

Biological characteristics and conservation genetics of the narrowly distributed rare plant *Cinnamomum chago* (Lauraceae)



Wenjing Dong, Xue Zhang, Yang Guansong, Liu Yang, Yuehua Wang^{**}, Shikang Shen^{*}

School of Life Sciences, Yunnan University, Kunming, 650091, People's Republic of China

ARTICLE INFO

Article history:

Received 6 May 2016

Received in revised form

31 August 2016

Accepted 2 September 2016

Available online 12 September 2016

(Editor: Weibang Sun)

Keywords:

Cinnamomum chago

Habitat

Biological characteristic

Genetic diversity

Conservation strategies

ABSTRACT

Cinnamomum chago (family Lauraceae) is an essential source of timber and oil. This plant is narrowly distributed in the western part of the Yunnan Province. In this study, the distribution, habitat, and biological characteristics of *C. chago* were examined through field investigation. The genetic diversity and the variation of the remnant populations were also studied using the inter-simple sequence repeat technique. Results showed that *C. chago* is mainly distributed in the upstream tributary mountains of Lancang River in Yunlong County of Yunnan Province. The species distribution exhibited a fragmented pattern with five isolated populations and high-frequency anthropogenic interference. A combination of morphological features (opposite leaves, pinnate leaf veins, absence of glandular fossa, large drupe, small punch, and pollen surface with triangular spike grain, with cushion bumps at the base) indicated that *C. chago* is a key phylogenetic taxon between the two sections of Asian *Cinnamomum* plants (Sect. *Camphora* (Trew) Meissn. and Sect. *Cinnamomum*). Analysis of the genetic diversity of *C. chago* indicated that it has a moderately high level of genetic diversity at the population and species levels (populations level: $N_e = 1.629$, $H = 0.348$, $I = 0.504$, and $PPB = 83.3\%$; species level: $N_e = 1.864$, $H = 0.460$, $I = 0.652$, and $PPB = 100\%$). Analysis of molecular variance revealed that 17% of the genetic variation was divided between the populations, whereas 83% was observed within the populations. Based on these results, we suggest the inclusion of *C. chago* in the Wild Plants with Extremely Small Populations in China. Moreover, the species should be given special attention and protection. Some strategies were proposed for the conservation of the *C. chago* populations.

Copyright © 2016 Kunming Institute of Botany, Chinese Academy of Sciences. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co., Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Cinnamomum chago is an extremely rare species from the genus *Cinnamomum* of the family Lauraceae. This species is not well known and has not yet been recorded in Flora of China (Wu, 1991; Li et al., 2008). *C. chago* is a morphologically distinct, large evergreen tree that grows approximately 25 m tall, with a trunk diameter of 40 cm; leaves opposite or nearly opposite; leaf blade green or yellow-green,

ovate-elliptic, 5–13 cm × 2.5–4.5 cm, leathery; venation pinnate with stout midrib, 7–9 pairs of lateral veins, axillary fossa absent or inconspicuous; petiole, 1.8 cm long; panicle terminal or sometimes axillary at the upper part of twig, 5–10 cm long; pedicels 5 mm long, glabrous; flowers yellow-green or redish at the upper part of the perianth lobes; perianth lobes elliptic, ca. 6 mm long; Fertile stamens 9, ca. 5 mm long; filaments pubescent; fruit subglobose or ovoid, green-yellow at maturity, 3 cm long and 2.5 cm in diameter, much bigger in this genus; Perianth cup is flat or discoid, 1.2–1.4 cm in diameter, leathery. Inside the flesh mesocarp, the hard leathery black-brown endocarp is pointed oval, with two hemispheric and white flesh cotyledons inside its embryo (Sun and Zhao, 1991). *C. chago* is considerably different from other *Cinnamomum* species because of the following: the perianth and the stamen are glabrous except the filament, the inner side of the perianth lobes is sparsely pubescent, the leaves are opposite but with pinnate venation, the fruit is bigger that grows approximately 3 cm long and 2.5 cm in diameter, and its typical discoid fruit cup. In the recently reported

* Corresponding author. School of Life Sciences, Yunnan University, No. 2 Green Lake North Road Kunming, Yunnan, 650091, People's Republic of China. Fax: +86 871 65031491.

