ORIGINAL ARTICLE

Polymorphisms of the *RET* Gene in Hirschsprung Disease, Anorectal Malformation and Intestinal Pseudo-obstruction in Taiwan

Trang-Tiau Wu,^{1,2} Tsui-Wei Tsai,³ Han Chang,⁴ Ching-Chyuan Su,⁵ Shuan-Yow Li,³ Hong-Shiee Lai,⁶ Chuan Li³*

Background/Purpose: Mutations in the receptor tyrosine kinase *RET* gene are associated with Hirschsprung disease (HD), which is also known as congenital intestinal aganglionosis. We found an association with specific alleles in five single nucleotide polymorphism (SNP) sites of the *RET* gene in our HD patients. **Methods:** We compared the association of specific *RET* SNP alleles in patients with severe GI disorders such as anorectal malformation (ARM) or pediatric intestinal pseudo-obstruction (IPO) to that in HD patients. Sixty-four HD, 23 ARM and 35 IPO patients were included. Genomic DNA extracted from blood samples was analyzed by polymerase chain reaction and DNA sequencing analysis.

Results: The allele distributions of all five *RET* SNPs in the HD patients deviated from those in the normal population (p < 0.05), whereas those of the ARM patients did not. The allele distributions of these *RET* SNPs in the IPO patients were all significantly different from those in the HD patients. Allele distributions of exon 2 and 13 in the IPO patients were also significantly different from those of the normal population. The frequencies of all the HD-predominant alleles were lower in the HD patients than the normal population, and were even lower in the IPO patients.

Conclusion: This study strengthens the association of specific *RET* SNP alleles with typical HD in Taiwan. [*J Formos Med Assoc* 2010;109(1):32–38]

Key Words: anorectal malformations, Hirschsprung disease, intestinal pseudo-obstruction, RET

Hirschsprung disease (HD) is the most common cause of neonatal intestinal obstruction, and is characterized by the absence of intramural ganglion cells in the nerve plexuses of the distal gut. HD is present in neonates or early childhood, with symptoms ranging from chronic constipation to acute ileus. It affects one in 5000 live newborns, with a male predominance (3:1 to 5:1).¹

The disease has a complex genetic etiology with susceptibility genes including members of the RET,^{2–5} or endothelin (*EDNRB*)-regulated signaling pathways,^{6,7} and *SOX-10*-mediated transcriptional regulation.⁸

Coding sequence mutations in the receptor tyrosine kinase gene *RET*, the major HD gene, can be identified in up to 50% of familial and

©2010 Elsevier & Formosan Medical Association

¹Department of Pediatric Surgery, Chung Shan Medical University Hospital, ²Institute of Medicine, ³Department of Biomedical Sciences, ⁴Department of Pathology, School of Medicine, Chung Shan Medical University, Taichung, ⁵Tian-Sheng Memorial Hospital, Tong Kang, Pin-Tong, and ⁶Department of Surgery, National Taiwan University Hospital and National Taiwan University College of Medicine, Taipei, Taiwan.

Received: January 12, 2009 Revised: May 7, 2009 ELSEVIER Accepted: June 9, 2009 *Correspondence to: Dr Chuan Li, Department of Biomedical Sciences, Chung Shan Medical University, 110 Section 1, Chein-Kuo North Road, Taichung, Taiwan. E-mail: cli@csmu.edu.tw 7–35% of sporadic cases in various populations with different penetrance.^{4,5,9–14} The rate of *RET* mutation in HD patients in Taiwan in our previous study was low (3.6%) compared with that reported in other populations. Nevertheless, we have demonstrated the association between HD and specific alleles in five single nucleotide polymorphism (SNP) sites in the *RET* coding regions.¹⁵

Anorectal malformation (ARM) caused by the absence or ectopic location of the anus is often associated with rectal ectasia, a state of massive dilation of the rectum and distal sigmoid colon. The incidence is approximately 1:5000 live births, with a male predominance. ARM can be primary at birth, with characteristic features such as dilated recto-sigmoid colon with a thin bowel wall, without hypertrophy of smooth muscles, or rectal ectasia with balloon-like rectum,16 colonic inertia,17 megarectum,18 and pseudo-Hirschsprung disease.¹⁹ As a result of intestinal obstruction and sepsis, many newborn ARM patients have to be treated surgically. Some ARM patients develop secondary rectal ectasia postoperatively as a result of bowel reaction to distal obstruction, or inadequate evacuation.²⁰ Despite its clinical relevance, the etiology and pathogenesis of typical HD and ARM associated with rectal ectasia are different.

