In vitro activities of acetic extracts from leaves of three forage legumes (Calliandra calotyrsus, Gliricidia sepium and Leucaena diversifolia) on Haemonchus contortus

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Objective: To assess ovicidal activity of three acetonic extracts from the leaves of three forage legume, Calliandra calotyrsus (C. calotyrsus), Gliricidia sepium (G. sepium) and Leucaena diversifolia (L. diversifolia) in vitro on Haemonchus contortus (H. contortus). Methods: Eggs were exposed for 24 hours to five different concentrations (0.075, 0.15, 0.3, 0.6 and 1.25 mg/mL) of acetic extracts at room temperature (24 °C). Distilled water and 0.4% Tween were used in the bioassay as negative controls. Results: The later did not affect embryonation and egg hatching of H. contortus. Conversely, significant effects were obtained with the acetonic extracts of leaves of all three plants and the maximum activity was observed with the highest concentration (1.25 mg/mL). The acetonic extract of G. sepium was found to be more active (2.9% and 0.0% for embryonation and egg hatching, respectively) than the other substances 16.5% and 33.5%, respectively for C. calothyrsus, 33.7% and 33.3%, respectively for L. diversifolia. Conclusions: These results suggest that the three forage legumes do possess ovicidal properties and further studies on larvae should be carried out.

1. Introduction

Small ruminants play an important role in food production for developing countries. Sheep or goat meat contributes 3.5% of animal proteins in these countries compared to 0.2% in the developed world[1]. In Africa where climatic factors such as humidity and temperature favour the development of parasites, 97% of small ruminants suffer from gastrointestinal helminthiases. Among these diseases, haemonchosis is the one which has a significant impact on productivity and leads to serious economic losses for small holders. In Kenya for example, the loss to the agricultural sector was recently estimated at US$ 26 million per year[2]. Commercial anthelmintics have been mainly used for some decades for the control of intestinal helminths. However, these synthetic compounds are on one hand not available in the tropics and sub-tropics and on the other hand their costs in the small local market are relatively high. In addition, the misuse of these substances by the farmers has led to the development of resistant organisms in many developed countries[3,4]. Therefore new approaches for helminth control are needed especially in countries of the South. Works carried out in this field to assess the in vitro anthelmintic activities of extracts of: Vernonia amygdalina and Annona senegalensis in Nigeria, Canthium manni (C. manni) in Cameroon, Melia azedarach in Brazil and Canthium moschata in Guadeloupe, respectively[5-9]. The discovery of natural new cheaper drugs with relatively low toxicity seems to be a desirable solution[10,11]. The present study aimed to evaluate in vitro the activities of acetic extracts of leaves of three forage legumes namely Calliandra calotyrsus (C. calotyrsus), Gliricidia sepium (G. sepium) and Leucaena diversifolia (L. diversifolia) on Haemonchus contortus (H. contortus), one of the most pathogenic gastrointestinal nematode of small ruminants. These forages are used to feed animals.

2. Materials and methods

2.1. Plant materials

Leaves of C. calotyrsus, G. sepium and L. diversifolia were collected in the experimental field of the Faculty of Agronomy and Agricultural Sciences of the University of Dschang. These specimens were dried separately in a
ventilating oven at a maximum temperature of 50 °C for two
days, ground and stored in airtight plastic bags for about a
week, at room temperature (25 °C) and relative humidity
67% for subsequent use in the laboratory.

2.2. Preparation of acetic extracts of leaves

Five hundred grams of each leaf powder were macerated
in 1.5 L of acetone which removed phenolic compounds
from the leaves[12]. Each mixture was daily stirred, and 72
hours later the acetic extracts were obtained[13]. A total
of 100 mg of a given concentrated extract was diluted with
0.4 mL of 0.4% tween. Then distilled water was added to the
solution to obtain a total volume of 20 mL. This resulted to a
5 mg/mL stock solution from which a series of dilutions were
made to obtain different final tested concentrations: 0.075,
0.15, 0.3, 0.6 and 1.25 mg/mL.

2.3. Reference drug

The reference drug, Mebendazole (MBZ), used in larvicidal
test was bought in a local pharmacy (Mebencol N. D.,
Colorama pharmaceutical Ltd). The product was diluted
in distilled water to obtain a stock solution with the same
concentration as with organic extracts. The controls used for
the bioassay were 0.4% tween and distilled water.

