



Data in Brief

Microarray gene expression profiling reveals potential mechanisms of tumor suppression by the class I HDAC-selective benzoylhydrazide inhibitors



Iqbal Mahmud, Daiqing Liao *

Department of Anatomy and Cell Biology, UF Health Cancer Center and UF Genetics Institute, University of Florida College of Medicine, Gainesville, FL 32610, United States

ARTICLE INFO

Article history:

Received 11 June 2015

Accepted 16 June 2015

Available online 18 June 2015

Keywords:

Microarray profiling

Class I histone deacetylase (HDAC) inhibitor

Pathway analysis

Cell cycle

Tumor suppression

ABSTRACT

Histone deacetylase (HDAC) inhibitors (HDIs) have therapeutic potentials for treating cancer and other diseases. Modulation of gene expression by HDIs is a major mechanism underlying their therapeutic effects. A novel class of HDIs with a previously undescribed benzoylhydrazide scaffold has been discovered through a high throughput screening campaign. Using microarray profiling of gene expression, we have previously demonstrated that treatment of breast cancer cells with a lead benzoylhydrazide HDI UF010 results in cell cycle arrest and apoptosis, likely through activation of tumor suppression pathways with concurrent inhibition of oncogenic pathways. In this brief report, we show methodological and analytical details and discuss additional pathways such as immune signaling that are affected by UF010. Raw and processed data from the microarray were deposited in NCBI's Gene Expression Omnibus (GEO) database under the accession number: GSE56823.

© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Specifications	
Organism/cell line/tissue	Human/MDA-MB-231/Breast, derived from metastatic site (pleural effusion)
Sex	Female (51 years adult)
Sequencer or array type	Affymetrix GeneChip Human Transcriptome Array 2.0
Data format	CEL files
Experimental factors	Cultured MDA-MB-231 cells exposed to DMSO control (n = 3) or benzoylhydrazide HDAC inhibitor UF010 (n = 3)
Experimental features	Assess effects of UF010 treatment on global gene expression in cancer cells
Consent	N/A
Sample source location	ATCC (www.atcc.org)

2. Experimental design, materials and methods

2.1. Experimental design

A novel class of small molecule HDIs with a benzoylhydrazide scaffold has been discovered recently [1]. They are specific to class I HDACs 1–3 and appear to exhibit fast-on/slow-off target-binding mechanism. Therefore, the new inhibitors are distinct chemically and mechanistically from known HDIs such as hydroxamic acids and benzamides. As histone deacetylation plays a major role in transcriptional regulation [2–4], we have assessed impact of the new HDIs on global gene expression. We used the triple-negative breast cancer cell line MDA-MB-231 and the benzoylhydrazide analog UF010 to interrogate effects of the new HDIs on gene expression. The experimental design is summarized in Fig. 1.

2.1.1. Cell culture and drug treatment

MDA-MB-231 cells were obtained from ATCC and cultured with Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% bovine calf serum, penicillin to 10 units/ml, and streptomycin to 10 µg/ml. Cells (500,000 cells per well) were seeded in a 6-well plate. At 24 h after seeding, dimethyl sulfoxide (DMSO) or UF010 was added. The final concentration for UF010 was 1 µM.

1. Direct link to deposited genomic data

<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE56823>.

* Corresponding author at: Department of Anatomy and Cell Biology, University of Florida College of Medicine, 1333 Center Drive, Gainesville, FL 32610, United States.
E-mail address: dliao@ufl.edu (D. Liao).

