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Control of Brain Development, Function, and Behavior by the Microbiome

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Animals share an intimate and life-long partnership with a myriad of resident microbial species, collectively referred to as the microbiota. Symbiotic microbes have been shown to regulate nutrition and metabolism and are critical for the development and function of the immune system. More recently, studies have suggested that gut bacteria can impact neurological outcomes—altering behavior and potentially affecting the onset and/or severity of nervous system disorders. In this review, we highlight emerging evidence that the microbiome extends its influence to the brain via various pathways connecting the gut to the central nervous system. While understanding and appreciation of a gut microbial impact on neurological function is nascent, unraveling gut-microbiome-brain connections holds the promise of transforming the neurosciences and revealing potentially novel etiologies for psychiatric and neurodegenerative disorders.

Introduction

Metazoans evolved in a world dominated by microbial life. Despite the long evolutionary history that has forged elaborate host-microbial symbioses over many millennia, it is only recently that science and society have begun to appreciate the inextricable connection between microbes and mammals. We are witnessing a groundswell of research that is describing and defining how gut bacteria (known as the microbiota) influence critical aspects of our physiology. The last decade of research has illuminated numerous complex interactions between the microbiota and the immune and metabolic systems, many of which have significant implications on human health. While the fascinating and profound mechanisms by which gut bacteria control immunity and metabolism has led to a modern renaissance in biomedical research, regulation of the nervous system by the microbiota had remained relatively unexplored until very recently (Mayer et al., 2014, 2015; Stilling et al., 2014b). How could simple gut microbes influence a complex and distant organ such as the brain? This seemingly improbable concept that specific microbes influence the behavior and neurological function of their hosts had, in fact, already been established. One prime example of "microbial mind control" is the development of aggression and hydrophobia in mammals infected with the rabies virus (Driver, 2014). Another well-known example of behavior modification occurs by Toxoplasma gondii, which alters the host rodents' fear response. Infected rodents lose their defensive behavior in the presence of feline predators and instead actually become sexually attracted to feline odors (House et al., 2011). This results in infected rodents being preyed upon more readily by cats and allows Toxoplasma to continue its lifecycle in the feline host (House et al., 2011). Further, a variety of parasitic microbes are capable of altering the locomotive behavior and environmental preferences of their hosts to the benefit of the microbe. For instance, the Spinochordodes tellinii parasite causes infected grasshopper hosts to not only jump more frequently but also seek an aquatic environment where the parasite emerges to mate and produce eggs (Biron et al., 2005). Temperature preference of the host can even be altered, such as observed during infection of stickleback fish by *Schistocephalus solidus*, which changes the hosts' preference from cooler waters to warmer waters where the parasite can grow more readily (Macnab and Barber, 2012). Other microbes can even alter host behavior to seek higher elevations, believed to allow the infected host to be noticed more easily by predators or to eventually fall and disperse onto susceptible hosts below (Maitland, 1994). More coercively still, microbes can influence the social behavior of their hosts, causing insects, such as ants, to become more or less social to the benefit of the parasite (Hughes, 2005). In fact, the sexually transmitted virus IIV-6/CrIV causes its cricket host (*Gryllus texensis*) to increase its desire to mate, causing its rate of mating to be significantly elevated and allowing for transmission between individual hosts (Adamo et al., 2014).

While all of the above examples most certainly represent pathogenic and/or parasitic relationships, they nonetheless raise the possibility that the indigenous microbes, which are in constant, life-long interaction with their human and animal hosts, could influence neurological function and behavior during development or within health and disease states. It is becoming increasingly recognized that psychiatric and neurological illnesses are often co-morbid with gastrointestinal (GI) pathology (Vandvik et al., 2004), including schizophrenia, autism, neurodegenerative diseases, and depression. Furthermore, recent observations have indicated that the commensal microbiota of the intestine do indeed alter aspects of their hosts' neurological function, leading to effects on mood and behavior, including depression, anxiety, social behavior, and mate choice (Table 1) (Bravo et al., 2011; Desbonnet et al., 2010; Foster and McVey Neufeld, 2013; Hsiao et al., 2013; Neufeld et al., 2011; Sharon et al., 2010). The intestinal microbiota are, however, well established to have a profound impact in shaping the host immune system, which itself may subsequently influence host behavior (Dantzer et al., 2008) and indirectly have effects on neurodegeneration and repair during the process of aging, neurological trauma, and disease. The precise mechanisms of how the intestinal microbes impact neurological function and behavior remain largely unknown, but are likely vast, varied, and complex.