2.4. Recovery of nematode eggs

Fresh eggs of *H. contortus* were obtained from the faeces of
sheep experimentally infected[14]. Briefly, 3 g of faeces were
collected, homogenised in a mortar, suspended in saturated
salt solution (0.4% NaCl), and cleaned of organic debris
by filtration through sieves (1 mm and 150 μm) into a 100
ml beaker. The contents of the latter were poured into four
tubes and centrifuged at 1 000 g for 5 min. The supernatant
was poured through a 45 μm sieve then, the retained
material on the sieve containing eggs was washed with tap
water to remove the salt solution. The sieve was then turned,
the opposite side was washed with tap water and eggs were
collected in a Petri dish (Ø=16 cm)[15].

2.5. Evaluation of ovicidal activity

The ovicidal efficacy test of the different acetone extracts
was performed using two different procedures. To assess
the effects of the extracts on fresh eggs, 1 mL of a diluted
solution containing about 48 parasite eggs was distributed
in each of 12 Petri dishes (35 mm Ø x 10 mm) and mixed
with the same volume of a specific plant extract or MBZ at different concentrations. The dishes were covered
and kept at room temperature for 24 hours, after which the
number of dead or immobilized larvae was determined under
a microscope (at 4x magnification). The percent mortality
(Me%) was calculated using Abbott’s formula[5]:

\[
\text{Mc} = \frac{\text{Mce} - \text{Mt}}{100 - \text{Mt}} \times 100
\]

Where Mc, Mce and Mt correspond to the corrected
mortality; the ones which were obtained during the test, and
registered in the negative control dishes, respectively. When
the rate in the latter diseases is less than 5%, Mc=Mce[19].

The larvicidal concentration 50 (LC50) was determined
using the regression line of the mortality expressed in probit
according to the decimal logarithm of the concentration. All
tests were repeated six times for each treatment and control.

2.6. Recovery of nematode larvae

3 mL of the egg suspension were poured on a filter paper
covering the bottom of the Petri dishes. The dishes were then
covered to maintain a high relative humidity (65%–
67%) to prevent from drying out, and stored at 24°C. After 3 and
4–5 days of incubation, L1 and L2 larvae were observable in
Petri dishes, respectively. They were differentiated using
their morphological features[16] and were concentrated with a
Baermann apparatus[17,18].

2.7. Evaluation of larvicidal activity

To test the effects of the extracts on L1 and L2 larvae, 1
mL of a solution containing about 30 parasite larvae was
distributed in each of 12 Petri dishes (35 mm Ø x 10 mm)
and mixed with the same volume of a specific plant extract
or MBZ at different concentrations. The dishes were covered
and kept at room temperature for 24 hours, after which the
number of dead or immobilized larvae was determined under
a microscope (at 4x magnification). The percent mortality
(Me%) was calculated using Abbott’s formula[5]:

\[
\text{Mc} = \frac{\text{Mce} - \text{Mt}}{100 - \text{Mt}} \times 100
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Where Mc, Mce and Mt correspond to the corrected
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The larvicidal concentration 50 (LC50) was determined
using the regression line of the mortality expressed in probit
according to the decimal logarithm of the concentration. All
tests were repeated six times for each treatment and control.

2.8. Data analysis

The mean percentage of embryonation, hatching and the
mean mortality rates were compared using the chi-square
test at the P<0.05 significance level.

3. Results

The effects of different acetone extracts of the leaves of
three forage legumes (*C. calothyrsus*, *G. sepium* and *L.
diversifolia*) were given in Table 1. The mean embryonation
rate of fresh eggs of *H. contortus* was all 96.97±2.62 and the
hatching rate were 100% in 0.4% tween and distilled water.
The values obtained in both embryonation and egg hatch
tests with various concentrations of determined extracts
differed significantly from negative controls. In general,
the mean embryonation rates decreased gradually with the
concentration of different acetone extracts. At 0.075 mg/mL,
the effect of *C. calothyrsus* leaf extract was similar to that
of the negative controls. Conversely, at the concentration
higher or equal to 0.3 mg/mL, the acetonic extract of *C.
calothyrsus*, *G. sepium*, *L. diversifolia* significantly reduced
(P<0.05) the embryonation of *H. contortus* and this activity
was stronger at the highest concentration. Thus, at 1.25 mg/
/mL the mean inhibition rate of embryonation was 100%.
Concerning the effects of various extracts on egg hatching
(Table 2), it was observed that, the hatching rate of L1, larvae
decreased with increasing concentrations of extracts as in the first experiment. *G. sepium* extract at concentration of 0.075 mg/mL inhibited ecibility for about 20% and this rate fell to 0% at concentration equal to or higher than 1.5 mg/mL. The extract of the later forage legume seem to be more active in this second experiment than the other two extracts. 0.4% tween and distilled water did not provoke mortality of L1 and L1 larvae of *H. contortus*. Conversely, with the mebendazole and the different acetonic extracts the mortality rates increased in concentration dependence. The later seems to be more efficient producing mortality rates of 44%–97%. Mortality due to the acetonic extracts of *C. colotyrsus* and *L. diversifolia* was stronger than that of *C. calotyrsus*. For concentrations higher than or equal to 0.3 mg/mL. At the highest tested concentration (1.25 mg/mL), the activity of the extracts of *G. sepium* and *L. diversifolia* was stronger than that of *C. calotyrsus*. For concentrations higher than or equal to 0.3 mg/mL, these three extracts inhibited significantly ($P<0.05$) larval survival of *H. contortus*. Meanwhile, the larvicidal activity of different extracts was similar to that of the positive control at 1.25 mg/mL. After the transformation of the mortality rate to probit, the regression lines obtained showed a dose dependent response according to the probit transformation of the concentration (mg/mL), we observe that there was a concentration dependent relationship with all the tested substances and the larvicidal concentrations 50 (LC$_{50}$) were 0.177, 0.463, 0.840 and 0.373 mg/mL for MBZ, *C. colotyrsus*, *G. sepium* and *L. diversifolia* respectively.

