

**ScienceDirect** 



# Current initiatives for the validation of analytical methods for botanicals<sup>\*</sup>

Paula N Brown and Patience Lister

The demand for validated analytical methods for botanicals has grown in response to the increasing consumer market for botanical supplements. Government initiatives to increase the availability of validated analytical methods and botanical reference material have led to the publication of numerous validation studies in scientific journals. Single laboratory validation and collaborative validation studies are structured to confirm a method's ruggedness and fit for purpose. The performance characteristics and statistical protocols followed throughout a validation study vary with the source of guidelines. Analytical techniques and priority methods are influenced by the need for fast-screening techniques, the limited availability of reference material, market value, and the prevalence of contaminants in botanical supplements.

#### Addresses

Natural Health and Food Products Research Group, British Columbia Institute of Technology, 3700 Willingdon Avenue, Burnaby, British Columbia V5G 3H2, Canada

Corresponding author: Brown, Paula N. (pbrown@bcit.ca, Paula\_Brown@bcit.ca)

Current Opinion in Biotechnology 2014, 25:124–128 This review comes from a themed issue on Analytical biotechnology

Edited by Frank L Jaksch and Savas Tay

For a complete overview see the Issue and the Editorial

Available online 18th December 2013

0958-1669 © 2013 The Authors. Published by Elsevier Ltd. Open access under CC BY-NC-ND license.

http://dx.doi.org/10.1016/j.copbio.2013.10.003

# Introduction

A growing consumer market for botanical supplements has surpassed the availability of reliable analytical methods to verify botanical identify, purity and strength. The lack of publicly available validated methods makes it difficult to assess product quality, both composition and stability, and has stymied scientific research on these products. This for validated methods is further driven by laws that require publicly available methods to enforce legal action against dietary supplements [1]. Initiatives have been taken in response to the need for validated analytical methods for botanical supplements. These involve collaborations between government, industry and private scientific organizations where scientists and industry members have been working to develop and validate standard analytical methods for dietary supplements [1]. Despite the U.S. Food and Drug Administration's (FDA) Current Good Manufacturing Practices (CGMP) for dietary supplements, the industry still suffers from botanical misidentification, product contamination and adulteration. The National Institutes of Health's (NIH) initiative to validate methods for priority dietary supplements drove AOAC International to adapt the traditional Official Methods process to include single laboratory validation (SLV). The scope of this review includes initiatives, guidance and current practice in the validation of analytical methods for botanicals.

#### Validation

Validation is an applied approach to verifying that a method is suitable and rugged enough to function as a quality control tool. AOAC International defines a validated method as a method that is fit for its intended purpose. The purpose may include quantifying a specific analyte in a product, confirming whether a product meets its specifications or regulations, identifying the presence of a nutrient or contaminant in a product, or identifying a product ingredient. Methods can be validated in a single laboratory or through a collaborative study in multiple laboratories (AOAC guidelines for single laboratory validation of chemical methods for dietary supplements and botanicals; URL: http://www.aoac.org/vmeth/SLV\_Guidelines\_Dietary%20Supplements.pdf).

#### Single laboratory validation

A Single Laboratory Validation (SLV) is the first step towards becoming an official method of analysis (OMA) through AOAC International. Once the method has passed a SLV, it is ready for a collaborative study between multiple laboratories. If the collaborative study is successful then the method may be considered for an OMA.

An analytical method should be fully developed and optimized before single laboratory validation. The purpose of the validation is to confirm performance parameters determined during development and should provide information on how it will perform under routine use. An unstable method may require re-validation [2]. If validation results do not meet the performance standards then the method may require further development and optimization. When possible, a validation should also be conducted as a collaborative study by multiple laboratories, on different instruments, reagents, and standards.

#### **Collaborative study**

The purpose of a collaborative study is to determine the reproducibility of performance characteristics when followed by different laboratories. Under AOAC International guidelines, this requires a minimum of 10 independent laboratories producing valid data for 12 replicates of each material. All samples are blinded and randomized [1]. Some methods are validated independently through small-scale inter-laboratory studies [3]. This can provide information on method ruggedness but will not lead to an OMA. A 20 laboratory collaborative study was recently conducted on the analysis of the mycotoxin ochratoxin A in licorice products. The method was considered successful based on meeting the LOD set out in EU legislation [4<sup>•</sup>].