** Corresponding author. School of Life Sciences, Yunnan University, No. 2 Green Lake North Road Kunming, Yunnan, 650091, People's Republic of China. Fax: +86 871 65031491.

E-mail addresses: wangyh58212@126.com (Y. Wang), yunda123456@126.com (S. Shen).

Peer review under responsibility of Editorial Office of Plant Diversity.

Cinnamomum phylogeny, including *C. chago* (we provided the material but the name was miswritten as *Cinnamomum chaogo*), supports the consideration of *C. chago* as a distinct species, and situates its phylogenetic status between the two sections of the Asian *Cinnamomum* plants (Sect. *Camphora* (Trew) Meissn. and Sect. *Cinnamomum*) (Huang et al., 2016).

Our field investigation found that *C. chago* is distributed along the Lancang River upstream of the tributary mountains in Yunlong County, Yunnan Province. It exhibits an extremely limited distribution with a fragmented pattern. Furthermore, no seedlings were found near the adult trees during our field survey. Although the species is narrowly distributed and has a low survival rate, its biological characteristics and conservation biology have been scarcely been reported, despite the fact that practical conservation strategies for endangered and rare plants are based on the understanding of biological characteristics and survival status of these plants (Sun et al., 2006; Shen et al., 2009). The present study thus investigated the habitat, biological characteristics, and genetic diversity of *C. chago*. Furthermore, corresponding conservation and utilization strategies have been proposed based on our results.

2. Materials and methods

2.1. Field investigation of the populations

To investigate the distribution and the habitat of *C. chago*, we reviewed several studies and herbarium records and then performed a comprehensive field investigation during the flowering and the fruiting seasons of *C. chago* in 2014 and 2015. During our field survey, a global positioning device (GPS) was used to record the precise location of the detected species. The altitude, latitude, and longitude of each site were also recorded.

2.2. Biological characteristics

We collected flowers and fruits during the flowering and the fruiting season, respectively, and then we immediately placed them in FAA to preserve their original characteristics. Using light microscopy, we observed the anatomy of *C. chago*. Scanning electron microscopy (SEM) was employed to observe pollen morphology. Thirty samples were selected randomly for observation of biological characteristics. These data were consolidated with observations from previous studies.

2.3. Analysis of genetic diversity

Our field investigation revealed that the five remaining *C. chago* populations are distributed in Yunlong and Yangbi County in Yunnan Province. In 2014 and 2015, 54 *C. chago* samples were collected from these populations (Table 1). The distance between the collected individual samples was at least 15 m. Fresh young leaves were removed from the shoots, and then dried in silica gel and stored at -20°C prior to DNA extraction. Detailed informations regarding the locations and the population codes of the samples are shown in Table 1.

Table 1
Characteristics of five sampled populations in *Cinnamomum chago*.

Population	Number	Longitude	Latitude	Altitude (m)
NMP	10	99° 16' 35.03"	25° 33' 46.79"	2357
DSB	8	99° 10' 24.24"	25° 45' 49.7"	2317
SBX	12	99° 56' 28.35"	25° 34' 13.27"	2249
XC	12	99° 56' 33.40"	25° 34' 8.23"	2296
LG	12	99° 55' 09.2"	25° 33' 08.9"	2310

NMP: NanMuPing; DSB: DaShiBa; SBX: ShunBiXiang; XC: XingCun; LG: LaGuo.

2.4. DNA extraction

DNA was extracted from dried leaves using a modified CTAB method (Doyle and Doyle, 1988). Purified DNA was detected by 1.0% agarose gel electrophoresis and stored at -20°C until use.

