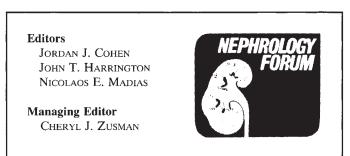
## NEPHROLOGY FORUM

# Extracellular calcium-sensing receptor: Implications for calcium and magnesium handling in the kidney

Principal discussant: STEVEN C. HEBERT

Brigham & Women's Hospital, and Harvard Medical School, Boston, Massachusetts, USA



Tufts University School of Medicine

#### **Case presentation**

A 48-year-old man presented with recurrent nephrolithiasis. He had passed two stones over the past seven years; a stone passed three years ago required ureterolithotomy. At that time, the serum calcium was 10.6 mg/dl (normal, 8.8–10.6 mg/dl) and the serum phosphorus was 2.9 mg/dl (normal, 2.5–4.5 mg/dl). Crystallographic analysis of that stone showed that it contained a predominance of calcium oxalate with a small amount of calcium phosphate. Since then the patient has generally felt well while taking no medications. He had noticed that he had been drinking more water over the previous few months and had had occasional nocturia. He had no family history of nephrolithiasis.

On physical examination he was alert and in no distress. Weight was 75 kg; blood pressure, 145/92 mm Hg; the rest of the examination was unremarkable. Serum chemistries were: calcium, 11.1 mg/dl; phosphorus, 3.2 mg/dl; alkaline phosphatase, 75 U/liter (normal, 15–95 U/liter); BUN, 15 mg/dl; creatinine, 1.0 mg/dl; uric acid, 7.5 mg/dl (normal, 2.6–8.1 mg/dl); sodium, 139 mEq/liter; potassium, 4.1 mEq/liter; chloride, 102 mEq/liter; and bicarbonate, 25 mEq/liter. A 24-hour urine collection revealed a total volume of 2750 ml; creatinine clearance, 80 ml/min; calcium, 350 mg/day; and uric acid, 600 mg/day (normal, < 750 mg/day). The scrum intact PTH was 70 pg/ml (normal, 10–55 pg/ml), 25-hydroxyvitamin D was normal, and serum 1,25(OH)<sub>2</sub>D was 65 pg/ml (normal, 10–60 pg/ml). Maximal urinary concentration after overnight water deprivation was 550 mOsm/kg H<sub>2</sub>O. An adenoma weighing 2 g was removed from the right parathyroid gland.

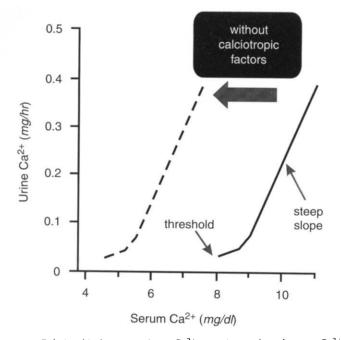
#### Discussion

DR. STEVEN C. HEBERT (Director, Laboratory of Molecular Physiology and Biophysics, Renal Division, Department of Medicine,

The Nephrology Forum is funded in part by grants from Amgen, Incorporated; Merck & Co., Incorporated; Marion Merrell Dow, Incorporated; Dialysis Clinic, Incorporated; and R & D Laboratories. Brigham & Women's Hospital, and Professor of Medicine, Harvard Medical School, Boston, Massachusetts, USA): In addition to providing a new target for the development of pharmaceutical agents that could be effective in altering PTH secretion, the molecular cloning of the extracellular Ca2+-sensing G-proteincoupled receptor from parathyroid, kidney, and brain (variously referred to as BoPCaR, RaKCaR, HuPCaR, HuKCaSR, and CaSR), which will be abbreviated in this discussion to CaR [1-6], has significantly expanded our understanding of calcium and magnesium metabolism. New insights into the roles played by this receptor in several organs, including kidney and brain, soon will be forthcoming. Some of this new information may well suggest hitherto unsuspected roles for the CaR and extracellular Ca2+ in the various functions of these organ systems. In this discussion, I will focus on the proposed function of the CaR in Ca<sup>2+</sup>, Mg<sup>2+</sup>, and water metabolism. Today's patient, who had primary hyperparathyroidism, illustrates some of the effects that increases in extracellular calcium can have on renal function [see Refs. 7, 8 for reviews].

The kidney provides the major route for mineral ion excretion from the body by adjusting the tubular reabsorption of divalent cations from the glomerular filtrate. The kidney therefore plays a key role in Ca<sup>2+</sup> and Mg<sup>2+</sup> homeostasis. The cellular mechanisms mediating mineral ion transport across nephron segments from proximal tubule to collecting duct have been reviewed elsewhere [9, 10] and will not be covered here in detail. Traditional views of renal mineral ion handling have focused on the important roles played by the calciotropic hormones, PTH and calcitonin, as well as vitamin D [9, 11-16]. As discussed by Kurokawa in a previous Nephrology Forum a decade or so ago [16], urinary calcium excretion  $[U_{Ca}]$  increases steeply when circulating Ca<sup>2+</sup> concentrations  $[P_{Ca}]$  rise beyond a certain threshold (Fig. 1; see Ref. 16 for a recent review). The inverse sigmoidal relationship between increasing extracellular Ca<sup>2+</sup> and PTH secretion from parathyroid cells is also quite steep; this relationship suggests the possible cooperative interactions of three or more Ca<sup>2+</sup> ions with the cation-sensing mechanism [8]. Both PTH and vitamin D are important modulators of the relationship between  $U_{Ca}$  and  $P_{Ca}$ , and the absence of either calciotropic factor (or both) significantly shifts the threshold (or "set point") for the curve to the left, such that urinary Ca<sup>2+</sup> loss is observed at lower circulating Ca<sup>2+</sup> concentrations (Fig. 1) [11]. The steepness of the relationship between  $U_{Ca}$  and  $P_{Ca}$  is not lost, however, even when both PTH and vitamin D are absent. This relationship indicates that one or more additional factors contribute to renal Ca<sup>2+</sup> excretion (Fig.

<sup>© 1996</sup> by the International Society of Nephrology



**Fig. 1.** Relationship between urinary  $Ca^{2+}$  excretion and total serum  $Ca^{2+}$  concentration. Urinary  $Ca^{2+}$  increases steeply in proportion to serum  $Ca^{2+}$  beyond a threshold concentration. The curves in the presence (solid line) or absence (dashed line) of the calciotropic factors vitamin D and PTH are shown. Note that absence of either calciotropic factor shifts the set point to the left without affecting the slope of the curves. (Adapted from Ref. 16.)

1). As I will discuss in this Forum, recent evidence suggests that extracellular  $Ca^{2+}$  itself, interacting with the newly cloned CaR in the kidney, provides a major part of this regulatory function.

The CaR plays crucial roles in regulating renal divalent mineral transport processes by both direct and indirect mechanisms. Parathyroid cells recognize remarkably small perturbations in the circulating concentration of Ca<sup>2+</sup> (or Mg<sup>2+</sup>) and then respond by altering the secretion of PTH. In fact, most of the "sensing" of extracellular Ca<sup>2+</sup> in parathyroid cells occurs over changes in free Ca<sup>2+</sup> concentration of approximately 0.25 mM; that is, PTH secretion versus  $[Ca^{2+}]_o$  is quite steep, with a  $K_{1/2} \approx 1.2$  mM; [8]. Recent molecular and genetic evidence has demonstrated that the cloned CaR, which is expressed on the surface of parathyroid cells, provides the principal mechanism for extracellular Ca<sup>2+</sup> "sensing" by the parathyroid gland [reviewed in Ref. 17]. In turn, PTH modulates the renal reabsorption of  $Ca^{2+}$  and  $Mg^{2+}$  as well as  $PO_4^{3-}$ . Thus the CaR, by providing a sensor for extracellular Ca2+-mediated modulation of calciotropic hormones, indirectly regulates renal divalent mineral handling. Moreover the kidney, like the parathyroid, is able to respond directly (that is, independently of changes in calciotropic hormones) to alterations in extracellular  $Ca^{2+}$  (or  $Mg^{2+}$ ) with the resultant modulation of mineral ion transport [see Refs. 8, 18-21 for reviews]. The cloning of the CaR from rat [2] and human [5] kidney and the expression of the CaR in renal epithelial cells [22] is consistent with a mechanism whereby extracellular calcium participates directly in the regulation of its own reabsorption through local, receptormediated actions of  $Ca^{2+}$  (and/or  $Mg^{2+}$ ) on the kidney. Evidence supporting this view will be described in the following sections.

