



# Genetics of liver disease: From pathophysiology to clinical practice

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## Summary

Paralleling the first 30 years of the *Journal of Hepatology* we have witnessed huge advances in our understanding of liver disease and physiology. Genetic advances have played no small part in that. Initial studies in the 1970s and 1980s identified the strong major histocompatibility complex associations in autoimmune liver diseases. During the 1990s, developments in genomic technologies drove the identification of genes responsible for Mendelian liver diseases. Over the last decade, genome-wide association studies have allowed for the dissection of the genetic susceptibility to complex liver disorders, in which also environmental co-factors play important roles. Findings have allowed the identification and elaboration of pathophysiological processes, have indicated the need for reclassification of liver diseases and have already pointed to new disease treatments. In the immediate future genetics will allow further stratification of liver diseases and contribute to personalized medicine. Challenges exist with regard to clinical implementation of rapidly developing technologies and interpretation of the wealth of accumulating genetic data. The historical perspective of genetics in liver

diseases illustrates the opportunities for future research and clinical care of our patients.

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## Introduction

Over the last 30 years, genetics has enhanced our understanding and management of liver disease (Table 1). The fundamental technologies available for genetic analysis encompass positional cloning of unknown disease genes, simple gene tests for known single nucleotide variants (SNV), genome-wide association studies (GWAS) that compare genotype frequencies across the whole genome between cases and controls to identify unknown genetic risk factors, and next-generation sequencing (NGS) of selected genes, all exons, or the whole genome in individual patients.

To illustrate the developments and opportunities of these genetic methods and technologies, we here pose and address five key questions that should help students, researchers, and health care professionals to understand the strides in the genetics of liver disease. Our answers cover both historical aspects and indicate how the advances in our understanding have led to a rapidly developing framework for diagnostic and therapeutic strategies. Rather than a comprehensive representation of the complete genetics of liver disease (for which comprehensive reviews can be found [41]), we aim to present the reader with key examples of the current state of the field and our associated reflections. We focus on germline mutations and deliberately omit the utility of genomics in the management of hepatocellular carcinoma and cholangiocarcinoma, which is also covered elsewhere [42] and in this special issue.

## Has genetics improved our understanding of the pathophysiology of liver diseases?

The first genes of monogenic diseases with predominant liver phenotypes that were mapped and cloned were the Wilson disease gene *ATP7B* (1993) and the haemochromatosis gene *HFE* (1996) (Table 1; Fig. 1). The Wilson disease gene was localized

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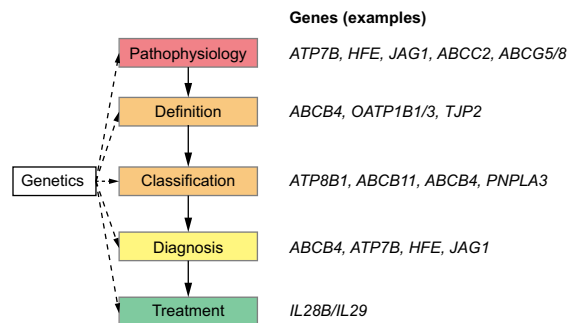
Abbreviations: SNV, single nucleotide variant; GWAS, genome-wide association study; NGS, next-generation sequencing; MHC, major histocompatibility complex; p, protein (amino acid position); SNP, single nucleotide polymorphism; ABC, ATP-binding cassette; PBC, primary biliary cirrhosis (cholangitis); PSC, primary biliary cholangitis; ICP, intrahepatic cholestasis of pregnancy; LPAC, low phospholipid-associated cholelithiasis; BSEP, bile salt export pump; OATP, organic anion transporting polypeptide; ASH, alcoholic steatohepatitis; NASH, non-alcoholic steatohepatitis; PASH, PNPLA3-associated steatohepatitis; TNF, tumour necrosis factor; IL, interleukin; IFN, interferon; HCV, hepatitis C virus; HSV, herpes simplex virus; DILI, drug-induced liver injury; HLA, human leukocyte antigen; WES, whole-exome sequencing; WGS, whole-genome sequencing.



**Key Points**

Different liver diseases are at different stages of the following sequence of steps:

- The first genetic discoveries identified the major loci for Mendelian diseases
- Numerous studies have identified susceptibility to complex diseases
- All diseases are actually complex with multiple genes contributing to the phenotype
- With the identification of different underlying genetic contributors to disease those diseases are being reclassified
- A better understanding of genetic aetiology will suggest new modes of treatment for disease
- A complete understanding of all the contributors to a individual's disease will allow personally tailored treatment



**Fig. 1. Scope of review.** With an emphasis on the developments of the last 30 years, the present article aims to discuss the implications of genetic studies in liver diseases onto relevant aspects of research and clinical hepatology. Topics covered range from descriptions of basic pathophysiology to practical aspects of diagnosis and treatment of the individual patient. The examples have been selected to illustrate the utility of genetic tools for specific aspects of clinical practice. For details, see text; for gene name abbreviations, see <http://www.ncbi.nlm.nih.gov/gene/>.

by linkage disequilibrium and haplotype analysis in more than 100 families [8]. Functional studies demonstrated that it encodes a P-type ATPase gene with metal binding regions similar to those found in prokaryotic heavy metal transporters [9]. Building on this discovery, subsequent studies were able to determine vast aspects of previously unknown aspects of copper transport and the pathophysiology of Wilson disease [43]. Using a similar strategy three years later, the *HFE* gene in the extended major histocompatibility complex (MHC) was shown to be mutated in patients with autosomal-recessive haemochromatosis [10]. Furthermore, additional types of non-*HFE* hereditary haemochromatosis were subsequently described, linked to mutations in the ferroportin gene (*SLC40A1*) [17,18], the transferrin receptor 2 gene (*TFR2*) [19], the hepcidin gene (*HAMP*) [20] and the hemojuvelin gene (*HJV*) [21], respectively. These genetic discoveries jointly paved the way for the full characterization of hepatic iron metabolism and its regulators [44], allowing for dissection of mechanisms responsible for the development of liver disease in the presence of detected mutations.

The mutation profile of disease genes varies. Hereditary haemochromatosis is most often caused by a predominant founder mutation (p.C282Y) which accounts for approximately 95% of patients [45]. Wilson disease shows a different genetic profile, with extensive allelic heterogeneity with more than 500 mutations described in the *ATP7B* gene to date. For this reason, genetic testing in haemochromatosis is undertaken via targeted genotyping of the predominant variants (always starting with p.C282Y), whereas in Wilson disease there is a need for gene sequencing and variant analysis to obtain genetic support for the diagnosis.

The first GWAS in hepatobiliary diseases was performed for gallstone disease and confirmed the candidacy of the hepatobiliary cholesterol transporter *ABCG5/G8* as a major susceptibility gene for gallstones worldwide, with p.D19H representing the likely causal single nucleotide polymorphism (SNP) [22,46]. Other rare loss-of-function SNVs in this transporter had been identified previously, in individual patients, to underlie the monogenic disease sitosterolemia, which is characterized by unrestricted intestinal absorption of both cholesterol and phytosterols such as sitosterol [47]. The adjacent, oppositely oriented *ABCG5/G8* genes encode two ATP-binding cassette (ABC) hemitransporters that are localized in the apical membranes of enterocytes and hepatocytes. The case also illustrates a

**Table 1. Present day technologies involved in the detection of disease relevant genetic variation.** These genomic technologies are principally deriving from a variety of sequencing techniques (targeting genes, the coding regions of the genome [exome] or the whole genome) and microarrays for genome-wide single nucleotide or copy number variant detection. Cost (price) and the availability of relevant bioinformatic tools also need considerations in making the ideal selection of method for investigating any observed trait.