Intestinal pseudo-obstruction (IPO) is a functional gastrointestinal (GI) disorder with symptoms including cramps, stomach pain, nausea, vomiting, bloating, fewer bowel movements than usual, and loose stools. Pediatric IPO refers to a heterogeneous group of disorders that are characterized by intestinal obstruction in the absence of mechanical evidence of obstruction.²¹ The etiology of IPO can involve abnormalities of the neurogenic or myogenic control mechanisms of gut motility. Although most IPO patients are referred to as sporadic, familial forms also have been described with either autosomal recessive or dominant transmission.^{22–24} Surgery usually plays a limited role in the treatment of IPO.

ARM and IPO patients represent a population with severe GI disorders other than HD, which is

frequently encountered in pediatric surgery departments. We thus analyzed the five *RET* SNP sites of our patients to further support the nonrandom association of the *RET* polymorphisms in HD patients in Taiwan.

Materials and Methods

Samples

Sixty-four HD patients were diagnosed at infancy and underwent a definite pull-through operation at Chung Shan Medical University Hospital and Affiliate Hospitals from 1976 to 2005. Fiftytwo patients had short-segment aganglionosis and 12 had long-segment aganglionosis, including two with total colon aganglionosis. Patients with Down syndrome or ARM were excluded from this group. Twenty-three ARM patients and 35 functional GI disorder patients with IPO were under medical treatment at Chung Shan Medical University Hospital from 1999 to 2006. The ARM patients included 14 with dilated rectal pouch and six with Down syndrome (three combined with dilated rectal pouch). The IPO patients suffered from severe constipation, but rectal biopsies revealed the presence of ganglion cells. There were four familial cases. Control DNA samples were obtained from normal individuals in Central or Southern Taiwan, who did not have HD, ARM or IPO. The study was approved by the local Institutional Review Board.

Polymerase chain reaction (PCR) and DNA sequence analysis

Genomic DNA was extracted from blood samples by the Puregene Genomic DNA Purification Kit (Gentra Systems, Big Lake, MN, USA), according to manufacturer's instruction. PCR amplification of the five *RET* exons (2, 7, 11, 13 and 15) was performed as described previously.¹⁵ Basically, 150 ng of genomic DNA was amplified in a 50-µL reaction that contained 20 pmol each primer and 2 U *Taq* DNA polymerase (Gene Pure Technology, Taichung, Taiwan). The PCR products were purified by a PCR-M[®] Clean Up System (Viogene, Taipei, Taiwan) and subjected to automated sequencing. For some patients or controls, polymorphisms in exons 2, 13, and 15 of the *RET* gene were examined by restriction enzyme cleavage as described previously.¹⁵ The PCR products of exon 2 or 15 were digested with *EagI* or *RsaI*, respectively, at 37 °C for 2–4 hours. The PCR products of exon 13 were cleaved with *TaqI* at 65 °C for 1.5 hours. DNA fragments were then analyzed by agarose gel electrophoresis.

Immunohistochemistry

Bowel tissue sections were obtained from a HD patient with typical rectosigmoid and an ARM patient with rectal ectasia. Immunohistochemical studies were performed on paraffin-embedded tissue sections. The slides were incubated at 50°C overnight, then soaked in xylene for 10 minutes for three times, and immersed in 100% ethanol for 5 minutes, 95% ethanol for 5 minutes, and 80% and 70% ethanol for 1 minute each. After rehydration for 5 minutes, the slides were boiled in citrate buffer (pH 6.1, Target Retrieval Solution; Dako, Carpinteria, CA, USA) for 25 minutes, rinsed with water and 1 × PBS, and treated with 3% hydrogen peroxide for 10 minutes. After that, the slides were incubated with anti-Neuronal Nuclei (NeuN) antibodies (1:100 dilution; Chemicon, Temecula, California, USA) at room temperature for 30 minutes and then biotinylated secondary antibody (Dako) for 30 minutes, and finally in a solution of streptavidinhorseradish peroxidase (LSAB2 System; Dako) for 30 minutes. Color was with the Substrate-Chromogen Solution (LSAB2 System; Dako) for 5 minutes, and counter-stained with Mayer hematoxylin (Merck, Darmstadt, Germany) for 1 minute. At last, the slides were dehydrated in 50%, 75% and 100% ethanol for 1 minute each, and sealed with mounting medium (CytosealTM 60 Thermo Scientific, Waltham, MA, USA).