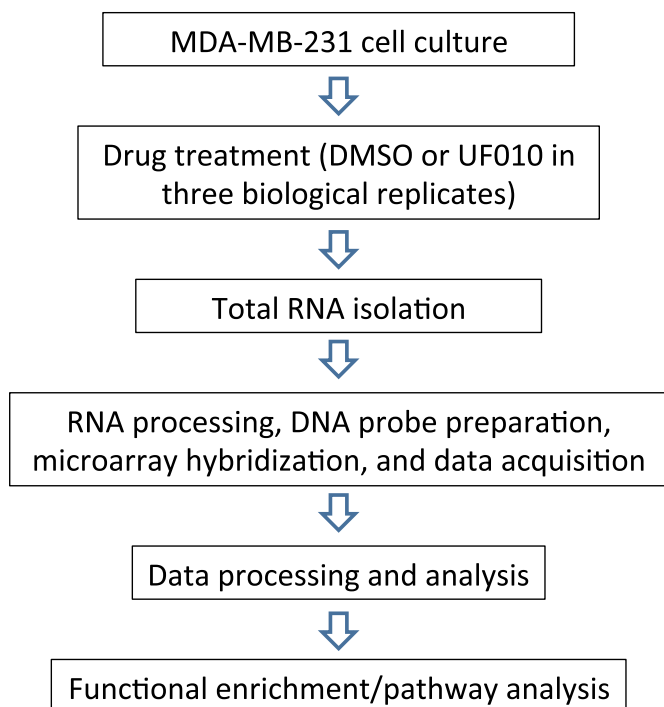


Fig. 1. A schematic diagram of gene expression profiling of MDA-MB-231 cells treated with a new HDI (UF010). MDA-MB-231 cells were cultured and then exposed to DMSO (control) or to UF010. Total RNAs were isolated from the treated cells and then processed for hybridization to microarray chips (Affymetrix GeneChip Human Transcriptome Array 2.0). The chips were scanned and data captured. Data were processed (background adjustment, summarization and normalization). The selected statistically significant genes in the experimental groups were analyzed for functional enrichments in certain pathways using Ingenuity Pathway Analysis (IPA) software.

2.1.2. RNA isolation and processing

Total RNAs from the treated cells were isolated using the RNeasy kit (Qiagen) and submitted to the Gene Expression Core of the University of Florida Interdisciplinary Center for Biotechnology Research. A NanoDrop Spectrophotometer (NanoDrop Technologies, Inc.) was used to determine RNA concentration and sample quality was assessed using an Agilent 2100 Bioanalyzer (Agilent Technologies, Inc.).

2.1.3. Microarray probe preparation, hybridization and data acquisition

The Ambion® WT Expression Kit, the GeneChip WT Terminal Labeling and Controls Kit (Affymetrix) were used for all microarray probe preparation following manufacturer's protocols. We used 200 ng of

total RNA as template for cDNA synthesis. The resulting cDNA was used as template for in vitro transcription (IVT) to generate antisense RNAs, which were then used to produce sense DNA. The sense strand DNA was fragmented, biotinylated, and hybridized with rotation at 45 °C for 16 h to microarray chips (GeneChip® Human Transcriptome Array 2.0, Affymetrix). The arrays were washed and stained with streptavidin-phycoerythrin (SAPE) with an Affymetrix Fluidics Station 450, and scanned using a GeneChip® 7G scanner (Affymetrix).

2.1.4. Microarray data quality control and analysis

The Affymetrix® Expression Console™ Software (Version 1.3) was used to generate.txt files for each RNA hybridization. All subsequent data analyses were performed in R 3.0.0 (<http://www.R-project.org/>). The Limma package [5] was used for background adjustment, summarization and quantile normalization. Normalization was made using the Robust Multichip Average (RMA) pre-normalization algorithm [6]. Data quality was assessed using various quality control measures. Specifically, density plots were generated to assess log-intensity distributions across a chip. The ideal distributions of the chips show no significant variation (Fig. 2A). An intensity boxplot (Fig. 2B) was used to compare the probe intensity levels between the arrays of the dataset. After normalization, the median lines are not significantly different from each other (Fig. 2B). A heatmap of the six samples was constructed to compare the UF010-treated cells with control (Fig. 2C). For each replicate array, each probe-set signal value from UF010-treated samples was compared to the probe-set signal value of DMSO-treated control samples to give gene expression ratios. Differentially expressed genes were identified using the Limma package with a Benjamini and Hochberg false discovery rate multiple testing correction. The statistically significant or differentially expressed genes were calculated using volcano plot analysis with a fold change (FC) threshold of >1.5 and a *p* value of <0.05.

2.1.5. Ingenuity pathway analysis

The Ingenuity Pathway Analysis (IPA) software (Ingenuity Systems Inc., Redwood City, CA) was used for functional enrichment analysis of the selected statistically significant genes in each of these experimental groups. The association between the genes in the dataset and a functional pathway were made using Fisher's exact test. Functional groups (or pathways) with a *P* value less than 0.05 and at least one focused molecule in a pathway were considered to be statistically significant.