#### **Reference material**

Chemical analysis requires reference points. Analytical methods for botanicals typically require reference materials with measurable physical properties that are used for comparison to the test materials. Chemical standards are a common form of reference material that can be purchased from chemical suppliers; however, if no reference material exists then a compound with similar properties can be used. As part of method development, reference materials should be assessed for identity, purity, stability, and storage conditions [1]. Botanical identification methods (BIM) require that the availability and identity of panel materials be verified [5"]. Some reference standards can be isolated in-house, as was done with baicalein-7-O-glucoside for the analysis of Semen oroxyli. Authors checked its purity by UV and NMR spectroscopy [6<sup>•</sup>].

Botanical reference materials, or voucher specimens, are preserved specimens that can be used to authenticate sample identification and can be sourced from herbariums [7,8]. When botanical material is collected from wild sources, a voucher specimen should be verified and saved for future reference [9]. Germplasm banks are another potential source of verified plant material [10<sup>\*</sup>].

# Performance characteristics

The performance characteristics of an analytical method include applicability (scope), selectivity, calibration, accuracy, repeatability precision, measurement uncertainty, variability, limit of detection (LOD), and limit of quantification (LOQ). They indicate the degree to which replicate measurements approach the 'true' values of a method's parameters. Other characteristics that should be measured during the method development and optimization stage include analyte stability, matrix effects, sensitivity, and ruggedness or robustness [1,9].

According to AOAC International, a chemical calibration curve should have six or more calibration points that span the relevant range. This practice was followed using six calibration levels ranging in concentration of 10–215  $\mu$ g/kg for validating a method for quantifying deoxynivale-nol-3-glucoside in processed cereal products [12]. An assessment was done to confirm the stability of cichoric acid during its extraction and analysis from Echinacea. This involved an exhaustive extraction procedure, storage at room temperature for six days, and 30 consecutive HPLC injections [13<sup>•</sup>].

Ruggedness is the degree to which a method's results can be reproduced under different conditions. The Youden Ruggedness Trial is a statistical tool used to identify how significantly each factor contributes to a method's variability. It involves small, deliberate changes to the procedure and then an assessment of the results [1]. The Youden Ruggedness Trial was used to examine the effect of seven parameters of an extraction method for cranberry anthocyanins. High and low parameters were examined for sample mass, sonication time, percent acid in extraction solvent, shaking time, sonication temperature, injection time, and centrifugation time [14<sup>••</sup>].

# **Statistical tools**

Current practices are now shifting over to design of experiment (DOE) for evaluating ruggedness. DOE is considered a time-efficient and cost-efficient technique used to simultaneously identify the effects of multiple factors on results.

Chemometrics is gaining significance as a data analysis tool and can be used for method development by identifying the effects of analytical conditions in factorial experiments. This tool was used in combination with DOE for the analysis of alkaloids in poppy straw. A 24 full factorial design was accomplished through 19 GC/ FID/MS method optimization experiments. Using DOE, the authors were able to identify the most effective parameters for rapid screening [15<sup>••</sup>]. Some authors forgo statistical methods for evaluating ruggedness and instead vary parameters individually to see which significantly affect results [16,17,19]. This can be a time consuming approach.

The Horwitz ratio (HorRat) is a statistical performance parameter that indicates the acceptability of method precision. It is a common criterion for validation of analytical methods under AOAC International protocols and has an acceptable value range of 0.3–1.3 for SLV data and 0.5-2.0 for collaborative study data (Definitions and calculations of HorRat values from intralaboratory data; URL: http://www.aoac.org/dietsupp6/Dietary-Supplement-web-site/HORRAT\_SLV.pdf). HorRat was a useful indicator of good overall method performance of a validated method for the analysis for Catechin and Epicatechin in cocoa [19].

# Leading organizations and government bodies

The ODS is a key government leader in the development and validation of analytical methods and reference materials for botanicals and dietary supplements. Goals of the ODS's Analytical Methods/Reference Materials Dietary Supplements Program (AMRM) are to build infrastructure for validation, support the development of validated methods, reference material, and calibration standards, and make validated methods and reference materials available to the community (Analytical methods/reference materials (AMRM) dietary supplements program description; URL: http://ods.od.nih.gov/ Research/AMRMProgramDescription.aspx).

