Hill coefficients CAF 2.14 ± 0.16, HF 2.53 ± 0.14, P < 0.05. Consistently, the results of real time PCR and western blot demonstrated that captopril significantly downregulated the expression of apamin sensitive SK channels (SK3 mRNA: CAF 2.10 ± 0.9, n = 6 vs HF 8.40 ± 2.10, n = 6; SK3 protein: CAF 0.40 ± 0.07, n = 6 vs HF 0.56 ± 0.09, n = 6).

CONCLUSIONS Captopril significantly downregulated the sensitivity of SK channels to [Ca<sup>2+</sup>] and the SK3 channels expression in HF, and reversed the SK channels remodeling.

**GW26-e1586**

AMPK attenuates proliferation of cardiac fibroblast via regulating TGF-β1/Smad pathways

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**OBJECTIVES** AMP-activated protein kinase (AMPK) exerts inhibitory effects on cardiac hypertrophy. However, the mechanism remains unclear. The aim of the present study was to investigate the effects of AMPK on angiotensin II (AngII) -induced proliferation of cardiac fibroblast and the mechanisms involved.

**METHODS** Proliferation of cardiac fibroblast was induced by angiotensin II (AngII). Cardiac fibroblasts were treated with the specific AMPK activator 5-aminoimidazole-4-carboxamide ribonucleoside (AICAR, 0.5 mmol/L) and the specific AMPK antagonist Compound C (1 umol/L), and then stimulated with AngII (1 umol/L). Cell proliferation and the DNA synthesis were measured by MTT assay and EdU incorporation assay. TGF-β1 and Smad2, 3, 4 mRNA and protein expression was detected using Real-Time PCR and western blot analysis.

**RESULTS** Activation of AMPK by AICAR could inhibit AngII-induced proliferation of cardiac fibroblasts, manifesting decreased DNA synthesis and collagen production (P < 0.05). Moreover, AngII significantly increased the mRNA and protein expression of TGF-β1 and Smad2, 3, 4 (P < 0.05). AMPK activation markedly reversed the elevated TGF-β1 and Smad2, 3, 4 mRNA and protein levels (P < 0.05). Furthermore, Treatment of proliferated cardiac fibroblasts with Compound C blunted the effects of AMPK on proliferation of cardiac fibroblasts and changes to the TGF-β1/Smad pathway (P < 0.05).

**CONCLUSIONS** AMPK activation could attenuate proliferation of cardiac fibroblast induced by AngII, which may be due to the inhibition of TGF-β1/Smad pathways.

**GW26-e4768**

Improved Recovery After Myocardial Ischemic Infarction by Copper Supplementation

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**OBJECTIVES** Depressed angiogenesis due to ischemic injury leads to myocardial infarction. Copper (Cu) is involved in angiogenesis and ischemia causes copper loss in the heart. The present study was undertaken to test the hypothesis that Cu supplementation improves myocardial angiogenesis, leading to regression of myocardial ischemic infarction in Rhesus monkey model.

**METHODS** Coronary artery ligation was used to produce myocardial infarction in Rhesus monkey model. This procedure effectively increased Cu concentrations in myocardial microvessels (SK3 mRNA: CAF 2.10 ± 0.9, n = 6 vs HF 8.40 ± 2.10, n = 6; SK3 protein: CAF 0.40 ± 0.07, n = 6 vs HF 0.56 ± 0.09, n = 6). Along with these changes, myocardial infarct size was significantly decreased and the density of myocardial microvessels was significantly increased. In addition, cardiac function was significantly recovered, as evidenced by increased ejection fraction (EF) values and decreased end-systolic volume (ESV) measured by echocardiography.

**CONCLUSIONS** This study thus demonstrated that Cu supplementation improved cardiac structural and functional recovery after ischemic infarction.

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**GW26-e5402**

Discovery of a new conduction substrate associated with atrioventricular node-anterior extension pathway

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**OBJECTIVES** The atrioventricular node(AVN) plays a role in conducting action potentials at an appropriate conduction velocity from atria to ventricles. Its complex anatomical structures and functional longitudinal dissociations are considered important in delayed conduction and AVN reentrant tachycardia (AVNRT). Inferior nodal extensions(INE) are part of the AVN. It is thought that these extensions may be involved in slow-pathway conduction and are part of the underlying circuitry that causes AVNRT. Some other conduction tissue shared the same origin layers with AVN distribute around the two valve annulus. The retro-aortic node defined as the enlargement of this node-like tissue is located on the right side of the atrium parallel with the aorta. The potential electrophysiological function of these node-like tissues is still unknown. Understanding their detailed anatomical structure, histological features and electrophysiological behavior are significant to clear the complex conduction characteristics of the AV junction.

**METHODS** Adult rats(n = 6), mice(n = 5) and rabbit(n = 5) were used. Serial sections from the entire AV junction were obtained. Masson’s trichrome stain was performed on AVN regions to assess for fibrous tissue. Three connexins(Cx) proteins including Cx43, Cx40 and Cx45 which dominate the electronic conduction of the heart and three main ion channels(Na<sub>1.5</sub>, Ca<sub>3.1</sub> and HCN4) participating the depolarization of myocardial cell were immunohistochemically labeled. Serial sections were used to reconstruct a 3D computational model of the anatomy of the AV junction which display the different histological features at different levels.

**RESULTS** There appears to be an anterior extension of the AVN which connect retroaortic node and AVN. The anterior node extension (ANE), compact node and INE express the same connexin isoforms. Na<sub>1.5</sub> labeling was abundant in the atrial and ventricular myocardium. Na<sub>1.5</sub> labeling presents at a reduced level in the compact node, ANE and INE. Ca<sub>3.1</sub> and HCN4 expression were mainly expressed in the Na<sub>1.5</sub> reduced area. Further, connections between the atria and inferior extension occur indirectly via small branches. However, this was distinct from the connection pattern that we observed between the atria and ANE, which was direct.

**CONCLUSIONS** We conclude that the retroaortic node connect with ANE forming ANE which suggests there would be direct electric conduction between them. Characteristics of these structures are connected among various species including rat, mouse and rabbit as we had proven. ANE, AVN and INE have nearly the same electronic level and action potential level structure basic that highlight they would have the same conduction properties, but there are different connection patterns in atrium between them that suggests there would be the subtle conduction velocity difference after accept the impulse, which provide a new location and substrate of conduction that may present new insights on mechanisms underlying normal AV conduction and AVNRT.

**GW26-e2274**

AVE 0991, Nonpeptide angiotensin-(1-7) analogue, modulates cardiac hypertrophy via reducing oxidative stress

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**OBJECTIVES** AVE 0991, the nonpeptide angiotensin-(1-7) (Ang-(1-7)) analog, is recognized as having beneficial cardiovascular effects. However, the mechanisms have not been fully elucidated. This study was designed to investigate the effects of AVE 0991 on cardiac hypertrophy and the mechanisms involved.

**METHODS** Mice were subjected to aortic banding (AB) to induce cardiac hypertrophy. After treatment with AVE 0991 (20 mg kg<sup>-1</sup> day<sup>-1</sup>) for four weeks, indices of cardiac hypertrophy and heart function were measured by echocardiography, histological analyses and quantitative...