2.5. ISSR PCR amplification

DNA samples were randomly selected from mixed DNA of each population. The samples were screened using 100 primers obtained from the University of British Columbia Biotechnology Lab (UBCBL). Each randomly selected sample that has a mixed DNA from each population was used to screen 100 primers obtained from the University of British Columbia Biotechnology Lab (UBCBL). Finally, the 14 primers that generated clear bands with high polymorphism were selected for PCR amplification using a final volume of 20 μl . The reaction mixture consisted of 40 ng of template DNA, 1.5 μl of dNTP, 2 μl of 10x buffer, 1 μl of primer, and 0.3 μl of Taq DNA polymerase. The reaction underwent an initial denaturation for 7 min at 94°C , followed by 45 cycles of PCR, which includes 30 s of denaturation at 94°C , 45 s of annealing at appropriate annealing temperature, 90 s of extension at 72°C , and a final extension step of 7 min at 72°C . Subsequently, 5 μl of amplified products were visualized on a 1.5% agarose gels after electrophoresis in $0.5 \times \text{TBE}$ at voltage 100 V for 80 min. The gels were photographed using a UV gel imaging system.

2.6. Data analysis

The individuals were scored for the presence (1) or absence (0) of amplified bands. The percentage of the polymorphic loci (PPB), effective number of alleles (N_e), Nei's genetic diversity (H), Shannon's information index (I), estimate of gene flow (N_m), total gene diversity (H_t), variability within populations (H_s), and coefficient of genetic differentiation (G_{st}) were calculated using POPGENE version 1.32 (Yeh et al., 1999) and GenAEx version 6.501 (Peakall and Smouse, 2012) with manual corrections.

Analysis of molecular variance (AMOVA) was conducted to calculate the extent of genetic variation between and within the two populations by using GenAEx version 6.501 (Peakall and Smouse, 2012).

We conducted Bayesian analysis of the population structures using STRUCTURE version 2.2 (Pritchard et al., 2000). A total of 20 independent runs were performed for each set with K ranging from 1 to 20, a burn-in of 1×10^5 iterations, and 1×10^5 subsequent Markov Chain Monte Carlo steps. The combination of the admixture and the correlated allele frequency models was also analyzed. The second-order rate of change in the log probability of the data with respect to the number of clusters (ΔK) was also used to estimate the number of genetic clusters (Evanno et al., 2005). The best-fit number of groupings was evaluated using ΔK through STRUCTURE HARVESTER version 0.6.8 (Earl and von Holdt, 2012).

3. Results

3.1. Distribution and habitat of *C. chago*

Investigating populations of *C. chago* during the flowering and fruiting seasons of 2014 and 2015, we found that this species is mainly distributed along the Lancang River upstream of the tributary mountains in Yunlong County, Yunnan Province. The species occurs in five isolated, fragmented populations: ShunBiXiang (SBX), XingCun (XC), LaGuo (LG), DaShiBa (DSB), NanMuPing (NMP) (Table 1). We sampled at least twelve individuals for each population where possible, though for DSB and NMP 8 and 10

individuals were sampled, respectively. SBX, XC, and LG populations of *C. chago* are distributed near local villages in YangBi County and are exposed to human activities. In contrast, the DSB and NMP populations are distributed in secondary evergreen broad-leaved forest in YunLong County at altitudes between 2100 m and 2400 m. All populations sampled have been subjected to high-frequency anthropogenic interference, particularly during the fruiting seasons when fruits are harvested by local villagers. Thus, the seeds were unable to grow into new seedlings.

