

Possible conformations involved in the binding of neurotensin, xenopsin and bradykinin molecules to mast cell receptors

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1. INTRODUCTION

The study of binding of neurotensin molecule (NT, tridecapeptide pGlu¹-Leu²-Tyr³-Glu⁴-Asn⁵-Lys⁶-Pro⁷-Arg⁸-Arg⁹-Pro¹⁰-Tyr¹¹-Ile¹²-Leu¹³), of its analogues and fragments, as well as of several other peptide bioregulators to rat mast cells [1-3] has shown that xenopsin (XE, octapeptide pGlu¹-Gly²-Lys³-Arg⁴-Pro⁵-Trp⁶-Ile⁷-Leu⁸) and bradykinin (BK, nonapeptide Arg¹-Pro²-Pro³-Gly⁴-Phe⁵-Ser⁶-Pro⁷-Phe⁸-Arg⁹) compete successfully with neurotensin for the binding, whereas 15 other peptides are devoid of this property [2]. A comparable and relatively high ($K_d \sim 100$ nM) level of specific binding to the same type of receptors observed for different peptides is a rather unusual example of peptide ligand-receptor interaction; it is legitimate to assume that one of the reasons responsible for such an effect is the similarity shared by some primary sequence elements and especially 'binding conformations' of NT, XE and BK molecules. The data concerning the three-dimensional structure of NT, XE and BK molecules necessary for their comparison can be provided by means of energy calculations; it must be noted that no experimental evidence as to the conformations of NT and XE molecules is available so far.

2. METHODS AND RESULTS

The calculated sets of low-energy backbone structures for XE and BK have been described in

[4] and [5], respectively. Given below are the calculation results obtained for the NT molecule; the calculation procedures and the potential functions used were as in [4-7]. The general calculation pattern was essentially as for α -MSH molecule in [6]. During the first two steps, the 'model decapeptide' Glu⁴-Ala⁵-Lys⁶-Pro⁷-Arg⁸-Arg⁹-Pro¹⁰-Ala¹¹-Ala¹²-Ala¹³ containing all ionogenic groups of the molecule and the N-terminal pentapeptide pGlu¹-Leu²-Tyr³-Glu⁴-Asn⁵ were considered. Finally energy calculations were performed for all possible combinations of low-energy backbone structures for the two fragments overlapping in positions 4 and 5 of the peptide sequence, conformations of the Glu⁴ sidechain being the same as determined according to the 'model decapeptide' calculations. This final step (energy calculations for ~ 410 structures of the whole NT molecule) was accompanied by the refinement of side chain packing for Leu², Tyr³, Asn⁵, Tyr¹¹, Ile¹² and Leu¹³ residues by means of a specially designed algorithm [8].

The calculation resulted in 5 types of low-energy backbone structures found for the 4-13 fragment, which with optimal space orientation of the 1-3 fragment meet the requirement $\Delta U = U - U_{\min} \leq 10$ kcal/mol. These structures are listed in table 1. Keeping in mind that the 9-13 fragment possesses the same level of affinity to rat mast cell receptors as NT molecule itself [3], one can note that only 4 backbone structure types of this fragment are presented in table 1, namely BRBLL, BRRBL, BBLLB and BRBLB.

