Cell, Vol. 110, 9–12, July 12, 2002, Copyright 2002 by Cell Press

p53: Good Cop/Bad Cop Minireview

Harvard Medical School with an unwanted long-term cost: aging. Boston, Massachusetts 02115 *The Bad Cop*

to a variety of cellular stresses, including DNA damage telomeric shortening, and oxidative stress. and oncogene activation, initiates a program of gene *DNA damage responses***. An age-related decrease of expression that blocks the proliferative expansion of DNA repair or increase in DNA damage has long been damaged cells. While the beneficial impact of the anti- thought to play a role in human aging (Figure 1). For cancer function of p53 is well established, several re- example, significant exposure to DNA damaging agents, cent papers suggest that p53 activation may in some such as chemo- or radiotherapy for malignancy, can circumstances act in a manner detrimental to the long- induce aging-like phenotypes such as impaired wound term homeostasis of the organism. Here, we discuss healing, early menopause, alopecia, and secondary cancer predisposition. Also, genetic conditions with proger- the significant participation of p53 in three non-mutually exclusive theories of human aging involving DNA oid features such as ataxia-telangectasia, Fanconi anedamage, telomere shortening, and oxidative stress. mia, and Werner and Bloom syndromes involve genes These "good cop/bad cop" functions of p53 appear to that sense DNA damage and/or participate in DNA repair place it at the nexus of two opposing forces, cancer processes. In mice, genetic deficiency of either nonand aging. By extension, this relationship implies that homologous end-joining (Ku80) or nucleotide excision repair (TTD), has been shown to engender signs of accel- therapies aimed to reduce cancer and postpone aging, and thereby increase longevity, will necessarily work erated aging (de Boer et al., 2002; Vogel et al., 1999).** either upstream or downstream, but not on the level **breaks and adducts) resulting from these genetic de- of, p53.**

tumor suppression is underscored by the fact that any tion, telomeric shortening is no longer an efficient antitation, reduced gene dosage or by p14^{ARF} inactivation
or MDM2 overexpression is associated with increased
fore, in this system, p53 determines whether short telo-

Norman E. Sharpless and Ronald A. DePinho¹ p53 is at work keeping us safe from cancer, the worst Department of Adult Oncology sort of cellular organized crime. Unfortunately, however, Dana-Farber Cancer Institute **inclusive and the set of the set of the set of the set of police force may erode civil liber-Department of Medicine and Genetics ties, so it now appears that excess p53 activity comes**

There are several, non-mutually exclusive theories of human aging, and it appears p53 has plausible links Activation of the p53 transcription factor in response to three of the most familiar: unrepaired DNA damage,

fects are sensed, at least in part, through p53. Thus, *The Good Cop*
We long-lived metazones find ourselves in a rough *Increasing amounts of DNA damage with advanced age,* **We long-lived metazoans find ourselves in a rough increasing amounts of DNA damage with advanced age,**

neighborhood. Each day, our component cells are bon-

bareans likely that p53 activation would play a role in

baread to background ionizing radiation and UV sun-

light, exposed to chemical mutagens such as aflatoxin,

li cancer mechanism, but rather fuels genomic instability **or MDM2 overexpression is associated with increased fore, in this system, p53 determines whether short telo** t meres will be sensed, leading to prematurely aged mice, **or ignored, leading to tumor-prone animals. It seems likely that p53 would play a similar role in human dis- ¹**

Figure 1. p53 Induces Aging in Response to Increased Metabolism, ROS, and DNA Damage

Telomere dysfunction engenders a bridgefusion-breakage cycle (BFB) that induces a DNA-damage, p53-dependent checkpoint response. Telomere shortening may also induce p53 through a DNA-damage-independent mechanism (dotted line). Impaired DNA repair and mutagen exposure lead to unrepaired DNA damage, but also may be associated with increased ROS (e.g., as a result of UV exposure or increased growth signaling). Increased metabolism from caloric excess and/ or insulin-like growth factor signaling induces ROS. The observation that Sir2 represses p53-mediated transactivation also suggests a DNA damage-independent link between metabolism and aging. Sir2 activity appears to be regulated by NAD levels and ROS levels as shown. p66SHC appears to be a downstream effector of p53 that mediates apoptosis and increases ROS levels. Loss of p66SHC leads to reduced levels of intracellular oxidants and extends murine lifespan, but *p66SHC***/ mice are not tumor prone.**

