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Molecular cloning and sequence determination of four different cDNA species coding for α -subunits of G proteins from *Xenopus laevis* oocytes

Juan Olate¹, Sixta Martinez², Patricia Purcell¹, Hugo Jorquera¹, Juan Codina³, Lutz Birnbaumer³ and Jorge E. Allende¹

¹Departamento de Bioquímica, Facultad de Medicina, Universidad de Chile, Casilla 70086, Santiago 7, Chile, ²Departamento de Química, Universidad Nacional, Bogotá, Colombia and ³Department of Cell Biology, Baylor College of Medicine, Texas Medical Center, Houston, TX 77030, USA

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A cDNA library preprared from Xenopus laevis oocytes in λ gt10 was screened with a mixture of three oligonucleotide probes designed to detect sequences found in different mammalian genes coding for α -subunits of G-proteins. In addition to a clone coding for a G α o-type subunit previously reported [(1989) FEBS Lett. 244, 188–192] four additional clones have been found coding for different G α protein subunits. By comparison with mammalian α -subunits, these oocyte cDNAs correspond to two closely related G α s-1a, to a G α i-1 and to a G α i-3 species. The derived amino acid sequences showed that both G α s species contain 379 residues, corresponding to the short species without the serine residue and with a calculated M_r of 42720. The G α i-1 gene encodes a 354 amino acid protein with an M_r of 39000 and the G α i-3 encodes an incomplete open reading frame of 345 residues, lacking the first 9 amino acid residues at the NH₂ terminus. All these G α -subunits showed high identity with their respective mammalian counterparts (75–80%), indicating a great degree of conservation through the evolution and the important cellular regulatory function that they play.

G-protein; cDNA cloning; Nucleotide sequence; Xenopus laevis oocyte

1. INTRODUCTION

The transduction of many external signals towards the interior of the cells involves trimeric proteins that bind guanine nucleotides and that are known as Gproteins [1]. There is a large family of these proteins since more than 16 different G-proteins have been isolated from different species and tissues [2-5]. Although the function of some of these G-proteins has been elucidated in particular signal transduction pathways, there are still many questions open as to the role that each one of these may play in different systems.

The Xenopus laevis oocyte has become a popular system for researchers studying the function of receptors and who have isolated mRNAs coding for these receptor proteins. The reason for this popularity is the fact that the oocyte microinjected with these mRNAs has shown itself to be capable of both translating these receptors and also of coupling the newly synthesized receptors to transducing systems. The microinjected oocyte thus acquires the capacity to respond physiologically to the agonist that binds to that particular receptor [6–9].

It has become important, therefore, to study the endogenous transducing systems of the oocyte in order to be able to determine the entities that participate in the mechanism of action of various signals and that couple to their respective effector systems. These considerations have induced us to clone the genes coding for different G-proteins that are expressed in this amphibian oocyte. In a previous communication [10], we reported the cloning of the cDNA coding for the oocyte $G\alpha$ otype subunit which showed a high degree of identity to the mammalian $G\alpha$ o.

In this report, we present the cloning and sequencing of four other different cDNAs from *Xenopus laevis* oocytes coding for α -subunits highly analogous to $G\alpha$ s, $G\alpha$ i-1 and $G\alpha$ i-3 of mammalian systems. These results indicate that this single cell type has at least 5 different types of G-proteins.

2. MATERIALS AND METHODS

2.1. cDNA library

A Xenopus laevis oocyte cDNA library constructed in the vector gt10 (kindly donated by Dr D.A. Melton of Harvard University) was utilized [11].

2.2. Screening of the cDNA library

Close to 2×10^5 recombinant plaques were screened by plaque hybridization [12] with three synthetic probes labeled at the 5'-end with ³²P. The probes used for this purpose and their respective se-

Correspondence address: J. Olate, Departamento de Bioquímica, Facultad de Medicina, Universidad de Chile, Casilla 70086, Santiago 7, Chile

quences were the same utilized in the previous work [10]. Phages from five positive lytic plaques from the first screening round were plaque-purified through secondary and tertiary screening. Four of these clones were fully sequenced. Hybridizations were done overnight at 40°C in a solution containing $6 \times SSC$, pH 6.8, 100 μ g of heat-denatured salmon sperm DNA per ml and 0.1% SDS. Filters were washed three times at 40°C in $6 \times SSC$ and then three times at 45° C for 15 min. Films were exposed overnight at room temperature.