	Mendelian diseases	Complex diseases
Degree of heritability	Relative sibling risk increased >200-1000 times compared with general population risk	Relative sibling risk increased 2-20 times compared with general population risk
Technology preferences	Whole-exome and whole-genome sequencing	Single nucleotide polymorphism arrays
Study design	Linkage analysis, <i>in silico</i> candidate variant filtering and prioritization, functional assessments of disease variants	Genome-wide case-control association analysis
Key challenges	Large number of candidate variants makes conclusive identification difficult, mechanistic support for causality needed and may require extensive studies	Low effect size of relevant variants means large numbers of patients and controls are needed to establish robust findings (typically thousands)

## Review

**Table 2. Four decades of genetic discoveries in liver diseases.** The table lists landmark articles on the genetic susceptibility to liver diseases. The listing is not comprehensive, but serves to show the development of the field over the timespan of the last 40 years. The attention of the field has shifted from the detection of the major histocompatibility complex (MHC) associations in autoimmune liver diseases via the elucidation of the genetics of Mendelian phenotypes to the broader dissection of the genetics of complex liver diseases by genome-wide association studies (GWAS).

Year	Landmark genetic publication	References
1972	MHC associations in autoimmune hepatitis	[1]
1979	MHC associations in primary biliary cirrhosis	[2]
1982	MHC associations in primary sclerosing cholangitis	[3]
1989	<i>CFTR</i> in cystic fibrosis	[4]
1992/95	<i>UGT1A1</i> in Gilbert and Crigler-Najjar syndromes	[5-7]
1993	<i>ATP7B</i> in Wilson disease	[8, 9]
1996	<i>HFE</i> in haemochromatosis (type 1)	[10]
1997	<i>ABCC2</i> in Dubin-Johnson syndrome	[11]
1997	<i>JAG1</i> in Alagille syndrome	[12,13]
1998	<i>ATP8B1</i> in PFIC (type 1)	[14]
1998	<i>ABCB11</i> in PFIC (type 2)	[15]
1998	<i>ABCB4</i> in PFIC (type 3)	[16]
2001	<i>SLC40A1</i> (ferroportin) in haemochromatosis (type 4)	[17,18]
2002	<i>TFR2</i> in haemochromatosis (type 3)	[19]
2003	<i>HAMP</i> in haemochromatosis (type 2B)	[20]
2004	<i>HJV</i> in haemochromatosis (type 2A)	[21]
2006	<i>NOTCH2</i> in Alagille syndrome	[12]
2007	<i>ABCG8</i> in gallstone disease (GWAS)	[22]
2008	<i>PNPLA3</i> in non-alcoholic fatty liver disease (GWAS)	[23]
2008	First GWAS on genetic factors associated with plasma liver enzyme activities	[24]
2009	First GWAS in primary biliary cirrhosis	[25]
2009	MHC associations in flucloxacillin DILI (GWAS)	[26]
2009	MHC associations in HBV clearance (GWAS)	[27]
2009	<i>IL28B</i> in HCV treatment response (GWAS)	[28]
2009	<i>IL28B</i> in spontaneous HCV clearance (GWAS)	[29]
2009	Genetic modifiers for <i>CFTR</i> -associated liver disease	[30]
2010	<i>PNPLA3</i> in alcoholic liver disease	[31]
2010	First GWAS in primary sclerosing cholangitis	[32]
2010	<i>ITPA</i> in ribavirin-induced anaemia (GWAS)	[33]
2010	First GWAS in hepatitis B-related hepatocellular carcinoma (GWAS)	[34]
2011	MHC associations in amoxicillin-clavulanate DILI (GWAS)	[35]
2012	<i>OATP1B1</i> and <i>OATP1B3</i> in Rotor syndrome	[36]
2012	First GWAS on liver fibrosis in chronic HCV infection	[37]
2014	First GWAS in autoimmune hepatitis	[38]
2014	<i>TJP2</i> in PFIC (type 4)	[39]
2014	MHC associations in hepatitis B vaccine response (GWAS)	[40]

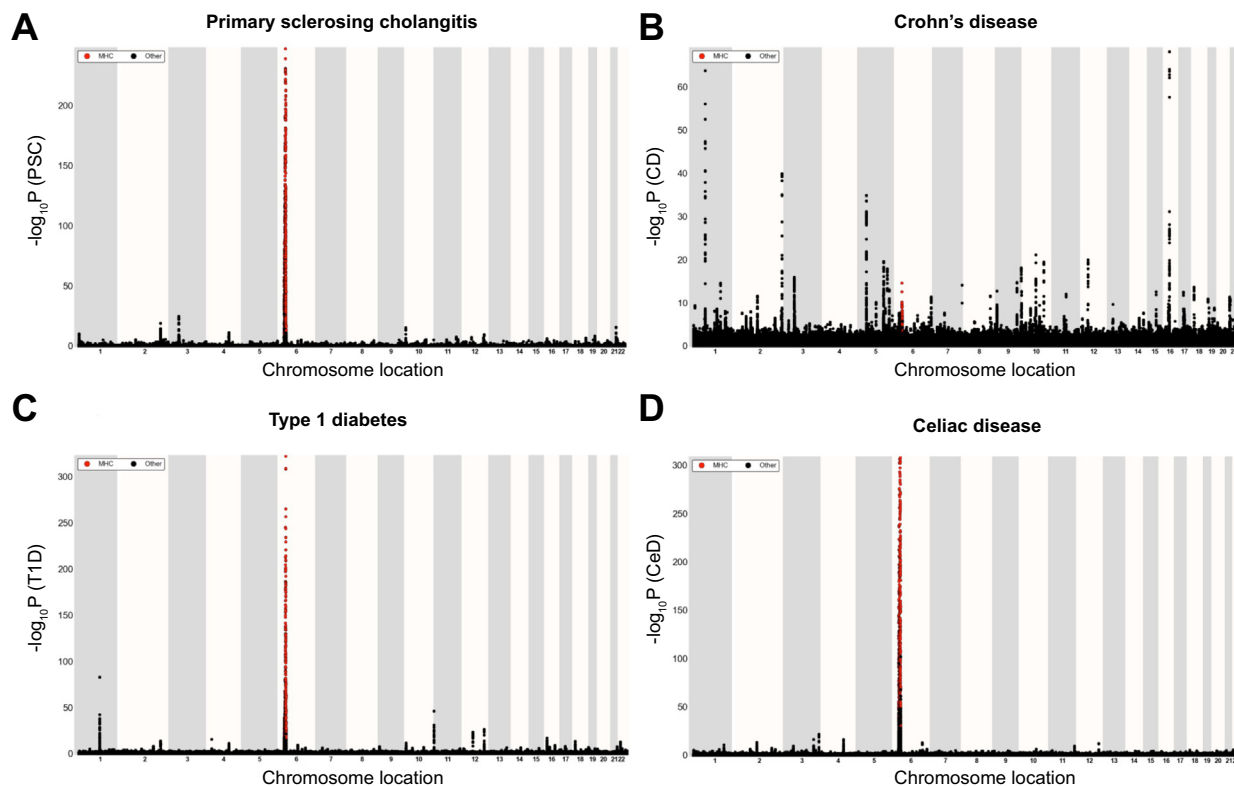
DILI, drug-induced liver injury; HBV, hepatitis B virus; HCV, hepatitis C virus; PFIC, progressive intrahepatic cholestasis. For gene name abbreviations, see <http://www.ncbi.nlm.nih.gov/gene/>.

fundamental difference between risk factors in monogenic vs. polygenic/complex traits (Table 2). For sitosterolemia, the SNVs appear to be sufficient to cause the observed phenotype. For risk alleles of SNPs detected by GWAS (that have an odds ratio of approximately 2), interacting genetic and environmental factors are required for gallstone development [46,48,49].

