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STATE OF THE ART: REVIEW OF LIPIDS AND PROGRESSIVE RENAL DISEASE

Recent advances in statins and the kidney

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Recent advances in statins and the kidney.

Background. Experimental and clinical studies have suggested a correlation between the progression of renal disease and dyslipidemia. Indeed, apolipoprotein B-containing lipoproteins have been demonstrated to be an independent risk factor for the progression of renal disease in humans. Interventional strategies in experimental models of renal disease have clearly demonstrated a beneficial effect on renal structure and function in a variety of models of renal disease. Investigations into the mechanisms whereby reduction of lipids by lipid-lowering agents benefits renal disease have suggested that the 3-hydroxy-3-methylglutaryl coenzyme reductase inhibitors, the so-called statin class of lipid-lowering agents, may have additional effects on the biology of inflammation that are germane to the progression of renal disease.

Methods. Both *in vivo* and *in vitro* studies that investigated secondary mechanisms of statin effects are reviewed. In addition, new studies that investigated the effects on novel cellular mechanisms are presented.

Results. Lipid-lowering agents appear to have biologically important effects in modulating a variety of intracellular signaling systems involved in cell proliferation, inflammatory responses that involve macrophage adhesion, recruitment, and maturation. In addition, the effects on fibrogenesis have been recently defined. These latter effects may influence not only the development of glomerulosclerosis, but also interstitial fibrosis. These potentially major effects of lipid-lowering agents appear to be related to the effects on intracellular synthesis of nonsterol isoprenoids, which are involved in prenylation of critical small molecular weight proteins involved in cell signal transduction.

Conclusions. In addition to the beneficial effects of the reduction in serum lipids, statins and other lipid-lowering agents may influence important intracellular pathways that are involved in the inflammatory and fibrogenic responses, which are common components of many forms of progressive renal injury.

Experimental evidence suggests that lipids are important modulators of progressive renal disease. In recent years, the inhibitors of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, so called "statins," have demonstrated beneficial effects in different models of progressive renal failure. Lovastatin ameliorated the ex-

tent of glomerular injury in 5/6 nephrectomy Sprague-Dawley rats [1], obese Zucker rats [2], Dahl-sensitive rats [3], guinea pigs (abstract; Gröne et al, *J Am Soc Nephrol* 3:739, 1992), and the puromycin aminonucleoside model of the nephrotic syndrome [4]. Lovastatin also retarded the development of polycystic kidney disease in the Han:SPRD rats [5], ischemic renal failure in high-cholesterol-loaded rats [6], and hypertension in spontaneously hypertensive rats [7]. Simvastatin suppressed the cell proliferation in rats with anti-Thy-1.1 nephritis [8]. Interestingly, some of the beneficial effects of statins can be seen independent of the cholesterol reduction (abstract; Kauffman et al, *FASEB J* 319:8, 1994) [4, 9]. These beneficial effects of statins on these *in vivo* animal models have been investigated using *in vitro* studies. In this regard, the effects of statins on the interaction of inflammatory cytokines, cell proliferation, and intracellular signaling pathways have recently been reported.

STATINS AND INFLAMMATORY CHEMOKINES

Macrophages participate in the uptake and metabolism of lipids in glomeruli, which may play a critical role in the pathogenesis of lipid-induced glomerulosclerosis [10-12]. The mechanisms whereby monocytes are recruited to glomeruli are poorly understood; however, mesangial cells stimulated by lipids [13] or inflammatory cytokines have been shown to produce important monocyte chemokines such as monocyte chemoattractant protein-1 (MCP-1) and macrophage-colony stimulating factor (M-CSF) [14, 15]. We have reported that lovastatin can reduce *in vitro* mesangial cell expression and the production of MCP-1 [16, 17] and M-CSF, as well as vascular cell adhesion molecule-1 (VCAM) and intracellular adhesion molecule-1 (ICAM-1), which are integral adhesion molecules [18]. Moreover, lovastatin inhibited the activation of transcription factor nuclear factor- κ B (NF- κ B), which plays a major role in the gene expression involved in mesangial cell inflammatory responses [19], suggesting that statins have modulating effects on the intracellular signaling pathways activated by inflammation.

Key words: fibrinogen, dyslipidemia, apolipoprotein, lipid-lowering agents.