2.3. cDNA sequence analysis

Nucleotide sequences were determined by using the M13mp19 and the dideoxynucleotide chain-termination method [13] as described in the 'Sequenase' booklet provided by the US Biochemical Corp. (USB).

3. RESULTS

3.1. Screening of the $\lambda gt10$ Xenopus laevis oocyte cDNA library

Using the same strategy described previously by Olate et al. we screened about 2×10^5 lysis plaques and five positive clones were obtained. All these clones were subjected to a secondary and tertiary screening, their DNAs purified and finally analyzed by nucleotide sequencing of the cDNA inserts following the strategy shown in Fig. 1.

3.2. Nucleotide sequences of the $G\alpha i$ -1 cDNA clone

Fig. 2 shows the nucleotide sequence of the cDNA encoding the G α i-1 type α -subunit. The sequence is 2759 nucleotides long and predicts an open reading frame of a 354 amino acid protein (M_r 40200), with a 5'-untranslated region of 184 nucleotides and a 3'-untranslated region of 1510 nucleotides. The sequence contains two poly(A) addition sequences, AATAA, at positions 1979 and 2539 and it ends with a 14 residue poly(A) tail. The deduced amino acid sequence showed a high degree of identity (85%) with the human G α i-1 [14].

3.3. Nucleotide sequence of the $G\alpha i$ -3 cDNA clone

Fig. 3 shows the nucleotide sequence analysis of an insert with great similarity to human $G\alpha$ i-3. The sequence is 2037 nucleotides long and it predicts a single open reading frame of 1035 nucleotides coding for a continuous sequence of a 345 amino acid protein. Since no ATG initiation codon was found, we assume that this sequence contains the partial sequence for a $G\alpha$ -type protein missing a short stretch of the NH₂-terminus. At the protein level, comparison of corresponding residues between the oocyte and human $G\alpha$ i



Fig. 1. Sequencing strategy and partial restriction endonuclease map for the *Xenopus laevis* $G\alpha$ cDNAs. The top scale indicates cDNA length in nucleotides. Open boxes show the open reading frames (ORF) for the different proteins. The thin black lines show the 5'- and 3'-untranslated regions of the mRNA. The arrows indicate extent and direction of sequencing obtained with the oligonucleotide primers. All the cDNA inserts were sequenced in their complete length after subcloning them into the M13mp19 vector. The numbers in parentheses correspond to the length of each cDNA. The restriction endonuclease sites are denoted by one letter. B, *Bam*H1; H, *Hind*III; N, *NcI*1; E, *Eco*R1; Bc, *BcI*1; S, *Sph*1; Bs, *Bst*1.

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AAC	AGG	TCA	AGA	GAA	TAT	CAG	стс	AAT	GAC	TCT	GCA	GCA	TAT	TAT	CTT	AAC	GAT	TTG	GAC		
Asn	Arg	Ser	Arg	Glu	Tyr	Gln	Leu	Asn	Asp	Ser	Ala	Ala	Tyr	Tyr	Leu	Asn	Asp	Leu	Asp		
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			-					2575													

Fig. 2. Nucleotide and predicted amino acid sequence of the cDNA insert for G α i-1. Numbers indicate the position of nucleotides or amino acid residues starting at the initiation codon. Sequences enclosed in open boxes correspond to the poly(A) addition sequences. Nucleotide sequence recognized by the common 27-base probe is underlined [10].

proteins indicated a high degree of identity (87%) with the human $G\alpha i$ -3 [15], so this protein was classified as a $G\alpha i$ -3 oocyte protein. The sequence has a 3'-untranslated region of 1002 nucleotides, it contains a poly(A) addition sequence at position 2010 followed by a stretch of 38 adenosines.