3.2. Biological characteristics

Previous research found that the leaves of *C. chago* are opposite or nearly opposite. The veins are pinnate and the midrib is burly and evident while the lateral veins are thin and have no glandular fossa. The flowers are bisexual, and green–white or yellowish. The perianth is glabrous or puberulent outside and densely pubescent inside. It has 12 stamens in total, with 4 regularly arranged whorls. There are fertile stamens 9 (of 1st, 2nd, and 3rd whorl) and regressive stamens 3 (of 4th whorl) (Sun and Zhao, 1991). Our field work confirmed these observations. We also noted that the pollen surface has small punch and triangular spike grain with cushion bumps at the base. The fruit is drupe, purple-black (color), globular, and large with a diameter of 2.5 cm when fully mature (Fig. 1). The seeds contain various nutrients, including sugar, protein, crude fat, and amino acids, and have been regarded as an ideal nut (unpublished data). The flowering time of *C. chago* varies slightly between different populations and microhabitats, whereas the period of flowering and fruiting is generally from April to October. Its special morphological feature combination (opposite leaves, pinnate leaf veins, no glandular fossa, drupe large, and pollen surface with triangular spike grain, with cushion bumps at the base, and small punch) indicated that *C. chago* is a key phylogenetic taxon between the two sections of Asian *Cinnamomum* plants (Sect. *Camphora* (Trew) Meissn. and Sect. *Cinnamomum*).

3.3. Genetic diversity of *C. chago*

A total of 109 bands were generated by the 14 selected primers that produced 6–13 bands each, producing an average of 7.8 bands. All bands were polymorphic and completely accounted (Table 2).

We observed a moderately high level of genetic diversity at the population and species level (population level: $N_e = 1.629$, $H = 0.348$, $I = 0.504$, and $PPB = 83.3\%$; species level: $N_e = 1.864$, $H = 0.460$, $I = 0.652$, and $PPB = 100\%$), as shown in Table 3. The genetic diversities within species (H_t) and within populations (H_s) were 0.4453 and 0.3485, respectively (Table 4). The genetic differentiation between the populations (G_{st}) was 0.2174. Based on the G_{st} value, the level of gene flow (N_m) was estimated at 1.7999 ($N_m > 1$). These results indicated high gene flow and low differentiation between the extant populations.

3.4. Genetic structure of *C. chago*

Analysis of molecular variance results revealed that 17% of the genetic variation was partitioned between the populations and 83% occurred within the populations based on inter-simple sequence repeat (ISSR) markers (Table 5). These results indicated low genetic variation levels between the five populations.

STRUCTURE analysis based on the ΔK method revealed that ΔK was 56.18 for $K = 5$ and ΔK was <56.18 for all of the values of K (ranging from 1 to 20, except 5) as shown in Fig. 2a and b. Therefore, the optimal ΔK for $K = 5$ provided strong evidence for the presence of five independent populations, which is consistent with the five natural populations sampled here (Fig. 2c).

4. Discussion

4.1. Population status of *C. chago*

Our field survey revealed that distribution of *C. chago* is restricted to the Lacang River upstream of the tributary mountains in Yunlong County, Yunnan. These remaining populations not only have a limited geographical reach but are fragmented and highly isolated. It is well known that habitat is essential for the long-term persistence and survival of endemic and rare species (Kalliovirta et al., 2006; Shen et al., 2009). Importantly, our field surveys revealed that all the populations of *C. chago* occur in unprotected areas outside of nature reserves and are frequently affected by human interference. The survival status of *C. chago* is similar to other critically endangered plants such as *Euryodendron excelsum* (Shen et al., 2009) and *Manglietiastrum sinicum* (Tian et al., 2002;



Fig. 1. Biological characteristic and field investigation of *Cinnamomum chago* populations. a. habitat; b. Plant morphological; c. flowers; d. pollen; e. fruit; f. seed; g. field work.

Table 2
Primers and their amplification polymorphism of amplified bands of *C. chago* populations.