GWAS for autoimmune liver diseases have led to the detection of several dozen risk genes in primary biliary cirrhosis (PBC) [25], primary sclerosing cholangitis (PSC) [32,50] and recently, autoimmune hepatitis (AIH) [37]. With few exceptions, the detected risk loci overlap with those of other autoimmune and immune-mediated diseases (Fig. 2) [51-53]. This means that the majority of the genetic risk factors for these phenotypes

involves an increasing susceptibility to autoimmunity rather than liver affections per se. A large fraction of the risk loci and the strong MHC associations found in both PBC, PSC, and AIH point to the pathophysiological importance of adaptive immune responses [46]. The overall genetic contribution to disease susceptibility in PBC, PSC, and AIH is relatively low, with the current risk gene pool representing less than 10% of the overall disease liability [51,52]. Larger studies will increase this fraction, but more than 50% of the susceptibility to these diseases is still likely to be of an environmental origin.

Individual whole-exome sequencing (WES) has been successfully used to identify unknown genetic variants for inherited diseases or conditions of indeterminate aetiology such



**Fig. 2. Genetic architecture of complex diseases.** Genetic studies have unambiguously pinpointed a role for adaptive immune responses in causing primary sclerosing cholangitis (PSC, panel A). The overall genetic architecture of PSC shows similar features as prototypical autoimmune diseases (e.g., type 1 diabetes, panel C) and only a partial overlap with that of inflammatory bowel disease (panel B). Diseases caused by distinct exogenous factors (e.g., drug-induced liver injury, celiac disease, panel D) show a similar genetic architecture, where a strong association with genes in the major histocompatibility complex (MHC, plotted in red) is the defining feature. Plots derived from data in reference [52]; the X axis shows chromosomal location, the Y axis shows association statistic for investigated variants (individual dots).

as acute liver failure in children [54]. Recently this has been illustrated in two families with unsuspected mitochondrial hepatopathies, where WES of germline DNA indicated homozygous mutations in *MPV17* and *SERAC1* as causative in paediatric cases of acute liver failure [55]. In fact, liver disease-targeted re-sequencing of genes has been implemented in several liver centres and we believe that it ultimately will be fully integrated into care and get a firm place in the diagnostic evaluation of patients with unexplained liver disease, in particular in the setting of a strong family history of similar afflictions. Due to the massive amounts of data being generated, establishing the pathophysiological connection for detected variants within a clinically relevant time frame remains a major challenge.

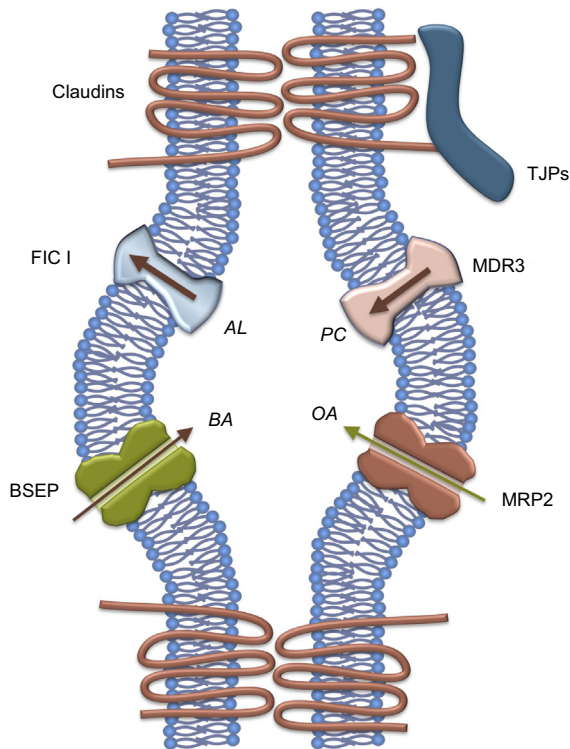
**Is genetics helpful for the definition of liver diseases and disease subtypes?**

Fifty years ago, Byler disease was first described in an Amish kindred [56]. The same disorder in non-Amish populations was termed Byler syndrome, which represented a heterogeneous group of autosomal-recessive liver diseases characterized by early onset of cholestasis that progresses to hepatic fibrosis, cirrhosis, and end-stage liver disease often before adulthood. In the past two decades basic research teams identified and cloned hepatobiliary transport proteins that are involved in the apical uptake and canalicular secretion of bile acids as well as other

endo- and xenobiotics (Fig. 3) [57,58]. The landmark study by Smit *et al.* [59] demonstrated that homozygous disruption of the murine *Mdr2* P-glycoprotein gene, now known as *Abcb4*, leads to a complete absence of phosphatidylcholine from bile and chronic cholangiopathy. Massive gene amplification and rearrangements also cause hepatocellular cancer in this model [60]. Studies in humans demonstrated that the phenotypic consequences of mutations of the orthologous human gene *MDR3/ABCB4* range from neonatal cholestasis to progressive familial intrahepatic cholestasis (PFIC) and cirrhosis in adults [16,61]; they can also result in intrahepatic cholestasis of pregnancy (ICP) [62] as well as intrahepatic and gallbladder cholesterol cholelithiasis, termed low phospholipid-associated cholelithiasis (LPAC) syndrome [63,64].

The related gene *ABCB11* encodes the bile salt export pump (BSEP) and was shown to be mutated in consanguineous families with PFIC but generally low serum  $\gamma$ -glutamyl transferase activities [15,65]. In contrast, patients with Byler disease and others were shown to carry mutations in the *ATP8B1 (FIC1)* gene [14,66] which encodes a P-type ATPase that flips phosphatidylserine from the outer to the inner leaflet of the hepatocanalicular membrane, thereby maintaining membrane integrity [67]. Studies comparing the phenotypic presentation and progression between BSEP and FIC1 deficiencies [68,69] reported that the former patients were more likely to develop neonatal cholestasis and manifested more severe hepatobiliary disease (as indicated by higher serum ALT activities and bile salt

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**Fig. 3. Major molecular proteins involved in genetic cholestasis.** Multidrug resistance-associated protein 2 (MRP2) transports organic anion (OA) conjugates, including bilirubin, and is mutated in Dubin–Johnson syndrome. Bile salt export pump (BSEP) is the major bile acid (BA) transporter. Multidrug resistance 3 (MDR3) flops phosphatidylcholine (PC) and familial intrahepatic cholestasis protein 1 (FIC1) flips aminophospholipids (AL). TJP proteins are involved in maintaining the stability of integral tight junction proteins, including claudins.

concentrations) as well as cholelithiasis and hepatocellular carcinoma [60,70], while the latter showed greater evidence of extrahepatic manifestations (malabsorption, pancreatitis, pneumonia, hearing loss). Most recently a fourth PFIC gene has been described. Mutations in *TJP2*, encoding tight junction protein 2 (or zona occludens 2), were shown to lead to early onset liver disease [38]. In fact, although all four genes were found to underlie PFIC, each has now been shown to be responsible for spectra with severe and mild phenotypes [22,66,71–73].