The homeostatic adjustments in urinary excretion of mineral

ions provided by calciotropic factors (mainly PTH and vitamin D) and the CaR are not without potential consequences on renal function. With increased loads of calcium (for example, from enhanced bone turnover or absorption from the intestinal tract, or from abnormalities of mineral ion reabsorption along the nephron), urinary calcium excretion can increase dramatically (Fig. 1). The continued formation of a concentrated urine during periods of increased urinary  $Ca^{2+}$  or  $Mg^{2+}$  loss could present a problem, because mineral ions can reach supersaturation levels in the terminal collecting duct; this supersaturation in turn increases the risk of nephrolithiasis, nephrocalcinosis, or both. Recently we have suggested that a "trade-off" of water conservation for Ca<sup>2+</sup> or  $Mg^{2+}$  loss operates to minimize the risk of stone formation under normal circumstances, especially during periods of increased mineral ion excretion [23]. The renal CaR appears to provide the crucial "sensing" mechanism in the distal nephron and collecting duct for integrating and balancing water and divalent mineral loss. Direct interactions of extracellular Ca<sup>2+</sup> with the renal CaR could explain in large part the disordered water metabolism (that is, nephrogenic diabetes insipidus) observed during pathologic states of hypercalcemia (for example, primary hyperparathyroidism or certain malignancies) [24, 25].

## The extracellular Ca<sup>2+</sup>-sensing receptor

Extracellular Ca<sup>2+</sup> modulates both PTH secretion and renal tubular divalent mineral and water transport processes by interacting with a specific receptor, the CaR. A wealth of indirect evidence has suggested the existence of a Ca<sup>2+</sup>-responsive, receptor-like mechanism in parathyroid cells; for instance, raising extracellular Ca2+ activated a number of second messenger systems in a fashion similar to that for other G-protein-coupled receptors, for example, activation of phospholipase C (PLC) with consequent accumulation of inositol 1,4,5-trisphosphate (IP<sub>3</sub>) [26] and release of  $Ca^{2+}$  from intracellular stores [27] [see Refs. 7, 8, 18 for reviews]. Moreover, second messenger systems in several nephron segments also are modulated by changes in extracellular  $Ca^{2+}$  [28–32]. The ability to express exogenous receptors in oocytes of the frog Xenopus laevis, and then to detect function of phosphoinositide (PI)-coupled receptors through electrophysiologic measurement of intracellular Ca2+-activated chloride channels, enabled Brown and coworkers to clone the complementary DNA (cDNA) encoding the Ca<sup>2+</sup>-sensing receptor from bovine parathyroid gland (BoPCaR) [1]. Subsequently, this receptor also was cloned from human parathyroid [3], rat [2] and human [5] kidney, and rat brain [4]. When these receptors were expressed in oocytes (by injecting the oocytes with synthetic mRNA transcribed from the cloned cDNAs), they behaved like the native  $Ca^{2+}$ -sensing receptor of the parathyroid gland [1–3]: the cloned CaR was activated by the same di- and trivalent cations and even polycations (for example, neomycin) as the native receptor [33-36]. This CaR represents the first example of a G-protein-coupled, cell-surface receptor in mammalian species that recognizes an inorganic ion rather than a molecule as its ligand [37]. The availability of these cloned genes and of antibodies against the receptor (produced from knowledge of the deduced amino acid sequences of the receptor proteins) has permitted studies of receptor localization, function, and regulation as well as identification of mutations of this receptor that produce inherited disorders of calcium metabolism. Much of this work is in progress, and over the next few years we can expect additional

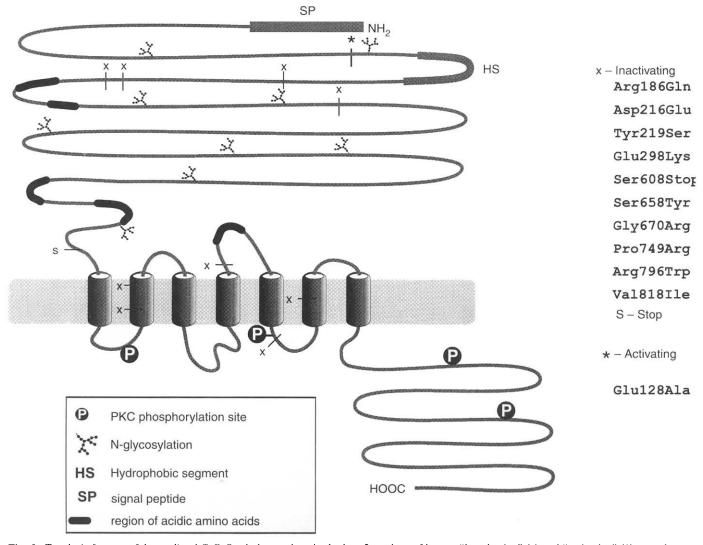


Fig. 2. Topologic features of the predicted CaR. Symbols are given in the key. Locations of known "inactivating" (x) and "activating" (\*) mutations are indicated.

information, as well as probably some surprises, on the roles of this receptor in endocrine glands, kidney, intestine, brain, and possibly other tissues and cells [see Refs. 23, 38].

As expected, the deduced amino acid sequence of the CaR shows the classic seven-transmembrane-spanning helical domain thus far found on all G-protein-coupled receptors (GPRs; Fig. 2) [39, 40]. The CaR is a novel receptor, however, in that it has a significant (but low) amino acid sequence similarity only with the G-protein-coupled (metabotropic) glutamate receptors, mGluRs, expressed in the central nervous system [41]. Conklin and Bourne have suggested that the extracellular ligand-binding domains of the CaR and the mGluRs have an overall structural organization [37] similar to that of bacterial periplasmic nutrient-binding proteins [42, 43]. These bacterial cell-surface proteins bind a variety of extracellular solutes (thus enabling cellular uptake), including organic nutrients as well as inorganic ions such as phosphate and nickel [43]. Thus the extracellular Ca<sup>2+</sup>-sensing receptor might have evolved from an ancient family of cell-surface

proteins binding essential extracellular solutes. This similarity also suggests the possibility of additional ion (solute)-sensing receptors.

The pharmacology of this CaR is unusual for a GPR in that it responds to its natural ligand, in this case  $Ca^{2+}$  (and  $Mg^{2+}$ ), only in the millimolar ion concentration range (recall that the pharmacologically relevant concentrations of other first messengers are usually in the picomolar to micromolar range). The former, however, is the physiologically relevant  $Ca^{2+}$  concentration range (0.75–2.0 mM free  $Ca^{2+}$ ) for extracellular fluid, as opposed to the  $Ca^{2+}$  cytosolic range  $[10^{-9}-10^{-6} \text{ M}]$ . In this regard, the large extracellular domain does not exhibit any of the known high-affinity  $Ca^{2+}$ -binding motifs. Instead it contains several regions rich in negatively charged (acidic) amino acids that probably mediate the low-affinity binding of cationic receptor agonists (for example,  $Ca^{2+}$ ,  $Mg^{2+}$ ,  $Gd^{3+}$ , neomycin) in a fashion similar to the acidic domains found on low-affinity calcium-binding proteins such as calsequestrin [44]. These negatively charged sites could

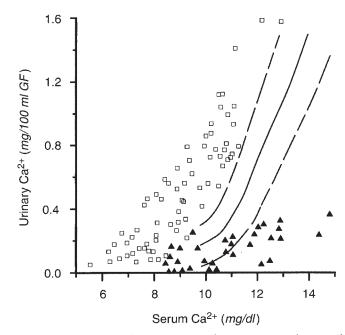
bind multiple  $Ca^{2+}$  ions on each receptor molecule, which might provide for cooperative cation interactions and the steep activity curve (Fig. 1).

### Inherited human diseases of extracellular Ca<sup>2+</sup> sensing

Elimination of a gene (and therefore the function of the protein encoded by the gene) through gene "knockouts," or overexpression of genes via production of transgenic animals, has provided powerful tools for defining the biologic role of a variety of genes. In certain cases, human genetic diseases have yielded examples of naturally occurring "knockouts" or "transgenics" by exhibiting mutations that inactivate or activate, respectively, the gene itself or the function provided by the encoded protein. Indeed, the clinical and physiologic relevance of the CaR has been established by the demonstration that mutations in it cause three inherited diseases of calcium metabolism. Two rare hypercalcemic disorders, familial hypocalciuric hypercalcemia (FHH) [45, 46] and neonatal severe hyperparathyroidism (NSHPT) result from inactivating mutations [47] when present in the heterozygous and homozygous ("knockout" equivalent) states, respectively [48]. One form of an autosomal dominant hypocalcemia [49] also results from a mutation in the CaR gene [50] and leads to expression of an overactivated receptor ("transgenic" equivalent).