Primer code	Sequence (5' to 3')	Scored bands	No. of polymorphic bands	Percentage of polymorphic bands
811	GAGAGAGAGAGAGAC	6	6	100%
815	CTCTCTCTCTCTCTG	7	7	100%
834	AGAGAGAGAGAGAGY	6	6	100%
836	AGAGAGAGAGAGAGYA	6	6	100%
840	GAGAGAGAGAGAGAYT	6	6	100%
841	GAGAGAGAGAGAGAYC	7	7	100%
843	GAGAGAGAGAGAGAYG	11	11	100%
853	TCTCTCTCTCTCTCRT	7	7	100%
854	TCTCTCTCTCTCTCRG	8	8	100%
855	ACACACACACACACYT	6	6	100%
857	ACACACACACACACYG	7	7	100%
873	GACAGACAGACAGACA	13	13	100%
880	GGAGAGGAGAGGAGA	11	11	100%
881	GGTGGGGTGGGGTG	8	8	100%
Mean		7.8	7.8	100%
Species level		109	109	100%

Table 3
Genetic diversity of *C. chago* populations based on ISSR markers.

Population	PPB(%)	N _a	N _e	H	I
SBX	75.23	1.752	1.583	0.320	0.460
XC	78.90	1.789	1.626	0.344	0.493
LG	74.31	1.743	1.529	0.300	0.438
NMP	92.66	1.927	1.711	0.389	0.562
DSB	95.41	1.954	1.693	0.389	0.567
Mean	83.30	1.833	1.629	0.348	0.504
Species level	100	2.000	1.864	0.460	0.652

Notes: N_a, observed number of alleles; N_e, effective number of alleles (Kimura and Crow (1964)); H, Nei's (1973) gene diversity; I, Shannon's Information index; P, the percentage of polymorphic loci.

Table 4
Nei's (1973) analysis of gene diversity in *C. chago* populations.

	H _t	H _s	G _{st}	N _m
Species level	0.4453	0.3485	0.2174	1.7999
Standard deviation	0.0034	0.0089		

Notes: H_t, total variability; H_s, variability within populations; G_{st}, coefficient of genetic differentiation; N_m, estimate of gene flow.

Table 5
Analysis of molecular variance (AMOVA) based on ISSR markers for five populations of *C. chago*.

Source of variation	df	Sum of squares	Variation components	Percentage of variation (%)
Among populations	4	228.120	3.625	17%
Within populations	49	886.417	18.090	83%
Total	53	1114.537	21.716	—

Sun et al., 2015). Our study also found that an additional threat to the survival and natural proliferation of the species is unrestrained harvesting of *C. chago* fruits for their nuts by local people. Thus, we suggest that this species be designated as a Plant Species with Extremely Small Populations (PSESP) in China and be given special attention and protection.

4.2. Genetic diversity of *C. chago*

The genetic diversity of a species in small populations is lower than that in large populations due to genetic drift and inbreeding (Willi et al., 2006; Li et al., 2012). As a result, rare and endangered

species with narrow geographical distributions harbor lower genetic diversity to a greater extent than similar species with broad geographical distributions (Hamrick and Godt, 1990). In the present study, genetic diversity within the *C. chago* species was detected using ISSR markers. However, our study showed that *C. chago* showed a high level of genetic diversity (N_e = 1.629, H = 0.348, I = 0.504, PPB = 83.3%) at the species level.

In general, the level of genetic diversity is influenced by several factors, including reproductive mode, biological traits and breeding system. Outcrossing species generally exhibit considerably higher levels of genetic diversity than selfing species (Hamrick and Godt, 1989; Nybom, 2004). The reproductive biology and breeding system of *C. chago* were not examined in this study, although, based on the high level of genetic diversity, we hypothesize that it is an outcrossing species. This hypothesis is also consistent with Rohwer's finding (1993) that plants from the family Lauraceae tend to be outcrossing.

High genetic variation enables a species to adapt to various environments (Zhao et al., 2012). High genetic diversity in the five remaining populations of *C. chago* indicate that the species is not endangered because of genetic factors (e.g. genetic diversity decline, genetic drift, and inbreeding). The main threat to this plant species may include habitat specialization, anthropogenic interference and its limited distribution. Regardless, the factors that lead to the "endangered" status of this species must be further elucidated.