Statistical analysis

The χ^2 test was used to determine the significance of the association of the allele frequencies between patients and normal control groups.

Results

RET gene polymorphisms in HD or ARM patients

We investigated 64 typical HD patients. The allele distributions of the five *RET* SNPs (c135G>A in exon 2, A45A; c1296G>A in exon 7, A432A; c2071G>A in exon 11, G691S; c2307T>G in exon 13, L769L; and c2712C>G in exon 15, S904S) in this HD population were very close to those in our previous study,¹⁵ with significant deviation (p<0.05) from those of the normal population (Table 1). The frequencies of allele A of c135G>A, allele G of c1296G>A, allele G of c2307T>G and allele C of c2712C>G were significantly higher in our HD patients than in the controls.

To investigate whether the association of *RET* SNPs was specific to HD patients, we investigated *RET* polymorphisms in ARM patients. No significant differences were detected in the allele distributions of these five *RET* SNPs between the ARM patients and normal controls. Statistically significant deviations (p<0.02) were detected in the allele frequencies of three *RET* SNPs (exons 7, 11, and 13) between HD and ARM patients (Table 1).

RET gene polymorphism in pediatric IPO patients

We determined the allele distributions of the RET SNPs in patients with IPO. There were significant differences in the allele frequencies of all five SNPs between patients with HD and IPO (p <0.001; Table 2). In addition, significant differences were also detected in the frequencies of exons 2 and 13 SNPs between the IPO patients and normal controls (p < 0.05). The frequency of the G allele of c135 (exon 2) in HD patients was 0.31, while the frequency increased to 0.46 and 0.61 in normal and IPO patients, respectively. For the T variant of c2307 (exon 13), the frequency in HD patients was 0.30, while the frequency increased to 0.47 and 0.64 in normal and IPO patients, respectively. It was apparent that the G allele of the c135 SNP and the T allele of the c2307 SNP were more frequent in the normal and IPO

| | | - | | | | | | | | | | | |
|-----|---------|------------|----------------------|------|------------|------------------|---------|-------------------|-------|--------------------|---------|--|--|
| | | | Allele frequency* | | | Statistics | | | | | | | |
| Exo | n Codon | Nucleotide | HD ARM (128) (46) | ARM | N (100) | HD <i>us</i> . N | | ARM <i>vs</i> . N | | ARM <i>vs</i> . HD | | | |
| | | change | | (46) | | χ^2 | р | χ^2 | р | χ^2 | р | | |
| 2 | A45A | c135G>A | G:0.31 | 0.43 | 0.46 | 5.19 | 0.023 | 0.081 | 0.776 | 2.24 | 0.135 | | |
| | | | A:0.69 | 0.57 | 0.54 | | | | | | | | |
| 7 | A432A | c1296G>A | G:0.97 | 0.80 | 0.77 | 21.24 | < 0.001 | 0.217 | 0.641 | 13.23 | < 0.001 | | |
| | | | A:0.03 | 0.20 | 0.23 | | | | | | | | |
| 11 | G691S | c2071G > A | G:0.98 | 0.89 | 0.91 | 4.98 | 0.026 | 0.127 | 0.772 | 5.61 | 0.018 | | |
| | | | A:0.02 | 0.11 | 0.09 | | | | | | | | |
| 13 | L769L | c2307T > G | T:0.30 | 0.54 | 0.47 | 7.19 | 0.017 | 0.681 | 0.409 | 8.91 | 0.003 | | |
| | | | G:0.70 | 0.46 | 0.53 | | | | | | | | |
| 15 | S904S | c2712C > G | C:0.99 | 0.96 | 0.92 | 7.71 | 0.005 | 0.660 | 0.418 | 2.54 | 0.111 | | |
| | | | G:0.01 | 0.04 | 0.08 | | | | | | | | |

Table 1. Allele frequencies of RET polymorphisms in Hirschsprung disease patients, anorectal malformation patients, and normal population

*The numbers in the parentheses are the numbers of the alleles included in the analyses. HD=Hirschsprung disease; ARM=anorectal malformation; N = normal population.

| obstruction patients and normal population | | | | | | | | | | |
|--|-------|----------------------|-------------|-------------|------------|------------|------------|------------------|---------|--|
| RET exon | Codon | Nucleotide change | Alle | le frequen | су* | | Statistics | | | |
| | | | HD (128) | IPO (70) | N (100) | HD us. IPO | | IPO <i>us.</i> N | | |
| | | | | | | χ^2 | р | χ^2 | р | |
| 2 | A45A | c135G>A | G:0.31 | 0.61 | 0.46 | 16.92 | < 0.001 | 3.93 | < 0.050 | |