The FHH gene had already been localized to the long arm of chromosome 3 in most [51, 52] but not all [51–54] families with FHH and NSHPT when the CaR was cloned. This CaR was an obvious candidate gene for FHH. Indeed, Pollak and coworkers demonstrated that the CaR gene mapped to the FHH disease locus and that point mutations (that is, single base changes) within this receptor gene were responsible for FHH in three families [47]. Other groups subsequently confirmed these results [54–56]. Mutations are scattered throughout the predicted protein (Fig. 2) [47, 55, 56] and apparently modify the structure and/or ligand-binding properties of the CaR. In one family, the mutation produces a stop codon that results in translation of a truncated and presumably biologically inactive receptor (Fig. 2) [56]. The genes involved in the families with the FHH phenotype not mapping to chromosome 3q are unknown [54].

In accord with the genetics, in-vivo and in-vitro studies have shown abnormal parathyroid and renal Ca2+ sensing in FHH and NSHPT. Parathyroid cells either show reduced sensitivity (FHH) or lack any PTH secretion responses (NSHPT) to increases in extracellular Ca<sup>2+</sup> [57-59]. Abnormal renal Ca<sup>2+</sup> sensing is suggested by the finding that, despite hypercalcemia, individuals with these disorders have reduced fractional renal clearance of Ca<sup>2+</sup> and Mg<sup>2+</sup> [45, 46, 60] and often exhibit frank hypocalciuria, that is, well under 100 mg/day. Moreover, individuals with FHH or NSHPT who have had parathyroidectomy, and thus have reduced or absent circulating PTH, continue to show markedly reduced renal calcium clearance, even during the intravenous infusion of sufficient calcium to induce frank hypercalcemia [60]. Figure 3 illustrates a comparison of urinary calcium  $(U_{Ca})$  as a function of serum calcium levels (P<sub>Ca</sub>) in hypoparathyroid "normal" and FHH individuals. Another feature of hypoparathyroid FHH shown in Figure 3 is the complete loss of the steep relationship between  $U_{Ca}$  and  $P_{Ca}$  (compare Figs. 1 and 3 and note that the curve is almost flat in the FHH individuals). These studies clearly demonstrate that the hypocalciuria observed in hypercalcemic FHH is PTH independent and secondary to an intrinsic alteration in renal handling of Ca<sup>2+</sup> somewhere along the nephron. More

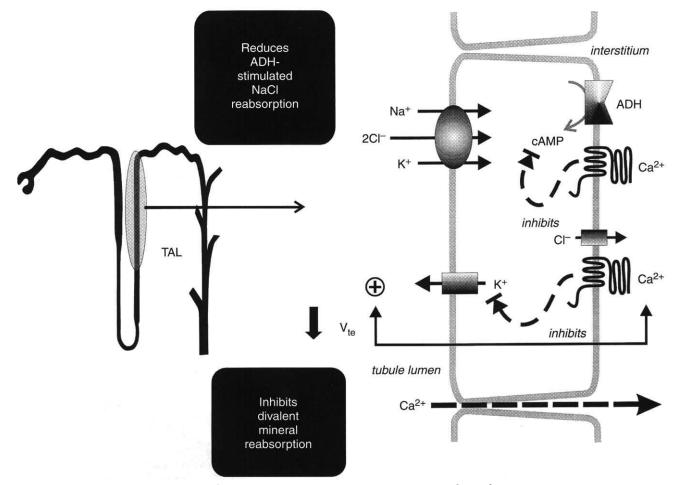


**Fig. 3.** Effect of calcium load on urinary calcium excretion in hypoparathyroid FHH patients ( $\blacktriangle$ ) and hypoparathyroid controls ( $\square$ ) during infusions of calcium. Urinary excretion of calcium is plotted as a function of total serum Ca<sup>2+</sup> concentration. In the subjects receiving calcium infusion, creatinine clearance did not change during the study. The solid line and dashed lines represent mean and 95% confidence intervals, respectively, from normoparathyroid control individuals. (Adapted from Refs. 60 and 90.)

than a decade ago, Attie et al suggested a defect in the ascending limb of the loop of Henle, because people with FHH exhibited an exaggerated calciuric response to the loop diuretic ethacrynic acid [60]. This response is consistent with the hypothesis that the CaR, which is decreased in FHH, influences  $Ca^{2+}$  reabsorption in the thick ascending limb (Fig. 4).

The renal clearance of  $Mg^{2+}$ , like that of  $Ca^{2+}$ , also is reduced in patients with FHH, and some individuals with NSHPT have overt hypermagnesemia even with a normal creatinine clearance [45, 46]. This finding suggests that the CaR in kidney also functions as a  $Mg^{2+}$  sensor. The apparent affinity of the CaR for  $Mg^{2+}$ , however, is too low for normal variations in the circulating  $Mg^{2+}$  concentration to influence this receptor [8]. Nevertheless, it is possible that the basolateral concentration of  $Mg^{2+}$ , to which the receptor might be exposed in the thick ascending limb (where  $Ca^{2+}$  and  $Mg^{2+}$  regulate their own reabsorption [10, 20, 61] and where both  $Ca^{2+}$  and  $Mg^{2+}$  are reabsorbed [10, 62] in the absence of water [63]), is actually higher than that in blood. In this regard, Attie and colleagues found that hypoparathyroid individuals with FHH and NSHPT exhibited a reduced  $Mg^{2+}$  [60].

Finally, unlike patients with hypercalcemia due to other causes, who commonly develop an ADH-resistant polyuria [25, 64–68], hypercalcemic individuals with FHH do not have polyuria and show normal maximal urinary concentrating ability after dehydration [24]. Their Ca<sup>2+</sup> "resistance," which results from a reduced number of normal Ca<sup>2+</sup>-sensing receptors, clearly diminishes the impact of hypercalcemia on loop of Henle or collecting duct functions, which are responsible for water handling.



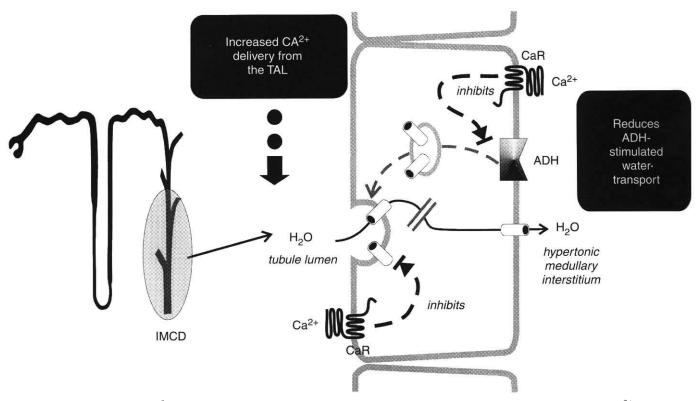
**Fig. 4.** Model for the role of the extracellular  $Ca^{2+}$ -sensing receptor in the regulation of NaCl and  $Ca^{2+}$  (Mg<sup>2+</sup>) reabsorption in the thick ascending limb (*TAL*). Increases in peritubular  $Ca^{2+}$  activate the CaR and reduce hormone-stimulated cyclic AMP production, which reduces net NaCl absorption and the lumen-positive transcriptical voltage,  $V_{tc}$ . Activation of the CaR also reduces activity of the apical K<sup>+</sup> channels and thereby reduces potassium recycling. The net effect of both CaR-mediated processes is a reduction in divalent mineral (Ca<sup>2+</sup> and Mg<sup>2+</sup>) transport (reabsorption) via the paracellular pathway.

#### CaR in the kidney

The Ca<sup>2+</sup>-sensing receptor has been localized within several segments of the rat nephron that subserve functions that are directly regulated by extracellular calcium. The receptor is most heavily expressed in the cortical thick ascending limb but also is present in the proximal tubule, medullary thick ascending limb, distal convoluted tubule, and along the entire collecting duct ([2], Riccardi D, unpublished observations). Previously described effects of extracellular  $Ca^{2+}$  (and  $Mg^{2+}$ ) on these segments of the nephron include the following: inhibition of the 1-hydroxylation of 25-hydroxyvitamin D in the proximal tubule independent of PTH [69, 70; see Refs. 7, 8 for discussions]; inhibition of NaCl transport in the thick ascending limb [68]; reduction in Ca<sup>2+</sup> and Mg<sup>2+</sup> reabsorption in the medullary thick ascending limb [10, 20, 21, 61, 71, 72]; pertussis-toxin-sensitive decrease in hormone-stimulated cAMP accumulation in the medullary thick ascending limb and cortical thick ascending limb [29, 31, 73]; inhibition of ADH's action in the collecting duct [28, 32, 74]; and the direct effect of  $Ca^{2+}$  on calbindin- $D_{28k}$  mRNA in the distal convoluted tubule (DCT) [75]. Studies are currently underway in several laboratories

to assess the role of the CaR as the mediator of the effects of extracellular  $Ca^{2+}$  on several aspects of renal function.