Genetic studies have also clarified the pathophysiology of Dubin–Johnson and Rotor syndrome, which are due to mutations of hepatocellular bilirubin transporter genes: mutations of the human canalicular multispecific organic anion transporter gene (now termed *ABCC2*) underlie Dubin–Johnson syndrome [11], whereas combined deficiency of the basolateral organic anion transporting polypeptide (OATP) 1B1 and OATP1B3 results in Rotor syndrome by interrupting conjugated bilirubin uptake into the liver [36]. In contrast, mutations in the promoter or exons of the bilirubin conjugating enzyme *UGT1A1* underlie hereditary unconjugated hyperbilirubinaemia, with the frequent and benign Gilbert–Meulengracht syndrome being due to a common promoter mutation; more severe mutations cause Crigler–Najjar syndrome in childhood. Unconjugated hyperbilirubinaemia is a known risk factor for gallstone disease, explaining the robust genetic association of the Gilbert–Meulengracht variant and cholelithiasis [74,75].

In 1997, Oda *et al.* [76] and Li *et al.* [77] demonstrated that another hereditary cholestatic liver disease, Alagille syndrome, is caused by mutations in the human homolog of *Jagged-1* (*JAG1*), which encodes a ligand in the Notch signalling pathway. Alagille syndrome is a rare autosomal-dominant disease with patients showing not only paucity of intrahepatic bile ducts, but characteristic dysmorphic facies and cardiac anomalies, notably peripheral pulmonary stenosis, along with posterior embryotoxon and notched or butterfly vertebrae as well as failure to thrive. In less than 1% of patients, the disease is caused by *NOTCH2* mutations [12,13], and the phenotypes are now known to be modified by other genes such as thrombospondin 2 [78].

### How might genetics contribute to the classification of liver diseases?

Usually fatty liver disease is attributed to alcohol or metabolic stress, resulting in the terms alcoholic steatohepatitis (ASH) or non-alcoholic steatohepatitis (NASH) [79]. However, ASH may present as acute alcoholic hepatitis or chronic steatohepatitis, and both alcohol and environmental factors (over-nutrition, physical inactivity) can contribute to liver disease. In addition, chemotherapy and other drugs might also cause steatohepatitis. Interestingly, only a subgroup of patients (10–20%) who drink alcohol and/or are overweight develop severe steatohepatitis, advanced fibrosis, and cirrhosis. Since 2008, GWAS-based studies have led to the identification of a major genetic determinant of fatty liver and steatohepatitis as well as progression to fibrosis, cirrhosis, and hepatocellular cancer. Romeo *et al.* [23] identified the amino acid substitution p.I148M of the lipid droplet-associated triglyceride lipase *PNPLA3* (also termed adiponutrin) to confer susceptibility to non-alcoholic fatty liver disease. It appears also to be the major genetic risk factor aggravating alcoholic liver disease [31,80]. This SNP is highly prevalent in European populations (risk allele frequency 21–28%) and confers a 3-fold risk for liver disease and NASH as well as a 12-fold risk for hepatocellular cancer in comparison to healthy controls [81]. The effect is modified by other genetic risk factors such as *TM6SF2* p.E167K [82] but the population-attributable risk for fatty liver disease is highest for *PNPLA3* p.I148M [81]. Therefore, we have proposed to use this novel genetic information to define a group of patients with fatty liver disease in whom the susceptibility gene *PNPLA3* appears to be a major driver of disease progression, often in combination with ethanol and Western diet but also without, hence the suggested name *PNPLA3*-associated steatohepatitis (“PASH”) [83]. Although the *PNPLA3* risk genotypes confers a high risk of NASH, the patients appear to be sensitive to the beneficial effects of lifestyle modification and dietary intervention should be encouraged to do so [84,85].

Common, conserved, aetiopathogenic pathways have been proposed across diseases, often related to failed resolution of responses to infections or stress and damage caused by the patient’s immune system [86]. Recently a new cytokine-based taxonomy has been proposed for chronic inflammatory diseases, comprising tumour necrosis factor (TNF)- $\alpha$ , interleukin (IL)-1, IL-6, IL-17, and IL-23 associated diseases, which are characterized by inflammation in different organs such as gut, skin, joints and brain [87]. Preclinical studies and randomised controlled trials with targeted cytokine intervention in humans indicate the existence of a cytokine hierarchy that was used to define these novel

disease groups. For example, IL-1 is predominantly involved when inflammasome activation is the dominant mechanism of inflammation, a situation that arises in gout and rare genetic autoinflammatory syndromes such as cryopyrin-associated periodic syndrome and familial Mediterranean fever. Hence, these disorders might be grouped together as IL-1-associated diseases. The fact that the accumulation of metabolic substrates such as urate, cholesterol, ceramide, and glucose can trigger the inflammasome provides the rationale for current therapeutic trials targeting IL-1 in metabolic liver diseases such as fatty liver disease and alcoholic hepatitis.

The excessive pleiotropy of autoimmune risk variants provides another example of the imperfect relationship between traditional disease classifications on one side and molecular entities on the other. For autoimmune liver diseases, there are shared features (e.g. *SH2B3* variants associating with PBC, PSC and AIH) as well as pathways predominantly associating more on either side of the spectrum (e.g. IL-2-related associations in PSC and IL-12-related associations in PBC) [53]. The importance of these observations as for clinical implementation is still not clear and it is important to acknowledge that phenomenology by means of clinical manifestations (e.g. sclerosing cholangitis) or traditional biochemistry (including autoantibodies) may still represent key biology upon which these apparently diverse associations converge. In Crohn's disease, several susceptibility genes (*ATG16L1*, *NOD2*, and *IRGM*) appear to involve Paneth cell dysfunction as a common denominator for ileal disease affection [88], exemplifying how the clinical features may still be as valid a representation of key pathophysiology as a gene based classification. As such, data on genetic background and genetic risk factors are likely never to replace disease definitions by medical history and clinical investigations, but may provide another layer of information with a strong potential to refine patient management.

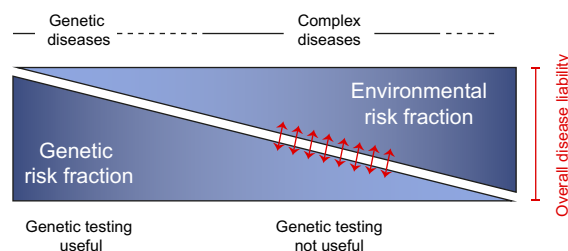
**Does genetics affect the treatment of liver disease?**

GWAS studies have led to the identification of variants in the *IL28B* gene encoding interferon (IFN)- $\lambda$ 3 that are associated with response to antiviral IFN-based therapy in patients with chronic hepatitis C virus (HCV) infection [28]. Additional studies showed that these polymorphisms are linked with a diplotype of the previously unknown IFN- $\lambda$ 4 (*IL29*) [89] that is induced by HCV and leads to an IFN-stimulated gene expression; a two-base pair insertion or the mutation p.P70S results in loss or reduced activity of IFN- $\lambda$ 4, respectively [90,91]. Of note, the mechanisms involving the IFN- $\lambda$  regulatory axis are not HCV-specific but also determine the outcome of infections with other viruses such as herpes simplex virus type 1 (HSV-1) infection causing vesicular oral and skin lesions, keratitis, or encephalitis [92]. This opens new insights and suggests integrative studies investigating the interaction between the innate immune response, including IFN- $\lambda$ 4, and T cells [93]. In light of the new potent direct antiviral agents, the clinical relevance of *IL28B* and *IL29* gene variation has diminished, but it also determines spontaneous clearance of HCV infection [29,94]. Of note, the gene variants are also associated with the severity of viral hepatitis, even with some contrasting findings [95], highlighting that the effects of gene polymorphisms can be pleiotropic, increasing the complexity of clinical phenotype, genetic background, and environmental factors [96]. Along the same line, variants of the NASH risk factors *PNPLA3* and *TM6SF2* have also been shown to modulate cardiovascular risk [97,98].