4.3. Genetic structure of *C. chago*

Genetic structure is affected by several factors, including breeding system, genetic drift, population size, seed dispersal, gene flow, evolutionary history, and natural selection (Hamrick and Godt, 1990). Our analyses of the genetic structure of *C. chago* showed that the 54 individuals formed 5 populations (Fig. 2c). Molecular variance analysis showed that genetic variation mostly occurred within populations (83%). The coefficients of genetic differentiation (G_{st}) and the gene flow (N_m) between the five extant populations were 0.2174 and 1.7999, respectively. These results indicate that the frequency of gene flow between the groups is sufficient to prevent differentiation between populations caused by genetic drift. This might be explained by long-distance dispersal of pollen and/or seeds which are known to result in low genetic differentiation and high gene flow between the populations of the same species (Yao et al., 2007; Zhao et al., 2012). At present, there has been little research on *C. chago* and the mechanisms of seed and pollen dispersal have not been elucidated for this species. Thus, the factors that facilitate frequent and continual gene flow between the

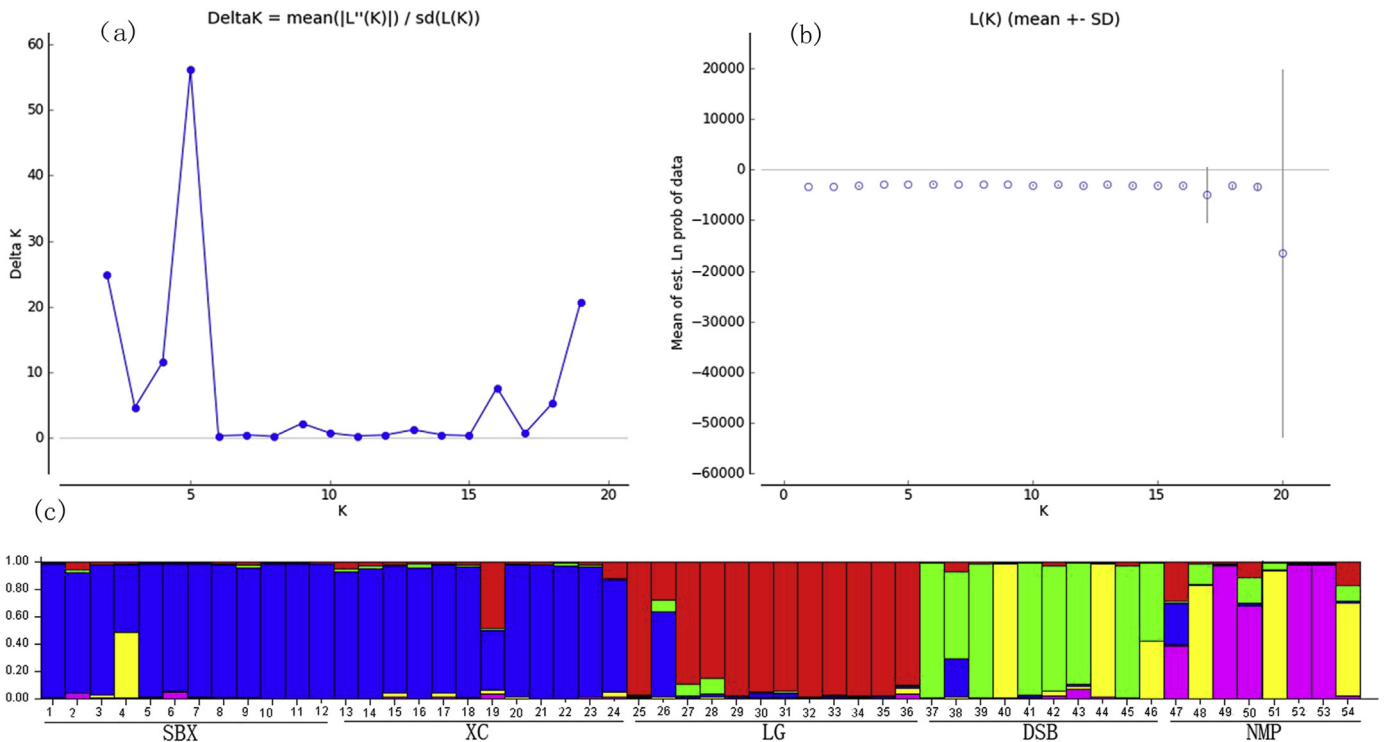


Fig. 2. Results of Bayesian model-based clustering STRUCTURE analysis of 54 individuals of *C. chago*. (a) The probability of the data $Lnp(D)$ ($\pm SD$) analysis the number of K clusters, and increase of $Lnp(D)$ given K , calculated as $(Lnp(D)k - Lnp(D)k-1)$. (b) Delta K values from the mean log-likelihood probabilities from STRUCTURE runs where inferred clusters (K) ranged from 1 to 20. (c) Estimated genetic clustering ($K = 5$) obtained with the STRUCTURE program for 54 individuals. Individuals are separated according to the population and black vertical line in the bar chart is population identifier.

five populations of *C. chago* are still unknown. In order to address these issues, future research on *C. chago* reproductive biology is needed.

4.4. Conservation implications

Endemic plant species with limited distribution face both internal and external threats to their survival (Zu et al., 1999). Our study indicates that while the distribution of *C. chago* populations are small, isolated and fragmented, this species still has a relatively high level of genetic diversity. According to Williamson and Werth (1999), the main threats to species with limited distribution but high genetic diversity are external, such as habitat fragmentation, geographical isolation, and human interference. We suggest intensified protection of the remaining populations of *C. chago in situ* and recommend prioritizing conservation of the DSB and NMP populations, which have the highest levels of genetic diversity. In addition, we suggest that local people be encouraged to grow more *C. chago* and to harvest more nuts. Seed collection for germplasm storage and artificial seedlings for *ex situ* conservation is also recommend. Population recovery and reintroductions of *C. chago* must also be conducted to facilitate recovery of wild populations.

Acknowledgments

This study was financially supported by grant 31560224 and 31360074 from the National Natural Science Foundation of China and grant 2015J002 from the Graduate Science of foundation projects of Yunnan Educational Committee.