3. Results

Analyses of the microarray dataset reveal that the benzoylhydrazide HDI UF010 induces gene activation and repression, consistent with findings by others that HDIs can up and downregulate gene expression

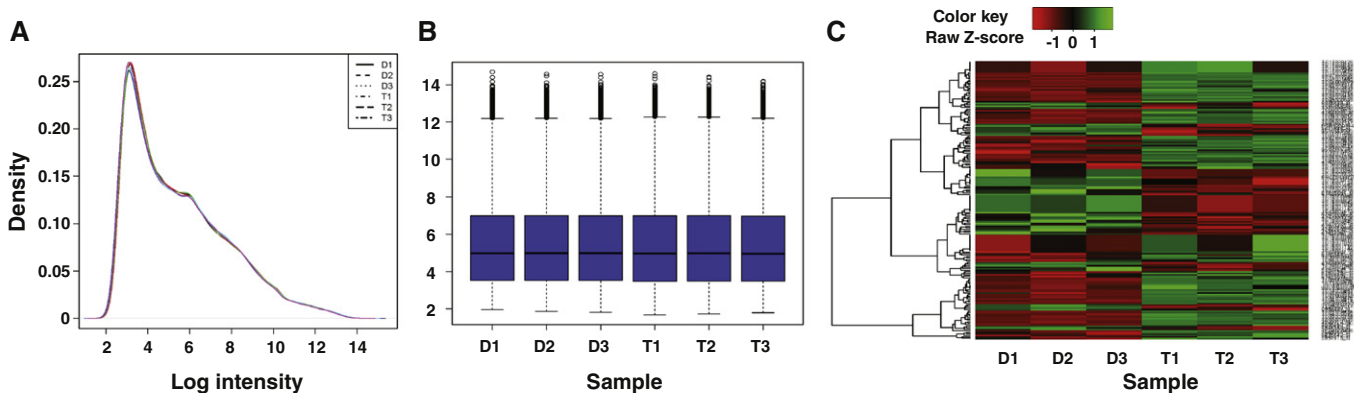


Fig. 2. Quality control assessments for the dataset. A, a plot of the log-intensity distribution in the experimental chips of the dataset. B, intensity box plot. The plot compares the probe intensity levels between the arrays of the dataset. Either end of the box represents the upper and lower quartile. The line in the middle of the box represents the median. C, a heatmap comparison of the six samples in the dataset. D, DMSO; T, UF010 treatment.

[7–12]. The expression of cell cycle regulators is specifically perturbed by UF010 with downregulation of genes that promote and upregulation of those that inhibit cell-cycle progression [1]. Induction of cell cycle arrest and cell death is a common mechanism of action by HDIs irrespective of cancer or inhibitor types [1,8,9]. Interestingly, UF010 treatment activates major histocompatibility complex (MHC) genes and other genes involved in immune response. Pathway analysis suggests that UF010 specifically induces B-cell receptor, IL-6 and IL-4 signaling. Activation of MHC and immune-related genes by other HDIs in prostate cancer cell lines has been reported [8].

4. Discussion

The class I HDACs 1–3 play important roles in gene regulation and are overexpressed in cancer [13–17]. HDACs 1 and 2 are closely related and assembled together in deacetylase complexes such as Sin3, NuRD and CoREST [18,19]. HDAC3 is found in the NCOR complex and depends on the deacetylase activation domain (DAD) of NCOR1 or NCOR2 and D-myo-inositol-(1,4,5,6)-tetrakisphosphate for catalyzing deacetylation reaction [20]. Thus, these HDACs may regulate gene expression in a different way and HDIs specific to an individual HDAC isoform may exert distinct impact on gene expression. Different chemical types of small molecule HDIs have been discovered. Hydroxamic acids are potent HDIs and exhibit strong Zn^{2+} -binding property, which limits their isoform selectivity. Nonetheless, isoform selectivity can be achieved through modifying the “cap” moiety that interacts with the residues in the rim outside the substrate tunnel [21]. Benzamide analogs occupy the catalytic center of HDACs and coordinate Zn^{2+} using the amine and carbonyl groups in a bidentate manner [22]. Benzamide derivatives with a five or six-membered aromatic ring “internal cavity” motif exhibit strong selectivity for HDACs 1 and 2 vs. HDAC3 [22], while HDAC3 selectivity is achieved by modifying the “cap” groups [21]. The benzoylhydrazide HDIs appear to use a linear aliphatic chain to bind the internal hydrophobic cavity of HDACs and they display notable selectivity for HDAC3, while inhibiting HDACs 1 and 2 with similar potency [1]. UF010 shows a fast-on/slow-off target binding mechanism. Whether the unique chemistry and pharmacology of the benzoylhydrazide HDIs can result in distinct modulation of gene expression and therapeutic effects remains to be determined.

Conflict of interest

None.

Acknowledgments

The authors thank the University of Florida Interdisciplinary Center for Biotechnology Research Gene Expression & Genotype Core (Dr. Yanping Zhang) and Bioinformatics Core (Dr. Jiqiang Yao) for microarray data acquisition and analysis. This work was supported by grants from the Bankhead-Coley Cancer Research Program, Florida Department of Health and the UF Health Cancer Center.