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STATINS AND CELL PROLIFERATION

We and others have reported that mesangial cells proliferate in response to various growth factors such as platelet-derived growth factor (PDGF), insulin, and insulin-like growth factor-1, as well as lipoproteins such as low-density lipoprotein (LDL), intermediate density lipoprotein (IDL), and very low-density lipoprotein (VLDL) [20–22]. *In vitro* experiments have shown that statins inhibit proliferation of cultured mesangial cells [23, 24], renal epithelial tubular cells [25], and vascular smooth muscle cells [26–28]. Although statins inhibit intracellular cholesterol synthesis, they also inhibit the formation of intermediate metabolites of the mevalonate pathway [23], particularly the nonsterol isoprenoids, which appear to be essential in cell replication.

INVOLVEMENT OF STATINS IN THE SIGNALING PATHWAYS

Mevalonate was reported to be an important product of HMG-CoA reductase for the progression of a cell cycle [29–31]. Recently, however, the important roles of the mevalonate metabolites such as farnesyl pyrophosphate (FPP) and geranylgeranyl pyrophosphate (GGPP) have been demonstrated to be indispensable anchors of small G proteins, such as p21ras, that allow it to bind to the cell membrane [32–34]. Binding of certain growth factors to its receptor induces p21ras activation in combination with Grb2-Sos-Shc binding to the receptor and p21ras. The signal is transmitted to the nucleus through a pathway that consists of Raf, MAPK-ERK kinase, and the mitogen-activated protein (MAP) kinase cascade. In the nucleus, the transcription factors such as activated protein-1 (AP-1), NF- κ B, NF-interleukin-6 (NF-IL-6), and Egr-1 are induced, as well as early response genes, and subsequent cell proliferation or differentiation occurs. Reduction of FPP and GGPP by statins may effectively block the p21ras-mediated mitogenic signaling event [19, 35]. Lovastatin suppressed the expression of c-fos and c-jun in conjunction with a reduction in membrane-bound p21ras. In addition, it has been shown in renal tubular epithelial cells to functionally impair AP-1 binding to DNA [25]. This could, in part, contribute to the reported beneficial effect of statins on reducing tubulointerstitial inflammation in models of progressive renal disease. Lovastatin has recently been shown to inhibit isoprenylation of p21RhoA, a small G protein, and thereby induce apoptosis by inhibiting mitotic and postmitotic signals in cultured mesangial cells [36]. Similarly, atorvastatin decreased isoprenylation of p21RhoB, a small G protein, and induced apoptosis in vascular smooth muscle cells in culture [37]. Thus, statins appear to regulate cellular signaling pathways involved in proliferation and apoptosis.

In recent years, the involvement of statins in the cell-

cycle regulation has received considerable attention. Lovastatin induced p27kip1, a cyclin-dependent kinase inhibitor, resulting in a cell-cycle arrest at G1 phase [38, 39]. These effects of statins on nuclear events may be explained by the observation that pravastatin inhibited geranylgeranylation of p21RhoA and p21RhoB, two small GTPase(s). Isoprenylation of these factors is essential for p27Kip1 degradation and subsequent cell-cycle progression from the G1 to S phase [40]. In our study, lovastatin reduced DNA synthesis by inducing p27Kip1 and suppressing cyclin D1 and E expression in cultured rat vascular smooth muscle cells [41].

Additional cellular effects of statins have been reported in recent studies. For example, simvastatin has been shown to improve vascular hemodynamic responses, suggesting a direct or indirect effect on endothelial-derived nitric oxide [42]. Atorvastatin and simvastatin were able to prevent the reduction in mRNA expression and protein levels of endothelial cell nitric oxide synthase after exposure to oxidized LDL [43]. Pravastatin [44] and lovastatin [45] have also been reported to improve vasomotor tone and endothelial function in patients with coronary artery disease in association with cholesterol reduction. The inhibitory effects of lovastatin and fluvastatin on platelet activation have been shown to reduce platelet aggregation via altered platelet lipid composition [46].

Thus, both *in vitro* and *in vivo* studies have demonstrated that statins may have important effects on the pathophysiology of progressive renal disease by modifying monocyte infiltration, mesangial cell proliferation, and mesangial matrix expansion, as well as tubulointerstitial inflammation and fibrosis. These effects appear to be related, at least in part, to inhibition of small G-protein isoprenylation involved in early gene products, transcription factors, and modulation of the cell-cycle regulatory proteins. Although these data need further study in humans, statins may be an important and beneficial therapeutic strategy for progressive renal diseases.