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Category	Attribute	Effect	Citation(s)
Behavioral	Anxiety	Reduced self-reported anxiety in humans treated with <i>L. helveticus</i> R0052 and <i>B. longum</i> R0175	Messaoudi et al., 2011
Behavioral	Anxiety-like behavior	Decreased anxiety-like behavior in GF mice (Swiss Webster, NIH Swiss, and NMRI)	Clarke et al, 2013; Diaz Heijtz et al., 2011; Neufeld et al., 2011; Selkrig et al., 2014
Behavioral	Anxiety-like behavior	Increased anxiety-like behavior in GF mice (BALB/c, C57BI6)	Bercik et al., 2011; Selkrig et al., 2014
3ehavioral	Anxiety-like behavior	Reduced anxiety-like behavior in rodents treated with <i>Bifidobacterium breve</i> 1205, <i>B. longum</i> 1714, <i>B. longum</i> R0175 <i>Lactobacillus helveticus</i> R0052, or <i>L. rhamnosus</i> JB-1	Bravo et al., 2011; Messaoudi et al., 2011; Savignac et al., 2014
3ehavioral	Depression	Reduced self-reported feelings of depression and aggression in humans treated with probiotics	Steenbergen et al., 2015
Behavioral	Depression-like behavior	Decreased depression-like behavior in mice treated with <i>B. infantis</i> or <i>L. rhamnosus</i> JB-1	Bravo et al., 2011; Desbonnet et al., 2010; Savignac et al., 2014; Savignac et al., 2015
Behavioral	Emotional processing	Reduced activation following emotional stimulus in humans treated with probiotic milk product	Tillisch et al., 2013
Behavioral	Social recognition	Reduced novel and familiar social recognition in GF mice	Desbonnet et al., 2014
Behavioral	Stereotyped behaviors and vocalizations	Restoration of social behaviors in <i>Bacteroides</i> fragilis-treated MIA mice	Hsiao et al., 2012
Behavioral	Stress response	Increased response to restraint stress in GF mice	Sudo et al., 2004
Hormonal	Corticosterone	Increased hypothalmic corticosterone in GF mice	Sudo et al., 2004
Hormonal	Cortisol	Reduced urinary cortisol in humans treated with L. helveticus R0052 and B. longum R0175	Messaoudi et al., 2011
Neurochemical	BBB	Decreased expression of tight junction proteins, and subsequent increase of BBB permeability	Braniste et al., 2014
Neurochemical	BDNF	Decreased hypothalmic BDNF in GF mice	Sudo et al., 2004
Neurochemical	Dopamine and GABA	Decreased serum levels of dopamine and GABA in GF mice	Matsumoto et al., 2012; Velagapudi et al., 2010
Neurochemical	G-CSF	Reduced serum levels of G-CSF in GF mice	Deshmukh et al., 2014
Neurochemical	Peripheral serotonin	Decreased peripheral and intestinal serotonin in GF mice, restored by colonization with spore forming bacteria	Wikoff et al., 2009; Yano et al., 2015
Neurochemical	Serotonin and Serotonin receptor	Decreased serotonin and receptor (5 $\mathrm{HT}_{1\mathrm{A}}$) in the amygdala and hippocampus of GF mice	Bercik et al., 2011; Clarke et al., 2013; Diaz Heijtz et al., 2011; Neufeld et al., 2011; Yano et al., 2015
Neurochemical	Serotonin, noradrenaline, and dopamine	Increased turnover of serotonin, noradrenaline, and dopamine in the striatum of GF mice	Diaz Heijtz et al., 2011

Interactions between a host and its microbiota are decidedly intricate. Intestinal microbes influence numerous aspects of metabolism, producing metabolic precursors to hormones and neurotransmitters or directly producing the active metabolites themselves (Lyte, 2014; Sharon et al., 2014) (Figure 1). Symbiotic bacteria additionally have the capability to influence the status of the systemic immune system, which may alter how the immune system subsequently interacts with the nervous system (Belkaid and Hand, 2014; Hooper et al., 2012; Round and Mazmanian, 2009) (Figure 1). Furthermore, the enteric nervous system (ENS) is directly connected to the central nervous system (CNS) through the vagus nerve, providing a direct neurochemical pathway for microbial-promoted signaling in the GI tract to be propagated to the brain (Forsythe et al., 2014) (Figure 1). Herein, we review the current understanding of how the intestinal microbiota influence behavior and neurological function during both health and disease. First, we will focus on how indigenous microbes shape mood and cognitive behaviors, as well as social behaviors. We will next discuss the physiological aspects that are modulated by signals derived from the microbiota. In particular, we will

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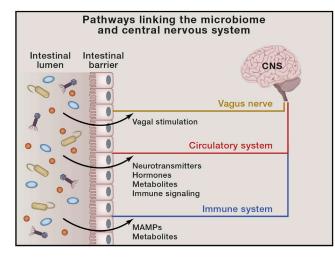


Figure 1. Pathways Linking the Microbiome and CNS

Signals from the intestinal microbiome may potentially traffic to the CNS via several mechanisms: (1) direct activation of the vagus nerve from the ENS to the CNS; (2) production of, or induction of, various metabolites that pass through the intestinal barrier and into the circulatory system, where they may cross the BBB to regulate neurological function; (3) and MAMPs (such as LPS, BLP, and PSA) and metabolites produced by the microbiota can signal to the immune system. Immune cells (and particularly their cytokines) can influence neurophysiology.

focus on how the commensal microbes directly and indirectly shape neurochemical and immunologic responses that can subsequently affect behaviors and other neurological functions (Table 1). With well-documented evidence that the microbiota shape immunity and metabolism, the impact of gut microbes on the nervous system represents an exciting new frontier for research with vast translational implications.

Role of the Microbiota in Mood and Individual Behaviors

One of the seminal studies on the influence of the commensal microbiota on neurological function observed that germ-free (GF) mice display an elevated response to restraint stress, the stress that occurs during forced immobilization (Sudo et al., 2004). In the absence of the microbiota, mice have substantially higher concentrations of corticosterone, a stress hormone in the hypothalamus, as well as reduced levels of brain-derived neurotrophic factor (BDNF; a protein that stimulates neurogenesis, stimulates synaptic growth, and modulates synaptic plasticity and transmission) (Lu et al., 2013; Sudo et al., 2004). Interestingly, this phenomenon can be partially reversed by re-colonization with a diverse microbiota in adulthood, suggesting that active signals from the microbiota play a critical role in brain development. In fact, colonization with a specific bacterial species, Bifidobacterium infantis, restores the defect (compared to monocolonization with Esherichia coli), demonstrating that rather than the general sensing of the bacteria within the population of indigenous microbes, signals from specific bacteria drive normal behavior (Sudo et al., 2004). This rigorous demonstration that the microbiota affects the hypothalamic-pituitary-adrenal axis (HPA axis) revealed bi-directional communication between the gut and the brain, with a significant effect on host behavior.

Conversely, in the absence of restraint stress, some strains of GF mice (Swiss Webster, NIH Swiss, and NMRI) display decreased anxiety-like behavior, in the form of increased exploration compared to specific pathogen-free (SPF) mice, which are colonized with a diverse microbial population. For instance, GF mice spend more time in the lighted section of a light-dark box, as well as the open arms of an elevated plus-maze, than their colonized counterparts, evidence of decreased anxietylike behavior, as mice typically desire to be in a closed, dark area (Clarke et al., 2013; Diaz Heijtz et al., 2011; Neufeld et al., 2011; Selkrig et al., 2014). Importantly, these behavioral effects can be restored to levels similar to conventionally raised mice by recolonizing the GF mice with a complete SPF microbiota. The effect of the microbiota is not necessarily consistent across all strains of mice. For instance, SPF BALB/c and C57Bl6 mice display increased anxiety behavior compared to other strains, such as NIH Swiss mice (Bercik et al., 2011; Selkrig et al., 2014). The microbiome present in BALB/c mice is significantly different than that present in NIH Swiss mice. When fecal microbiota from either strain are transplanted into GF mice of the counterpart strain, the behavior of the parent strain is transferred (Bercik et al., 2011). This interesting finding implicates that signals derived from the microbiota can drive behaviors in the host. Further, phenotypic changes following fecal microbiota transplants in adult mice reveal that anxiety can be actively modulated by the microbial population and are not necessarily developmental in origin. It is therefore interesting to contemplate that active modulation of the population or function of the gut microbiota, such as through probiotic supplementation, may act to modify host stress and anxiety behaviors as well. In fact, there is an active and growing line of observations of the effect of probiotic species on the behavior of the host.