The model for the thick ascending limb shown in Figure 4 proposes that elevated levels of peritubular  $Ca^{2+}$  (or  $Mg^{2+}$ ) reduce NaCl reabsorption and the magnitude of the transepithelial voltage, and hence  $Ca^{2+}$  and  $Mg^{2+}$  reabsorption, via a Ca<sup>2+</sup>-sensing, receptor-dependent mechanism. In the thick ascending limb (TAL), the lumen positive potential, V<sub>te</sub>, is the driving force for Ca<sup>2+</sup> and Mg<sup>2+</sup> transport via the paracellular pathway [76-79]. In turn, V<sub>te</sub> is directly related to the magnitude of net NaCl absorption [63], which is stimulated by the integrated action of several hormones including ADH and the calciotropic hormones PTH and calcitonin [62, 63, 80]. Preliminary results from my own laboratory suggest that the CaR is expressed on basolateral membranes of TAL cells (unpublished observations). Thus peritubular Ca<sup>2+</sup>-mediated activation of the CaR could influence divalent mineral reabsorption either by reducing hormone-stimulated cyclic AMP production or by directly modulating the activity of the salt transporters themselves. In fact, increases in extracellular Ca2+ mediate pertussus-toxin-sensitive



**Fig. 5.** Role of the extracellular  $Ca^{2+}$ -sensing receptor in the regulation of water reabsorption in the terminal collecting duct. Peritubular  $Ca^{2+}$  can influence ADH-stimulated cAMP production. In addition, luminal  $Ca^{2+}$  reduces ADH's ability to stimulate the activity of aquaporin-2 (APQ2) water channels in the apical membrane. The net effect of these CaR-mediated processes is a reduction in ADH-stimulated water permeability of the collecting duct.

reductions in cyclic AMP generated by these hormones [29, 31, 73], and the CaR likely mediates this effect. Could extracellular Ca<sup>2+</sup> via the CaR also modulate any of the transporters involved in NaCl absorption by the TAL? This possibility was recently examined by Wang and coworkers [81, 82], who assessed the effects of extracellular Ca<sup>2+</sup> on the 70 pS inwardly rectifying K<sup>+</sup> channel, which provides the major apical membrane potassium permeability of TAL cells [83]. Recycling of K<sup>+</sup> via this channel not only supplies cation for Na<sup>+</sup>-K<sup>+</sup>-2Cl<sup>-</sup> cotransport but also is crucial to the generation of  $V_{te}$  [63, 83]. Wang et al showed that increasing extracellular Ca<sup>2+</sup> (as well as neomycin, a polyvalent organic cation that is a potent agonist of the CaR [1, 2, 8]) reversibly reduced K<sup>+</sup> channel activity via a P450 oxygenase metabolite [82]. This reduction in the apical K<sup>+</sup> conductance of the TAL would be expected to reduce both NaCl and  $Ca^{2+}$  (or Mg<sup>2+</sup>) reabsorption and markedly increase urinary Ca<sup>2+</sup> excretion similar to that which occurs with the administration of furosemide [84, 85].

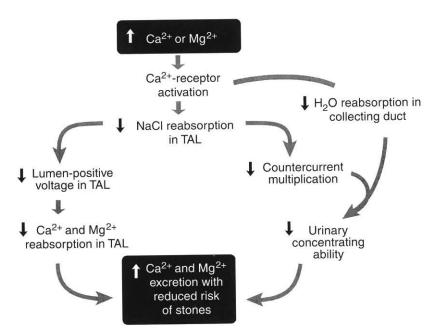
It is also possible that extracellular  $Ca^{2+}$  might modulate NaCl and/or  $Ca^{2+}$  transport in the DCT during hypercalcemia via a direct interaction of extracellular  $Ca^{2+}$  with the  $Ca^{2+}$ -sensing receptor. Thiazide-type diuretics, which block the apical Na<sup>+</sup>-Cl<sup>-</sup> cotransporter in the DCT, dissociate Na<sup>+</sup> and Ca<sup>2+</sup> reabsorption by this nephron segment [86, 87]; this dissociation has been attributed to effects of cellular Na<sup>+</sup> uptake on either cellular Ca<sup>2+</sup> entry [88] or exit [86] mechanisms. This inverse relationship between Ca<sup>2+</sup> and Na<sup>+</sup> transport in DCT cells [88, 89] could account for reduced Ca<sup>2+</sup> reabsorption by the DCT during hypercalcemia. In other words, the increased delivery of Na<sup>+</sup> to

the DCT from the TAL would tend to decrease the ability of the DCT to reabsorb  $Ca^{2+}$  and hence help promote the calciuria. Clearly, this issue requires further study.

Several studies also suggest a potential role for the CaR in water handling by the collecting duct. These actions are depicted in Figure 5. Previous studies had suggested an effect of peritubular Ca2+ on modulating vasopressin-stimulated increases in water permeability of the cortical collecting duct [28, 32]. Moreover, a recent preliminary study also suggests an important role for tubular urine Ca<sup>2+</sup> concentration in regulating the water permeability of the inner medullary collecting duct (IMCD) from rat kidney [74]. Baum and colleagues have shown that the CaR and aquaporin-2 co-localize to apical membranes of the IMCD and that raising the Ca<sup>2+</sup> concentration in tubular fluid reversibly reduces the magnitude of ADH-stimulated water permeability of this terminal nephron segment. Thus, in the collecting duct there may be a Ca<sup>2+</sup>-sensing function via the CaR from both basal (interstitial or circulating) and tubular urinary sides. More studies of the effects of extracellular Ca<sup>2+</sup> on water and other solute transport mechanisms in the collecting duct are required before we can fully understand the extent of the regulatory roles played by the CaR in these processes.

## Summary of the integrated effects of the CaR on divalent mineral and water excretion

Let me summarize the proposed role of the renal extracellular  $Ca^{2+}$ -sensing receptor in integrating divalent mineral and water handling by the kidney (Fig. 6). The net effect of increased



**Fig. 6.** Proposed central role of the renal extracellular  $Ca^{2+}$ -sensing receptor (CaR) in integrating divalent mineral and water handling by the kidney. With physiologic transient hypercalcemia, this CaR-mediated reduction in concentrating ability would provide a regulatory mechanism ensuring that the increased delivery of Ca<sup>2+</sup> and Mg<sup>2+</sup> from the thick ascending limb (TAL) to the collecting duct would be excreted in a more dilute urine, thereby decreasing the risk of crystal/stone formation. A similar mechanism could account for the nephrogenic DI commonly observed in patients with pathologic or chronic hypercalcemia.

basolateral (peritubular) Ca<sup>2+</sup> on the TAL is a "loop"-diureticlike action. This effect not only would reduce NaCl reabsorption and hence countercurrent multiplication, but also would decrease the lumen-positive  $V_{te}$  in TAL, which is the driving force for  $Ca^{2+}$ and Mg<sup>2+</sup> transport via the paracellular pathway. An increased load of Ca<sup>2+</sup> (and/or Mg<sup>2+</sup>) then would be delivered to the collecting duct. Direct effects of extracellular Ca<sup>2+</sup>, acting via the CaR to reduce the ADH-stimulated water permeability of the collecting ducts would add further to the reduction in urine concentration brought about by effects of extracellular Ca2+ in the TAL. Particularly important would be the potential direct influence of the tubular urine Ca2+ concentration itself, as it would help to integrate directly changes in distal Ca<sup>2+</sup> delivery with the magnitude of water reabsorption in the terminal collecting duct. Thus the kidney has a built-in divalent mineral regulatory mechanism (that is, the CaR) that helps ensure that Ca<sup>2+</sup> and Mg<sup>2+</sup> are excreted at concentrations below saturation. During pathologic or chronic hypercalcemia this mechanism could account for the nephrogenic diabetes insipidus commonly seen with this electrolyte disorder [25]. As demonstrated by the case presentation, despite this integrated mechanism, stone formation still occurs under certain circumstances.

#### Questions and answers

DR. NICOLAOS E. MADIAS (*Chief, Division of Nephrology, New* England Medical Center, Boston, Massachusetts): Two other cations that come to mind in terms of possible interaction with the  $Ca^{2+}$ -sensing receptor are lithium and aluminum. Lithium is a monovalent cation that occasionally leads to hyperparathyroidism. On the other hand, in-vitro work indicates that aluminum, a trivalent cation, inhibits PTH release from parathyroid cells, and aluminum-related bone disease is often associated with decreased PTH. Do these cations bind to the receptor?