Table 1
Low-energy structures of neurotensin

| Residue | Angle | Molecular backbone conformation ^a | | | | | Residue | Angle | Molecular backbone conformation ^a | | | | |
|-------------------|----------|--|------|------|------|------|-----------------------------|--------|--|-------|-------|-------|------|
| | | 1 | 2 | 3 | 4 | 5 | | | 1 | 2 | 3 | 4 | 5 |
| pGlu ¹ | ψ | -32 | 151 | 151 | -36 | 146 | Arg ⁸ | ϕ | -114 | -133 | -128 | -140 | -138 |
| Leu ² | ϕ | -112 | -123 | -119 | -138 | -133 | ψ | 129 | 127 | 129 | 153 | 141 | |
| | ψ | 141 | 133 | 132 | 129 | 131 | χ_1 | -141 | -139 | -148 | -77 | -76 | |
| | χ_1 | -77 | -161 | -161 | -160 | -160 | χ_2 | -91 | -94 | -107 | 160 | 154 | |
| | χ_2 | 98 | 80 | 81 | 100 | 100 | χ_3 | 112 | 104 | 109 | -164 | -168 | |
| | | | | | | | χ_4 | 139 | 88 | 92 | -161 | 179 | |
| Tyr ³ | ϕ | 53 | -132 | -134 | -137 | -136 | Arg ⁹ | ϕ | -123 | -128 | -122 | -137 | -177 |
| | ψ | 110 | 139 | 142 | 137 | 133 | ψ | 151 | 149 | 137 | 138 | 127 | |
| | χ_1 | 165 | -58 | -61 | -60 | -60 | χ_1 | -57 | -53 | -57 | 57 | 43 | |
| | χ_2 | -101 | -80 | -81 | -80 | 99 | χ_2 | 173 | 172 | 152 | -166 | -152 | |
| | χ_3 | 100 | -80 | -80 | 80 | 80 | χ_3 | -65 | -61 | -66 | -179 | -179 | |
| Glu ⁴ | ϕ | 44 | 46 | 51 | 56 | 56 | χ_4 | -86 | -94 | -98 | 116 | 140 | |
| | ψ | 34 | 32 | 29 | 51 | 53 | Pro ¹⁰ | ψ | -28 | -24 | -15 | 127 | -31 |
| | χ_1 | -75 | -77 | -77 | -159 | -159 | Tyr ¹¹ | ϕ | -130 | -127 | -99 | 60 | -115 |
| | χ_2 | -174 | 179 | -177 | -145 | -144 | ψ | 146 | 137 | -44 | 134 | 145 | |
| | χ_3 | -96 | -99 | -97 | 120 | 138 | χ_1 | 43 | 61 | -4 | -60 | 40 | |
| Asn ⁵ | ϕ | 42 | 37 | 40 | 43 | 43 | χ_2 | 85 | 97 | 101 | 100 | 82 | |
| | ψ | 62 | 71 | 62 | 37 | 38 | χ_3 | 99 | 100 | 80 | 80 | 80 | |
| | χ_1 | -164 | -60 | -60 | 60 | 59 | Ile ¹² | ϕ | 39 | 37 | -135 | 45 | 33 |
| | χ_2 | 81 | -80 | -80 | 80 | 81 | ψ | 95 | 98 | 98 | 96 | 90 | |
| Lys ⁶ | ϕ | -129 | -136 | -135 | -148 | -147 | χ_1 | -49 | -58 | 56 | -59 | -42 | |
| | ψ | 132 | 130 | 131 | 140 | 133 | χ_2 | 122 | 100 | 120 | 100 | 120 | |
| | χ_1 | -78 | -79 | -75 | -77 | -75 | Leu ¹³ | ϕ | 48 | 50 | 26 | -96 | -104 |
| | χ_2 | 154 | 161 | 163 | 161 | 162 | ψ | 91 | 96 | 93 | 140 | 145 | |
| | χ_3 | -178 | -166 | -164 | -159 | -158 | χ_1 | -78 | -161 | -79 | -80 | -178 | |
| | χ_4 | 171 | 176 | 175 | 79 | 77 | χ_2 | 99 | 100 | 100 | 100 | 81 | |
| Pro ⁷ | ψ | -35 | -46 | -37 | -102 | 104 | $\Delta U(\text{kcal/mol})$ | 0.0 | 0.41 | 10.12 | 10.21 | 10.28 | |

^a Symbols B, R and L are used to denote local energy minima of the peptide backbone in B ($\phi \sim -140^\circ$, $\psi \sim 140^\circ$), R ($\phi \sim -60^\circ$, $\psi \sim -60^\circ$), L ($\phi \sim 60^\circ$, $\psi \sim 60^\circ$) and H ($\phi \sim 80^\circ$, $\psi \sim -80^\circ$) regions of potential maps: (1) RBLLLRBBRBL; (2) BBLLLRBBRBL; (3) BBLLLRBBRBL; (4) RBLLBBBBBL; (5) BBLLBBBBRBL