0.54

0.77

0.23

0.91

0.09

0.47

0.53

0.92

0.08

27.51

12.31

22.25

13.76

0.39

0.71

0.29

0.84

0.16

0.64

0.36

0.87

0.13

A:0.69

G:0.97

A:0.03

G:0.98

A:0.02

T:0.30

G:0.70

C:0.99

G:0.01

of RET polymorphisms in Hirschsprung disease patients, intestinal pseudo

*The numbers in the parentheses are the numbers of the alleles included in the analyses. HD=Hirschsprung disease; IPO=intestinal pseudo-obstruction; N = normal population.

populations than that in the HD patients. Although there are no statistically significant difference in the other sites (exon 7, 11, and 15) among the three groups, the frequencies of the HD-predominant alleles showed a decrease order of HD patients > normal population>IPO patients in exon 7 (0.97 > 0.77 > 0.71), exon 11 (0.98 > 0.91 > 0.84)and exon 15 (0.99 > 0.92 > 0.87), respectively.

c1296G>A

c2071G > A

c2307T > G

c2712C > G

Immunohistochemistry of tissues from HD and ARM patients

< 0.001

< 0.001

< 0.001

< 0.001

0.68

1.78

4.96

1.08

0.411

0.181

< 0.050

0.300

A low incidence of ARM associated with typical HD has been reported.^{26,27} To confirm our classification of patients as typical HD or ARM patients associated with rectal ectasia, we performed immunohistochemical studies. NeuN has been used as a neuron-specific marker for the

7

11

13

15

A432A

G691S

L769L

S904S



Figure. NeuN immunoreactivity in tissue sections from a typical Hirschsprung disease patient and an anorectal malformation patient associated with rectal ectasia. (A) Normal and (B) abnormal distal segments of a typical Hirschsprung disease patient. (C) Normal and (D) abnormal rectal ectasia segments, along with the distal pouch of an anorectal malformation patient with rectal ectasia. White arrows indicate positions of the myenteric plexuses in these sections (original magnification, 200×).

detection of ganglion cells in colonic plexuses.²⁸ Immunostaining results showed that mature large ganglion cells were present in the myenteric plexuses of the normal segments (Figure A) but was absent in those of abnormal distal segments in a typical HD patients (Figure B). Conversely, NeuN immunostaining results of an ARM patient associated with rectal ectasia showed mature large ganglion cells in the myenteric plexuses of normal segments (Figure C) and in those of the abnormal segments from rectal ectasia or the distal pouch (Figure D).

Discussion

RET coding region mutations are only found in a small fraction of typical HD patients but the association of variants or haplotypes has been shown in different populations.^{29–32} We have reported previously the association of five different

RET SNPs with typical HD in Taiwan.¹⁵ The result was very similar to that in the Chinese population in Hong Kong³² but different from other Asian populations in Korea³³ and Thailand.³⁴ We further analyzed these polymorphic sites in 23 ARM patients but did not detect any significant difference (p > 0.05) between the allele distributions of all five RET SNPs in patients and the normal population. However, the allele distributions of exon 2 and 15 SNPs in ARM patients did not differ significantly from those in HD patients. This was probably the result of the limited sample size of the ARM patients. Neuron-specific NeuN immunohistochemical analysis confirmed our clinical classification, and excluded the possibility of misclassification of the patients.

Pediatric IPO might arise from neuropathy or myopathy.²¹ IPO patients included in the present study were characterized by signs and symptoms of intestinal obstruction without mechanical evidence of obstruction. Consequently, typical HD patients were excluded from this group of patients. The RET SNPs in these IPO patients and their allele distributions were all significantly different from those in the HD patients (p < 0.001). Besides, for these IPO patients, allele distributions at the polymorphic sites of exons 2 and 13 were also significantly different (p < 0.05) from those of the normal control population. The frequencies of the HDpredominant A allele of c135 (at exon 2) and the G allele of c2307 (exon 13) was the highest in IPO patients and the lowest in HD patients. Even though the allele distributions in the other three SNP sites did not differ significantly between IPO patients and the normal population, the frequencies of the HD-predominant alleles in these three sites similar trends (HD patients>normal population > IPO patients). The composition of the 35 IPO patients was rather heterogeneous and included three patients born to Vietnamese mothers and one to a Korean father. Thus, the association of specific RET alleles with our IPO patients was not because the patients were from a subpopulation with common ancestors.