eases associated with critical telomeric shortening, as mutations known to increase the murine lifespan is loss of p66SHC further evidenced by the impact of p53 in cell culture , a protein that appears to be involved in both models of telomere dysfunction (Karlseder et al., 1999). the production of and the response to ROS (Migliaccio

of diseases like ataxia-telangectasia (associated with dent apoptotic response to oxidative stress is impaired, mutation of *ATM***) and Werner syndrome (associated and intracellular oxidants and oxidation-induced DNA** with mutation in *WRN*) results from impaired DNA repair damage are reduced (Migliaccio et al., 1999; Nemoto **and Finkel, 2002; Trinei et al., 2002). p66SHC and/or altered telomere metabolism, since both ATM appears to and WRN have been linked to processes of DNA repair be stabilized by p53, whereas p53 phosphorylation and** and telomere maintenance. Interestingly, the deficiency acetylation are not affected in $p66^{8HC-I}$ cells, and **in DNA repair and progeria are dissociated in** *Atm***-defi-** *p66SHC***/ mice are not tumor prone (Trinei et al., 2002). cient mice that display reduced DNA repair yet do not These observations suggest that not only could p53 be manifest the dramatic premature aging phenotype seen a sensor of ROS, but also that p53 might regulate ROS** in AT patients. It could be that the biochemical function levels (and therefore aging) through p66^{SHC}. **of ATM is partially fulfilled by some other protein in A further link between metabolism and aging has now the mouse, or alternatively that differences in telomere been suggested by the observation that p53 interacts reserve may influence the phenotype. Along these lines, with, and is deacetylated by, the human ortholog of mouse telomeres are much longer than those in humans, yeast Sir2 (Langley et al., 2002; Luo et al., 2001; Vaziri and the moderate degree of telomeric shortening evi- et al., 2001). Sir2 expression attenuates the ability of dent in** *Atm***-deficient mice would not be sufficient to p53 to transactivate target genes and to induce growth activate p53. It will be of interest therefore, to determine arrest and apoptosis, while a dominant-negative form if the effects of ATM loss would be altered in mice with of Sir2 potentiates p53-mediated apoptosis in response shortened human-like telomeres, and thereby under- to oxidative stress. It is not clear if this effect of Sir2 stand whether the progeroid features seen in AT patients requires deactylation of p53 directly, or if Sir2 is tethered** result from altered telomere metabolism, impaired DNA by p53 to the promoters of p53-responsive genes func**repair, or both. Whichever theory of aging prevails in tioning to repress target gene transcription through the this disease, however, it seems likely that p53 will figure established histone deactylase activity of Sir2. The relaprominently as an important downstream mediator. tionship between p53 and Sir2 is particularly intriguing,**

worms, flies, yeast, and mammals have linked oxidative whose overexpression extends the longevity of yeast, stress, metabolism, and aging (reviewed in Guarente and whose function is required for lifespan extension and Kenyon, 2000). Caloric restriction, or mutations that by caloric restriction in this system (Lin et al., 2000). All decrease glucose metabolism, will extend the lifespan of this has led to speculation that the dependence of Sir2 of many species, and it has been suggested that in on NAD concentration links metabolism and oxidative general a lower metabolic rate will decrease the produc- stress to the activity of this enzyme (Guarente, 2001). tion of toxic reactive oxygen species (ROS). In mammals In this model, free NAD is relatively scarce in times of at least, the response to oxidative stress appears to caloric excess because it is sequestered for electron involve p53. While it has long been thought that p53 transport within the glycolytic pathway. In times of calomight sense free-radical-induced DNA damage, two di-

<u>ric restriction</u>, however, NAD would be relatively abun**rect links between ROS, metabolism, and p53 have re- dant because of decreased glycolysis, which then would cently been forged (Figure 1). One of the few genetic induce Sir2 activity and limit p53 activity. Of note, this**