AOAC International is a not-for profit organization standards body that develops program foundation and technical aspects of analytical methods. They coordinate scientific studies, evaluate results, give official sanction to applicable methods, and disseminate the methods or performance data to the public (Help the dietary supplement community create a reliable compendium of collaborative studied methods; URL: http://www.aoac.org/ dietsupp6/Dietary-Supplement-web-site/DSHomePage2.html). The AOAC's Presidential Task Force on Dietary Supplements identifies priority ingredients and selects methods for validation. They rank ingredients based on funding, market share of the ingredient, availability of methods to validate, related clinical trials, and ingredient safety concerns. AOAC's expert review panel (ERP) assists in this process by peer-reviewing the quality of existing methods and supporting data. The ERP recently reviewed methods for antioxidant activity of botanicals. They were evaluated against AOAC International's standard method performance requirements (SMPRs) and an ORAC method was approved as First Action Official Method [20].

Other organizations that provide SLV guidance include International Conference on Harmonization (ICH) and Brazil's Agencia Nacional de Vigilancia sanitaria (ANVISA) [21,22].

# **Priority methods and botanicals**

The ODS publishes a list of ODS-supported validated methods and botanical ingredients. Methods that are currently under development and awaiting validation include those for black cohosh, general botanicals, glycosides, pesticides, and valerian (Analytical Methods for Dietary Supplements; URL: http://ods.od.nih.gov/

#### Research/AMRMAnalyticalMethods.aspx#umethods).

AOAC's ERP prioritizes methods and analytes of interest for validation. In 2009 the ERP selected pyridoxine as the main analyte of interest after reviewing over 50 vitamin B6 methods [23<sup>••</sup>].

Ginseng is one of the top selling dietary supplements on the market and has been assessed in various clinical trials. This led to the execution of SLV and interlaboratory studies of an accurate and reliable method to identify and quantify ginsenosides in different ginseng species [24<sup>••</sup>,25<sup>•</sup>]. This method is now on tract to becoming an OMA by AOAC Int. OMAs can be extended to the analysis of new matrices, but should be validated to confirm applicability. Authors extended AOAC's Official Method 991.31 for the determination of Aflatoxins in corn, raw peanuts, and peanut butter to analysis of aflatoxins in botanical roots [26<sup>•</sup>].

Pesticide analysis is an important aspect of identifying contaminants and adulterants in botanicals. Although matrix effects pose challenges, the high presence of pesticides in botanicals has led to validation studies [27,28,29°]. The growing consumer interest in gluten-free foods has also driven the need to validate analytical test methods for gluten [30°].

Standardization of plant material is a critical aspect of botanical supplement quality. As a first step towards standardizing oenothein B in plants from the Oenotheraceae family, authors developed and validated a HPLC-DAD-MS method for its quantification [31]. In response to the FDA's request that supplement manufacturers adopt new technologies for measuring process attributes, authors validated a NIR method for quantification of chlorogenic acid in Lonicera japonica [32]. Interest in the bioavailability of botanical compounds has led to the validation of analytical methods on biological matrices, such as plasma [33]. In preparation for a Phase I clinical trial, a UHPLC/MS/MS method was validated for the quantification of prenylflavonoids in human serum. The method provides an accurate and precise way to measure clinical samples and support product efficacy [39<sup>••</sup>].

# **Analytical techniques**

Fast screening techniques and simple sample preparation are growing priorities intended to make methods more convenient and cost-effective [34<sup>•</sup>]. Authors combined this need with the interest in alkaloid analysis to develop and validate a rapid-screening method for simultaneous alkaloid analysis in poppy straw [35]. An HPTLC-densitometric method was developed and validated to meet the need for a high throughput procedure for routine quantification of primulasaponins [36<sup>•</sup>]. Derivatization procedures, which can increase sample preparation or run times, are being replaced by faster techniques when possible [37<sup>•</sup>].

Another motivation for increasing the efficiency of analytical methods is to conserve plant material. A 6-min UHPLC-QTOF-MS method was developed and validated for high-throughput analysis of huperzine A in a species of club moss. The authors validated the microscale extraction protocol and short analytical method with the goal of conserving plant resources [38<sup>•</sup>].

#### Conclusion

This review shows that the increasing consumer use of botanical supplements has led to initiatives and support for validating analytical methods to verify their identity, purity and strength. Through government, academic and industry initiatives, validated analytical methods and reliable reference material are becoming more prevalent and available to the public. Fast screening techniques and efficient use of limited study material are priorities in method design and optimization. Commercial value as well as the prevalence of pesticides and allergens in botanicals is leading the direction of validated analytical methods.

#### Acknowledgement

The authors would like to thank Grace Lai for her assistance in preparing the annotations.

#### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- · of special interest
- •• of outstanding interest
- International AOAC: Guidelines for dietary supplements and botanicals: Appendix K. AOAC Off Methods Anal 2012:1-32.
- Murphy CJ, MacNeil JD, Capar SG: Best practices for singlelaboratory validation of chemical methods for trace elements in foods. Part I—Background and general considerations. J AOAC Int 2013, 96:190-203.
- Kolberg DI, Mack D, Anastassiades M, Hetmanski MT, Fussell RJ, Meijer T, Mol HG: Development and independent laboratory validation of a simple method for the determination of paraquat and diquat in potato, cereals and pulses. Anal Bioanal Chem 2012, 404:2465-2474.
- Lerda D, Ambrosio M, Kunsagi Z, Stroka J: Determination of ochratoxin A in licorice and licorice extracts by highperformance liquid chromatography couples with fluorescence detection: collaborative study. J AOAC Int 2013, 96:331-340.

20 laboratories participated in a collaborative study on the determination of ochratoxin A in licorice root and extracts by HPLC. Recoveries ranged from 84 to 88% and the method identified as fit-for-purpose by the European Committee for Standardization and AOAC International.

- 5. AOAC International: AOAC international guidelines for
- validation of botanical identification methods. J AOAC Int 2012, 95:268-272.

This document provides technical guidelines for conducting AOAC validation studies on botanical identification methods. It covers relevant terms, definitions and validation study guidelines. This document could be used as a direct reference for designing SLV and collaborative studies.

 Yang X, Miao J, Ma X, Xiao P: Establishment and validation of an
 HPLC quantification method for quality control of semen oroxyli. J AOAC Int 2013, 96:27-32. Authors optimized and validated a HPLC method for the quantification of four flavonoids in *Semen oroxyli*. Validation parameters were assessed according to ICH guidelines. This paper describes validation results and data analysis in detail, and provides a good example of the SLV process.

- Baccarin T, Muceneeki RS, Bresolin TM, Yunes RA, Malheiros A, Lucinda-Silva RM: Development and validation of an HPLC-PDA method for the determination of myrsinoic acid B in the extracts of Rapanea ferruginea Mez. Talanta 2011, 85:1221-1224.
- Fucina G, Block LC, Baccarin T, Ribeiro TR, Quintao NL, Filho VC, Bresolin TM: Development and validation of a stability indicative HPLC-PDA method for kaurenoic acid in spray dried extracts of Sphagneticola trilobata (L.) Pruski, Asteraceae. Talanta 2012, 101:530-536.
- Nugroho A, Lim S, Lee CM, Choi JS, Park HJ, Lim SC: Simultaneous quantitative determination and validation of quercetin glycosides with peroxynitrite-scavenging effects from Saussurea grandifolia. J Pharm Biomed Anal 2012, 61:247-251.
- Pereira AB, Verissimo TM, Oliveira MA, Araujo IA, Alves RJ,
  Braga FC: Development and validation of an HPLC–DAD method for quantification of bornesitol in extracts from hancornia speciosa leaves after derivatization with ptoluenesulfonyl chloride. *J Chromatogr B* 2012, 887–888:133-137.

This article describes a validation study of a simple HPLC–DAD method for quantifying bornesitol in *Hancornia speciosa* leaves. Using a chemical derivatization reaction, this is the first HPLC–DAD protocol for the analysis of bornesitol in a botanical matrix.

- Suman M, Bergamini E, Catellani EB, Manzitti A: Development and validation of a liquid chrromatography/linear ion trap mass spectrometry method for the quantitative determination of deoxynivalenol-3-glucose in processed cereald products. Food Chem 2013, 136:1568-1576.
- Brown PN, Chan M, Paley L: Determination of major phenolic
  compounds in echinacea spp. raw material and finished products by high-performance liquid chromatography with ultraviolet detection: single-laboratory validation matrix extension. J AOAC Int 2011, 94:1400-1410.

This article describes a SLV study on caftaric acid, chlorogenic acid, cichoric acid, cynarin, and echinacoside from in *Echinacea* spp. Authors used a HPLC method quantify these phenolic compounds in products and raw materials and conducted studies to verify the stability of cichoric acid during extraction.

 Brown PB, Shipley PR: Determination of anthocyanins in
 cranberry fruit and cranberry fruit products by highperformance liquid chromatographywith ultraviolet detection: single-laboratory validation. J AOAC Int 2011, 94:459-466.