Drug-induced liver injury (DILI) represents a clinically diverse spectrum. Hyman J. Zimmerman discriminated direct and idiosyncratic hepatotoxicity [99]. Whereas direct hepatotoxic drugs lead to dose-related liver injury that is reproducible in animal models, idiosyncratic liver injury shows an unpredictable outcome and is rare. The clinical manifestation of idiosyncratic DILI is acute hepatitis, and to date the diagnosis has been made by exclusion [100]. Idiosyncratic DILI cases are most often caused by antibiotics and generic drugs that have been approved decades ago. In terms of aetiology, DILI is not a single, uncommon disease but rather many, rare diseases converging on a similar presentation of immune-mediated liver injury. A GWAS for a common type of DILI caused by amoxicillin-clavulanate exposure showed the strongest finding in the MHC region, with signals localizing to human leukocyte antigen (HLA) class I and II genes. Although the respective HLA genotypes (HLA-A\*02:01 and DQB1\*06:02) have limited utility as predictive biomarkers because of low positive-predictive values, they can post hoc serve to assess causality in individual severe cases. Furthermore, the MHC associations point to the involvement of adaptive immunity, likely representing immune reactions to the drug itself, to drug metabolites, or to drug-generated neoantigens (protein adducts), any of which can trigger immunity in otherwise susceptible individuals. Such mechanisms might even be amendable to early treatment with steroids or other types of immunosuppression [101,102]. To date, additional GWAS have indicated only a limited contribution of common genetic variants outside of the MHC to risk of DILI [103].

**What are the future hurdles and perspectives for translation of genetic research into clinics?**

The best strategy for the transformation of the large number of genetic findings into novel therapeutic opportunities for complex liver diseases is not known. Almost 40 years after the discovery of the gene defect in cystic fibrosis [4], genetic information provides information on the future disease course of individual patients, and variant-specific treatment is being deployed [104]. In PBC, an attempt was made to target the IL-12-related genetic findings by ustekinumab, a monoclonal antibody which targets the p40 subunit of IL-12 and IL-23. The study was terminated due to lack of efficacy (<https://clinicaltrials.gov/ct2/show/NCT01389973>) and serves to exemplify the point that genetic findings do not



**Fig. 4. Distinguishing Mendelian from complex liver diseases.** For complex phenotypes, the contribution from genetics to overall disease liability is limited (typically less than 50%). In addition, only a fraction (typically 10–20%) of the genetic susceptibility is known. In both Mendelian and complex disease manifestations the gene findings serve clues as to the underlying pathophysiology. Only for the case of Mendelian liver affections do genetic findings have clinically useful predictive power. Modified from [109].

## Review

necessarily identify therapeutic targets. It is also clear that for the complex phenotypes, genetic findings are also not useful for disease prediction (genetic testing) (Fig. 4) [105,106]. Both these aspects pertain to the lack of knowledge on interacting environmental (and other genetic) factors and the genuine function of the relevant gene products. In our opinion, the main purpose of genetics in complex phenotypes is to specify experimental research into disease pathophysiology ("pathway detection"). A challenge in forming relevant hypotheses for experimental and translational research following the application of GWAS is that most of the research into the relevant biology has been performed without the prior knowledge of the liver disease associations. Surprising and novel insights are likely to arise from correctly designed experiments on relevant models and human material, exemplified by the unravelling of IL-29 in HCV clearance [89]. These efforts are now urgently needed and will be the source of genuine clinical relevance and therapeutic applications – which so far are lacking for the gene findings per se.

Evident from the above discussions, for monogenic and even oligogenic liver diseases, genetic testing is already relevant. Here, the obstacles to clinical implementation are that of access to technology and associated expertise. WES has now been used extensively, and successfully, for research. Whole-genome sequencing (WGS) has advantages and disadvantages in comparison to WES. There is complete genomic coverage, sample preparation is easier, and there is less bias in the generated sequence [107]. Yield, in terms of a conclusive diagnosis is therefore higher [108]. The costs of WGS are decreasing rapidly, but major hurdles to widespread use of either technique for routine diagnostics remains in the interpretation of data. A number of commercial and mainly research-oriented software packages are now available, but none are perfect and there is no integration with the regular patient information systems. The most critical issue remains however the huge number of variants found in each individual. Within these variants lie not only the "cause" of Mendelian diseases, but, as discussed above, disease-modifiers and predictors of treatment response. Since disease severity varies even for "classic" monogenic liver diseases (Table 1), it is difficult to tell the difference between a susceptibility allele and a disease modifier, and ultimately every disease is more or less complex (Fig. 4). We simply do not yet have the knowledge and tools to implement most of the new data in clinical practice. The only way we will be able to approach this information, and thereby get closer to a vision of "personalized medicine", is through large cohort studies incorporating detailed clinical and molecular phenotyping alongside the WGS data. Such efforts are needed now more than ever.

### Conflict of interest

The authors declared that they do not have anything to disclose regarding funding or conflict of interest with respect to this manuscript.