3.4. Nucleotide sequence of a $G\alpha s$ -type cDNA

Fig. 4 shows the nucleotide and predicted amino acid

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61	-	-	-	-	-	-	-	-	Arg 30	Glu	Ala	Ala	Glu	Arg	Ser	Lys	Met	Ile	Asp
ÇĞG	AAC	CTT	CGG	GAA	GAT	GGG	GAA	AAG	GCA	TCC	AAG	GAG	GTG	AAA	CTG	CTG	CTA	CTC	GGT
Arg 121	Asn	Leu	Arg	Glu	Asp	Gly	Glu	Lys	Ala 50	Ser	Lys	Glu	Val	Lys	Leu	Leu	Leu	Leu	Gly
GCT	GGT	GAG	TCT	GGG	<u>888</u>	AGC	ACC	ATT	GTG	AAG	CAA	ATG	AAA	ATT	ATC	CAT	GAG	GAT	GGA
ALA 181	Gly	Glu	Ser	Gly	Lys	Ser	Thr	Ile	Val	Lys	Gln	Net	Lys	Ile	Ile	His	Glu	Asp	Gly
TAC	TCC	GAG	GAA	GAA	TGC	CGG	CAG	TAC	AAA	GTG	GTC	GŤG	TAC	AGT	AAC	ACT	ATT	CAG	TCA
Tyr 201	Ser	Glu	Glu	Glu	Cys	Arg	Gln	Tyr	Lys	Val	Val	Val	Tyr	Ser	Asn	The	Ĩle	Gln	Ser
ATC	ATC	GCT	ATA	ATC	CGA	GCC	ATG	GGA	AGG	CTA	AGG	ATT	GAT	TTT	GGA	GAT	GTG	GCT	100 AGA
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GCT	GAT	GAT	GCT	CGA	CAG	CTC	TTT	GTG	110 TTG	GCC	AGT	AGT	GCT	GAG	GAG	GGA	GTT	ATC:	120 TCT
Ala	Asp	Asp	Ala	Arg	Gln	Leu	Phe	Val	Leu	Ala	Ser	Ser	Ala	Glu	Glu	Gly	Val	Met	Ser
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AAT	CGG	ATG	CAT	GAA	AGC	ATG	AAA	TTG	TTT	GAT	AGC	ATC	TGC	AAC	AAC	AAG	tgg	TTC	ATA
ASN 781	Arg	met	HIS	618	Ser	flet	Lys	Leu	270	Asp	Ser	lle	Cys	Asn	Asn	Lys	Trp	Phe	11e 280
GAC	ACC	TCA	ATC	ATC	CTC	TTC	CTT	AAC	AAA	AAG	GAC	CTG	TTT	GAA	GAA	AAG	ATC	TCC	AGG
Asp Rui	Thr	Ser	Ile	Ile	Leu	Phe	Leu	Åsn	1.ys 290	Lys	Asp	Leu	Phe	Glu	Glu	Lys	Ile	Ser	Arg
AGC	cec	CTT	ACT	ATT	TCC	TAC	CCA	GAA	TAC	TCA	ccc	тст	лас	ACC	TAT	GAA	GAA	GCT	GCA
Ser	Pro	Leu	Thr	Ile	Cys	Tyr	Pro	Glu	Tyr	Ser	Gly	Ser	Asn	Thr	Tyr	Glu	Glu	Ala	Ala
GCC	TAC	ATT	CAG	tgc	CAG	TTT	GAG	GAC	TTG	AAC	CGG	AGG	AAA	GAC	ACA	AAG	GAA	ATA	TAC
Ala ac+	Tyr	Ile	61 n	Cye	Gln	Phe	Glu	Asp	Leu	Asn	٨rg	Arg	Lys	Азр	Thr	Lys	Glu	11e	Tyr
ACA	CAT	ттс	ACA	TGT	GCC	ACG	GAT	ACC	AAG	AAC	GTT	CAG	TTT	GTA	TTT	GAT	GCA	GTC	ACA
Thr	His	Phe	Thr	Cys	Ala	Thr	Asp	Thr	Lys	Asn	Val	Gln	Phe	Val	Phe	Asp	Ala	Val	Thr
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Fig. 3. Nucleotide and predicted amino acid sequence of the cDNA insert for G α i-3. Numbers indicate the position of nucleotides or amino acid residues starting at the hypothetical initiation codon. Sequence enclosed in an open box correspond to the poly(A) addition sequence. Nucleotide sequence recognized by the common 27-base probe is underlined [10].