This article describes the development and SLV study of a HPLC–UV method for the quantification of five major anthocyanins in cranberry. Authors follow AOAC's SLV guidelines and provide detailed descriptions of stability studies and a Youden Ruggedness Trial. This article could be used as a reference for SLV design.

 Acevska J, Stefkov G, Petkovska R, Kulevanova A: Chemometric
 approach for development, optimization, and validation of different chromatographic methods for separation of opium alkaloids. Anal Bioanal Chem 2012, 403:1117-1129.

The authors used a chemometric approach to examine the effects of a full factorial linear design of experiments with chromatographic conditions. The optimized GC/FID/MS method was validated as a fast screening technique for six opium alkaloids from *Papaver somniferum* L.

- Marinho AF, Barbosa-Filho JM, Oliveira EJ: A validated method for the simultaneous quantification of bioactive alkaloid markers in the leaf ethanolic extract of *Cissampelos* sympodialis eichl: a phenological variation study. *Phytochem Anal* 2012, 23:426-432.
- Tiwari N, Luqman S, Masood N, Gupta MM: Validated high performance thin layer chromatographic method for simultaneous quantification of major iridoids in *Vitex trifolia* and their antioxidant studies. *J Pharm Biomed Anal* 2012, 61:207-214.
- Machonis PR, Jones MA, Schaneberg BT, Kwik-Uribe CL: Method for the determination of catechin and epicatechin enantiomers in cocoa-based ingredients and products by

high-performance liquid chromatography: single-laboratory validation. J AOAC Int 2012, 95:500-507

- 20. Harnley JM: Expert review panel approves first action methods for antioxidants in foods. J AOAC Int 2012, 95:1555-1556.
- 21. Olszewska MA: New validated hi-performance liquid chromatographic method for simultaneous analysis of ten flavonoid aglycones in plant extracts using a C18 fused-core column and acetonitrile-tetrahydrofuran gradient. J Sep Sci 2012, 35:2174-2183
- 22. Moraes AC, Bertanha CS, Gimenez VM, Groppo M, Silva ML, Cunha WR, Pauletti PM: Development and validation of a highperformance liquid chromatography method for quantification of egonol and homoegonol in Styrax species. Biomed Chromatogr 2011, 26:869-874.
- 23. Goldschmidt RJ. Wavne RW: Determination of pyridoxine in
- dietary supplements by liquid chromatography with UV, fluorescence, and mass spectrometric detection: single-laboratory validation. J of AOAC Int 2013, 96:265-275.

Authors conducted a SLV study on the analysis of pyridoxine in dietary supplements by LC with UV, fluorescence or MS detection. The method was designed for widespread use and adaptability to other chromatographic approaches.

- Brown PN: Determination of ginsenoside content in Asian and 24.
- North American ginseng raw materials and finished products by high-performance liquid chromatography: singlelaboratory validation. J AOAC Int 2011, 94:1391-1399.

Using base hydrolysis to convert ginsenosides into nonmalonylated counterparts, the authors validated a HPLC method for the identification and quantification of six ginsenosides in Asian and North American ginseng species according to AOAC guidelines. Quantitative determinations were performed with eight test materials by two analysts over three days (n = 12). RSDr values ranged from 1.11 to 7.61%.

- 25. Brown PN, Yu R: Determination of ginsenoside content in
- Panax ginseng C.A. Meyer and Panax quinquefolius L. root materials and finished products by high-performance liquid chromatography with ultraviolet absorbance detection: interlaboratory study. J AOAC Int 2013, 96:12-19.

14 laboratories participated in an inter-collaborative study on a validated method for the HPLC-UAD of ginsenosides in Asian and North American ginseng species. The AOAC International Interlaboratory Study Workbook was used to statistically evaluate the individual and total ginsenosides reported for each matrix. Acceptable results were achieved for total ginsenosides in whole root and powder extract; however, both finished products were considered inconsistently reproducible possibly due to incomplete hydrolysis.

- Weaver CM, Trucksess MW: Determination of aflatoxins in 26
- botanical roots by a modification of AOAC Official Method 991.31: single-laboratory validation. J AOAC Int 2010, 93: 184-189.

This article describes the modification of AOAC Official Method 991.31 for use in determining aflatoxins in botanical roots.