### References

- [1] Mackay IR, Morris PJ. Association of autoimmune active chronic hepatitis with HL-A1.8. *Lancet* 1972;2:793–795.
- [2] Ercilla G, Pares A, Arriaga F, Bruguera M, Castillo R, Rodes J, et al. Primary biliary cirrhosis associated with HLA-DRw3. *Tissue Antigens* 1979;14:449–452.
- [3] Schruppf E, Fausa O, Forre O, Dobloug JH, Ritland S, Thorsby E. HLA antigens and immunoregulatory T cells in ulcerative colitis associated with hepatobiliary disease. *Scand J Gastroenterol* 1982;17:187–191.
- [4] Kerem B, Rommens JM, Buchanan JA, Markiewicz D, Cox TK, Chakravarti A, et al. Identification of the cystic fibrosis gene: genetic analysis. *Science* 1989;245:1073–1080.
- [5] Koiwai O, Nishizawa M, Hasada K, Aono S, Adachi Y, Mamiya N, et al. Gilbert's syndrome is caused by a heterozygous missense mutation in the gene for bilirubin UDP-glucuronosyltransferase. *Hum Mol Genet* 1995;4:1183–1186.
- [6] Bosma PJ, Chowdhury JR, Huang TJ, Lahiri P, Elferink RP, Van Es HH, et al. Mechanisms of inherited deficiencies of multiple UDP-glucuronosyltransferase isoforms in two patients with Crigler–Najjar syndrome, type I. *FASEB J* 1992;6:2859–2863.
- [7] Moghrabi N, Clarke DJ, Boxer M, Burchell B. Identification of an A-to-G missense mutation in exon 2 of the UGT1 gene complex that causes Crigler–Najjar syndrome type 2. *Genomics* 1993;18:171–173.
- [8] Petrukhin K, Fischer SG, Pirastu M, Tanzi RE, Chernov I, Devoto M, et al. Mapping, cloning and genetic characterization of the region containing the Wilson disease gene. *Nat Genet* 1993;5:338–343.
- [9] Bull PC, Thomas GR, Rommens JM, Forbes JR, Cox DW. The Wilson disease gene is a putative copper transporting P-type ATPase similar to the Menkes gene. *Nat Genet* 1993;5:327–337.
- [10] Feder JN, Gnirke A, Thomas W, Tsuchihashi Z, Ruddy DA, Basava A, et al. A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat Genet* 1996;13:399–408.
- [11] Paulusma CC, Kool M, Bosma PJ, Scheffer GL, ter Borg F, Scheper RJ, et al. A mutation in the human canalicular multispecific organic anion transporter gene causes the Dubin–Johnson syndrome. *Hepatology* 1997;25:1539–1542.
- [12] McDaniell R, Warthen DM, Sanchez-Lara PA, Pai A, Krantz ID, Piccoli DA, et al. NOTCH2 mutations cause Alagille syndrome, a heterogeneous disorder of the notch signaling pathway. *Am J Hum Genet* 2006;79:169–173.
- [13] Kamath BM, Bauer RC, Loomes KM, Chao G, Gerfen J, Hutchinson A, et al. NOTCH2 mutations in Alagille syndrome. *J Med Genet* 2012;49:138–144.
- [14] Bull LN, van Eijk MJ, Pawlikowska L, DeYoung JA, Juijn JA, Liao M, et al. A gene encoding a P-type ATPase mutated in two forms of hereditary cholestasis. *Nat Genet* 1998;18:219–224.
- [15] Strautnieks SS, Bull LN, Knisely AS, Kocoshis SA, Dahl N, Arnell H, et al. A gene encoding a liver-specific ABC transporter is mutated in progressive familial intrahepatic cholestasis. *Nat Genet* 1998;20:233–238.
- [16] De Vree JM, Jacquemin E, Sturm E, Cresteil D, Bosma PJ, Aten J, et al. Mutations in the MDR3 gene cause progressive familial intrahepatic cholestasis. *Proc Natl Acad Sci U S A* 1998;95:282–287.
- [17] Montosi G, Donovan A, Totaro A, Garuti C, Pignatti E, Cassanelli S, et al. Autosomal-dominant hemochromatosis is associated with a mutation in the ferroportin (SLC11A3) gene. *J Clin Invest* 2001;108:619–623.
- [18] Njajou OT, Vaessen N, Joosse M, Berghuis B, van Dongen JW, Breuning MH, et al. A mutation in SLC11A3 is associated with autosomal dominant hemochromatosis. *Nat Genet* 2001;28:213–214.
- [19] Girelli D, Bozzini C, Roetto A, Alberti F, Daraio F, Colombari R, et al. Clinical and pathologic findings in hemochromatosis type 3 due to a novel mutation in transferrin receptor 2 gene. *Gastroenterology* 2002;122:1295–1302.
- [20] Roetto A, Papanikolaou G, Politou M, Alberti F, Girelli D, Christakis J, et al. Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat Genet* 2003;33:21–22.
- [21] Papanikolaou G, Samuels ME, Ludwig EH, MacDonald ML, Franchini PL, Dube MP, et al. Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat Genet* 2004;36:77–82.
- [22] Buch S, Schafmayer C, Völzke H, Becker C, Franke A, von Eller-Eberstein H, et al. A genome-wide association scan identifies the hepatic cholesterol transporter ABCG8 as a susceptibility factor for human gallstone disease. *Nat Genet* 2007;39:995–999.
- [23] Romeo S, Kozlitina J, Xing C, Pertsemlidis A, Cox D, Pennacchio LA, et al. Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet* 2008;40:1461–1465.
- [24] Yuan X, Waterworth D, Perry JR, Lim N, Song K, Chambers JC, et al. Population-based genome-wide association studies reveal six loci influencing plasma levels of liver enzymes. *Am J Hum Genet* 2008;83:520–528.
- [25] Hirschfeld GM, Liu X, Xu C, Lu Y, Xie G, Lu Y, et al. Primary biliary cirrhosis associated with HLA, IL12A, and IL12RB2 variants. *N Engl J Med* 2009;360:2544–2555.