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APPENDIX

Abbreviations used in the paper are: AP-1, activator protein-1; Egr-1, early growth response factor-1; ERK, extracellular signal-regulated protein; Grb2, growth factor receptor-bound protein 2; HMG-CoA, 3-hydroxy-methylglutaryl coenzyme A; ICAM-1, intracellular adhesion molecule-1; LDL, low density lipoprotein; MAPK, mitogen activated protein kinase; MCP-1, monocyte chemoattractant protein-1; M-CSF, macrophage-colony stimulating factor; MEK, MAPK-ERK kinase; NF- κ B, nuclear factor- κ B; PDGF, platelet-derived growth factor; Shc, SH2 containing protein; Sos, son of sevenless; VCAM-1, vascular cell adhesion molecule-1.

REFERENCES

- KASISKE BL, O'DONNELL MP, GARVIS WJ, KEANE WF: Pharmacologic treatment of hyperlipidemia reduces glomerular injury in rat 5/6 nephrectomy model of chronic renal failure. *Circ Res* 62:367-374, 1988
- O'DONNELL MP, KASISKE BL, KIM Y, SCHMITZ PG, KEANE WF: Lovastatin retards the progression of established glomerular disease in obese Zucker rats. *Am J Kidney Dis* 22:83-89, 1993
- O'DONNELL MP, KASISKE BL, KATZ SA, SCHMITZ PG, KEANE WF: Lovastatin but not enalapril reduces glomerular injury in Dahl salt-sensitive rats. *Hypertension* 20:651-658, 1992
- HARRIS KP, PURKERSON ML, YATES J, KLAHR S: Lovastatin ameliorates the development of glomerulosclerosis and uremia in experimental nephrotic syndrome. *Am J Kidney Dis* 15:16-23, 1990
- GILE RD, COWLEY BD JR, GATTONE VH II, O'DONNELL MP, SWAN SK, GRANTHAM JJ: Effect of lovastatin on the development of polycystic kidney disease in the Han:SPRD rat. *Am J Kidney Dis* 26:501-507, 1995
- IAINA A, BENYAMIN G, LEVTOV O, GETTER R, SERBAN I, WOLLMAN Y, RUBINSTEIN A, CABILI S, PEER G, BLUM M: Effect of chronic cholesterol loading in the development of acute ischemic renal failure in rats. *Renal Fail* 16:117-123, 1994
- JIANG J, ROMAN RJ: Lovastatin prevents development of hypertension in spontaneously hypertensive rats. *Hypertension* 30:968-974, 1997
- YOSHIMURA A, INUI K, NEMOTO T, UDA S, SUGENOYA Y, WATANABE S, YOKOTA N, TAIRA T, IWASAKI S, IDEURA T: Simvastatin suppresses glomerular cell proliferation and macrophage infiltration in rats with mesangial proliferative nephritis. *J Am Soc Nephrol* 9:2027-2039, 1998
- KASISKE BL, O'DONNELL MP, KIM Y, ATLURU D, KEANE WF: Cholesterol synthesis inhibitors inhibit more than cholesterol synthesis. *Kidney Int* 45(Suppl 45):S51-S53, 1994
- KEANE WF, KASISKE BL, O'DONNELL MP, KIM Y: The role of altered lipid metabolism in the progression of renal disease: Experimental evidence. *Am J Kidney Dis* 17(Suppl 1):38-42, 1991
- GUIJARRO C, KEANE WF: Lipid abnormalities and changes in plasma proteins in glomerular diseases and chronic renal failure. *Curr Opin Nephrol Hypertens* 2:372-379, 1993
- PESEK-DIAMOND I, DING G, FRYE J, DIAMOND JR: Macrophages mediate adverse effects of cholesterol feeding in experimental nephrosis. *Am J Physiol* 263:F776-F783, 1992
- ROVIN BH, TAN LC: LDL stimulates mesangial fibronectin production and chemoattractant expression. *Kidney Int* 43:218-225, 1993
- SATRIANO JA, HORA K, SHAN Z, STANLEY ER, MORI T, SCHLONDORFF D: Regulation of monocyte chemoattractant protein-1 and macrophage colony-stimulating factor-1 by IFN- γ , tumor necrosis factor- α , IgG aggregates, and cAMP in mouse mesangial cells. *J Immunol* 150:1971-1978, 1993
- MORI T, BARTOCCI A, SATRIANO J, ZUCKERMAN A, STANLEY R, SANTIAGO A, SCHLONDORFF D: Mouse mesangial cells produce colony-stimulating factor-1 (CSF-1) and express the CSF-1 receptor. *J Immunol* 144:4697-4702, 1990
- KIM SY, GUIJARRO C, O'DONNELL MP, KASISKE BL, KIM Y, KEANE WF: Human mesangial cell production of monocyte chemoattractant protein-1: Modulation by lovastatin. *Kidney Int* 48:363-371, 1995
- PARK YS, GUIJARRO C, KIM Y, MASSY ZA, KASISKE BL, KEANE WF, O'DONNELL MP: Lovastatin reduces glomerular macrophage influx and expression of monocyte chemoattractant protein-1 mRNA in nephrotic rats. *Am J Kidney Dis* 31:190-194, 1998
- GUIJARRO C, KEANE WF: Effects of lipids on the pathogenesis of progressive renal failure: Role of 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors in the prevention of glomerulosclerosis. *Miner Electrolyte Metab* 22:147-152, 1996