DR. HEBERT: Ed Brown showed years ago that lithium induces abnormal calcium-regulated release of PTH from dispersed bovine parathyroid cells. Since the calcium-sensing receptor mediates the regulation of PTH secretion, when you reduce calcium sensing, PTH secretion increases and hyperparathyroidism results. Some patients on lithium will have a disease like primary hyperparathyroidism, which is most often reversible by eliminating the lithium. In some limited studies we've begun to look at that. It's not surprising that monovalent cations have some effects on this calcium sensor. The binding of calcium to the receptor is thought to involve acidic negatively charged amino acids mainly located in the large extracellular domain, and other cations could alter calcium-binding interactions.

DR. MADIAS: Isn't it somewhat surprising that a system like calcium homeostasis that is so tightly regulated would have a receptor recognized by several ligands other than  $Ca^{2+}$  and exhibiting such a substrate promiscuity?

DR. HEBERT: The promiscuity may be a consequence of physiologically appropriate millimolar affinity of this receptor for calcium. This sensor responds to small variations in calcium in a virtual "sea" of calcium (and other ions). If it were a high-affinity calcium ion receptor, all calcium-binding sites would be fully saturated at normal circulating calcium concentrations, and the receptor would be continuously activated and unaffected by changes in circulating calcium. Multiple (low-affinity) calciumion-binding sites may function in a cooperative fashion to account, at least in part, for the steep relationship between extracellular ionic calcium and PTH release or renal calcium excretion. The nature of the calcium-binding sites thus might account for the ability of other ions to interact with these sites. Consequently, other cations (inorganic ions or organic drugs) could interfere with calcium sensing.

DR. BESS DAWSON-HUGHES (USDA Human Nutrition Research Center on Aging, Tufts University School of Medicine, Boston): Do you believe that the calcium sensor has anything to do with the hypertension that is associated with hyperparathyroidism?

DR. HEBERT: That's an interesting question, but we don't have an answer for it. If we had to speculate, one of the areas we would offer as playing a potential role would be the effects of the receptor in the central nervous system. I discussed one locus, the subfornical organ, but the receptor is also expressed in other regions of the brain. Could there be a role for the calcium receptor in the central regulation of blood pressure? It would be premature to speculate too much other than to say that it's possible that there could be many effects of calcium on brain and possibly vascular function. Some of these cells might be responding to local changes in calcium that may be related to neuronal activity. Local changes in calcium could provide important signals to groups of functioning neurons in the central nervous system, including those involved in blood pressure control.

DR. JEFFREY TATRO (*Division of Endocrinology, New England Medical Center*): Is there evidence for other variants of the receptor, or is there only a single gene?

DR. HEBERT: So far, we think it's a single gene. There is another potential calcium-sensing molecule that has been identified by Lundgren et al, which is a member of the LDL receptor family, a GP330-like molecule that is quite interesting [91]. It's clear that from the FFH-NSHPT studies that the receptor that we have cloned is the primary sensor in the parathyroid gland. This does not exclude the possibility that associated molecules or other receptors can modulate (or provide) ion sensing in certain other tissues. We certainly can't exclude the possibility that other genes are involved in ion sensing. One could postulate sensors for potassium, phosphate, sodium, or protons, for example. Another aspect of this that makes such a speculation rational is the fact that if one looks at the potential structure of the extracellular domain, one can find a similarity between the extracellular domain of the metabotropic glutamate receptors and the calciumsensing receptor. Several investigators have studied the structural homologies of the extracellular domain with other proteins. They identified significant similarities between the extracellular domain of the metabotropic glutamate receptors and a group of proteins that are called periplasmic nutrient-binding proteins found in bacteria with cell walls. These small molecules, many of which have been crystallized, have known structures. These proteins are free-floating in the periplasm between the bacterial cell membrane and the cell wall. As nutrients enter through pores in the cell wall, these specific proteins bind these nutrients and complex with bacterial permeases. The latter provides for transport of the nutrient into the bacterial cell. Many different bacterial nutrientbinding proteins are known, and several of the nutrients include ions. Thus, if these proteins and the calcium-sensing receptor have a common evolutionary origin, there could exist receptors in humans for other ions or nutrients.

DR. ANDREW J. KING (*Division of Nephrology, New England Medical Center*): A frequent clinical finding in patients who are hypercalcemic is severe volume depletion. Do you think the mechanism by which that occurs relates to inhibition of the potassium channel by the calcium receptor?

DR. HEBERT: Yes, we think that the calcium receptor will be proven to be a major regulator of salt, as well as of divalent mineral reabsorption, in the thick limb. Calcium-mediated activation of the receptor leading to inhibition of the apical potassium channel would produce an effect similar to the response to furosemide on thick limb function. To that extent, the patient would lose sodium. In fact, one can detect small increases in sodium excretion with transient hypercalcemia.

Dr. KING: Is the receptor sensitive to monovalent cations such as sodium?

DR. HEBERT: Probably. As I said earlier, we expect that a number of monovalent ions alter calcium ion interactions with the receptor.

DR. MADIAS: In vitro, dispersed parathyroid cells exhibit substantial heterogeneity in their PTH response to changes in the  $Ca^{2+}$  concentration of the medium [92, 93]. Is it known whether the  $Ca^{2+}$ -sensing receptor is involved in this variability, for example due to different complement of receptors?

DR. HEBERT: You do see differences in staining among groups of cells in the parathyroid. This is particularly prominent if one looks at parathyroid tissue from primary and uremic secondary hyperparathyroidism [94]. In secondary hyperparathyroidism, the gland contains what looks like islands of cells that have a normal amount of receptor surrounded by many cells with a very low complement of receptor; this finding probably reflects the clonal nature of the cellular hypertrophy.

DR. AJAY K. SINGH (*Division of Nephrology, New England Medical Center*): You suggested that activation of the calciumsensing receptor due to high prevailing plasma levels of calcium and/or magnesium modulates urinary excretion of the corresponding divalent cation. Could you elaborate on calcium-sensing receptor function when there is hypocalcemia with a normal, or perhaps even an elevated, magnesium level or the converse, a low magnesium level with a normal calcium?

DR. HEBERT: That's an important question. As I discussed, it takes a much higher concentration of magnesium than calcium to activate the receptor. That partially answers the question you asked, but it raises an additional question: is this receptor a magnesium sensor, at least in certain tissues? Can the calcium receptor sense normal concentrations of magnesium in the kidney? Quamme and others have shown over the last two decades that, in the thick ascending limb, hypermagnesemia in rats dramatically affects loop function with regard to sodium chloride reabsorption as well as magnesium and calcium reabsorption [20, 21]. Thus extracellular magnesium is clearly involved somehow in modulating thick ascending limb function. It is possible that in the thick ascending limb, where calcium and magnesium reabsorption normally occur, that is, ions moving from urinary space to the basal side of the cell in an epithelium where there is virtually no water movement, the calcium and/or magnesium concentration on the basal side of the cell increases to levels that could activate or influence the receptor. In this way, magnesium reabsorption by the thick limb could influence both calcium and magnesium reabsorption.

DR. MADIAS: I'm trying to think of some other clinical disorders in which the  $Ca^{2+}$ -sensing receptor might be dysregulated. One disorder that comes to mind is familial idiopathic hypercalciuria. Another is Bartter's syndrome. Could you speculate on these possibilities?

DR. HEBERT: We've thought along these lines as well and are currently investigating the possible involvement of receptor dysregulation in idiopathic hypercalciuria and Bartter's. Lifton's group at Yale has made significant advances in the identification of genes involved in autosomal recessive forms of Bartter's syndrome, including the Gitelman's variant of Bartter's syndrome [95, 96]; these forms are characterized by inherited hypokalemic alkalosis as well as alterations in divalent mineral excretion. Gitelman's disease is caused by mutations in the thiazide-sensitive Na<sup>+</sup>-Cl<sup>-</sup> cotransporter that presumably result in loss of function of the cotransporter. We (unpublished observations) and Lemmink and coworkers [97] also have found linkage and/or mutations in the thiazide-sensitive Na<sup>+</sup>-Cl<sup>-</sup> cotransporter in other families with the Gitelman's phenotype, thus confirming the findings by Lifton's group. Gitelman's disease thus can be regarded as a dysfunction of salt transport in the distal convoluted tubule. Individuals with Gitelman's disease are characterized by hypomagnesemia (with hypermagnesuria) and hypocalciuria. Because Na<sup>+</sup>-Cl<sup>-</sup> cotransporter activity presumably is reduced in this disease, the reduced excretion of calcium can be explained by this functional defect that mimics thiazide use. More recently Lifton's group identified mutations in the loop-diuretic-sensitive Na<sup>+</sup>-K<sup>+</sup>-2Cl<sup>-</sup> cotransporter as the molecular basis of Bartter's syndrome in certain families, whose affected members are characterized by hypokalemic alkalosis, severe volume depletion, and hypercalciuria [96]. Thus, at least in some patients with familial Bartter's syndrome, the abnormal gene is the furosemide (bumetanide)-sensitive  $Na^+-K^+-2Cl^-$  cotransporter in the thick ascending limb of Henle (TAL), and this disease can be thought of as a dysfunction of salt transport in the TAL. Loss of function of Na<sup>+</sup>-K<sup>+</sup>-2Cl<sup>-</sup> cotransport in the TAL would decrease calcium and magnesium reabsorption and increase their loss in the urine; this effect mimics the response to furosemide. It is possible that mutations in other genes may be identified in other families with the autosomal recessive form of Bartter's syndrome.