Debate exists as to whether the progeroid features et al., 1999). In $p66^{8HC-I}$ cells and mice, the p53-depen-

*Oxidative Stress***. Several lines of evidence from as** *Sir2* **encodes an NAD-dependent histone deacetylase**

DNA damage, as opposed to the aforementioned aging essary, the problem seems simpler. For example, our theories. However, it is not clear at present if Sir2 is present new understanding of the downside of p53 actipredominantly regulated under physiologic conditions vation might support the transient use of agents that by NAD levels, ROS, or both. Nonetheless, these recent prevent the induction of p53 or attenuate its function in data clearly establish an interaction between p53 and specific clinical circumstances, for example to protect Sir2, a protein known to modify longevity in lower organ-

non-diseased tissues in patients undergoing chemo**isms, and imply that Sir2 and NAD levels could couple or radiotherapy. In fact, Gudkov and coworkers have oxidative stress and metabolism with p53 activity in developed a small molecule inhibitor of p53, the use of mammals (Langley et al., 2002; Luo et al., 2001; Vaziri which protects mice from the toxicity of radiotherapy, et al., 2001). suggesting that many of the side effects of cancer treat-**

in sensing DNA damage, telomeric shortening, and oxi- al., 1999). Likewise, several adjuncts to modern oncodative stress (and in particular, its activity is regulated logic therapy (e.g., the administration of amifostine or by the latter), and it seems clear that p53, acting as autologous stem cell transplants) may work at the moa "bad cop," contributes to mammalian aging. Direct lecular level by sparing healthy tissue the effects of p53 evidence for this has been suggested by the generation activation. Of course, anticancer therapies that do not of a hyperfunctional allele of p53 in the mouse germline induce p53 have long been sought in medical oncology, (Tyner et al., 2002). These animals resist cancer, but but the success rate for identification of such comdevelop an accelerated aging phenotype highly reminis- pounds has been low. Despite the promise of receptor cent of that seen in animals with DNA repair defects or tyrosine kinase inhibitors, anti-angiogenesis therapy, and tumor vaccines, it is sobering to realize that nearly all telomere dysfunction. As these mice presumably have normal regulation of p53, with only increased transacti- curative regimens currently in use for advanced cancer vation of p53-dependent genes, this observation sug- depend at least in part on the use of DNA damaging gests that the level of p53 function determines the onset agents. of aging phenotypes (and therefore sets longevity) in a In healthy individuals, however, the postponement of direct way. This result is of particular importance as it **suggests that some aspects of mammalian aging are pect, as the pharmacologic long-term inhibition of p53 not directly the consequence of ROS or damage to spe- function would be complicated by increased tumorigen**esis. As $p66^{8HC-I}$ mice do not appear to be tumor prone, cific loci or genes per se, but rather stem from the organ-**Pelicci and colleagues have suggested it might be possi- ismal response to these factors. The conclusion suggested by these disparate lines of data is that p53, while ble to extend longevity by targeting this downstream** cracking down on neoplasia, also limits the repair and
regeneration of normal tissues.
function (Trinei et al., 2002). However, if p66^{SHC} inhibition

C. elegans, whose life spans are generally not limited
by tumorigenesis. Also, the p53 homologs of C. elegans
are and the sected to increase tumorigenesis, despite attenuated
and Drosophila are predominantly mediators of shown that this de facto anticancer mechanism comes
with an attendant cost of accelerated aging, indepen-
develop pharmacologic inhibitors of oxidative stress,
dent of the stimulus of p53 induction (Typer et al. 2002)
DNA dent of the stimulus of p53 induction (Tyner et al., 2002).

Therefore, it seems fair to say that some, but certainly

not all, aspects of human aging stem from the efforts

Neeping the "bad cop" off the streets.