sequence for one of the G α s cDNA (clone 6A1). The sequence is 1321 nucleotides long and predicts an open reading frame of 379 amin acid residues (M_r 42720) and the protein presents more similarity (90%) to the short species of human G α s-1a [16]. At the nucleotide level, the 5'-non-coding region contains 80 nucleotides and a small 3'-non-coding region of 101 residues containing a poly(A) addition sequence at position 1226 and a poly(A) tail of 13 adenosines. Clone G α s 12B2 is very similar to clone 6A1 at the nucleotide level, showing only 42 differences which generate different restriction sites (see *Eco*RI site in Fig. 1) and 12 different amino acid residues (Fig. 5). -80

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61		•		-			-		30	-									40
GAG	ACC	AAC	AAG	AAG	ATC	GAG	AAG	CAG	CTG	CAG	AAG	GAC	AAG	CAG	GTG	TAT	AGG	6CG	ACG
Glu	Thr	Asn	Lys	Lys	Ile	Glu	Lys	Gln	Lou	Gln	Lys	Asp	Lys	Gln	Val	Tyr	Arg	Ala	Thr
121									50										60
CAC	AGG	CTT	CTG	CTG	CTC	GGT	GCT	GGA	GAG	TCT	GGT	AAA	AGC	TCA	ATT	GTG	AAG	CAG	ATG
H18	Arg	Leu	Leu	Leu	Leu	era	vra	GTÀ	Glu	Ser	ery	Lys	Ser	Ser	TTe	var	ràs	GIN	Met
181	ATC	CTG	CAC	CTC		CCA	TTO		CCA	GAC	GAG	***		400		070		CAT	80 878
Are	Tle	Leu	His	Val	Asn	GIV	Phe	Asn	Ala	Glu	Glu	LVS	Lars	The	LVR	Val	611	Aen	Tle
241								,	90			2,0	2,5		2,0		010		100
AAG	AAT	AAT	ATT	AAG	GAA	GCT	ATA	GAG	ACA	ATA	GTT	ACG	GCA	ATG	GGC	AAC	CTC	TCT	ccc
Lys	Asn	Asn	Ile	Lys	Glu	Ala	Ile	Glu	Thr	Ile	Val	Thr	Ala	Met	Gly	Asn	Leu	Ser	Pro
301									110										120
CCG	GTG	GAG	CTA	GTG	AAT	CCV	GAA	AAC	CAG	TTC	CGA	ATT	GAC	TAC	ATC	CTC	AAC	CTA	ccc
Pro	Val	Glu	Leu	Val	Asn	Pro	Glu	Åsn	Gln	Phe	Arg	Ile	Asp	Tyr	lle	Leu	Asn	Leu	Pro
361	-		~~~	-	-		-		130	_		~ • •	~ • •					-	140
Aen	Tim	Lue	APD	Phe	GIN	Pho	Son	Doo	646	Pho	TAT	GAA	CAC	ACA	744	ACG	CTC	TGG	CAG
421	- ,-	2,0	nop	7.00	010	1116	0.01	110	150	I ne	191	Gru	n19	101.	цув	Int.	rea	тŗр	160
GAC	GAG	GGG	GTG	AGG	GCG	TGC	TAC	GAG	CGA	тсс	AAC	GAG	TAC	CAG	CTG	ATT	GAC	TGC	GCA
Asp	Glu	Gly	Val	Arg	Ala	Cys	Tyr	Glu	Arg	Ser	Asn	Glu	Tyr	Gln	Leu	Ile	Asp	Cvs	Ala
481									170								•	•	180
CAG	TAT	TTT	CTA	GAC	AAA	ATT	GAC	ATT	GTG	AAA	CAG	AAC	GAC	TAC	ACG	CCT	AGT	GAT	CAG
