

3059-Pos Board B164**Cardiac Myofibroblast-Myocyte Gap Junction Coupling Promotes After Depolarizations**

Thao P. Nguyen, Yuanfang Xie, Alan Garfinkel, Zhilin Qu, James N. Weiss. Tissue fibrosis, as seen in diseased and aged hearts, promotes ventricular arrhythmias. We hypothesize that de novo gap junction coupling between myofibroblasts, which proliferate following cardiac stress injury, with neighboring ventricular myocytes may facilitate arrhythmia triggers such as early afterdepolarizations (EADs). A novel hybrid computational and biological approach was employed: virtual fibroblasts with programmable properties embedded in the dynamic clamp were coupled to a real patch-clamped rabbit ventricular myocyte exposed to oxidative (0.1 mM H₂O₂) or ionic (2.7 mM K⁺) stressors. A virtual gap junction current of programmable conductance was added in real time to myocyte currents and their combined effects on myocyte action potentials and EAD genesis were evaluated. Exposure of myocytes to basal conditions failed to induce EADs, whereas exposure to stress led to 'bradycardia-dependent' EADs that occurred only during slow, but not during fast, pacing. However, when stressed myocytes were then coupled to a virtual fibroblast, EADs emerged independently of pacing rates. Fibroblast coupling alone to unstressed myocytes failed to induce EADs at any pacing rate. The virtual gap junction current has 2 components, but the earlier transient outward component was most critical for EAD generation; EADs disappeared when the virtual fibroblast was uncoupled from the myocyte during the initial 100 ms of the AP, but not when uncoupled for all but the first 100 ms of the AP. Our findings demonstrate that gap junction coupling of ventricular myocytes to myofibroblasts may directly induce EADs and the probability of EAD induction correlates with the myofibroblast-myocyte gap junction coupling strength. Elucidating the mechanism of myofibroblast-induced arrhythmogenesis may suggest new therapeutic strategies for preventing ventricular arrhythmias based on inhibiting fibroblast proliferation and/or uncoupling fibroblasts from myocytes.

3060-Pos Board B165**Targeted Inhibition of Connexin 43 Hemichannels Blunts Ca²⁺-Induced Intercellular Dyssynchrony and ATP Efflux in HL-1 Cardiomyocyte Syncytia**

Nicole C. Silvester, Hala Jundi, Archana Jayanthi, Steven R. Barberini-Jammaers, W Howard Evans, Christopher H. George. Gap junctions (GJ) are essential conduits that underpin cell-to-cell coupling between cardiomyocytes and are formed from the coalescence of connexin (Cx) hemichannels from two close opposed cells. However, there is evidence supporting a (patho)physiologic role for 'unpaired' Cx hemichannels as they traffic through the plasma membrane (PM) to the GJ. We used beating HL-1 cardiomyocyte monolayers as a model system for GJ intercellular communication (GJIC) and to investigate the functional role of Cx hemichannels in syncytial behaviour. GJIC was quantified as an index of intercellular Ca²⁺ release synchrony and was reconciled with detailed spatio-temporal analysis of intracellular Ca²⁺ signalling. Ouabain-evoked Ca²⁺ perturbation inhibited GJIC in a dose-dependent manner and was associated with reduced cell viability. We hypothesised that the intercellular dyssynchrony and cell death linked to ouabain-induced Ca²⁺ dysfunction was exacerbated by aberrant Cx hemichannel opening that may also compromise cellular metabolism. Consistent with this concept, the magnitude of intracellular Ca²⁺ flux dysfunction correlated with ATP release from cells. Trans-PM ATP leak was attenuated by Gap20, a peptide corresponding to an intracellular loop motif in Cx43. Moreover, Gap20 reduced intracellular Ca²⁺ perturbation, improved intercellular synchrony and reduced cell death in ouabain-treated syncytia. The lack of efficacy of a peptide corresponding to a similar epitope in Cx26, and also following the conjugation of Gap20 to a high molecular weight dextran confirmed i) the specificity of this approach and ii) that peptide bioactivity is dependent on its entry into cells and interaction with the intracellular face of Cx. Our data provides evidence that altered cellular Ca²⁺ homeostasis opens Cx hemichannels and that this may accelerate the metabolic deterioration of cardiomyocytes and exacerbate cardiac dysfunction in situ.