Although dysfunction in the Ca<sup>2+</sup>-sensing receptor in the TAL would be expected to alter TAL function, I think that it is unlikely that the Ca<sup>2+</sup>-sensing receptor gene is directly involved in the molecular basis of Bartter's syndrome, because one would expect not only abnormalities in divalent handling by the kidney but also altered parathyroid-PTH secretory function. The potential role of the Ca<sup>2+</sup>-sensing receptor in familial idiopathic hypercalciuria is currently under investigation.

DR. MADIAS: Is any work being done on the regulation of the receptor, and are there any recognized hormonal effects on the receptor?

DR. HEBERT: To date, we have not seen any effector systems that have a significant impact or influence on receptor density.

DR. MADIAS: Is the  $Ca^{2+}$ -sensing receptor involved in other calcium-related phenomena, such as renal hemodynamics, renin release, calcitonin secretion, or bone resorption?

DR. HEBERT: This is a very interesting and important question. We and others are looking at these issues, and answers may be forthcoming.

Reprint requests to Dr. S. Hebert, Renal Division, Brigham & Women's Hospital, 75 Francis Street, Boston, Massachusetts 02115, USA

#### Acknowledgments

The Principal Discussant acknowledges that this work is the result of continuing collaboration with Dr. Edward Brown. The author also acknowledges the collaboration and support of the many postdoctoral fellows in Drs. Hebert's and Brown's laboratories who have worked on projects related to the CaR. Dr. Hebert also wishes to thank Dr. H. W. Harris, Jr. for helpful discussions. Both Drs. Hebert and Brown are supported by grant DK48330 from the NIH (NIDDK), and grants from NPS Pharmaceuticals, Inc., and The St. Giles Foundation.

#### References

1. BROWN EM, GAMBA G, RICCARDI D, LOMBARDI M, BUTTERS R, KIFOR O, SUN A, HEDIGER MA, LYTTON J, HEBERT SC: Cloning and characterization of an extracellular  $Ca^{2+}$ -sensing receptor from bovine parathyroid. *Nature* 366:575–580, 1993

- RICCARDI D, PARK J, LEE W-S, GAMBA G, BROWN EM, HEBERT SC: Cloning and functional expression of a rat kidney extracellular calcium/polyvalent cation-sensing receptor. *Proc Natl Acad Sci USA* 92:131-135, 1995
- GARRETT JE, CAPUANO IV, HAMMERLAND LG, HUNG BCP, BROWN EM, HEBERT SC, NEMETH EF, FULLER F: Molecular cloning and functional expression of human parathyroid calcium receptor cDNAs. *J Biol Chem* 270:12919–12925, 1995
- RUAT M, MOLLIVER ME, SNOWMAN AM, SNYDER SH: Calcium sensing receptor: Molecular cloning in rat and localization to nerve terminals. *Proc Natl Acad Sci USA* 92:3161–3165, 1995
- AIDA K, KOISHI S, TAWATA M, ONAYA T: Molecular cloning of a putative Ca<sup>2+</sup>-sensing receptor cDNA from human kidney. *Biochem Biophys Res Comm* 214:524–529, 1995
- BROWN EM, VASSILEV PM, HEBERT SC: Calcium ions as extracellular first messengers. Cell 83:679–682, 1995
- BROWN EM: Kidney and bone: physiological and pathophysiological relationships, in *Handbook of Physiology: Renal Physiology*, edited by WINDHAGER EE, New York, Oxford University Press, 1992, p 1841
- BROWN EM: Extracellular Ca<sup>2+</sup> sensing, regulation of parathyroid cell function, and role of Ca<sup>2+</sup> and other ions as extracellular (first) messengers. *Physiol Rev* 71:371–411, 1991
- FRIEDMAN PA, GESEK FA: Cellular calcium transport in renal epithelia: Measurement, mechanisms, and regulation. *Physiol Rev* 75:429– 471, 1995
- DE ROUFFIGNAC C, QUAMME G: Renal magnesium handling and its hormonal control. *Physiol Rev* 74:305–322, 1994
- 11. KUROKAWA K: The kidney and calcium homeostasis. *Kidney Int* 44:S97-S105, 1994
- STEWART AF, BROADUS AE: Mineral metabolism, in *Endocrinology* and Metabolism, edited by BAXTER FP, BROADUS AE, FROHMAN LA, New York, McGraw-Hill, 1987, p 1317
- AURBACH GD, MARX SJ, SPIEGEL AM: Parathyroid hormone, calcitonin, and calciferols, in *Textbook of Endocrinology*, edited by WILSON J, FOSTER DW, Philadelphia, Saunders, 1985, p 1137
- 14. PARFITT AM, KLEERKOPER M: The divalent cation homeostatic system: physiology and metabolism of calcium, phosphorus, magnesium, and bone, in *Clinical Disorders of Fluid and Electrolyte Metabolism*, edited by MAXWELL MH, KLEEMAN CR, New York, McGraw-Hill, 1980, p 269
- ROUSE D, SUKI WN: Renal control of extracellular calcium. *Kidney Int* 38:700-708, 1995
- KUROKAWA K: Nephrology Forum: Calcium-regulating hormones and the kidney. *Kidney Int* 32:760–771, 1987
- BROWN EM, POLLACK M, SEIDMAN CE, SEIDMAN JG, CHOU Y-HW, RICCARDI D, HEBERT SC: Calcium-sensing cell-surface receptors. N Engl J Med 333:234-240, 1995
- NEMETH EF: Ca<sup>2+</sup> receptor-dependent regulation of cellular function. News Physiol Sci 10:1-5, 1995
- LAU K, BOURDEAU JE: Parathyroid hormone action in calcium transport in the distal nephron. Curr Opin Nephrol Hypertens 4:55-63, 1995
- QUAMME GA: Control of magnesium transport in the thick ascending limb. Am J Physiol (Renal Fluid Electrol Physiol) 256:F197–F210, 1989
- QUAMME GA, DIRKS JH: Magnesium transport in the nephron. Am J Physiol (Renal Fluid Electrol Physiol) 239:F393–F401, 1980
- RICCARDI D, PLOTKIN MD, LEE W-S, LEE K, SEGRE GV, BROWN EM, HEBERT SC: Colocalization of the Ca<sup>2+</sup>-sensing receptor and PTH/ PTHrP receptor in rat kidney (*abstract*). J Am Soc Nephrol 6:954, 1995
- BROWN EM, HEBERT SC: A cloned Ca<sup>2+</sup>-sensing receptor: a mediator of direct effects of extracellular Ca<sup>2+</sup> on renal function? J Am Soc Nephrol 6:1530-1540, 1995
- MARX SJ, ATTIE MF, STOCK JL, SPIEGEL AM, LEVINE MA: Maximal urine-concentrating ability: familial hypocalciuric hypercalcemia versus typical primary hyperparathyroidism. J Clin Endocrinol Metab 52:736-740, 1981
- GILL JR JR, BARTTER FC: On the impairment of renal concentrating ability in prolonged hypercalcemia and hypercalciuria in man. J Clin Invest 40:716-722, 1961
- 26. BROWN E, ENYEDI P, LEBOFF M, ROTBERG J, PRESTON J, CHEN C:

High extracellular Ca<sup>2+</sup> and Mg<sup>2+</sup> stimulate accumulation of inositol