544	IJT	rne	Leu	лар	Lya	TTe	veb	11e	Val	Lys	Gin	Asn	Asp	Tyr	Thr	Pro	Ser	Asp	Gln
GAC	TTC	070	004	TCC	404		000	400	190	~~		-	~			-			200
Aan	Leu	Len	Ang	Cvs	Ano	Val	Len	The	Sev	Gly		The	C1.	The	AAG Lute	TTT	CAG	GIG	GAC
601			·~ 6	0,0	-14 K		199.0	Int.	210	GTÀ	116	7 ne	GTU	Inr	гуя	rne	GTU	vai	ASP 220
AAA	GTC	AAT	TTC	CAC	ATG	TTC	GAT	GTT	GGA	GGC	CAG	CGA	GAT	GAG	CGC	AGG	AAG	TGG	ATC
Lys	Val	Asn	Phe	His	Met	Phe	Asp	Val	Gly	Gly	Gln	Arg	Asp	Glu	Arg	Arg	Lys	Tro	Ile
661									230				•		Ū				240
CAG	TGC	TTT	AAC	GAT	GTC	ACA	GCT	ATA	ATC	TTC	GTT	GTA	GCC	AGC	AGC	AGC	TAC	AAC	ATG
61n 721	Cys	Pne	Asn	лар	Val	Thr	Ala	Ile	Ile	Phe	Val	Val	Ala	Ser	Ser	Ser	Tyr	Asn	Met
GTG	ATC	200	GAG	GAC	447	CAC	100		200	010	~**	~~							260
Val	Ile	Arg	Glu	Asp	Asn	His	Thr	Asn	Ane	Len	Gin	Glu	Ala	Lan	Ann	Len	Phe	AAG	AGT
781				•					270		•					Dea	t ne	цуа	280
ATC	TGG	AAC	AAT	AGG	TGG	CTA	CGG	ACC	ATT	TCA	GTC	ATT	CTC	TTC	стс	AAT	***	CAA	GAT
lle	Ттр	Asn	λ s n	٨rg	Тгр	Leu	Arg	Thr	Ile	Ser	Val	Ile	Leu	Phe	Leu	Asn	Lys	Gln	Asp
841	-								290										300
CTG	CTC	GCT	GAA	AAG	GTT	AAT	GCT	GGG	AAA	TCT	AAA	ATA	GAG	GAC	TAC	TTC	ССТ	GAG	TTT
0.00	Lieu	UTG	GTU	гув	var	Asn	лта	GTÀ	Lys	Ser	Lys	Ile	Glu	Asp	Tyr	Phe	Pro	Glu	Phe
GCC	CGA	TAC	ACĆ	ACC	CCA	GAT	GAT	GCA	ACT	004	C44	ore	000	C40	-	0.00	~~~	-	320
Ala	Arg	Tyr	Thr	Thr	Pro	Asp	Asp	Ala	Thr	Pno	Glu	Val	Glv	Glu	Ann	Dun	LOA	GTC Val	ACA
961	-	-				•			330				. .,	0.14	unh	110	'nв	Var	380
AGG	GCC	AAG	TAT	TTT	ATT	AGA	GAT	GAG	TTT	СТТ	AGA	ATC	AGC	ACA	GCC	AGT	GGG	GAC	GGC
Arg	Ala	Lys	Туг	Phe	Ile	Arg	Asp	Glu	Phe	Leu	Arg	Ile	Ser	Thr	Ala	Ser	Gly	Asp	Gly
1021	CAT	TAT	-	-	0.00		-		350								-	-	360
Arg	His	Tur	Core .	TAC	CCT Pro-	UAT .	TC	ACA	TGT	GCA	GTG	GAT	ACA	GAA	AAC	ATC	CGA	AGG	GTT
1081		- 7 -	-ya	191.	.10	112	LUG	111,	370	AIA	vai	Asp	Thr	Glu	Asn	Ile	Arg	Arg	Val
TTT	AAT	GAT	TGT	CGG	GAC	ATT	ATC	CAA	AGG	ATG	CAC	076	CGC		TAT	GAR		-	-
Phe	Asn	Asp	Cys	Arg	Asp	Ile	Ile	Gln	Arg	Net	His	Leu	Arg	Gln	Tvr	Glu	Len	116	LOW
1141					-				5				~ 0		- , -			Jeu	STOP
TGGA	GAGC	AC T	TTTT	GTTT	T TG	GTTC	cccc	ccc	CCCC	стс	CTTO	TGTT	°CC G	GACT	ACAA	A TI	ссто	ATT	
пляб	~ ^^^	AA A	CCTA	متجب	القرر	AAAA	****	A											
								12.	4.										