In one such probiotic supplementation study, it was observed that mice given the probiotic bacterium Lactobacillus rhamnosus (JB-1) over a 28 day period displayed decreased anxiety-like behavior in the elevated plus maze and open-field test (Bravo et al., 2011). Additionally, the effect of L. rhamnosus on anxiety behavior is ameliorated in mice that have been vagotomized and lack connectivity of the vagus nerve between the ENS and the CNS (Bravo et al., 2011). This would suggest that microbial signals can be directed by the vagus nerve to alter CNS outputs (such as behavior), further implicating an active role by the microbiota in mediating neurological functions. In a similar fashion, treatment with B. longum 1714 and B. breve 1205 was observed to decrease anxiety-like behaviors to the same extent as the pharmaceutical anti-anxiety medication escitalopram (Savignac et al., 2014). Other studies in rats utilizing a probiotic cocktail of L. helveticus R0052 and B. longum R0175 found that treatment with these microbes over a 30 day period resulted in decreased anxiety in an electric shock model, further suggesting that the functionality of the gut microbiota influences anxiety-like behaviors (Messaoudi et al., 2011). Together, these studies provide credence to the intriguing hypothesis that active modulation of the intestinal microbiota, through probiotic supplementation, can have drastic effects on behavior, perhaps with therapeutic potential.

Anxiety is not the only behavior that is modulated by signals from symbiotic bacteria. The same study that utilized the probiotic *L. rhamnosus* (JB-1) to treat mice also observed that these mice displayed decreased depression-like behaviors, particularly in the forced swim test (Bravo et al., 2011). Probiotic treatment resulted in animals spending less time immobilized when introduced into a water-filled cylinder, rather than attempt to swim. In a similar fashion, a separate study utilized the maternal separation model of depression in rats, in which pups are removed from their mothers for a period of time each day until weaning (Vetulani, 2013). This study noted that the depression-like behaviors induced in this model could be reduced upon treatment with B. infantis (Desbonnet et al., 2010). Strikingly, B. infantis treatment resulted in a similar reduction in depression-like behavior in the forced swim test as was observed upon treatment with the pharmaceutical antidepressant citalopram, a selective serotonin reuptake inhibitor (SSRI) (Desbonnet et al., 2010; Savignac et al., 2014, 2015). These surprising findings further suggest that the composition and/or function of the microbiota actively modulates behavior in adult animals.

Intriguingly, probiotic supplementation, or even diet alone, are observed to have an impact on both anxiety and depressive behavior in mice, as well as on learning and memory (Li et al., 2009; Pyndt Jørgensen et al., 2014; Savignac et al., 2015). While some studies have also demonstrated that specific diets alter the composition of the microbiome and have effects on behaviors, whether the microbial composition itself is causative for the observed depressive-behavior and learning deficits observed in these studies is not clear (Li et al., 2009; Pyndt Jørgensen et al., 2014). Functional studies utilizing transplantation of dietaltered microbiomes would be essential to establish a microbial role. Nonetheless, such observations serve to pave the way toward understanding the role that the composition and functionality of indigenous microbes may play in shaping mood and behavior in the host.

While most studies have utilized animal models, work is being performed to understand whether the microbiota have similar roles in shaping human neurological function. A recent human study was performed to address whether the consumption of a fermented, probiotic milk product (containing B. animalis subsp Lactis, Streptococcus thermophilus, L. bulgaricus, and Lactococcus lactis subsp Lactis.) could have an effect on the brain response to emotional stimuli. Using fMRI, Mayer and colleagues found that probiotic consumption over the span of 4 weeks could affect the processing of emotion (Tillisch et al., 2013). Specifically, those regions of the brain involved in emotional processing, including the primary interoceptive and somatosensory regions, were less activated following emotional stimulation (emotional faces task) in those individuals who had consumed this probiotic product (Tillisch et al., 2013). Similarly, a separate study observed that probiotic consumption reduced self-reported feelings of sadness and aggressive thoughts (Steenbergen et al., 2015). Importantly, while these particular studies were small and did not address exactly how the bacterial cocktail mediated changes in functional brain activity and mood, they are nonetheless strongly suggestive that the microbiota can actively alter some functional aspects of mood in humans. A larger human study of 55 individuals examined whether probiotic consumption could influence anxiety (through self-assessment questionnaires) (Messaoudi et al., 2011). Administration of L. helveticus R0052 and B. longum R0175 resulted in decreased self-reported anxiety and, notably, also resulted in decreased urinary cortisol (Messaoudi et al., 2011). Together, these studies

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further the concept that the microbiota effects neurological function in humans and ultimately influences mood and behavior. However, additional work with increased cohort sizes, using a crossover study design and clinical assessment, are needed to validate these seminal observations for a gut-microbiome-brain connection in humans.

Interestingly, it has been observed in both humans and mice that probiotic supplementation with fermented dairy products does not necessarily alter the composition of the gut microbiome (McNulty et al., 2011). Instead, the transcriptional state and/or metabolic activity of the microbiota are altered (McNulty et al., 2011). It is therefore interesting to consider that these behavioral and neurological changes may not necessarily be a direct function of the specific species of bacteria within the probiotic treatment; rather, microbial-mediated effects on emotion may be due to broader functionality of the community of symbiotic bacteria within the gut. Continued study of how the presence of specific bacterial species, particularly low abundance niche species, influences physiology but does not alter the overall population of gut microbes, will be critical to understanding the function of the microbiota and how they can be effectively harnessed and developed as a potential therapeutic modality for behavioral disorders.