3. DISCUSSION

The 9–13 fragment responsible for the binding of NT molecule to mast cell receptors contains a ...Pro-X... sequence (X is an aromatic amino acid residue) which is common for NT, XE and BK molecules (see table 2). The data shown in table 2 allow us to assume that the possible 'binding site'

for NT molecule can be restricted to the 9–12 fragment only, since the shortening of the peptide chain due to omission of one C-terminal residue (BK) affects the affinity only insignificantly whereas that involving two residues (AT) leads to a complete lack of affinity. On the other hand, our previous energy calculations for XE [4] and BK [5] indicate that the 'binding site' conformations of

Table 2
Comparison of amino acid sequences of NT, XE, BK and AT molecules

| | | |
|--------------------------|------|--|
| Neurotensin | (NT) | pGlu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Arg-Pro-Tyr-Ile-Leu |
| Xenopsin | (XE) | pGlu-Gly-Lys-Arg-Pro-Trp-Ile-Leu |
| Bradykinin | (BK) | Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg |
| Angiotensin ^a | (AT) | Asp-Arg-Val-Tyr-Ile-His-Pro-Phe |

^a The only one among 15 peptides discussed in [2] (NT, XE and BK are not included) having a Pro-X sequence in its molecule

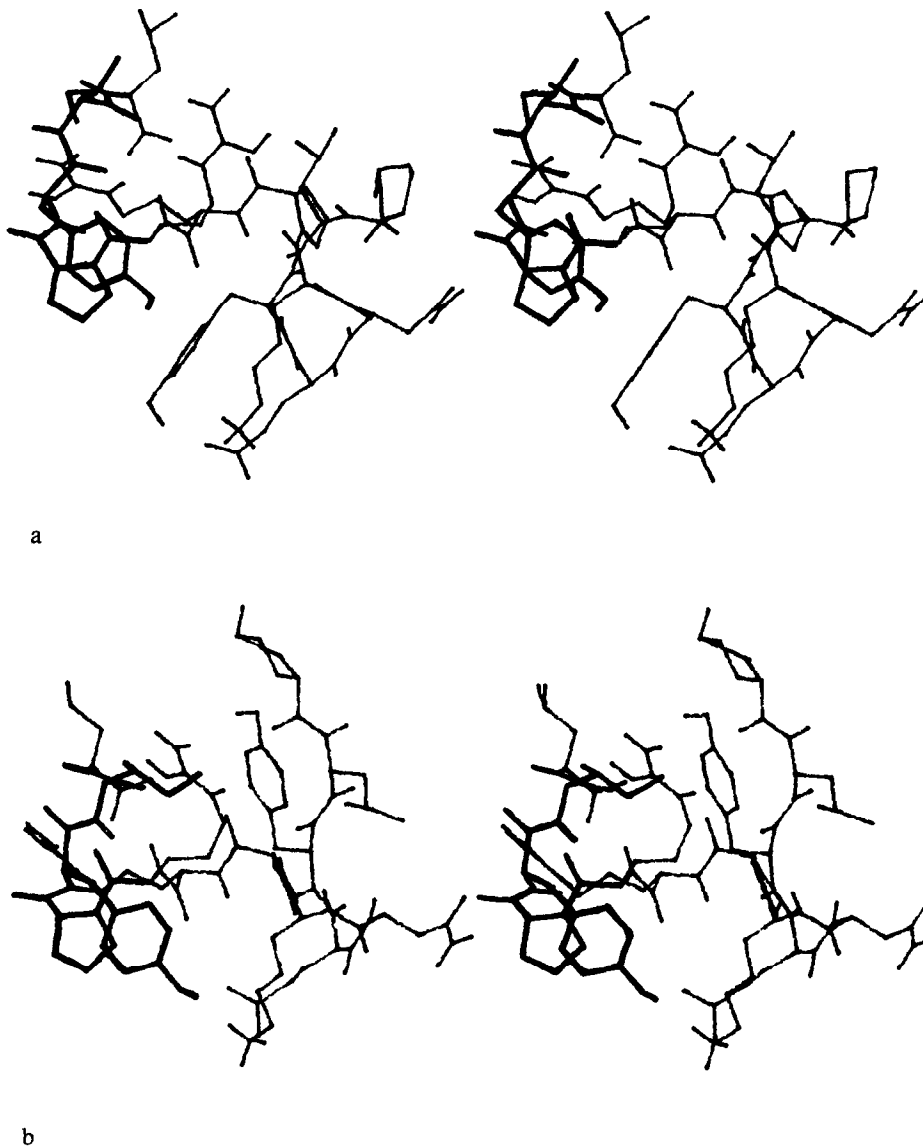


Fig.1. A stereo-view of neurotensin structures 1 and 3 (table 1): (—), site in the molecule directly involved in the binding to rat mast cells; (a) conformation of RBL type; (b) conformation of RRB type.