- GUIJARRO C, KIM Y, SCHOONOVER CM, MASSY ZA, O'DONNELL MP, KASISKE BL, KEANE WF, KASHTAN CE: Lovastatin inhibits lipopolysaccharide-induced NF- κ B activation in human mesangial cells. *Nephrol Dial Transplant* 11:990-996, 1996
- KEANE WF, O'DONNELL MP, KASISKE BL, KIM Y: Oxidative modification of low-density lipoproteins by mesangial cells. *J Am Soc Nephrol* 4:187-194, 1993
- NISHIDA Y, YORIOKA N, ODA H, YAMAKIDO M: Effect of lipoproteins on cultured human mesangial cells. *Am J Kidney Dis* 29:919-930, 1997
- NISHIDA Y, YORIOKA N, ODA H, ASAKIMORI Y, AMIMOTO D, YAMAKIDO M: Intermediate-density lipoprotein is a DNA synthesis stimulation factor in cultured human mesangial cells. *Nephron* 73:334-335, 1996
- O'DONNELL MP, KASISKE BL, KIM Y, ATLURU D, KEANE WF: Lovastatin inhibits proliferation of rat mesangial cells. *J Clin Invest* 91:83-87, 1993
- GRANDALIANO G, BISWAS P, CHOUDHURY GG, ABBOD HE: Simvastatin inhibits PDGF-induced DNA synthesis in human glomerular mesangial cells. *Kidney Int* 44:503-508, 1993
- VRTOVNIK F, COUETTE S, PRIÉ D, LALLEMAND D, FRIEDLANDER G: Lovastatin-induced inhibition of renal epithelial tubular cell proliferation involves a p21ras activated, AP-1-dependent pathway. *Kidney Int* 52:1016-1027, 1997
- ROGLER G, LACKNER KJ, SCHMITZ G: Effects of fluvastatin on growth of porcine and human vascular smooth muscle cells in vitro. *Am J Cardiol* 76:114A-116A, 1995
- MUNRO E, PATEL M, CHAN P, BETTERIDGE L, CLUNN G, GALLAGHER K, HUGHES A, SCHACHTER M, WOLFE J, SEVER P: Inhibition of human vascular smooth muscle cell proliferation by lovastatin: The role of isoprenoid intermediates of cholesterol synthesis. *Eur J Clin Invest* 24:766-772, 1994
- HIDAKA Y, EDA T, YONEMOTO M, KAMEI T: Inhibition of cultured vascular smooth muscle cell migration by simvastatin (MK-733). *Atherosclerosis* 95:87-94, 1992
- CHAKRABARTI R, ENGLEMAN EG: Interrelationships between mevalonate metabolism and the mitogenic signaling pathway in T lymphocyte proliferation. *J Biol Chem* 266:12216-12222, 1991
- GRIECO D, BEG ZH, ROMANO A, BIFULCO M, ALOJ SM: Cell cycle progression and 3-hydroxy-3-methylglutaryl coenzyme A reductase are regulated by thyrotropin in FRTL-5 rat thyroid cells. *J Biol Chem* 265:19343-19350, 1990
- BARBU V, DAUTRY F: Mevalonate deprivation alters the induction of fos and myc by growth factors. *Oncogene* 5:1077-1080, 1990
- FUKADA Y, TAKAO T, OHGURO H, YOSHIZAWA T, AKINO T, SHIMONISHI Y: Farnesylated g-subunit of photoreceptor G protein indispensable for GTP-binding. *Nature* 346:658-660, 1990
- QIU MS, PITTS AF, WINTERS TR, GREEN SH: Ras isoprenylation is required for ras-induced but not for NGF-induced neuronal differentiation of PC12 cells. *J Cell Biol* 115:795-808, 1991
- REISS Y, STRADLEY SJ, GIERASCH LM, BROWN MS, GOLDSTEIN JL: Sequence requirement for peptide recognition by rat brain p21ras protein farnesyltransferase. *Proc Natl Acad Sci USA* 88:732-736, 1991
- ISHIKAWA S, KAWASUMI M, SAITO T: Simvastatin inhibits the cellular signaling and proliferative action of arginine vasopressin in cultured rat glomerular mesangial cells. *Endocrinology* 136:1954-1961, 1995
- GHOSH PM, MOTT GE, GHOSH-CHOUDHURY N, RADNIK RA, STAPLETON ML, GHIDONI JJ, KREISBERG JI: Lovastatin induces apoptosis by inhibiting mitotic and post-mitotic events in cultured mesangial cells. *Biochim Biophys Acta* 1359:13-24, 1997
- GUIJARRO C, BLANCO-COLIO LM, ORTEGO M, ALONSO C, ORTIZ A, PLAZA JJ, DÍAZ C, HERNÁNDEZ G, EDIGO J: 3-Hydroxy-3-methylglutaryl coenzyme A reductase and isoprenylation inhibitors induce apoptosis of vascular smooth muscle cells in culture. *Circ Res* 83:490-500, 1998
- HENGST L, DULIC V, SLINGERLAND JM, LEES E, REED SI: A cell cycle-regulated inhibitor of cyclin-dependent kinases. *Proc Natl Acad Sci USA* 91:5291-5295, 1994
- HENGST L, REED SI: Translational control of p27Kip1 accumulation during the cell cycle. *Science* 271:1861-1864, 1996
- HIRAI A, NAKAMURA S, NOGUCHI Y, YASUDA T, KITAGAWA M, TATSUNO I, OEDA T, TAHARA K, TERANO T, NARUMIYA S, KOHN LD, SAITO Y: Geranylgeranylated rho small GTPase(s) are essential for the degradation of p27Kip1 and facilitate the progression from G1 to S phase in growth-stimulated rat FRTL-5 cells. *J Biol Chem* 272:13-16, 1997
- ODA H, KASISKE BL, O'DONNELL MP, KEANE WF: Effects of lova-