One well-established physiological function of the microbiota is the generation of essential nutrients for host physiology, such as vitamins and other cofactors (Gordon et al., 2012). Therefore, it is tempting to speculate that perhaps these microbes could subsequently alter satiety, influencing how much food the host ingests. While there is no unequivocal evidence demonstrating that the microbiota directly influence appetite and satiety, there are intriguing indirect observations. Upon ingestion of fermentable complex carbohydrates, the microbiota metabolize fiber into short-chain fatty acids (SCFAs) such as acetate, butyrate, and propionate (Miller and Wolin, 1979). Interestingly, SCFAs produced in the GI tract are trafficked not only into the serum, but also are capable of crossing the blood-brain barrier (BBB) (Conn et al., 1983; Mitchell et al., 2011). Once in the brain, one particular SCFA, acetate, has been observed to enact physiological changes in the hypothalamus. Here, acetate alters the level of the neurotransmitters glutamate, glutamine, and γ-aminobutyric acid (GABA) as well as increases anorectic neuropeptide expression (Frost et al., 2014), which together act as hormone signals to reduce appetite (Sobrino Crespo et al., 2014). In total, production of acetate leads to suppression of appetite. In fact, a small study in humans found that fermentation of complex carbohydrates to SCFAs by the microbiota was directly correlated with the sensation of satiety (Cani et al., 2009), providing a basis for the hypothesis that the symbiotic microbes may be capable of modulating host appetite. Indeed, changes to the microbiome, due to a specific genetic mutation in mice, induce a metabolic syndrome, likely through changes in feeding behaviors rather than alterations to metabolism (Vijay-Kumar et al., 2010).

Microbiota Shape Social Behaviors

Numerous facets of social behavior are also altered by the presence, composition, and functionality of the microbiota (Mayer et al., 2015; Stilling et al., 2014b). GF mice are significantly socially impaired compared to SPF colonized counterparts (Desbonnet et al., 2014). GF mice do not seek out other mice (both

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new and familiar) as readily as mice harboring a diverse microbial consortium (Desbonnet et al., 2014), suggesting that the microbiota affect these social behaviors. Surprisingly, social avoidance could be restored through recolonizing adult mice with a complete microbiota. Conversely, the defect in social cognition (that is, the ability to recognize familiar versus unfamiliar mice) was not restored following colonization. Thus, gut bacteria have differential effects on both developmental aspects, as well as active processes that occur in adulthood, which ultimately shape long-term behavioral traits (Desbonnet et al., 2014).

One particularly interesting example of microbiome-controlled social behavior is that of sexual mate preference in the fruit fly, Drosophila melanogaster. Flies that have been colonized with L. plantarum prefer to mate only with similarly colonized flies, and not with flies colonized by other bacterial species (Sharon et al., 2010). This is due to the increased production of certain pheromones whose precursors are produced by L. plantarum within these flies. As such, this provides a strong, mechanistic example to begin to understand how the microbiome can influence the extremely complex social behavior of mate preference (Sharon et al., 2010). Building on this, an interesting study of social behaviors in hyenas found that individual social groups of these animals harbored distinct microbiota within their scent glands (Theis et al., 2012, 2013). Scent-gland dwelling symbiotic bacteria are known to produce volatile compounds, and each unique microbial community produces different ratios of these compounds (Theis et al., 2012, 2013). As distinct social groups had similar microbiomes, individuals likely are recognized as belonging to a particular social group directly due to the composition of the microbes within their glands. These studies provide support for the notion that symbiotic bacteria affect how a host interacts in social settings, modulating how the host perceives novel versus familiar individuals for both mate choice and social grouping. The ramifications of this microbial influence on social behaviors has not gone unnoticed by evolutionary biologists (Montiel-Castro et al., 2014; Rosenberg et al., 2010; Stilling et al., 2014a). If gut bacteria modulate neurological function leading to the choice of mate, the microbiome may therefore play a significant role in driving the evolution of their hosts. By influencing how individuals interact and undergo vertical genetic transfer, the microbiota could ultimately be a critical contributing factor in the evolution of metazoan species.

The Link between Gut Bacteria and Disorders Involving Social Impairment

Autism spectrum disorder (ASD) comprises a set of complex neurodevelopmental disabilities characterized by repetitive/stereotypic behaviors and deficits in communication and social interaction. Intriguingly, a significant subset of ASD children exhibit GI complications, including constipation, increased intestinal permeability, and altered composition of the intestinal microbiome (Kang et al., 2013; Mulle et al., 2013; Rosenfeld, 2015). Mice displaying autism-like behaviors have a significantly altered microbiome compared to neurotypical controls; have an increase in the permeability of the colon; and, correlatively, display differences in serum metabolites (Hsiao et al., 2013). Surprisingly, treatment with a single organism, *Bacteroides fragilis*, was able to restore the intestinal permeability defects in a mouse model of ASD (Hsiao et al., 2013). While treatment with this probiotic did not restore the overall composition of the microbial population, *B. fragilis* treatment could restore levels of a small number of specific species (Hsiao et al., 2013). Notably, *B. fragilis* treatment rescued some behavioral defects, including stereotyped behavior (compulsive marble burying), communication deficits (ultrasonic vocalizations), and anxiety behaviors (open-field exploration) (Hsiao et al., 2013).

B. fragilis has been shown to augment the development and function of the immune system (Mazmanian et al., 2008; Ochoa-Repáraz et al., 2010; Round and Mazmanian, 2010); however, treatment with B. fragilis did not restore several aspects of immune dysfunction in an animal model of autism (Hsiao et al., 2012, 2013). Instead, levels of serum metabolites found to be altered in mice with ASD-related behaviors were restored to normal levels. Administration to healthy animals of a specific serum metabolite that was elevated in mice with behavioral deficits, namely 4-ethylphenyl sulfate (4EPS), was sufficient to induce anxiety-like behavior. As 4EPS is predicted to be of microbial origin, this remarkably demonstrates that defined molecules from the microbiome can impact behavior in mammals. Similarly, another study demonstrated that mice with features of ASD display a significantly altered microbiome and increased intestinal inflammation (de Theije et al., 2014). Although still correlative, these data together suggest that certain neurodevelopmental disorders, such as autism, may have microbial etiologies, a hypothesis that will require further validation in both animal models and human trials. The notion that the gut microbiome constitutes an environmental risk factor for autism is supported by epidemiologic studies showing a rapid increase in ASD diagnoses over the past few decades, suggesting that genetics alone cannot explain many cases of disease. If true, targeted repair of an altered microbiome through interventions including probiotics, prebiotics, or diet may represent natural, safe, and effective treatments for neurological disorders such as autism.