Fig. 4. Nucleotide and predicted amino acid sequence of the cDNA for G α s (clone 6A1). Numbers indicate the position of nucleotides or amino acid residues starting at the initiation codon. Sequence enclosed by the open box corresponds to the poly(A) addition sequence. Nucleotide sequence recognized by the common 27-base probe is underlined [10] and the *Eco*RI site is indicated by a line over the sequence GAATTC.

3.5. Comparison of the deduced amino acid sequences of the Xenopus laevis oocyte $G\alpha$ subunits

Fig. 5 shows the alignment of the predicted protein sequences of each of the cDNA clones reported here. The sequences share great similarity among themselves and with their mammalian counterparts. The regions of identity are enclosed in boxes. Also the figure shows the sequences labelled A, C, E and G which are supposedly involved in the binding of GTP.

4. DISCUSSION

The results presented in this communication and in the previous publication [10] demonstrate that one single cell type, the amphibian oocyte, contains at the mRNA level at least 5 different types of α G-proteins.

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Since our search has not been exhaustive, it is quite possible that there are more different α -subunits not yet detected.

The analysis of the nucleotide and deduced amino acid sequences allows us to classify the oocyte subunits according to their analogy to the structure of mammalian G-protein α -subunits. Thus it has been established that two of the isolated clones correspond to α s-type subunits, and the other two clones correspond to α i-1 and α i-3 subunits.

The two α s clones correspond to the 'short' version of the mammalian α s which arises through alternative splicing and elimination of exon 3, present in the 'long' α s [17]. Another variant found in mammals is that the short version can be found with or without a serine in position 72. The two oocyte clones are short versions without this serine. This is to our knowledge the first time that two different α s coding genes have been found in the same cell type and differ in other characteristics than those mentioned above. Since the 5'- and 3'-non-coding regions of the two clones are quite similar, it seems probable that they arose through a rather recent gene duplication. The small number of differences that account for 12 amino acid changes are all scattered through the polypeptide and therefore cannot be generated through alternative splicing. One of the changes alters an EcoRI site, a finding which may be useful for future analysis of the function of these proteins. One of the most interesting differences between the two α s clones is the conservative change of serine-178 present in the 6A1 clone for threonine in the clone 12B2. On the basis of the results obtained with mutations done in rats, the region encompassing residues 192–196 in long species of the α s subunits has been proposed to interact with the catalytic subunit of adenylyl cyclase and any mutation in this region could cause a change in its activity [18]. It is interesting to note that in this highly conserved region, all the α subunits that are not stimulatory (non- α s-type subunits) contain a threenine in position 178 (short α s species) while all previously sequenced α s subunits have a serine in this position. The only other exception to this latter generality is the short α s-type found in rat olfactory neurons which contains a threonine in position 180 [19]. Interestingly, in the same tissue, olfactory neuroepithelium, a different long α s species from nonneuronal origin was found by the same group and this contained a serine in position 193 [20]. The presence of these two types of α s subunits in the oocyte suggests that they may have different functions in this cell.

Several G-protein regions share considerable similarity with the guanine nucleotide binding regions of elongation factor Tu and p21^{ras} [21]. Consistent with this role, these regions (Fig. 5, labeled A, C, E and G) are all highly conserved within the four oocyte Gproteins. Other regions, as the one implicated in receptor, $G\beta\gamma$, and effector protein interaction [22] are not



Fig. 5. Alignment of the amino acid sequences of the oocyte $G\alpha$ subunits. Amino acid sequences are presented by the standard one-letter abbreviation code. The oocytes $G\alpha$ s, $G\alpha$ i-1, $G\alpha$ i-3 and the already published $G\alpha$ o [10] are shown. Amino acid regions that are identical are enclosed by open boxes. The arrows indicate the arginine and cysteine residues that are ADP-ribosylated by *Cholera* and *Pertussis* toxins, respectively. The 12 amino acid differences between clones 12B2 and 6A1 of $G\alpha$ s are enclosed by circles. The amino acid regions that participate in the binding of GTP are overlined. The residues marked by asterisks correspond to the $G\alpha$ s region proposed to be important for the adenylyl cyclase activation [18].

as conserved as the guanine nucleotide binding regions, reflecting their role in the independent modulation of different signal transduction pathways.

The proteins encoded by $G\alpha i$ -1 and $G\alpha i$ -3 cDNAs are potential substrates for pertussis toxin (PTX)-catalyzed ADP-ribosylation because they have a cysteine residue at the appropriate site near the carboxyl-terminus. Also the two $G\alpha$ s clones contain the arginine that is modified by cholera toxin (Fig. 5, arrowheads).

Currently, we are expressing these proteins in *E. coli* and in an in vitro system in order to study some of their G-protein properties and functions.

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