### Table 1

<table>
<thead>
<tr>
<th>Group (0.4 %)</th>
<th>Embryonation</th>
<th>Hatching of eggs</th>
</tr>
</thead>
<tbody>
<tr>
<td>TW (0.4 %)</td>
<td>96.97±2.62</td>
<td>100.00±0.00</td>
</tr>
<tr>
<td>DW</td>
<td>96.97±2.62</td>
<td>100.00±0.00</td>
</tr>
<tr>
<td>Me 0.075 mg/mL</td>
<td>56.05±8.00</td>
<td>66.99±12.46</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>40.82±11.08</td>
<td>59.77±16.95</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>40.69±17.34</td>
<td>60.61±9.46</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>21.22±15.79</td>
<td>41.28±35.75</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>20.88±8.78</td>
<td>10.00±17.32</td>
</tr>
<tr>
<td>Cc 0.075 mg/mL</td>
<td>58.67±33.01</td>
<td>68.98±21.62</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>42.87±9.12</td>
<td>55.37±5.57</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>29.89±26.91</td>
<td>55.97±17.02</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>22.13±20.06</td>
<td>52.36±11.33</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>16.49±14.51</td>
<td>33.54±6.37</td>
</tr>
<tr>
<td>Gs 0.075 mg/mL</td>
<td>46.65±8.33</td>
<td>19.95±7.56</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>33.44±11.01</td>
<td>14.17±7.00</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>30.21±12.19</td>
<td>0.00±6.00</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>15.79±6.14</td>
<td>0.00±6.00</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>2.09±3.61</td>
<td>0.00±6.00</td>
</tr>
<tr>
<td>Ld 0.075 mg/mL</td>
<td>62.07±29.58</td>
<td>70.75±4.17</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>56.83±15.04</td>
<td>71.98±11.34</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>54.93±7.45</td>
<td>63.69±2.45</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>41.83±12.19</td>
<td>49.80±19.03</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>33.72±15.59</td>
<td>33.29±13.97</td>
</tr>
</tbody>
</table>

Legend: TW (0.4%) = tween, DW= Distilled water, Me = mebendazole, Cc = *Calliandra colotyrsus*, Gs = *Gliricidia sepium*, Ld = *Leucaena diversifolia*.

On the other hand, the larvicidal effect on L2 larvae increases proportionally with the increased of the concentration. At all tested concentrations, the acetonic extract inhibited significantly ($P<0.05$) the survival of L2 larvae but, no statistical difference was observed between the activity of the extracts and that of MBZ. After the transformation of the mortality rate to probit, the regression lines obtained showed a dose dependent response according to the probit transformation of the concentrations, and the LC$_{50}$ were 0.218, 0.226, 0.286 and 0.117 mg/mL for MBZ, *C. calotyrsus*, *L. diversifolia* and *G. sepium*. These findings showed that, the different substances seem to be more active on the L1 larvae than on the L2 one. It was also noticed that, the extract of *G. sepium* was more efficient on L2.