Microbiome-Mediated Alterations to Neurophysiology

The observation that GF or probiotic-treated mice have altered behavior raises numerous interesting questions related to gutbrain communication. Do microbially derived signals act directly on the nervous system, or indirectly via the immune, endocrine, or metabolic systems and/or other pathways? Is the influence of the microbiome due to developmental effects, or is the microbiome-brain axis an actively modulated process? What are the neurophysiological changes in the brain that arise due to alterations to the microbiome, which may underlie behavioral effects? Mechanisms and consequences for long distance interoceptive communication between gut bacteria and the brain are likely to be context specific and not mutually exclusive.

In some cases, there is evidence that microbiota-mediated outcomes are required to occur during a specific time frame of development and subsequently have irreversible downstream neurological affects (Borre et al., 2014). In other examples, neurological function can be actively modulated by signals from the microbiome. For example, recolonizing adult GF mice with a complete microbiota restores their anxiety-like behavior to that of SPF mice, demonstrating that control of

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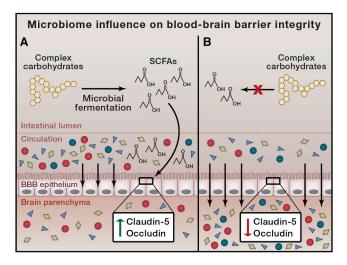


Figure 2. Microbiome Influence on BBB Integrity

Intestinal microbes are capable of fermenting complex carbohydrates into SCFAs.

(A) Microbially produced SCFAs signal to endothelial cells that create the BBB and increase production of the tight junction proteins claudin-5 and occludin. This leads to a tight and selective barrier, preventing undesired metabolites from entering the brain parenchyma.

(B) In the absence of microbial fermentation, no SCFA signaling occurs, and tight junction proteins are repressed. This leads to increased permeability of the BBB and a loss of the selective barrier to serum metabolites.

anxiety behavior can occur through an active and constant process between signals from the microbiome and the CNS (Clarke et al., 2013). However, GF mice also display lower levels of BDNF, serotonin (5-HT; 5-hydroxytryptamine), and specific 5-HT receptors (for instance, $5HT_{1A}$) in regions such as the amygdala and hippocampus (Bercik et al., 2011; Clarke et al., 2013; Diaz Heijtz et al., 2011; Neufeld et al., 2011). The levels of these host molecules are not restored upon re-colonization of adult mice, suggesting that certain phenotypes are likely programmed by the microbiota during fetal development or in adolescence (Clarke et al., 2013).

Other processes are more actively modulated by the microbiome. It has also been observed that GF mice display an increased rate of turnover of noradrenaline, dopamine, and 5-HT in the striatum region of the brain (Diaz Heijtz et al., 2011). A high rate of turnover may subsequently have an effect on steady-state levels of these neurotransmitters. Turnover of norepinephrine and dopamine specifically may be responsible for the increase in motor activity that is well documented in GF mice (Diaz Heijtz et al., 2011), as these neurotransmitters have roles in increasing blood flow to muscle and central motor control, respectively.

While alterations to the population or function of the microbiome may result in differential production of metabolites that enter the periphery, it is unknown if these molecules could cross the BBB and influence neurological function. Intriguingly, GF mice have significantly increased permeability of the BBB, both during fetal development and in adulthood (Braniste et al., 2014) (Figure 2). This increase in permeability is due to lowered expression of endothelial tight junction proteins, specifically occludin and claudin-5, in the absence of the microbiota. Interestingly, mono-colonization with either *Clostridium tyrobutyricum* or B. thetaiotaomicron could restore BBB integrity (and production of tight junction expression), even in adult mice. As these species produce SCFAs from fermentation of complex carbohydrates in the gut, it was subsequently observed that the SCFA butyrate was sufficient to restore BBB integrity (Braniste et al., 2014) (Figure 2). This suggests that metabolic signals that are derived from the gut microbiota actively and constantly influence the physiological status of the BBB, a site quite distant from their origin. However, it is unknown whether other SCFAs or microbial-derived signals, or even microbial species, may also play a role in influencing BBB permeability. This finding has important implications in other physiological processes: it indicates that serum metabolites that normally do not cross into the brain parenchyma may potentially cross the BBB based on the status of the microbiota, providing a mechanism for gut microbes to control concentrations of numerous metabolites that can act directly on neurological systems (Figure 2).

Regulation of Neurotransmitter Levels by the Gut Microbiota

While it has been observed that changes to neurophysiology can be mediated by the microbiome, the precise mechanism by which this influence occurs is still unclear. Although altering the permeability of the BBB would change the flux of serum metabolites into and out of the brain, there are potentially more direct ways in which the microbiome can alter neurological function. One of the most direct mechanisms could be through controlling the concentration of various neurotransmitters, both in the brain and in the periphery. For example, 5-HT levels in peripheral serum are decreased in the absence of the gut microbiota (Wikoff et al., 2009; Yano et al., 2015). This decrease corresponds to lower levels of 5-HT metabolites and precursors in the intestinal luminal contents and urine (Marcobal et al., 2013; Matsumoto et al., 2012). The vast majority of 5-HT in the body (~90%) is produced by enterochromaffin cells in the gut (Gershon, 2013). Release of 5-HT by enterochromaffin cells is necessary for modulating colonic motility (Fukumoto et al., 2003). It has recently been demonstrated that microbial-derived SCFAs are capable of inducing 5-HT production by enterochromaffin cells, in vitro and in animals (Reigstad et al., 2014; Yano et al., 2015). Additionally, 5-HT is not known to cross the BBB, and therefore the microbiome control of 5-HT turnover in the brain may instead occur through alterations in 5-HT precursor levels (O'Mahony et al., 2015; Sharon et al., 2014).