Decreased RET function caused by RET mutations is associated with HD. On the contrary, gain of function mutations in RET cause multiple endocrine neoplasia type 2 or medullary thyroid carcinoma.³⁵ Four SNPs in exons 2, 7, 13, and 15 that showed a strong association with HD in this study did not change the encoded amino acids. Association of reduced RET protein expression has been reported for a few variants or haplotypes of HD.^{31,36} Whether the association of HD with RET SNPs results from closely associated functioning variants, or the SNP per se is responsible for the reduced RET protein function is still under investigation. It has not been reported that the RET gene is involved in IPO. One possible explanation of the putative association of specific RET alleles in our IPO patients is that variants associated with specific RET SNPs might affect the normal RET-related signaling in the enteric nervous system, even though neuron development might not be permanently defective as in HD. Balanced allele distribution for normal RET protein function might be required for normal intestinal motility.

In summary, neither ARM nor IPO patients showed an association of specific alleles in the *RET* coding regions as in typical HD. The results strengthen the association of specific *RET* SNP alleles with typical HD in Taiwan. Furthermore, specific *RET* alleles in exon 2 and 13 SNPs appear to be associated significantly with IPO, which provides a new aspect for considering the role of RET-related signaling in regulation of enteric movements.

Acknowledgments

This study was supported in part by the National Science Council (NSC 89-2314-B040-036, 89-2745-P040-002, and 90-2745-P040-002) and Chung Shan Medical University (CSMC 89-OM-A038). We thank the patients and their parents involved in this study. We appreciate the help received from Mr L.C. Yang for conducting immunohistochemical experiments and Ms Y.T. Shen for preparation of the manuscript.

References

- Amiel J, Sproat-Emison E, Garcia-Barcelo M, et al. Hirschsprung disease, associated syndromes and genetics: a review. J Med Genet 2008;45:1–14.
- Doray B, Salomon R, Amiel J, et al. Mutation of the RET ligand, neurturin, supports multigenic inheritance in Hirschsprung disease. *Hum Mol Genet* 1998;7: 1449–52.
- Angrist M, Bolk S, Halushka M, et al. Germline mutations in glial cell line-derived neurotrophic factor (GDNF) and RET in a Hirschsprung disease patient. *Nat Genet* 1996;14:341–4.
- Edery P, Lyonnet S, Mulligan LM, et al. Mutations of the RET proto-oncogene in Hirschsprung's disease. *Nature* 1994;367:378–80.
- Romeo G, Ronchetto P, Luo Y, et al. Point mutations affecting the tyrosine kinase domain of the RET proto-oncogene in Hirschsprung's disease. *Nature* 1994;367:377–8.
- 6. Hofstra RM, Osinga J, Tan-Sindhunata G, et al. A homozygous mutation in the endothelin-3 gene associated with a combined Waardenburg type 2 and Hirschsprung

phenotype (Shah-Waardenburg syndrome). *Nat Genet* 1996;12:445–7.

- Puffenberger EG, Hosoda K, Washington SS, et al. A missense mutation of the endothelin-B receptor gene in multigenic Hirschsprung's disease. *Cell* 1994;79:1257–66.
- Pingault V, Bondurand N, Kuhlbrodt K, et al. SOX10 mutations in patients with Waardenburg–Hirschsprung disease. *Nat Genet* 1998;18:171–3.
- Garcia-Barcelo M, Sham MH, Lee WS, et al. Highly recurrent RET mutations and novel mutations in genes of the receptor tyrosine kinase and endothelin receptor B pathways in Chinese patients with sporadic Hirschsprung disease. *Clin Chem* 2004;50:93–100.
- Sakai T, Nirasawa Y, Itoh Y, et al. Japanese patients with sporadic Hirschsprung: mutation analysis of the receptor tyrosine kinase proto-oncogene, endothelin-B receptor, endothelin-3, glial cell line-derived neurotrophic factor and neurturin genes: a comparison with similar studies. *Eur J Pediatr* 2000;159:160–7.
- 11. Svensson PJ, Molander ML, Eng C, et al. Low frequency of RET mutations in Hirschsprung disease in Sweden. *Clin Genet* 1998;54:39–44.
- Seri M, Yin L, Barone V, et al. Frequency of RET mutations in long- and short-segment Hirschsprung disease. *Hum Mutat* 1997;9:243–9.
- Attie T, Pelet A, Edery P, et al. Diversity of RET protooncogene mutations in familial and sporadic Hirschsprung disease. *Hum Mol Genet* 1995;4:1381–6.
- Angrist M, Bolk S, Thiel B, et al. Mutation analysis of the RET receptor tyrosine kinase in Hirschsprung disease. *Hum Mol Genet* 1995;4:821–30.
- Wu TT, Tsai TW, Chu CT, et al. Low RET mutation frequency and polymorphism analysis of the RET and EDNRB genes in patients with Hirschsprung disease in Taiwan. *J Hum Genet* 2005;50:168–74.
- Cloutier R, Archambault H, D'Amours C, et al. Focal ectasia of the terminal bowel accompanying low anal deformities. J Pediatr Surg 1987;22:758–60.
- Partridge JP, Gough MH. Congenital abnormalities of the anus and rectum. Br J Surg 1961;49:37–50.
- Powell RW, Sherman JO, Raffensperger JG. Megarectum: a rare complication of imperforate anus repair and its surgical correction by endorectal pullthrough. *J Pediatr Surg* 1982;17:786–95.
- 19. Ravitch MM. Pseudo Hirschsprung's disease. *Ann Surg* 1958;147:781–95.
- Brent L, Stephens FD. Primary rectal ectasia. A quantitative study of smooth muscle cells in normal and hypertrophied human bowel. *Prog Pediatr Surg* 1976;9: 41–62.