Table 3

Comparison of the complete sets of low-energy backbone conformations for NT, BK [5] and XE [4] molecules

| | | |
|------------|----------------|----|
| NT: | RBLLBRB BRBL L | * |
| | BBLLBRB BRBL L | * |
| | BBLLBRB BRRB L | ** |
| | RBLLBBB BBLL B | |
| | BBLLBBB BRBL B | * |
| BK: | BBBHB BRBB | |
| | BBRRR BRBL | * |
| | BBBHB BRRB | ** |
| | BBBHR BRBB | |
| | BBRRR BBBB | |
| | BBBHR BRRB | ** |
| | BBBHB BRRL | |
| | BBBHR BRBL | * |
| | BBRRB LBBB | |
| | BBBHR BBLB | |
| | BBRRB LRBB | |
| | BBRLR BBBB | |
| XE: | BRB BRBL L | * |
| | BHR BRBL B | * |
| | BRB BBLL B | |
| | BRB BRRB L | ** |
| | BBL LRRB B | |
| | RRB LRRB B | |
| BRB BRBL B | * | |

By one and two asterisks are marked conformations with the three-dimensional structure of the 'binding site' in all of three sets of low-energy backbone conformations

BRBL and BRRB types appear to be common for all three peptides in question (table 3).

However, the biological data obtained for various NT analogues with single replacements for amino acid enantiomers demonstrate that only

analogues with replacements in positions 10–12 exhibit a decreased level of affinity to receptor [3]. Therefore, the backbone conformation of B type in position 9 (which is sterically hindered for D-Arg residue) cannot represent the 'conformational' factor affecting the tightness of ligand–receptor binding. Thus, it may be concluded that tripeptides Pro¹⁰–Tyr¹¹–Ile¹² (NT), Pro⁵–Trp⁶–Ile⁷ (XE) and Pro⁷–Phe⁸–Arg⁹ (BK) can possibly be the fragments providing a certain complementarity of NT, XE and BK molecules to the 'common' binding site of rat mast cell receptors, their common 'binding conformation' being that of RBL or RRB type (fig.1a,b). It can be added finally, that the general shape of the two proposed 'binding conformation' types are essentially the same (fig.1): they differ only in the twist by ~180° of the peptide bond plane connecting the second and third residues of the binding site of the molecule.

REFERENCES

- [1] Lazarus, L.H., Perrin, M.H. and Brown, M.R. (1977) *J. Biol. Chem.* 252, 7174–7179.
- [2] Lazarus, L.H., Perrin, M.H. and Rivier, J.E. (1977) *J. Biol. Chem.* 252, 7180–7183.
- [3] Rivier, J.E., Lazarus, L.H., Perrin, M.H. and Brown, M.R. (1977) *J. Med. Chem.* 20, 1409–1412.
- [4] Nikiforovich, G.V., Betinsh, J.R., Podinsh, L.U. and Chipens, G.I. (1981) *Stud. Biophys.* 85, 107–113.
- [5] Nikiforovich, G.V. and Podinsh, L.U. (1982) *Bioorg. Khim.* 8, 453–461.
- [6] Nikiforovich, G.V., Shenderovich, M.D. and Chipens, G.I. (1981) *FEBS Lett.* 126, 180–182.
- [7] Nikiforovich, G.V., Leonova, V.I., Galaktionov, S.G. and Chipens, G.I. (1979) *Int. J. Peptide Protein Res.* 13, 363–373.
- [8] Nikiforovich, G.V., Shenderovich, M.D. and Balodis, J. (1981) *Bioorg. Khim.* 7, 179–188.