- 27. Lozano A, Rajski L, Belmonte-Valles N, Ucles A, Ucles S, Mezcua M, Fernandez-Alba AR: Pesticide analysis in teas and chamomile by liquid chromatography and gas chromatography tandem mass spectrometry using a modified QuEChERS method: validation and pilot survey in real samples. J Chromatogr A 2012, 1268:109-122.
- 28. Munitz MS, Resnik SL, Montti MI: Method development and validation for boscalid in blueberries by solid-phase microextraction gas chromatography, and their degradation kinetics. Food Chem 2013, 136:1399-1404.
- 29. Ruiz I, Morales A, Oliva J, Barba A: Validation of an analytical method for the quantification of pyrethrins on lemons and apricots using high-performance liquid chromatography/ mass spectrometry. *J Environ Sci Health B* 2011, 46:530-534.
   This article describes the validation of a LC-ESI-MS method for the

quantification of the pesticide pyrethrins in select fruit species. Validation parameters were based on SANCO guidelines.

30. Lupo A, Roebuck C, Walsh A, Mozola M, Abouzied M: Validation study of the Veratox R5 Rapid ELISA for detection of gliadin. AOAC Int 2013, 96:121-132

This article describes an AOAC International guided validation study on the Veratox R5 Giadin test kit for detection of gluten. This paper provides a detailed description of the validation results and analysis.

- 31. Granica S, Bazylki A, Kiss AK: Determination of macrocyclic ellagitannin oenothein B in plant material by HPLC-DAD-MS: method development and validation. Phytochem Anal 2012, 23:582-587
- 32. Wu Z, Xu B, Du M, Sui C, Sui C, Shi X, Qiao Y: Validation of a NIR quantification method for the determination of chlorogenic acid in Lonicera japonica solution in ethanol precipitation process. J Pharm Biomed Anal 2012, 62:1-6.
- 33. Rubio L, Serra A, Macia A, Borras X, Romero MP, Motilva MJ: Validation of determination of plasma metabolites derived from thyme bioactive compounds by improved liquid chromatography coupled to tandem mass spectrometry. J Chromatogr B 2012, 905:75-84.
- 34. Schoedl K, Forneck A, Sulyok M, Schuhmacher R: Optimization,
- in-house validation, and application of a liquid chromatography-tandem mass spectrometry (LC–MS/MS)based method for the quantification of selected polyphenolic compounds in leaves of grapevine (Vitis vinifera L.). J Agric Food Chem 2011, 59:10787-10794.

This paper describes an independent validation of a LC-MS/MS method for quantification of select polyphenols in grapevine leaves. Authors identify the method as simple and fast, and describe its use in identifying differences between plant species.

- 35. Acevska J, Dimitrovska A, Stefkov G, Brezovska K, Karapandzova M, Kulevanova S: Development and validation of a reversed-phase HPLC method for determination of alkaloids from Papaver somniferum L. J AOAC Int 2012, 95:399-405
- 36. Coran SA, Mulas S: Validated determination of
- primulasaponins in primula root by a high-performance-thinlayer-chromatography densitometric approach. J Pharm Biomed Anal 2012, **70**:647-651.

This article describes a novel HPTLC-densitometric method for separating and quantifying primulasaponin I and II in different matrices. Following validation guidelines for pharmaceutical analysis, the authors designed and validated the method for use in high-throughput routine applications.

37. Wang H, Duan JA, Guo S, Qian D, Shang E: Development and

validation of a hydrophilic interaction UHPLC-TQ-MS/MS method for absolute and relative quantification of amino acids in Sophora alopecuroides L. J Sep Sci 2013. (Epub ahead of print: 1-28).

This article describes the validation study for a hydrophilic interaction method for quantification of amino acids using ultra high performance liquid chromatography with tandem mass spectrometry. The authors claim that this method is more efficient than those with a derivatization step.

- 38. Cuthbertson D, Piljac-Zegarac J, Lange BM: Validation of a
- microscale extraction and high-throughput UHPLC-QTOF-MS analysis method for huperzine A in huperzia. Biomed Chromatogr 2012, 26:1191-1195.

This article describes the development and validation of a new UHPLC-QTOF-MS method for analysis of HupA from Huperzia. This method is ideal for rapid quantification of HupA in Huperzia regardless of plant availability.

- 39. Yuan Y, Qiu X, Nikolic D, Dahl JH, Breemen RB: Method
- development and validation for ultra-high-pressure LC/MS/ MS determination of hop prenylflavonoids in human serum. J AOAC Int 2012, 95:1744-1749.

This article describes the development and validation of a UPLC-MS/MS method for clinical analysis of hop prenylflavonoids in blood serum. It provides an example of the method validation process when applied to botanical chemicals in a biological fluid matrix.