- [26] Daly AK, Donaldson PT, Bhatnagar P, Shen Y, Pe'er I, Floratos A, et al. HLA-B\*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. *Nat Genet* 2009;41:816–819.
- [27] Kamatani Y, Wattanapokayakit S, Ochi H, Kawaguchi T, Takahashi A, Hosono N, et al. A genome-wide association study identifies variants in the HLA-DP locus associated with chronic hepatitis B in Asians. *Nat Genet* 2009;41:591–595.
- [28] Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, et al. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 2009;461:399–401.
- [29] Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, et al. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 2009;461:798–801.
- [30] Bartlett JR, Friedman KJ, Ling SC, Pace RG, Bell SC, Bourke B, et al. Genetic modifiers of liver disease in cystic fibrosis. *JAMA* 2009;302:1076–1083.
- [31] Tian C, Stokowski RP, Kershenovich D, Ballinger DG, Hinds DA. Variant in PNLPA3 is associated with alcoholic liver disease. *Nat Genet* 2010;42:21–23.
- [32] Karlsen TH, Franke A, Melum E, Kaser A, Hov JR, Balschun T, et al. Genome-wide association analysis in primary sclerosing cholangitis. *Gastroenterology* 2010;138:1102–1111.
- [33] Fellay J, Thompson AJ, Ge D, Gumbs CE, Urban TJ, Shianna KV, et al. ITPA gene variants protect against anaemia in patients treated for chronic hepatitis C. *Nature* 2010;464:405–408.
- [34] Zhang H, Zhai Y, Hu Z, Wu C, Qian J, Jia W, et al. Genome-wide association study identifies 1p36.22 as a new susceptibility locus for hepatocellular carcinoma in chronic hepatitis B virus carriers. *Nat Genet* 2010;42:755–758.
- [35] Lucena MI, Molokhia M, Shen Y, Urban TJ, Aithal GP, Andrade RJ, et al. Susceptibility to amoxicillin-clavulanate-induced liver injury is influenced by multiple HLA class I and II alleles. *Gastroenterology* 2011;141:338–347.
- [36] Van de Steeg E, Stranecky V, Hartmannova H, Noskova L, Hrebicek M, Wagenaar E, et al. Complete OATP1B1 and OATP1B3 deficiency causes human Rotor syndrome by interrupting conjugated bilirubin reuptake into the liver. *J Clin Invest* 2012;122:519–528.
- [37] Patin E, Kutalik Z, Guernon J, Bibert S, Nalpas B, Jouanguy E, et al. Genome-wide association study identifies variants associated with progression of liver fibrosis from HCV infection. *Gastroenterology* 2012;143:1244–1252.
- [38] De Boer YS, van Gerven NM, Zwiers A, Verwer BJ, van Hoek B, van Erpecum KJ, et al. Genome-wide association study identifies variants associated with autoimmune hepatitis type 1. *Gastroenterology* 2014;147:e445.
- [39] Sambrotta M, Strautnieks S, Papouli E, Rushton P, Clark BE, Parry DA, et al. Mutations in TJP2 cause progressive cholestatic liver disease. *Nat Genet* 2014;46:326–328.
- [40] Pan L, Zhang L, Zhang W, Wu X, Li Y, Yan B, et al. A genome-wide association study identifies polymorphisms in the HLA-DR region associated with non-response to hepatitis B vaccination in Chinese Han populations. *Hum Mol Genet* 2014;23:2210–2219.
- [41] Lammert F. Hereditary liver diseases. Preface. *Best Pract Res Clin Gastroenterol* 2010;24:529.
- [42] Pinyol R, Nault JC, Quetglas IM, Zucman-Rossi J, Llovet JM. Molecular profiling of liver tumors: classification and clinical translation for decision making. *Semin Liver Dis* 2014;34:363–375.
- [43] Rosencrantz R, Schilsky M. Wilson disease: pathogenesis and clinical considerations in diagnosis and treatment. *Semin Liver Dis* 2011;31:245–259.
- [44] Hentze MW, Muckenthaler MU, Andrews NC. Balancing acts: molecular control of mammalian iron metabolism. *Cell* 2004;117:285–297.
- [45] Aguilar-Martinez P, Grandchamp B, Cunat S, Cadet E, Blanc F, Nourrit M, et al. Iron overload in HFE C282Y heterozygotes at first genetic testing: a strategy for identifying rare HFE variants. *Haematologica* 2011;96:507–514.
- [46] Hirschfield GM, Chapman RW, Karlsen TH, Lammert F, Lazaridis KN, Mason AL. The genetics of complex cholestatic disorders. *Gastroenterology* 2013;144:1357–1374.
- [47] Berge KE, Tian H, Graf GA, Yu L, Grishin NV, Schultz J, et al. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. *Science* 2000;290:1771–1775.
- [48] Buil A, Brown AA, Lappalainen T, Vinuela A, Davies MN, Zheng HF, et al. Gene-gene and gene-environment interactions detected by transcriptome sequence analysis in twins. *Nat Genet* 2015;47:88–91.
- [49] Henriksen EK, Melum E, Karlsen TH. Update on primary sclerosing cholangitis genetics. *Curr Opin Gastroenterol* 2014;30:310–319.
- [50] Folseraas T, Melum E, Rausch P, Juran BD, Ellinghaus E, Shiryaev A, et al. Extended analysis of a genome-wide association study in primary sclerosing cholangitis detects multiple novel risk loci. *J Hepatol* 2012;57:366–375.
- [51] Liu JZ, Almarri MA, Gaffney DJ, Mells GF, Jostins L, Cordell HJ, et al. Dense fine-mapping study identifies new susceptibility loci for primary biliary cirrhosis. *Nat Genet* 2012;44:1137–1141.
- [52] Liu JZ, Hov JR, Folseraas T, Ellinghaus E, Rushbrook SM, Doncheva NT, et al. Dense genotyping of immune-related disease regions identifies nine new risk loci for primary sclerosing cholangitis. *Nat Genet* 2013;45:670–675.
- [53] Hirschfield GM, Karlsen TH. Genetic risks link autoimmune hepatitis to other autoimmune liver disease. *Gastroenterology* 2014;147:270–273.
- [54] Wright CF, Brice P, Stewart A, Burton H. Realising the benefits of genetics for health. *Lancet* 2010;376:1370–1371.
- [55] Vilarinho S, Choi M, Jain D, Malhotra A, Kulkarni S, Pashankar D, et al. Individual exome analysis in diagnosis and management of paediatric liver failure of indeterminate aetiology. *J Hepatol* 2014;61:1056–1063.
- [56] Clayton RJ, Iber FL, Ruebner BH, McKusick VA. Byler disease. Fatal familial intrahepatic cholestasis in an Amish kindred. *Am J Dis Child* 1969;117:112–124.
- [57] Hagenbuch B, Meier PJ. Molecular cloning, chromosomal localization, and functional characterization of a human liver Na<sup>+</sup>/bile acid cotransporter. *J Clin Invest* 1994;93:1326–1331.
- [58] Gerloff T, Stieger B, Hagenbuch B, Madon J, Landmann L, Roth J, et al. The sister of P-glycoprotein represents the canalicular bile salt export pump of mammalian liver. *J Biol Chem* 1998;273:10046–10050.
- [59] Smit JJ, Schinkel AH, Oude Elferink RP, Groen AK, Wagenaar E, van Deemter L, et al. Homozygous disruption of the murine mdr2 P-glycoprotein gene leads to a complete absence of phospholipid from bile and to liver disease. *Cell* 1993;75:451–462.
- [60] Iannelli F, Collino A, Sinha S, Radaelli E, Nicoli P, D'Antiga L, et al. Massive gene amplification drives paediatric hepatocellular carcinoma caused by bile salt export pump deficiency. *Nat Commun* 2014;5:3850.
- [61] Jacquemin E, De Vree JM, Cresteil D, Sokal EM, Sturm E, Dumont M, et al. The wide spectrum of multidrug resistance 3 deficiency: from neonatal cholestasis to cirrhosis of adulthood. *Gastroenterology* 2001;120:1448–1458.
- [62] Jacquemin E, Cresteil D, Manouvrier S, Boute O, Hadchouel M. Heterozygous non-sense mutation of the MDR3 gene in familial intrahepatic cholestasis of pregnancy. *Lancet* 1999;353:210–211.
- [63] Rosmorduc O, Hermelin B, Poupon R. MDR3 gene defect in adults with symptomatic intrahepatic and gallbladder cholesterol cholelithiasis. *Gastroenterology* 2001;120:1459–1467.
- [64] Poupon R, Rosmorduc O, Boelle PY, Chretien Y, Corpechot C, Chazouilleres O, et al. Genotype-phenotype relationships in the low-phospholipid-associated cholelithiasis syndrome: a study of 156 consecutive patients. *Hepatology* 2013;58:1105–1110.
- [65] Strautnieks SS, Byrne JA, Pawlikowska L, Cebecauerova D, Rayner A, Dutton L, et al. Severe bile salt export pump deficiency: 82 different ABCB11 mutations in 109 families. *Gastroenterology* 2008;134:1203–1214.
- [66] Klomp LW, Vargas JC, van Mil SW, Pawlikowska L, Strautnieks SS, van Eijk MJ, et al. Characterization of mutations in ATP8B1 associated with hereditary cholestasis. *Hepatology* 2004;40:27–38.
- [67] Groen A, Romero MR, Kunne C, Hoosdally SJ, Dixon PH, Wooding C, et al. Complementary functions of the flippase ATP8B1 and the floppase ABCB4 in maintaining canalicular membrane integrity. *Gastroenterology* 2011;141:e1921–e1924.
- [68] Pawlikowska L, Strautnieks S, Jankowska I, Czubkowski P, Emerick K, Antoniou A, et al. Differences in presentation and progression between severe FIC1 and BSEP deficiencies. *J Hepatol* 2010;53:170–178.
- [69] Davit-Spraul A, Fabre M, Branchereau S, Baussan C, Gonzales E, Stieger B, et al. ATP8B1 and ABCB11 analysis in 62 children with normal gamma-glutamyl transferase progressive familial intrahepatic cholestasis (PFIC): phenotypic differences between PFIC1 and PFIC2 and natural history. *Hepatology* 2010;51:1645–1655.
- [70] Knisely AS, Strautnieks SS, Meier Y, Stieger B, Byrne JA, Portmann BC, et al. Hepatocellular carcinoma in ten children under five years of age with bile salt export pump deficiency. *Hepatology* 2006;44:478–486.
- [71] Van Mil SW, van der Woerd WL, van der Brugge G, Sturm E, Jansen PL, Bull LN, et al. Benign recurrent intrahepatic cholestasis type 2 is caused by mutations in ABCB11. *Gastroenterology* 2004;127:379–384.
- [72] Carlton VE, Harris BZ, Puffenberger EG, Batta AK, Knisely AS, Robinson DL, et al. Complex inheritance of familial hypercholesterolemia with associated mutations in TJP2 and BAAT. *Nat Genet* 2003;34:91–96.