In particular, the essential amino acid tryptophan is a central precursor to 5-HT synthesis. Tryptophan itself is generated by the intestinal microbiota, and tryptophan present in the periphery is capable of crossing the BBB, where it can then participate in 5-HT synthesis (O'Mahony et al., 2015; Sharon et al., 2014). Nonetheless, even local stimulation and production of 5-HT in the GI tract would have important effects on host physiology, since 5-HT modulates GI motility (Berger et al., 2009). Intriguingly, 5-HT levels in the gut can be restored in GF mice following colonization with a defined cocktail of spore-forming gut bacteria (Yano et al., 2015), a process that regulates platelet aggregation and blood clotting. Non-SCFA metabolites produced by this microbial community stimulate gut enterochromaffin cells to produce 5-HT, compensating for a defect in platelet activity and coagulation (a process known to be regulated by 5-HT).

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Consequences of 5-HT regulation by bacteria in the intestine on the concentration of 5-HT in the brain and on behavior in mice remain unknown; however, the prevalent usage of SSRIs, which increase the concentration of 5-HT (serotonin) at the synapse, warrants interest in the potential of developing probiotic therapies as an alternative treatment for major depressive disorder and anxiety.

5-HT is not the only neurotransmitter whose concentrations are influenced by the microbiome. Serum levels of other neurotransmitters are also decreased in the absence of the gut microbiota. Dopamine and GABA are decreased in the serum of GF mice, and specific precursors and metabolites of these are also altered in the intestine (Matsumoto et al., 2012; Velagapudi et al., 2010). Exactly how gut bacteria alter the levels of these neurotransmitters remains to be determined. Direct signals from the microbiota to neurotransmitter-producing cells, such as the enterochromaffin cells or even to enteric neurons and glia, may regulate neurotransmitter production. While such signaling to neurotransmitter-producing cells is one way in which the microbiota may influence neurotransmitter concentrations, gut-dwelling bacteria also directly produce small molecules with potential to act as neurotransmitters. In turn, these may act as signals to GI cells, or make their way to the periphery and potentially the brain, and ultimately influence neurological function.

It has been known for many years that specific species of gut bacteria are capable of producing small molecules such as serotonin, dopamine, norepinephrine, epinephrine, GABA, and acetylcholine, possible bioactive neuropeptides (Wall et al., 2014). However, it is unknown whether the microbially derived molecules can act directly on host receptors as neurotransmitters. Recently, it has been observed that the specific human gut microbes C. sporogenes and Ruminococcus gnavus are capable of producing the neurotransmitter tryptamine through decarboxylation of tryptophan (Williams et al., 2014). In the brain, tryptamine plays a role in the inhibitory response to 5-HT, through its action on the trace amine-associated receptor, and may modulate mood and appetite (Zucchi et al., 2006). In the gut, tryptamine can also induce enterochromaffin cells to release 5-HT. While tryptamine can in fact cross the BBB from serum, it is not known whether gut microbiota-produced tryptamine is trafficked from the GI tract to the CNS and subsequently influences neurological function.

Another potential neurotransmitter that is produced by bacteria present in the gut is tyramine. Multiple microorganisms, including L. brevis and Enterococcus species, are capable of decarboxylating tyrosine to tyramine (Lucas et al., 2003). Since tyramine has been shown to modulate motor function in worms, as well as trace amine-associated receptors in mammals (Zucchi et al., 2006), it raises the possibility that gut-microbiotaproduced tyramine may act as a modulator of neurological function as well. Similarly, 5-HT is produced by a number of gut microbes in vitro, specifically many lactic acid bacteria (Ozogul et al., 2012), and other human intestinal bacteria have been shown to produce GABA (Minuk, 1986). While GABA and 5-HT are not known to cross the BBB, intestinally derived (and therefore likely microbiota-derived) production of these molecules may instead act locally on the vagus nerve or through signaling via the periphery (Barrett et al., 2012). In the intestine, GABA is important for modulating motility, emptying, and secretion in the intestine; in the periphery, it controls aspects of stress and thermoregulation (Hyland and Cryan, 2010; Li et al., 2012; Paredes and Agmo, 1992), suggesting that signals from the microbiota may contribute to these diverse neurophysiological functions.

Contrary to conventional wisdom, most neurotransmitters are found in the gut at levels equal to or exceeding those in the brain. Furthermore, the proportion of total body levels of various neurotransmitters is greater in the gut than the brain. While most of the current data for microbial modulation of neurotransmitter production and/or levels is available for the gut and periphery, local neurotransmitter regulation by gut bacteria may have long distance effects on the brain. Neurotransmitters or other molecules derived from gut microbes have the potential to modulate activity of the vagus nerve, the primary nerve connecting the ENS to the CNS, and subsequently influence brain function (Bravo et al., 2011; Goehler et al., 2005). It is also possible that microbially derived metabolites, which can act as precursors to neurotransmitter production (such as tryptophan) and may cross through the intestinal barrier and the BBB, could subsequently influence both systemic and CNS neurotransmitter concentrations. It is important to note, however, that many of these studies describing neurotransmitter production by intestinal bacteria have been performed in vitro, and thus, it is not known precisely whether these bacteria utilize such metabolic pathways in vivo and, if so, when these specific gut microbes produce these compounds. Nevertheless, it is an interesting prospect that the intestinal microbiota act to produce neurotransmitters and directly modulate the nervous system (Lyte, 2013). Such findings open up exciting possibilities in understanding how the microbiome may affect function of the nervous system through the creation of bioactive metabolites that are capable of modulating CNS activity via several modes of gut-brain connection.