References

- [1] Y. Wang, R.L. Stowe, C.E. Pinello, G. Tian, F. Madoux, D. Li, L.Y. Zhao, J.L. Li, Y. Wang, Y. Wang, H. Ma, P. Hodder, W.R. Roush, D. Liao, Identification of histone deacetylase inhibitors with benzoylhydrazide scaffold that selectively inhibit class I histone deacetylases. *Chem. Biol.* 22 (2015) 273–284 (<http://www.ncbi.nlm.nih.gov/pubmed/25699604>).
- [2] C.A. Hassig, T.C. Fleischer, A.N. Billin, S.L. Schreiber, D.E. Ayer, Histone deacetylase activity is required for full transcriptional repression by mSin3A. *Cell* 89 (1997) 341–347 (<http://www.ncbi.nlm.nih.gov/pubmed/9150133>).
- [3] L. Nagy, H.Y. Kao, D. Chakravarti, R.J. Lin, C.A. Hassig, D.E. Ayer, S.L. Schreiber, R.M. Evans, Nuclear receptor repression mediated by a complex containing SMRT, mSin3A, and histone deacetylase. *Cell* 89 (1997) 373–380 (<http://www.ncbi.nlm.nih.gov/pubmed/9150137>).
- [4] Z. Wang, C. Zang, K. Cui, D.E. Schones, A. Barski, W. Peng, K. Zhao, Genome-wide mapping of HATs and HDACs reveals distinct functions in active and inactive genes. *Cell* 138 (2009) 1019–1031 (<http://www.ncbi.nlm.nih.gov/pubmed/19698979>).
- [5] G.K. Smyth, Linear models and empirical Bayes methods for assessing differential expression in microarray experiments. *Stat. Appl. Genet. Mol. Biol.* 3 (2004) (Article 3. <http://www.ncbi.nlm.nih.gov/pubmed/16646809>).
- [6] R.A. Irizarry, B. Hobbs, F. Collin, Y.D. Beazer-Barclay, K.J. Antonellis, U. Scherf, T.P. Speed, Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* 4 (2003) 249–264 (<http://www.ncbi.nlm.nih.gov/pubmed/12925520>).
- [7] Y.J. Kim, C.B. Greer, K.R. Cecchini, L.N. Harris, D.P. Tuck, T.H. Kim, HDAC inhibitors induce transcriptional repression of high copy number genes in breast cancer through elongation blockade. *Oncogene* 32 (2013) 2828–2835 (<http://www.ncbi.nlm.nih.gov/pubmed/23435418>).
- [8] M.S. Kortenhorst, M.D. Wissing, R. Rodriguez, S.K. Kachhap, J.J. Jans, P. Van der Groep, H.M. Verheul, A. Gupta, P.O. Aiyetan, E. van der Wall, M.A. Carducci, P.J. Van Diest, L. Marchionni, Analysis of the genomic response of human prostate cancer cells to histone deacetylase inhibitors. *Epigenetics* 8 (2013) 907–920 (<http://www.ncbi.nlm.nih.gov/pubmed/23880963>).
- [9] J.E. Bolden, W. Shi, K. Jankowski, C.Y. Kan, L. Cluse, B.P. Martin, K.L. MacKenzie, G.K. Smyth, R.W. Johnstone, HDAC inhibitors induce tumor-cell-selective pro-apoptotic transcriptional responses. *Cell Death Dis.* 4 (2013) e519 (<http://www.ncbi.nlm.nih.gov/pubmed/23449455>).
- [10] Y. Huang, S.N. Vasilatos, L. Boric, P.G. Shaw, N.E. Davidson, Inhibitors of histone demethylation and histone deacetylation cooperate in regulating gene expression and inhibiting growth in human breast cancer cells. *Breast Cancer Res. Treat.* 131 (2012) 777–789 (<http://www.ncbi.nlm.nih.gov/pubmed/21452019>).
- [11] T. Roger, J. Lugin, D. Le Roy, G. Goy, M. Mombelli, T. Koessler, X.C. Ding, A.L. Chanson, M.K. Reymond, I. Miconnet, J. Schrenzel, P. Francois, T. Calandra, Histone deacetylase inhibitors impair innate immune responses to Toll-like receptor agonists and to infection. *Blood* 117 (2011) 1205–1217 (<http://www.ncbi.nlm.nih.gov/pubmed/20956800>).