Neeping **of p53 to inhibit tumor growth. Selected Reading It remains to be seen whether this new understanding**

of the dual role of p53 in cancer and aging will translate Chin, L., Artandi, S.E., Shen, Q., Tam, A., Lee, S.L., Gottlieb, G.J., into therapeutic advances. In patients with cancer, Greider, C.W., and DePinho, R.A. (1999). Cell *97***, 527–538.**

model of the role of p53 in aging does not depend upon where p53 activation is desirable and perhaps even nec-**As these data suggest, p53 is of principal importance ment stem from p53's "bad cop" functions (Komarov et**

Conclusions
Conclusions **in humans induces some unappreciated toxicity, the**
The "good con'had con" analogy of p53 raises the ques. remaining approaches to this problem appear to be up-The "good cop/bad cop" analogy of p53 raises the ques-
tion of the extent to which aging depends on p53 per
se. An extreme interpretation would be that all human
aging is an untoward result of an anticancer mechanism.
This

de Boer, J., Andressoo, J.O., de Wit, J., Huijmans, J., Beems, R.B., van Steeg, H., Weeda, G., van der Horst, G.T., van Leeuwen, W., Themmen, A.P., et al. (2002). Science *296***, 1276–1279.**

Greenberg, R.A., Chin, L., Femino, A., Lee, K.H., Gottlieb, G.J., Singer, R.H., Greider, C.W., and DePinho, R.A. (1999). Cell *97***, 515–525.**

Guarente, L. (2001). Trends Genet. *17***, 391–392.**

Guarente, L., and Kenyon, C. (2000). Nature *408***, 255–262.**

Hahn, W.C., and Weinberg, R.A. (2002). Nat. Rev. Cancer *2***, 331–341.**

Karlseder, J., Broccoli, D., Dai, Y., Hardy, S., and de Lange, T. (1999). Science *283***, 1321–1325.**

Komarov, P.G., Komarova, E.A., Kondratov, R.V., Christov-Tselkov, K., Coon, J.S., Chernov, M.V., and Gudkov, A.V. (1999). Science *285***, 1733–1737.**

Langley, E., Pearson, M., Faretta, M., Bauer, U.M., Frye, R.A., Minucci, S., Pelicci, P.G., and Kouzarides, T. (2002). EMBO J. *21***, 2383–2396.**

Lin, S.J., Defossez, P.A., and Guarente, L. (2000). Science *289***, 2126– 2128.**

Luo, J., Nikolaev, A.Y., Imai, S., Chen, D., Su, F., Shiloh, A., Guarente, L., and Gu, W. (2001). Cell *107***, 137–148.**

Migliaccio, E., Giorgio, M., Mele, S., Pelicci, G., Reboldi, P., Pandolfi, P.P., Lanfrancone, L., and Pelicci, P.G. (1999). Nature *402***, 309–313.**

Mori, H., Colman, S.M., Xiao, Z., Ford, A.M., Healy, L.E., Donaldson, C., Hows, J.M., Navarrete, C., and Greaves, M. (2002). Proc. Natl. Acad. Sci. USA *12***, 8242–8247.**

Nemoto, S., and Finkel, T. (2002). Science *295***, 2450–2452.**

Rudolph, K.L., Chang, S., Lee, H.W., Blasco, M., Gottlieb, G.J., Greider, C., and DePinho, R.A. (1999). Cell *96***, 701–712.**

Trinei, M., Giorgio, M., Cicalese, A., Barozzi, S., Ventura, A., Migliaccio, E., Milia, E., Padura, I.M., Raker, V.A., Maccarana, M., et al. (2002). Oncogene *21***, 3872–3878.**

Tyner, S.D., Venkatachalam, S., Choi, J., Jones, S., Ghebranious, N., Igelmann, H., Lu, X., Soron, G., Cooper, B., Brayton, C., et al. (2002). Nature *415***, 45–53.**

Vaziri, H., Dessain, S.K., Ng Eaton, E., Imai, S.I., Frye, R.A., Pandita, T.K., Guarente, L., and Weinberg, R.A. (2001). Cell *107***, 149–159.**

Vogel, H., Lim, D.S., Karsenty, G., Finegold, M., and Hasty, P. (1999). Proc. Natl. Acad. Sci. USA *96***, 10770–10775.**

Vulliamy, T., Marrone, A., Goldman, F., Dearlove, A., Bessler, M., Mason, P.J., and Dokal, I. (2001). Nature *413***, 432–435.**