electrophysiological mechanisms for antiarrhythmic efficacy of resveratrol, a red wine antioxidant, Eur. J. Pharmacol. 554 (2007) 196–204.
- [209] C.J. Ng, D.M. Shih, S.Y. Hama, N. Villa, M. Navab, S.T. Reddy, The paraoxonase gene family and atherosclerosis, Free Radic. Biol. Med. 38 (2005) 153–163.
- [210] M. Aviram, E. Hardak, J. Vaya, S. Mahmood, S. Milo, A. Hoffman, S. Billicke, D. Draganov, M. Rosenblat, Human serum paraoxonase (PON1) Q and R selectively decrease lipid peroxides in human coronary and carotid atherosclerotic lesions: PON1 esterase and peroxidase like activity, Circulation 101 (2000) 2510–2517.
- [211] T.M. Van Himbergen, L.J.H. Van Tits, M. Roest, A.F.H. Stalenhoef, The story of PO N1: how an organophosphate hydrolyzing enzyme is becoming a player in cardiovascular medicine, Neth. J. Med. 64 (2006) 34–38.
- [212] E.B. Schmidt, W. Koenig, N. Khuseyinova, J.H. Christensen, Lipoprotein-associated phospholipase A2 concentrations in plasma are associated with the extent of coronary artery disease and correlate to adipose tissue levels of marine n-3 fatty acids, Atherosclerosis 196 (2008) 420–424.
- [213] E.M. Hjerkinn, M. Abdelnoor, L. Breivik, L. Bergengen, I. Ellingsen, I. Seljeflot, O. Aase, T. Ole Klemsdal, I. Hjermann, H. Arnesen, Effect of diet or very long chain omega-3 fatty acids on progression of atherosclerosis, evaluated by carotid plaques, intima-media thickness and by pulse wave propagation in elderly men with hypercholesterolaemia, Eur. J. Cardiovasc. Prev. Rehabil. 13 (2006) 325–333.
- [214] C. Mindrescu, R.P. Gupta, E.V. Hermance, M.C. DeVoe, V.R. Soma, J.T. Coppola, C.S. Staniloae, Omega-3 fatty acids plus rosuvastatin improves endothelial function in South Asians with dyslipidemia, Vasc. Health Risk Manag. 4 (2008) 1439–1447.
- [215] V. Schiano, E. Laurenzano, G. Brevetti, J.I. De Maio, S. Lanero, F. Scopacasa, M. Chiariello, Omega-3 polyunsaturated fatty acid in peripheral arterial disease: effect on lipid pattern, disease severity, inflammation profile, and endothelial function, Clin. Nutr. 27 (2008) 241–247.
- [216] M. Cloarec, P. Caillard, J.C. Provost, J.M. Dever, Y. Elbeze, N. Zamaria, GliSODin, a vegetal sod with gliadin, as preventative agent vs. atherosclerosis, as confirmed with carotid ultrasound-B imaging, Eur. Ann. Allergy Clin. Immunol. 39 (2007) 45–50.
- [217] P. Bernatchez, A. Sharma, P.M. Bauer, E. Marin, W.C. Sessa, A noninhibitory mutant of the caveolin-1 scaffolding domain enhances eNOS-derived NO synthesis and vasodilation in mice, J. Clin. Invest. 121 (2011) 3747–3755.
- [218] K.M. Channon, Tetrahydrobiopterin: a vascular redox target to improve endothelial function, Curr. Vasc. Pharmacol. 10 (2012) 705–708.
- [219] B. Viollet, F. Andreelli, AMP-activated protein kinase and metabolic control, Handb. Exp. Pharmacol. 203 (2011) 303–330.
- [220] C. Szabó, J.G. Mabley, S.M. Moeller, R. Shimanovich, P. Pacher, L. Virag, F.G. Soriano, J.H. Van Duzer, W. Williams, A.L. Salzman, J.T. Groves, FP 15, a novel potent peroxynitrite decomposition catalyst: in vitro cytoprotective actions and protection against diabetes mellitus and diabetic cardiovascular complications, Mol. Med. 8 (2002) 571–580.
- [221] T. Radovits, L. Seres, D. Gero, L.N. Lin, CJ. Beller, S.H. Chen, J. Zotkina, I. Berger, J.T. Groves, C. Szabó, G. Szabó, The peroxynitrite decomposition catalyst FP15 improves ageing-associated cardiac and vascular dysfunction, Mech. Ageing Dev. 128 (2007) 173–181.
- [222] S. Wolfrum, A. Dendorfer, Y. Rikitake, T.J. Stalker, Y. Gong, R. Scalia, P. Dominiak, J.K. Liao, Inhibition of Rho-kinase leads to rapid activation of phosphatidylinositol 3-kinase/protein kinase Akt and cardiovascular protection, Arterioscler. Thromb. Vasc. Biol. 24 (2004) 1842–1847.
- [223] R. Arita, Y. Hata, S. Nakao, T. Kita, M. Miura, S. Kawahara, S. Zandi, L. Almulki, F. Tayyari, H. Shimokawa, A. Hafezi-Moghadam, T. Ishibashi, Rho kinase inhibition by fasudil ameliorates diabetes-induced microvascular damage, Diabetes 58 (2009) 215–226.
- [224] G.R. Drummond, S. Selemidis, K.K. Griendling, C.G. Sobey, Combating oxidative stress in vascular disease: NADPH oxidases as therapeutic targets, Nat. Rev. Drug Discov. 10 (2011) 453–471.
- [225] J.D. Symons, Opportunity "Nox": a novel approach to preventing endothelial dysfunction in the context of insulin resistance, Diabetes 62 (2013) 1818–1820.
- [226] N. Lanati, E. Emanuele, N. Brondino, D. Geroldi, Soluble RAGE-modulating drugs: state-of-the-art and future perspectives for targeting vascular inflammation, Curr. Vasc. Pharmacol. 8 (2010) 86–92.
- [227] H. Duplain, R. Burcelin, C. Sartori, S. Cook, M. Egli, M. Lepori, P. Vollenweider, T. Pedrazzini, P. Nicod, B. Thorens, U. Scherrer, Insulin resistance, hyperlipidemia, and hypertension in mice lacking endothelial nitric oxide synthase, Circulation 104 (2001) 342–345.
- [228] S. Kashiwagi, D.N. Atochin, Q. Li, M. Schleicher, T. Pong, W.C. Sessa, P.L. Huang, eNOS phosphorylation on serine 1176 affects insulin sensitivity and adiposity, Biochem. Biophys. Res. Commun. 431 (2013) 284–290.
- [229] O. Rozenberg, M. Shiner, M. Aviram, T. Hayek, Paraoxonase1 (PON1) attenuates diabetes development in mice through its antioxidative properties, Free Radic. Biol. Med. 44 (2008) 1951–1959.
- [230] M. Rosenblat, M. Aviram, Paraoxonases role in the prevention of cardiovascular diseases, Biofactors 35 (2009) 98–104.