- 21. Rudolph CD, Hyman PE, Altschuler SM, et al. Diagnosis and treatment of chronic intestinal pseudo-obstruction in children: report of consensus workshop. *J Pediatr Gastroenterol Nutr* 1997;24:102–12.
- 22. Di Lorenzo C. Pseudo-obstruction: current approaches. Gastroenterology 1999;116:980–7.
- 23. Coulie B, Camilleri M. Intestinal pseudo-obstruction. Annu Rev Med 1999;50:37–55.
- 24. Stanghellini V, Corinaldesi R, Barbara L. Pseudo-obstruction syndromes. *Baillieres Clin Gastroenterol* 1988;2:225–54.
- 25. Gimm O, Neuberg DS, Marsh DJ, et al. Over-representation of a germline RET sequence variant in patients with sporadic medullary thyroid carcinoma and somatic RET codon 918 mutation. *Oncogene* 1999;18:1369–73.
- 26. Hasse W. Associated malformation with anal and rectal atresiae. *Prog Pediatr Surg* 1976;9:99–103.
- 27. Kiesewetter WB, Sukarochana K, Sieber WK. The frequency of aganglionosis associated with imperforate anus. *Surgery* 1965;58:877–80.
- Yang S, Donner LR. Detection of ganglion cells in the colonic plexuses by immunostaining for neuron-specific marker NeuN: an aid for the diagnosis of Hirschsprung disease. *Appl Immunohistochem Mol Morphol* 2002;10:218–20.
- 29. Borrego S, Ruiz A, Saez ME, et al. RET genotypes comprising specific haplotypes of polymorphic variants predispose to isolated Hirschsprung disease. *J Med Genet* 2000;37:572–8.
- Gabriel SB, Salomon R, Pelet A, et al. Segregation at three loci explains familial and population risk in Hirschsprung disease. *Nat Genet* 2002;31:89–93.
- Fitze G, Appelt H, Konig IR, et al. Functional haplotypes of the RET proto-oncogene promoter are associated with Hirschsprung disease (HSCR). *Hum Mol Genet* 2003;12: 3207–14.
- Garcia-Barcelo MM, Sham MH, Lui VC, et al. Chinese patients with sporadic Hirschsprung's disease are predominantly represented by a single RET haplotype. J Med Genet 2003;40:e122.
- Kim JH, Yoon KO, Kim JK, et al. Novel mutations of RET gene in Korean patients with sporadic Hirschsprung's disease. J Pediatr Surg 2006;41:1250–4.
- 34. Sangkhathat S, Kusafuka T, Chengkriwate P, et al. Mutations and polymorphisms of Hirschsprung disease candidate genes in Thai patients. *J Hum Genet* 2006;51:1126–32.
- 35. Plaza-Menacho I, Burzynski GM, de Groot JW, et al. Current concepts in RET-related genetics, signaling and therapeutics. *Trends Genet* 2006;22:627–36.
- Emison ES, McCallion AS, Kashuk CS, et al. A common sex-dependent mutation in a RET enhancer underlies Hirschsprung disease risk. *Nature* 2005;434:857–63.