References

- Doyle, J.J., Doyle, J.L., 1988. Isolation of plant DNA from fresh tissue. *Focus* 12, 13–15.
- Earl, D.A., vonHoldt, B.M., 2012. STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conserv. Genet. Res.* 4, 359–361.
- Evanno, G., Regnaut, S., Goudet, J., 2005. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Mol. Ecol.* 14, 2611–2620.
- Hamrick, J.L., Godt, M.J.W., 1989. Allozyme diversity in plant species. In: Brown, H.D., Clegg, M.T., Kahler, A.L. (Eds.), *Plant Population Genetics, Breeding and Genetic Resources*. Sinauer Associates, Inc, Sunderland, MA, pp. 43–46.
- Hamrick, J.L., Godt, M.J., 1990. *Plant Population Genetics, Breeding, and Genetic Resources*. Sinauer, Sunderland, MA, pp. 43–63.
- Huang, J.F., Li, L., van der Werff, H., et al., 2016. Origins and evolution of cinnamon and camphor: a phylogenetic and historical biogeographical analysis of the *Cinnamomum* group (Lauraceae). *Mol. Phylogenet. Evol.* 96, 33–44.
- Kalliovirta, M., Rytteri, T., Heikkinen, R.K., 2006. Population structure of a threatened plant, *Pulsatilla patens*, in boreal forests: modeling relationships to overgrowth and site closure. *Biodivers. Conserv.* 15, 3095–3108.
- Kimura, M., Crow, J.F., 1964. The number of alleles that can be maintained in a finite population. *Genetics* 49, 725–738.
- Li, H.W., Li, J., Huang, P.H., Wei, F.N., Cui, H.B., van der Werff, H., 2008. Lauraceae. In: Wu, Z.Y., Raven, P.H., Hong, D.Y. (Eds.), *Flora of China*, vol. 7. Science Press and Missouri Botanical Garden Press, Beijing, China, St. Louis, Missouri, USA.
- Li, Y.Y., Guan, S.M., Yang, S.Z., et al., 2012. Genetic decline and inbreeding depression in an extremely rare tree. *Conserv. Genet.* 13, 343–347.
- Nei, M., 1973. Analysis of gene diversity in subdivided populations. *Proc. Natl. Acad. Sci. USA* 70, 3321–3323.
- Nyblom, H., 2004. Comparison of different nuclear DNA markers for estimating intraspecific genetic diversity in plants. *Mol. Ecol.* 13, 1143–1155.
- Peakall, R., Smouse, P.E., 2012. GenA1ex 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. *Bioinformatics* 28, 2537–2539.
- Pritchard, J.K., Stephens, M., Donnelly, P., 2000. Inference of population structure using multilocus genotype data. *Genetics* 155, 945–959.
- Rohwer, J.G., 1993. Lauraceae. In: Kubitzki, K., Rohwer, J.G., Brittrich, V. (Eds.), *The Families and Genera of Flowering Plants*. Springer-Verlag, Berlin, pp. 426–437.
- Shen, S.K., Wang, Y.H., Wang, B.Y., et al., 2009. Distribution, stand characteristics and habitat of a critically endangered plant *Euryodendron excelsum* H T Chang (Theaceae): implications for conservation. *Plant Spec. Biol.* 24, 133–138.

- Sun, B.X., Zhao, H.L., 1991. A new species of cinnamomum from Yunnan. *Journal of Yunnan University* 13, 93–94.
- Sun, W., Zhou, Y., Han, C., et al., 2006. Status and conservation of *Trigonobalanus doichangensis* (Fagaceae). *Biodivers. Conserv.* 15, 1303–1318.
- Sun, W.B., Ma, Y., Chen, G., et al., 2015. Rescuing *Magnolia sinica* (Magnoliaceae), a critically endangered species endemic to Yunnan, China. *Oryx* 1–4.
- Tian, K., Zhang, G., Cheng, X., et al., 2002. The habitat fragility of *Manglietiastrum sinicum*. *Acta Bot. Yunnanica* 25, 551–556.
- Williamson, P.S., Werth, C.R., 1999. Levels and patterns of genetic variation in the endangered species *Abronia macrocarpa* (Nyctaginaceae). *Am. J. Bot.* 86, 293–301.
- Willi, Y., Van Buskirk, J., Hoffmann, A.A., 2006. Limits to the adaptive potential of small populations. *Annu. Rev. Ecol. Evol. Syst.* 37, 433–458.
- Wu, Z.Y., 1991. *Flora of Yunnan*. Science Press, Beijing.
- Yao, X.H., Ye, Q.G., Kang, M., et al., 2007. Microsatellite analysis reveals interpopulation differentiation and gene flow in endangered tree *Changiosyax dolichocarpa* (Styracaceae) with fragmented distribution in central China. *New Phytol.* 176, 472–480.
- Yeh, F.C., Yang, R.C., Boyle, T., 1999. POPGENE VERSION 1.31: Microsoft Window-based Free Software for Population Genetic Analysis. <ftp://ftp.microsoft.com/Softlib/HPGL/EXE>.
- Zhao, X.F., Ma, Y.P., Sun, W.B., et al., 2012. High genetic diversity and low differentiation of *Michelia coriacea* (Magnoliaceae), a critically endangered endemic in southeast Yunnan, China. *Int. J. Mol. Sci.* 13, 4396–4411.
- Zu, Y.G., Zhang, W.H., Yan, X.F., 1999. *Conservation Biology of the Endangered Plant Adenophora lobophylla* Hong. Science Press, Beijing.