Microbial Control of Neurological Function by the Immune System

Interactions between the microbiota and the nervous system may be indirect. Growing data indicate that the peripheral immune system can influence neurological function and behavior. In fact, immune signaling has been shown to cause or allow progression of certain neurological disorders, including neurodegenerative diseases and psychological illness such as anxiety and depression. One of the better-known examples of immune-mediated effects on neurological function is sickness behavior (Figure 3). This behavior is characterized by appetite suppression, decreased motor activity, loss of social interaction, and reduced cognition (Dantzer et al., 2008). Microbial-associated molecular patterns (MAMPs), such as lipopolysaccharide (LPS), bacterial lipoprotein (BLP), flagellin, and CpG DNA, among others, activate various cells of the immune system, particularly innate immune cells such as macrophages, neutrophils, and dendritic cells. Once activated, these cells produce numerous pro-inflammatory cytokines, such as IL-1 α , IL-1 β , TNF α , and IL-6, which make their way to the brain by crossing the BBB via both diffusion and cytokine transporters. Once in the brain, these cytokines act on receptors expressed by neurons and glial cells, particularly microglia (brain-resident, innate immune phagocytes), altering their activation status and physiology

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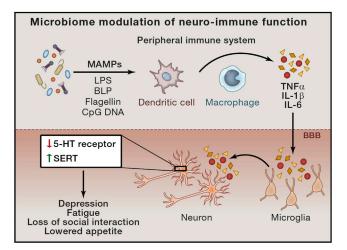


Figure 3. Microbiome Modulation of Neuro-Immune Function MAMPs derived from the intestinal microbiome can drive various aspects of immune function in the periphery. Cytokine signals, such as $TNF\alpha$, IL-1 β , and IL-6, can cross the BBB and trigger their production by the microglia. Interaction of these cytokines with neurons influences physiology and leads to sickness behavior and depression.

(Dantzer et al., 2000) (Figure 3). In the periphery, these cytokines are capable of acting on receptors in afferent nerves, promoting alterations in the signals leading from distant, peripheral body sites to the CNS. Introduction of purified IL-1 β and TNF α into the brain or the periphery, as well as systemic treatment with LPS (which induces pro-inflammatory gene expression in the brain and periphery) is sufficient to mediate sickness behavior, while IL-6 treatment is only capable of promoting fever, but not behavioral attributes (Breder et al., 1994; van Dam et al., 1992) (Figure 3). In further support of pro-inflammatory cytokines mediating the onset of this behavior, treatment with anti-inflammatory mediators, such as IGF-1 and IL-10, prevents sickness behavior (Bluthé et al., 1999, 2006). In fact, the role of pro-inflammatory cytokines in modulating human behavior is exemplified by the finding that one-third of patients who have been treated with IL-2 and IFN α (used as immunotherapy against cancer and certain viral infections) develop major depressive disorder (Raison et al., 2006).

Additionally, immune dysregulation is observed in both human cases and animal models of ASD (Ashwood et al., 2006; Hsiao, 2013; Hsiao et al., 2012; Mead and Ashwood, 2015). In the maternal immune activation (MIA) model of autism, a decrease in regulatory T cells and an increase in pro-inflammatory innate monocytes are observed in mice that display ASD symptoms. Bone marrow transplantation from healthy donors (and subsequent restoration of immune function) is capable of rescuing behavioral defects in this model (Hsiao et al., 2012). However, given the role of the microbiota in modulating behaviors in this model (Hsiao et al., 2013), it is interesting to consider that irradiation itself may have also restored behavioral functions due to an effect on the composition of the microbiome and/or by "resetting" the immune system. Nonetheless, these data provide evidence for a link between immune dysfunction and complex social and behavioral disorders.

Exactly how pro-inflammatory cytokine signaling can mediate alterations in neurological functions that affect behavior and behavioral disorders is beginning to be revealed. Sickness behavior and depressive disorders, for instance, are linked to alterations in 5-HT signaling. In this vein, IL-1 β or TNF α treatment stimulate 5-HT uptake through upregulation of the serotonin transporter (SERT), decreasing the concentration of 5-HT available to signal in the synapase (Zhu et al., 2006). At the same time, these cytokines decrease the level of 5-HT receptor (5-HT_{2A}) present on neurons, escalating the loss of 5-HT signaling and likely mediating the alterations to behavior (Cai et al., 2005) (Figure 3).

While pro-inflammatory cytokine signaling can be driven by pathogenic microbes, some microbiota-derived signals induce non-inflammatory cytokine pathways. For instance, in the absence of a complete microbiota, mice exhibit a lower plasma concentration of the cytokine granulocyte colony stimulating factor (G-CSF) (Deshmukh et al., 2014). Plasma G-CSF is capable of crossing the BBB and acts to stimulate neurogenesis in the brain (Zhao et al., 2007). Thus, through stimulation of G-CSF production, the microbiota may influence the rate of neurogenesis. This could have effects not only on normal neurodevelopmental processes where G-CSF signaling plays a role, but also in injury and neurodegenerative disease. G-CSF is a protective factor following ischemic injury (Shyu et al., 2004) as well as protective and therapeutic in certain models of both Parkinson's and Alzheimer's diseases (Meuer et al., 2006; Prakash et al., 2013). Therefore, it is exciting to think that this may be one way in which alterations to the microbiome could modulate the outcome of these neurodegenerative diseases. Or more intriguingly, introduction of microbes that specifically promote G-CSF production may represent a future avenue of microbiota-mediated therapy to combat neurodegenerative illnesses.

Future Directions and Conclusions

While the role of the microbiome in influencing numerous aspects of metabolic and immunologic aspects is well established, how indigenous microbes modulate neurological function during health and disease is only now becoming appreciated. Here, we have described emerging evidence for the behavioral and neurophysiological conditions in animal models and human studies that have been linked to the microbiome. Rapid and sustained growth of research on the gut-microbiome-brain connection may lead to discoveries that prompt a reconsideration of potential environmental influences on numerous neurological diseases whose causes have remained enigmatic and where treatment options are limited.