- [12] M.J. LaBonte, P.M. Wilson, W. Fazzzone, S. Groshen, H.J. Lenz, R.D. Ladner, DNA microarray profiling of genes differentially regulated by the histone deacetylase inhibitors vorinostat and LBH589 in colon cancer cell lines. *BMC Med. Genet.* 2 (2009) 67 (<http://www.ncbi.nlm.nih.gov/pubmed/19948057>).
- [13] B.M. Muller, J. Jana, A. Kasajima, A. Lehmann, J. Prinzler, J. Budczies, K.J. Winzer, M. Dietel, W. Weichert, C. Denkert, Differential expression of histone deacetylases HDAC1, 2 and 3 in human breast cancer—overexpression of HDAC2 and HDAC3 is associated with clinicopathological indicators of disease progression. *BMC Cancer* 13 (2013) 215 (<http://www.ncbi.nlm.nih.gov/pubmed/23627572>).
- [14] W. Weichert, C. Denkert, A. Noske, S. Darb-Esfahani, M. Dietel, S.E. Kalloger, D.G. Huntsman, M. Kobel, Expression of class I histone deacetylases indicates poor prognosis in endometrioid subtypes of ovarian and endometrial carcinomas. *Neoplasia* 10 (2008) 1021–1027 (<http://www.ncbi.nlm.nih.gov/pubmed/18714364>).
- [15] W. Weichert, A. Roske, V. Gekeler, T. Beckers, M.P. Ebert, M. Pross, M. Dietel, C. Denkert, C. Rocken, Association of patterns of class I histone deacetylase expression with patient prognosis in gastric cancer: a retrospective analysis. *Lancet Oncol.* 9 (2008) 139–148 (<http://www.ncbi.nlm.nih.gov/pubmed/18207460>).
- [16] W. Weichert, A. Roske, V. Gekeler, T. Beckers, C. Stephan, K. Jung, F.R. Fritzsche, S. Niesporek, C. Denkert, M. Dietel, G. Kristiansen, Histone deacetylases 1, 2 and 3 are highly expressed in prostate cancer and HDAC2 expression is associated with shorter PSA relapse time after radical prostatectomy. *Br. J. Cancer* 98 (2008) 604–610 (<http://www.ncbi.nlm.nih.gov/pubmed/18212746>).
- [17] W. Weichert, A. Roske, S. Niesporek, A. Noske, A.C. Buckendahl, M. Dietel, V. Gekeler, M. Boehm, T. Beckers, C. Denkert, Class I histone deacetylase expression has independent prognostic impact in human colorectal cancer: specific role of class I histone deacetylases in vitro and in vivo. *Clin. Cancer Res.* 14 (2008) 1669–1677 (<http://www.ncbi.nlm.nih.gov/pubmed/18347167>).
- [18] X.J. Yang, E. Seto, The Rpd3/Hda1 family of lysine deacetylases: from bacteria and yeast to mice and men. *Nat. Rev. Mol. Cell Biol.* 9 (2008) 206–218 (<http://www.ncbi.nlm.nih.gov/pubmed/18292778>).
- [19] G.P. Delcuve, D.H. Khan, J.R. Davie, Roles of histone deacetylases in epigenetic regulation: emerging paradigms from studies with inhibitors. *Clin. Epigenetics* 4 (2012) 5 (<http://www.ncbi.nlm.nih.gov/pubmed/22414492>).
- [20] P.J. Watson, L. Fairall, G.M. Santos, J.W. Schwabe, Structure of HDAC3 bound to co-repressor and inositol tetraphosphate. *Nature* 481 (2012) 335–340 (<http://www.ncbi.nlm.nih.gov/pubmed/22230954>).
- [21] F.F. Wagner, M. Weiwer, M.C. Lewis, E.B. Holson, Small molecule inhibitors of zinc-dependent histone deacetylases. *Neurotherapeutics* 10 (2013) 589–604 (<http://www.ncbi.nlm.nih.gov/pubmed/24101253>).
- [22] B.E. Lauffer, R. Mintzer, R. Fong, S. Mukund, C. Tam, I. Zilberleyb, B. Flicke, A. Ritscher, G. Fedorowicz, R. Vallero, D.F. Ortwin, J. Gunzner, Z. Modrusan, L. Neumann, C.M. Koth, P.J. Lupardus, J.S. Kaminker, C.E. Heise, P. Steiner, Histone deacetylase (HDAC) inhibitor kinetic rate constants correlate with cellular histone acetylation but not transcription and cell viability. *J. Biol. Chem.* 288 (2013) 26926–26943 (<http://www.ncbi.nlm.nih.gov/pubmed/23897821>).