- statin on expression of cell cycle regulatory proteins in vascular smooth muscle cells. *Kidney Int* 56(Suppl 71):S-202–S-205, 1999
42. O'DRISCOLL G, GREEN D, TAYLOR RR: Simvastatin, an HMG-coenzyme A reductase inhibitor, improves endothelial function within 1 month. *Circulation* 95:1126–1131, 1997
 43. HERNÁNDEZ-PERERA O, PEREZ-SALA D, NAVARRO-ANTOLIN J, SANCHEZ-PASCUALA R, HERNANDEZ G, DÍAZ C, LAMAS S: Effects of the 3-hydroxy-3-methylglutaryl-CoA reductase inhibitors, atorvastatin and simvastatin, on the expression of endothelin-1 and endothelial nitric oxide synthase in vascular endothelial cells. *J Clin Invest* 101:2711–2719, 1998
 44. EGASHIRA K, HIROOKA Y, KAI H, SUGIMACHI M, SUZUKI S, INOU T, TAKESHITA A: Reduction in serum cholesterol with pravastatin improves endothelium-dependent coronary vasomotion in patients with hypercholesterolemia. *Circulation* 89:2519–2524, 1994
 45. TREASURE CB, KLEIN JL, WEINTRAUB WS, TALLEY JD, STILLABOWER ME, KOSINSKI AS, ZHANG J, BOCCUZZI SJ, CEDARHOLM JC, ALEXANDER RW: Beneficial effects of cholesterol-lowering therapy on the coronary endothelium in patients with coronary artery disease. *N Engl J Med* 332:481–487, 1995
 46. OSAMAH H, MIRA R, SORINA S, SHLOMO K, MICHAEL A: Reduced platelet aggregation after fluvastatin therapy is associated with altered platelet lipid composition and drug binding to the platelets. *Br J Clin Pharmacol* 44:77–83, 1997