## Review

- [73] Stapelbroek JM, van Erpecum KJ, Klomp LW, Houwen RH. Liver disease associated with canalicular transport defects: current and future therapies. *J Hepatol* 2010;52:258–271.
- [74] Wasmuth HE, Keppeler H, Herrmann U, Schirin-Sokhan R, Barker M, Lammert F. Coinheritance of Gilbert syndrome-associated UGT1A1 mutation increases gallstone risk in cystic fibrosis. *Hepatology* 2006;43:738–741.
- [75] Buch S, Schafmayer C, Völzke H, Seeger M, Miquel JF, Sookoian SC, et al. Loci from a genome-wide analysis of bilirubin levels are associated with gallstone risk and composition. *Gastroenterology* 2010;139:e1942.
- [76] Oda T, Elkahloun AG, Pike BL, Okajima K, Krantz ID, Genin A, et al. Mutations in the human Jagged1 gene are responsible for Alagille syndrome. *Nat Genet* 1997;16:235–242.
- [77] Li L, Krantz ID, Deng Y, Genin A, Banta AB, Collins CC, et al. Alagille syndrome is caused by mutations in human Jagged1, which encodes a ligand for Notch1. *Nat Genet* 1997;16:243–251.
- [78] Loomes KM, Tsai E, Underkoffler LA, Grochowski C, Falsey AM, Kamath BM, et al. A genome-wide association study identifies THBS2 as a candidate modifier of liver disease severity in Alagille syndrome. *Hepatology* 2014;60:252A.
- [79] Nobili V, Svegliati-Baroni G, Alisi A, Miele L, Valenti L, Vajro P. A 360-degree overview of paediatric NAFLD: recent insights. *J Hepatol* 2013;58:1218–1229.
- [80] Stickel F, Buch S, Lau K, Meyer zu Schwabedissen H, Berg T, Ridinger M, et al. Genetic variation in the PNPLA3 gene is associated with alcoholic liver injury in caucasians. *Hepatology* 2011;53:86–95.
- [81] Krawczyk M, Stokes CS, Romeo S, Lammert F. HCC and liver disease risks in homozygous PNPLA3 p. I148M carriers approach monogenic inheritance. *J Hepatol* 2015;62:980–981.
- [82] Liu YL, Reeves HL, Burt AD, Tiniakos D, McPherson S, Leathart JB, et al. TM6SF2 rs58542926 influences hepatic fibrosis progression in patients with non-alcoholic fatty liver disease. *Nat Commun* 2014;5:4309.
- [83] Krawczyk M, Portincasa P, Lammert F. PNPLA3-associated steatohepatitis: toward a gene-based classification of fatty liver disease. *Semin Liver Dis* 2013;33:369–379.
- [84] Sevastianova K, Kotronen A, Gastaldelli A, Perttilä J, Hakkarainen A, Lundbom J, et al. Genetic variation in PNPLA3 (adiponutrin) confers sensitivity to weight loss-induced decrease in liver fat in humans. *Am J Clin Nutr* 2011;94:104–111.
- [85] Shen J, Wong GL, Chan HL, Chan RS, Chan HY, Chu WC, et al. PNPLA3 gene polymorphism and response to lifestyle modification in patients with nonalcoholic fatty liver disease. *J Gastroenterol Hepatol* 2015;30:139–146.
- [86] Medzhitov R, Schneider DS, Soares MP. Disease tolerance as a defense strategy. *Science* 2012;335:936–941.
- [87] Schett G, Elewaut D, McInnes IB, Dayer JM, Neurath MF. How cytokine networks fuel inflammation: toward a cytokine-based disease taxonomy. *Nat Med* 2013;19:822–824.
- [88] Adolph TE, Tomczak MF, Niederreiter L, Ko HJ, Bock J, Martinez-Naves E, et al. Paneth cells as a site of origin for intestinal inflammation. *Nature* 2013;503:272–276.
- [89] Hamming OJ, Terczynska-Dyla E, Vieyres G, Dijkman R, Jorgensen SE, Akhtar H, et al. Interferon lambda 4 signals via the IFNlambda receptor to regulate antiviral activity against HCV and coronaviruses. *EMBO J* 2013;32:3055–3065.
- [90] Prokunina-Olsson L, Muchmore B, Tang W, Pfeiffer RM, Park H, Dickensheets H, et al. A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus. *Nat Genet* 2013;45:164–171.
- [91] Bibert S, Roger T, Calandra T, Bochud M, Cerny A, Semmo N, et al. IL28B expression depends on a novel TT/-G polymorphism which improves HCV clearance prediction. *J Exp Med* 2013;210:1109–1116.
- [92] Aka PV, Kuniholm MH, Pfeiffer RM, Wang AS, Tang W, Chen S, et al. Association of the IFNL4-DeltaG allele with impaired spontaneous clearance of hepatitis C virus. *J Infect Dis* 2014;209:350–354.
- [93] Griffiths SJ, Koegl M, Boutell C, Zenner HL, Crump CM, Pica F, et al. A systematic analysis of host factors reveals a Med23-interferon-lambda regulatory axis against herpes simplex virus type 1 replication. *PLoS Pathog* 2013;9:e1003514.
- [94] Estrabaud E, Vidaud M, Marcellin P, Asselah T. Genomics and HCV infection: progression of fibrosis and treatment response. *J Hepatol* 2012;57:1110–1125.
- [95] Abe H, Ochi H, Maekawa T, Hayes CN, Tsuge M, Miki D, et al. Common variation of IL28 affects gamma-GTP levels and inflammation of the liver in chronically infected hepatitis C virus patients. *J Hepatol* 2010;53:439–443.
- [96] Krawczyk M, Müllenbach R, Weber SN, Zimmer V, Lammert F. Genome-wide association studies and genetic risk assessment of liver diseases. *Nat Rev Gastroenterol Hepatol* 2010;7:669–681.
- [97] Holmen OL, Zhang H, Fan Y, Hovelson DH, Schmidt EM, Zhou W, et al. Systematic evaluation of coding variation identifies a candidate causal variant in TM6SF2 influencing total cholesterol and myocardial infarction risk. *Nat Genet* 2014;46:345–351.
- [98] Kahali B, Liu YL, Daly AK, Day CP, Anstee QM, Speliotes EK. TM6SF2: catch-22 in the fight against nonalcoholic fatty liver disease and cardiovascular disease? *Gastroenterology* 2015 [epub].
- [99] Zimmerman HJ. Hepatotoxicity: the adverse effects of drugs and other chemicals on the liver. Lippincott Williams & Wilkins; 1999, 789.
- [100] Chalasani NP, Hayashi PH, Bonkovsky HL, Navarro VJ, Lee WM, Fontana RJ. ACG Clinical Guideline: the diagnosis and management of idiosyncratic drug-induced liver injury. *Am J Gastroenterol* 2014;109:950–966, quiz 967.
- [101] Weiler-Normann C, Schramm C. Drug induced liver injury and its relationship to autoimmune hepatitis. *J Hepatol* 2011;55:747–749.
- [102] Urban TJ, Daly AK, Aithal GP. Genetic basis of drug-induced liver injury: present and future. *Semin Liver Dis* 2014;34:123–133.
- [103] Urban TJ, Shen Y, Stolz A, Chalasani N, Fontana RJ, Rochon J, et al. Limited contribution of common genetic variants to risk for liver injury due to a variety of drugs. *Pharmacogenet Genomics* 2012;22:784–795.
- [104] Cutting GR. Cystic fibrosis genetics: from molecular understanding to clinical application. *Nat Rev Genet* 2015;16:45–56.
- [105] Clayton DG. Prediction and interaction in complex disease genetics: experience in type 1 diabetes. *PLoS Genet* 2009;5:e1000540.
- [106] Karlsen TH, Melum E, Franke A. The utility of genome-wide association studies in hepatology. *Hepatology* 2010;51:1833–1842.
- [107] Meynert AM, Ansari M, FitzPatrick DR, Taylor MS. Variant detection sensitivity and biases in whole genome and exome sequencing. *BMC Bioinformatics* 2014;15:247.
- [108] Gilissen C, Hehir-Kwa JY, Thung DT, van de Vorst M, van Bon BW, Willemssen MH, et al. Genome sequencing identifies major causes of severe intellectual disability. *Nature* 2014;511:344–347.
- [109] Mells GF, Kaser A, Karlsen TH. Novel insights into autoimmune liver diseases provided by genome-wide association studies. *J Autoimmun* 2013;46:41–54.