- phosphates in bovine parathyroid cells. *FEBS Lett* 218:113–118, 1987
  27. NEMETH EF, SCARPA A: Cytosolic Ca<sup>2+</sup> and the regulation of secretion in parathyroid cells. *FEBS Lett* 203:15–19, 1986
- 28. JONES SM, FRINDT G, WINDHAGER EE: Effect of peritubular [Ca] or ionomycin on hydrosmotic response of CCTs to ADH or cAMP. Am J Physiol (Renal Fluid Electrolyte Physiol) 254:F240-F253, 1988 29. Такагсні К, Кигокаwa К: High Ca<sup>2+</sup> inhibits peptide hormone-
- dependent cAMP production specifically in thick ascending limbs of Henle. Miner Electrolyte Metab 12:342-346, 1986
- 30. KUSANO E, MURAYAMA N, WERNESS JL, CHRISTENSEN S, HOMMA S, YUSUFI ANK, DOUSA TP: Effects of calcium on the vasopressinsensitive cAMP metabolism in medullary tubules. Am J Physiol (Renal Fluid Electrolyte Physiol) 249:F956-F966, 1985
- 31. TAKAICHI K, UCHIDA S, KUROKAWA K: High Ca<sup>2+</sup> inhibits AVPdependent cAMP production in thick ascending limbs of Henle. Am J Physiol (Renal Fluid Electrolyte Physiol) 250:F770-F776, 1986
- 32. DILLINGHAM MA, DIXON BS, ANDERSON RJ: Calcium modulates vasopressin effect in rabbit cortical collecting tubule. Am J Physiol (Renal Fluid Electrolyte Physiol) 252:F115-F121, 1987
- 33. BROWN EM, FULEIHAN GE-H, CHEN CJ, KIFOR O: A comparison of the effects of divalent and trivalent cations on parathyroid hormone release, 3', 5'-cyclic-adenosine monophosphate accumulation, and the levels of inositol phosphates in bovine parathyroid cells. Endocrinology 127:1064-1071, 1990
- 34. NEMETH EF: Regulation of cytosolic calcium by extracellular divalent cations in C-cells and parathyroid cells. Cell Calcium 11:323-327, 1990
- 35. BROWN EM, BUTTERS R, KATZ C, KIFOR O: Neomycin mimics the effects of high extracellular calcium concentrations on parathyroid function in dispersed bovine parathyroid cells. Endocrinology 128: 3047-3054, 1991
- 36. RIDEFELT P, HELLMAN P, WALLFELT C, AKERSTRÖM G, RASTAD J, GYLFE E: Neomycin interacts with Ca<sup>2+</sup> sensing of normal and adenomatous parathyroid cells. Mol Cell Endocrinol 83:211-218, 1992
- 37. CONKLIN BR, BOURNE HR: Marriage of the flytrap and the serpent. Nature 67:22, 1994
- 38. HEBERT SC, BROWN EM: The extracellular calcium receptor. Curr Opin Cell Biol 7:484-492, 1995
- 39. BOCKAERT J: G proteins, G-protein-coupled receptors: Structure, function, and interactions. Curr Opin Neurobiol 1:1132-1142, 1991
- 40. JACKSON T: Structure and function of G protein coupled receptors. Pharmacol Ther 50:425-442, 1991
- 41. NAKANISHI S: Molecular diversity of glutamate receptors and implications for brain function. Science 258:597-603, 1992
- 42. O'HARA PJ, SHEPPARD PO, THOGERSEN H, VENEZIA D, HALDEMAN BA, MCGRANE V, HOUAMED KM, THOMSEN C, GILBERT TL, MUL-VIHILL ER: The ligand-binding domain in metabotropic glutamate receptors is related to bacterial periplasmic binding proteins. Neuron 11:41-52, 1993
- 43. TAM R, SAIER MH JR: Structural, functional, and evolutionary relationships among extracellular solute-binding receptors of bacteria. Microbiol Rev 57:320-346, 1993
- 44. FLIEGEL L, OHNISHI M, CARPENTER MR, KHANNA VK, REITHMEIER RAF, MACLENNAN DH: Amino acid sequence of rabbit fast-twitch skeletal muscle calsequestrin deduced from cDNA and peptide sequencing. Proc Natl Acad Sci USA 84:1167-1171, 1987
- 45. LAW WM, HEATH H III: Familial benign hypercalcemia (hypocalciuric hypercalcemia). Clinical and pathogenetic studies in 21 families. Ann Intern Med 102:511-519, 1985
- 46. MARX SJ, ATTIE MF, LEVINE MA, SPIEGEL AM, DOWNS RW JR, LASKER RD: The hypocalciuric or benign variant of familial hypercalcemia: clinical and biochemical features in fifteen kindreds. Medicine (Baltimore) 60:235-242, 1981
- 47. POLLAK MR, BROWN EM, CHOU Y-HW, HEBERT SC, MARX SJ, STEINMANN B, LEVI T, SEIDMAN CE, SIEDMAN JG: Mutations in the Ca<sup>2+</sup>-sensing receptor gene cause familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. Cell 75:1297-1303, 1993
- 48. POLLAK MR, CHOU Y-HW, MARX SJ, STEINMANN B, COLE DEC, BRANDI ML, PAPAPOULOS SE, MENKO FH, HENDY GN, BROWN EM, SEIDMAN CE, SEIDMAN JG: Familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. Effect of mutant gene dosage on phenotype. J Clin Invest 93:1108-1112, 1994

- 49. ESTEP HL, MISTRY Z, BURKE PK: Familial idiopathic hypocalcemia (abstract). 63rd Annu Mtg Endocrine Soc, 1981, p 275
- POLLAK MR, BROWN EM, ESTEP HL, MCLAINE PN, KIFOR O, PARK J, 50. HEBERT SC, SEIDMAN CE, SEIDMAN JG: Autosomal dominant hypocalcaemia caused by a  $Ca^{2+}$ -sensing receptor gene mutation. *Nature* Genet 8:303-307, 1994
- 51. CHOU Y-H W, BROWN EB, LEVI T, CROWE G, ATKINSON AB, ARNQVIST HJ, TOSS G, FULEIHAN GE-H, SEIDMAN JG, SEIDMAN CE: The gene responsible for familial hypocalciuric hypercalcemia maps to chromosome 3q in four unrelated families. Nature Genet 1:295-300, 1992
- 52. HEATH H III, JACKSON CE, OTTERUD B, LEPPERT MF: Genetic linkage analysis in familial benign (hypocalciuric) hypercalcemia: evidence for locus heterogeneity. Am J Hum Genet 53:193-200, 1993
- 53. TRUMP D, WHYTE MP, WOODING C, PANG JT, KOCHER D, THAKKER RV: Linkage studies in a kindred with hereditary hypercalcemia and increasing parathyroid hormone levels indicate genetic heterogeneity (abstract), J Bone Miner Res 8:S167, 1993
- 54. HEATH H III: Familial benign hypercalcemia-From clinical description to molecular genetics. West J Med 160:554-561, 1994
- 55. PEARCE SHS, TRUMP D, WOODING C, BESSER GM, CHEW SL, HEATH DA, HUGHES IA, THAKKER RV: Four novel mutations in the calciumsensing receptor gene associated with familial benign (hypocalciuric) hypercalcemia (abstract). J Bone Miner Res 9:S145, 1994
- 56. HEATH H III, ODELBERG S, BROWN D, HILL VM, ROBERTSON M, JACKSON CE, TEH BT, HAYWARD N, LARSSON C, BUIST N, GARRETT J, LEPPERT M: Sequence analysis of the parathyroid cell calcium receptor (CaR) gene in familial benign hypercalcemia (FBH): multiplicity of mutations? (abstract) J Bone Miner Res 9:S414, 1994
- 57. MARX SJ, LASKER RD, BROWN EB, FITZPATRICK LA, SWEENEY NB, GOLDBLOOM RB, GILLIS DA, COLE DEC: Secretory dysfunction in parathyroid cells from neonate with severe primary hyperparathyroidism. J Clin Endocrinol Metab 62:445-449, 1986
- 58. KHOSLA S, EBELING PR, FIREK AF, BURRITT MM, KAO PC, HEATH H III: Calcium infusion suggests a "set-point" abnormality of parathyroid gland function in familial benign hypercalcemia and more complex disturbances in primary hyperparathyroidism. J Clin Endocrinol Metab 76:715-720, 1993
- 59. COOPER L, WERTHEIMER J, LEVEY R, BROWN E, LEBOFF M, WILKIN-SON R, ANAST CS: Severe primary hyperparathyroidism in a neonate with two hypercalcemic parents: management with parathyroidectomy and heterotopic autotransplantation. Pediatrics 78:263-268, 1986
- 60. ATTIE MF, GILL JR JR, STOCK JL, SPIEGEL AM, DOWNS RW JR, LEVINE MA, MARX SJ: Urinary calcium excretion in familial hypocalciuric hypercalcemia. Persistence of relative hypocalciuria after induction of hypoparathyroidism. J Clin Invest 72:667-676, 1983
- 61. QUAMME GA, DIRKS JH: Intraluminal and contraluminal magnesium on magnesium and calcium transfer in the rat nephron. Am J Physiol (Renal Fluid Electrolyte Physiol) 238:F187-F198, 1980
- 62. DE ROUFFIGNAC C, DISTEFANO A, WITTNER M, ROINEL N, ELALOUF JM: Consequences of differential effects of ADH and other peptide hormones on thick ascending limb of mammalian kidney. Am J Physiol (Regulatory Integrative Comp Physiol) 260:R1023-R1035, 1991
- 63. HEBERT SC, ANDREOLI TE: Control of NaCl transport in the thick ascending limb. Am J Physiol (Renal Fluid Electrolyte Physiol) 246: F745-F756, 1984
- 64. GUIGNARD J-P, JONES NF, BARRACLOUGH MA: Effect of brief hypercalcaemia of free water reabsorption during solute diuresis: evidence for impairment of sodium transport in Henle's loop. Clin Sci 39:337-347, 1970
- 65. MANITIUS A, LEVITIN H, BECK D, EPSTEIN FH: On the mechanism of impairment of renal concentrating ability in hypercalcemia. J Clin Invest 39:693-697, 1960
- 66. BECK D, LEVITIN H, EPSTEIN FH: Effect of intravenous infusions of calcium on renal concentrating ability. Am J Physiol 197:1118-1120, 1959
- 67. BECK N, SINGH H, REED SW, MURDAUGH HV, DAVIS BB: Pathogenic role of cyclic AMP in the impairment of urinary concentrating ability in acute hypercalcemia. J Clin Invest 54:1049-1055, 1974
- SUKI WN, EKNOYAN G, RECTOR FC JR, SELDIN DW: The renal 68. diluting and concentrating mechanism in hypercalcemia. Nephron 6:50-61.1969
- 69. TRECHSEL U, EISMAN JA, FISCHER JA, BONJOUR J-P, FLEISCH H:

Calcium-dependent, parathyroid hormone-independent regulation of 1,25-dihydroxyvitamin D. *Am J Physiol (Endocrinol Metab)* 239:E119–E124, 1980

- WEISINGER JR, FARUS MJ, LANGMAN CB, BUSHINSKY DA: Regulation of 1,25-dihydroxyvitamin D<sub>3</sub> by calcium in the parathyroidectomized, parathyroid hormone replete rat. J Bone Miner Res 4:929–935, 1989
- SHAREGHI GR, AGUS ZS: Magnesium transport in the cortical thick ascending limb of Henle's loop of the rabbit. J Clin Invest 69:759–769, 1982
- QUAMME GA: Effect of hypercalcemia on renal tubular handling of calcium and magnesium. Can J Physiol Pharmacol 60:1275–1280, 1982
- TAKAICHI K, KUROKAWA K: Inhibitory guanosine triphosphate-binding protein-mediated regulation of vasopressin action in isolated single medullary tubules of mouse kidney. J Clin Invest 82:1437–1444, 1988
- 74. BAUM MA, CHATTOPADHYAY N, BROWN EM, RUDDY MK, HOSSELET C, RICCARDI D, HEBERT S, HARRIS HW: Perinatal expression of aquaporins-2 and 3 (APQ2, APQ3) and calcium receptor (RaKCaR) in developing rat kidney collecting duct (CD) (abstract). J Am Soc Nephrol 6:319, 1995
- CLEMENS TL, MCGLADE SA, GARRETT KP, CRAVISO GL, HENDY GN: Extracellular calcium modulates vitamin D-dependent calbindin-D28k gene expression in chick kidney cells. *Endocrinology* 124:1582– 1584, 1989
- 76. DISTEFANO A, DE ROUFFIGNAC C, WITTNER M: Transepithelial Ca<sup>2+</sup> and Mg<sup>2+</sup> transport in the cortical thick ascending limb of Henle's loop of the mouse is a voltage-dependent process. *Renal Physiol Biochem* 16:157–166, 1993
- 77. MANDON B, SIGA E, ROINEL N, DE ROUFFIGNAC C:  $Ca^{2+}$ ,  $Mg^{2+}$  and  $K^+$  transport in the cortical and medullary thick ascending limb of the rat nephron: influence of transpithelial voltage. *Pflugers Arch* 424: 558–560, 1993
- BOURDEAU JE, BURG MB: Voltage dependence of calcium transport in the thick ascending limb of Henle's loop. Am J Physiol (Renal Fluid Electrolyte Physiol) 236:F357–F364, 1979
- FRIEDMAN PA: Renal calcium transport: sites and insights. News Physiol Sci 3:17-21, 1988
- DE ROUFFIGNAC C, ELALOUF J-M, ROINEL N: Physiological control of the urinary concentrating mechanism by peptide hormones. *Kidney Int* 31:611–620, 1987
- WANG W, LU M, HEBERT SC: P450-metabolite of arachidonic acid (AA) mediates the Ca<sup>2+</sup>-induced inhibition of the apical 70 pS K<sup>+</sup> channel in thick ascending limb (TAL) (*abstract*). J Am Soc Nephrol 6:355, 1995
- 82. WANG W, LU M, HEBERT SC: P450 metabolites mediate extracellular Ca<sup>2+</sup>-induced inhibition of apical K<sup>+</sup> channels in the thick ascending limb of the rat kidney. Am J Physiol (Cell Physiol), in press
- Wang W: Two types of K<sup>+</sup> channel in TAL of rat kidney. Am J Physiol (Renal Fluid Electrolyte Physiol) 267:F599-F605, 1994
- 84. EDWARDS BR, BAER PG, SUTTON RAL, DIRKS JH: Micropuncture

study of diuretic effects on sodium and calcium reabsorption in the dog nephron. J Clin Invest 52:2418-2427, 1973

- WANG T, WANG W, KLEIN-ROBBENHAAR J, GIEBISCH G: Effects of glyburide on renal tubule transport and potassium-channel activity. *Renal Physiol Biochem* 18:169–182, 1995
- COSTANZO LS, WINDHAGER EE: Calcium and sodium transport by the distal convoluted tubule of the rat. Am J Physiol (Renal Fluid Electrolyte Physiol) 235:F492-F506, 1978
- COSTANZO LS: Comparison of calcium and sodium transport in early and late distal tubules: effect of amiloride. Am J Physiol (Renal Fluid Electrolyte Physiol) 246:F937–F945, 1984
- GESEK FA, FRIEDMAN PA: Mechanism of calcium transport stimulated by chlorothiazide in mouse distal convoluted tubule cells. J Clin Invest 90:429-438, 1992
- BRUNETTE MG, MAILLOUX J, LAJEUNESSE D: Calcium transport through the luminal membrane of the distal tubule. I. Interrelationship with sodium. *Kidney Int* 41:281–288, 1992
- 90. PEACOCK M, ROBERTSON WG, NORDIN BEC: Relation between serum and urinary calcium with particular reference to parathyroid activity. *Lancet* 1:384–386, 1969
- 91. LUNDGREN S, HJÄLM G, HELLMAN PBE, JUHLIN C, RASTAD J, KLARESKOG L, AKERSTRÖM G, RASK L: A protein involved in calcium sensing of human parathyroid and placental cytotrophoblast cells belongs to the LDL-receptor protein. *Exp Cell Res* 212:344–350, 1994
- 92. SUN F, RITCHIE C, HASSAGER C, MAERCKLEIN P, FITZPATRICK L: Heterogeneous response to calcium by individual parathyroid cells. *J Clin Invest* 91:595-601, 1993
- CLARKE BL, HASSAGER C, FITZPATRICK LA: Regulation of parathyroid hormone release by protein kinase-C independent of extracellular calcium in bovine parathyroid cells. *Endocrinology* 132:1168–1175, 1993
- 94. KIFOR O, MOORE FJ JR, WANG P, GOLDSTEIN M, VASSILEV P, KIFOR I, HEBERT SC, BROWN EM: Reduced expression of the extracellular Ca<sup>2+</sup>-sensing receptor in primary and uremic secondary hyperparathyroidism. *J Endocrinol Metab* 81:1598–1606, 1995
- 95. SIMON DB, NELSON-WILLIAMS C, BIA MJ, ELLISON D, KARET FE, MOLINA AM, VAARA I, IWATA F, CUSHNER HM, KOOLEN M, GAINZA FJ, GITELMAN HJ, LIFTON RP: Gitelman's variant of Bartter's syndrome, inherited hypokalemic alkalosis, is caused by mutations in the thiazide-sensitive Na-Cl cotransporter. Nat Genet 12:1–8, 1996
- 96. SIMON DB, KARET FE, HAMDAN JM, DIPIETRO A, SANJAD SA, LIFTON RP: Bartter's syndrome, hypokalemic alkalosis with hypercalciuria, is caused by mutations in the Na-K-2Cl cotransporter NKCC2. Nat Genet 13:183–188, 1996
- 97. LEMMINK HH, VAN DEN HEUVEL LPWJ, VAN DIJK HA, MERKX GFM, SMILDE TJ, TASCHNER PEM, MONNENS LAH, HEBERT SC, KNOERS NVAM: Linkage of Gitclman syndrome to the human thiazidesensitive sodium-chloride cotransporter (hTSC) gene with identification of mutations in three Dutch families. *Pediatr Nephrol*, in press