3061-Pos Board B166**Binding Kinetics of Inter-Connexon Interaction**

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Gap junctions are pairs of hexameric half-channels called connexons, which coaxially dock to connect two adjacent cells, mediating both adhesion and channeling between cells in many types of tissue. Connexons are formed of six connexin proteins (Cx). Gap junctions form by interdigitating the two extracellular loops of each connexin. While gap junction structure and function has been widely characterized using different techniques, the binding affinity of the inter-connexon interaction remains unknown. The goal of this work was

to determine the binding affinity of gap junctions using dynamic force spectroscopy atomic force microscopy (AFM). Among the residues that mediate inter-connexon interaction, an exposed stretch of conserved amino acids 'NTVD' within the extracellular loop 2 (E2) has been identified. For dynamic force spectroscopy, we covalently linked mimetic peptides 'NTVD' that mimic loop E2 of Cx26 to the AFM tip, while Cx26 two-dimensional (2D) crystals were immobilized on a mica substrate. We report the first characterization of the binding strength of the gap junction interaction. Force curves at various retraction speeds were acquired to determine the dissociation kinetics of the peptide-Cx26 interaction, while adhesion probability measurements at different contact times revealed the binding kinetics. The relatively fast intrinsic dissociation rate (k_{off}) inferred a rather dynamic inter-connexon interaction, while the slow association rate (k_{on}) probably reflects the restricted mobility and degrees of freedom of the connexons in the densely packed organization observed in native gap junction plaques and the reduced flexibility and dimensions of the extracellular loops. Our results suggest that gap junction formation may occur before plaque formation.

3062-Pos Board B167**A Stochastic Model of Voltage-Gating of Connexin-Based Gap Junction Channels Containing Fast and Slow Gates**

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Connexins (Cxs), a family of membrane proteins, form gap junction (GJ) channels that provide a direct pathway for electrical and metabolic cell-cell interaction. Each hemichannel in the GJ channel contains fast and slow gates sensitive to transjunctional voltage (V_j). The fast gate operates between open and residual states with conductances of $\gamma_{F,o}$ and $\gamma_{F,res}$, while the slow gate operates between open ($\gamma_{S,o}$) and fully closed ($\gamma_{S,closed}=0$) states; all γ s rectify but at different degree. We developed a stochastic 16-state model (S16SM), which extends earlier reported S4SM (Paulauskas et al., 2009) and accounts operation of four gates in series, instead of two, to describe the gating properties of homotypic and heterotypic GJ channels. Operation of each gate depends on the state of three other gates in series do to their effect on the fraction of V_j that falls across the gate (V_G) and is determined by equilibrium constants, $K_i = e^{A_i \cdot (II_i \cdot V_G - V_0)}$, where A_i characterizes the sensitivity to voltage, V_0 is the voltage for $K_i=1$ and II_i is gating polarity. S16SM allows to simulate kinetics of junctional current and junctional conductance (g_j) dependence on V_j for several frequently used experimental protocols: 1) consecutive V_j steps rising in amplitude, 2) slowly rising V_j ramps, and 3) series of V_j steps of high frequency. In addition, we have developed *universal* V_j protocol simulating freely selected forms of V_j consisting of an unlimited number of consecutively combined pulses and ramps of variable durations and amplitudes. The model was used to evaluate parameters of fast and slow gates of homo- and heterotypic GJs for experimentally measured g_j - V_j dependencies under normal/control and pathological conditions. The proposed S16SM was also used to evaluate gating properties of unapposed hemichannels residing in the non-junctional plasma membrane.

Voltage-gated K Channels - Permeation**3063-Pos Board B168****Ion Conduction in a Shaker Potassium Channel Mutant Having an Unusually High Single Channel Conductance**

David Naranjo, Ignacio Valencia, Cristian Moscoso, Katherine Stack, Valeria Márquez, Ariela Vergara, Marcos Sotomayor, Fernando González-Nilo.

K-channels are endowed of a highly K⁺ selective pore but are very diverse in single channel conductance, ranging from 3 to 300 pS. Because the selectivity filter is so conserved among K-channels, the origin of this diversity must be searched for in other parts of the pore. We studied the single-channel behavior of the Shaker K-channel with the P475D point mutation located at its internal entrance. In 100 mM K⁺ solution, this variant has a unitary conductance 8-10 fold larger than wild type (Sukhareva et al. 2003. J. Gen. Physiol. 122:541). We did single channel recording of the variants expressed in *Xenopus* oocytes and molecular dynamics simulations of channels modeled by homology with the Kv1.2-Kv2.1 chimera structure (Long et al. 2007. Nature. 450:376). Single channel conductance was measured in the interval -100/+150 mV between 50-1000 mM KMES. Below 300 mM KMES, single channel currents exhibit significant inward rectification, but at 1000 mM the I-V relation is nearly symmetrical. This channel is blocked by Mg²⁺ ions in a voltage independent fashion ($z\delta \sim$ zero) at 50 mM KMES, but at 1000 mM KMES, $z\delta$ approaches ~ 0.5 . These results suggest that internal K⁺ ions lock in Mg²⁺ inside the pore. To test, indirectly, the physical dimensions of the channel internal entrance, we