### Table 2

<table>
<thead>
<tr>
<th>Group</th>
<th>$L_1$ larvae</th>
<th>$L_2$ larvae</th>
</tr>
</thead>
<tbody>
<tr>
<td>TW (0.4%)</td>
<td>0.00±0.00</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>DW</td>
<td>0.00±0.00</td>
<td>0.00±0.00</td>
</tr>
<tr>
<td>Me 0.075 mg/mL</td>
<td>44.93±35.54</td>
<td>50.66±18.13</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>66.33±13.08</td>
<td>53.65±5.82</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>80.76±12.23</td>
<td>83.65±26.91</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>87.28±4.73</td>
<td>97.53±4.28</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>96.56±5.96</td>
<td>100.00±0.00</td>
</tr>
<tr>
<td>Cc 0.075 mg/mL</td>
<td>37.05±21.27</td>
<td>40.75±26.13</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>40.32±27.95</td>
<td>49.11±20.68</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>49.91±3.71</td>
<td>83.99±22.95</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>65.65±17.44</td>
<td>100.00±0.00</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>79.08±18.29</td>
<td>100.00±0.00</td>
</tr>
<tr>
<td>Gs 0.075 mg/mL</td>
<td>8.60±2.74</td>
<td>59.91±11.67</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>13.15±11.39</td>
<td>72.78±15.69</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>39.52±30.78</td>
<td>81.49±15.66</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>43.11±8.48</td>
<td>94.44±9.62</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>92.06±4.46</td>
<td>98.04±3.39</td>
</tr>
<tr>
<td>Ld 0.075 mg/mL</td>
<td>29.66±17.35</td>
<td>41.33±34.88</td>
</tr>
<tr>
<td>0.15 mg/mL</td>
<td>39.87±31.02</td>
<td>41.11±30.97</td>
</tr>
<tr>
<td>0.30 mg/mL</td>
<td>63.92±12.11</td>
<td>66.39±8.99</td>
</tr>
<tr>
<td>0.60 mg/mL</td>
<td>72.83±15.03</td>
<td>94.50±4.97</td>
</tr>
<tr>
<td>1.25 mg/mL</td>
<td>94.50±4.97</td>
<td>90.96±8.32</td>
</tr>
</tbody>
</table>

Legend: TW (0.4%) = tween, DW= Distilled water, Me = mebendazole, Cc = *Calliandra colotyrsus*, Gs = *Gliricidia sepium*, Ld = *Leucaena diversifolia*.

### 4. Discussion

Results in the present study are similar to the report from Houznzangbe–Adote et al.[20]. It indicates that, the active compounds in the extract penetrates the egg shell and prevents egg development or paralyse the first stage larvae within the egg as is the case with levamisol. The unidentified mass in un—embryonated egg may be as a result of segmentation inhibition of the egg. Overall, the leaf extracts of the three forage legumes negatively affected the egg of *H. contortus*. It was shown that, the acetonic extract of *G. sepium* was more active on the embryogenic development than the extracts of the other leaves. In the same manner, this effect was observed on embryonated egg compared to the effect of the other extracts and MBZ. The same findings were observed on *H. polygyrus* and *Ancylostoma caninum* with the extract of the stem bark of the shrub named *C. mannii*, a medicinal plant used traditionally by the people of Dschang, Western Region of Cameroon, Central Africa[6,7].
These observations were also shown in Guadeloupe[9]. The activity of the three extracts may be attributed to phenolic compounds and particularly to tannins present in the leaves. These substances were identified in the leaves of the three forage legumes.

At higher concentration, the extracts significantly inhibited the development of L1 and L2 larvae. Similar findings were reported that Hedysanum coronarium extracts which inhibited the migration of H. contortus larvae[21,22]. They also reported the activity of tannins on Strongylus edentatus larvae. The larvicidal activity observed in this work with different acetonic extracts may therefore be attributed to phenolic compounds in general or to tannins in particular. These substances are able to penetrate the cuticle of the nematode and to prevent the absorption of the glucose, or block post synaptic receptors thus paralysing the larvae. Tannins may also induce the release of gamma-aminobutyric acid (GABA) which could block transmission of nerve impulses or could act in decoupling the phosphorylation oxidative reaction which led to the energy exhorition of the larva[23]. In the same manner, tannins absorbed by insects bind on the intestinal mucosa and led to autolysis[24]. These findings can be extrapolated on nematode larvae. Also, tannins may bind on the cuticle of the larvae and led to dead. Extracts were more active on the L1 larvae. These findings confirm the fact that, these life cycle–stages are more sensible than the L2 one[25]. In conclusion, the different acetonic extracts tested in this work exhibited in vitro negative effects on larvae of H. contortus parasite of small ruminants. The extracts were more active on L1 larvae and the LD50 obtained in this study were very low compared to that obtained with the extracts of C. mannii on Heliogmosomoides polygyrus larvae.[7]

In conclusion, acetonic extract of C. calothyrsus, G. sepium and L. diversifolia exhibited in vitro negative effects on eggs and larvae of H. contortus parasite of small ruminants. In addition to their high nutritive value on animal feeding, the leaves of the three forage legume can be used for the control of intestinal nematode of ruminants. However, further experiment incorporating in vitro and toxicological studies should be carried out.

Conflict of interest statement

We declare that we have no conflict of interest.

References