Several seminal reports now show that animals lacking microbiota have significantly altered brain development and behavior compared to colonized counterparts, highlighting the stark importance of host-microbial symbiosis. GF animals provide a valuable model to determine not only the precise physiological processes that the microbiota influence, but also the extent of the effects. Along with probiotic or antibiotic treatment of colonized animals, GF models provide an experimental platform to reconstitute biological systems with defined communities or single microbial species, and from a variety of donor sources (e.g., genetically engineered mice, humans, etc.) allowing for the discovery and functional characterization of organisms and molecules that impact the nervous system. Identification of microbes (either single species or consortia) that modulate these systems

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will advance efforts to explore the nature of the microbial-derived signals and host pathways that influence specific neurophysiological function. Genomic and genetic approaches to study newly discovered organisms, alongside metabolomic analyses to identify products or compounds from symbiotic bacteria, may provide critical insight into how particular microbial molecules alter host neurophysiology. Accordingly, identification of bioactive microbial signals may serve as a tool for the discovery of currently unidentified host pathways or novel activities of known pathways that influence behavior and neurological function, similar to how the study of bacterial pathogenesis has uncovered the intricacies of host immune system pathways by first identifying those microbial signals that modulate them.

It will be critical to differentiate whether the influence of the microbiome on a host process is developmental or active in nature. That is, are the signals from the microbiota that alter neurological function important at a specific time during development (Borre et al., 2014), or can phenotypes be actively modulated in fully developed animals? Are certain aspects of behavior mediated by signals that are derived from the maternal microbiome, or instead, can fluctuations to the composition and function of an adult microbiome also contribute to neurological function? Given that the majority of brain development occurs in utero, maternal signals are likely to play a significant role. While the womb was believed to be sterile, recent controversial observations have called this into question and suggest microbially derived products may interact directly with the developing fetus (Aagaard et al., 2014; Borre et al., 2014; Funkhouser and Bordenstein, 2013; Jiménez et al., 2008). Furthermore, the physiological influence on maternal systems may have indirect effects on fetal development, such as through metabolic and/or immune pathways (PrabhuDas et al., 2015). Several studies have recently shed light on the relevance of the maternal-fetal interaction and the important contribution of microbiome dynamics in the first few years of life. There is likely not a single answer, and specific processes will likely have distinct roles for the microbiome in different contexts.

Perhaps one of the most pressing areas of research in the field is understanding the physiological consequences of altered microbiome populations that correlate with certain disease states. Do these changed populations enact a physiological effect that drives the disease they are correlated with? In other words, are alterations to the microbiome causal to a given condition? Or instead, are changes to gut bacterial composition simply a consequence of the disease state? For many neurological diseases that lack a strong genetic component, environmental factors are thought to play a critical role. Further study may uncover the microbiome as an important environmental factor that may be an etiological agent of disease, instigating effects that have lasting consequences on the initiation and/or progression of neurological illness. Future studies addressing these questions will add critical relevance to correlative associations described to date between the microbiome and disease of the nervous system.

As we begin to understand how microbial-derived processes influence the brain and behavior during health and disease, we may perhaps begin to rationally design microbiota-based therapeutics for neurological disorders. For example, understanding the signals that drive neurotransmitter production by intestinal bacteria (either directly or indirectly through the production of precursors) may provide a foundation for therapies in diseases that are currently treated by pharmacological alteration of neurotransmitter levels. This could include a range of disorders, from neurodegenerative diseases like Parkinson's disease, which is treated with oral L-DOPA to stimulate dopamine levels, to depression, which is commonly treated with SSRIs that increase the concentration of 5-HT available to signal in the synapse. Additionally, understanding how the microbiota modulate immune responses at distal sites may lead to understanding how we can specifically alter gut microbes to influence immunemediated pathologies in the brain, such as those that occur during stroke, seizures, and neurodegenerative diseases.

While microbiologists have understood for decades that microbes can alter the hosts' behavior in various invertebrate and vertebrate systems, it is only in the last 5 years that other scientific communities have begun to appreciate the potential scale and depth by which commensal microbes affect complex neurological function in mammals. A handful of molecular mechanisms that control these interactions have been identified; however, the field is poised to make great strides in understanding this new paradigm based on growing awareness of the gut-microbiome-brain axis. Multi-disciplinary collaborations between microbiologists, immunologists, neuroscientists and bioinformaticians will continue to drive discovery into how symbiotic microbes function to shape our brains and behaviors and how we may exploit these organisms to combat neurological diseases. Neurodevelopmental, psychiatric, and neurodegenerative disorders represent some of the most serious medical and societal burdens of our time; yet the etiologies of most disorders of the brain remain unknown, and therapies are largely either ineffective or have severe side effects. New concepts and therapeutic modalities are desperately needed to explain and address many neurological conditions. It appears that future discoveries in the neurosciences will not solely rely on studying the brain, but also surprisingly through exploration of a forgotten organ with similar size and complexity-the human microbiome.

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REFERENCES

Aagaard, K., Ma, J., Antony, K.M., Ganu, R., Petrosino, J., and Versalovic, J. (2014). The placenta harbors a unique microbiome. Sci. Trans. Med. *6*, 237ra265.

Adamo, S.A., Kovalko, I., Easy, R.H., and Stoltz, D. (2014). A viral aphrodisiac in the cricket Gryllus texensis. J. Exp. Biol. 217, 1970–1976.

Ashwood, P., Wills, S., and Van de Water, J. (2006). The immune response in autism: a new frontier for autism research. J. Leukoc. Biol. *80*, 1–15.

Barrett, E., Ross, R.P., O'Toole, P.W., Fitzgerald, G.F., and Stanton, C. (2012). γ -Aminobutyric acid production by culturable bacteria from the human intestine. J. Appl. Microbiol. *113*, 411–417.

Belkaid, Y., and Hand, T.W. (2014). Role of the microbiota in immunity and inflammation. Cell *157*, 121–141.

Bercik, P., Denou, E., Collins, J., Jackson, W., Lu, J., Jury, J., Deng, Y., Blennerhassett, P., Macri, J., McCoy, K.D., et al. (2011). The intestinal microbiota affect central levels of brain-derived neurotropic factor and behavior in mice. Gastroenterology *141*, 599–609.

Berger, M., Gray, J.A., and Roth, B.L. (2009). The expanded biology of serotonin. Annu. Rev. Med. 60, 355–366.

Biron, D.G., Marché, L., Ponton, F., Loxdale, H.D., Galéotti, N., Renault, L., Joly, C., and Thomas, F. (2005). Behavioural manipulation in a grasshopper harbouring hairworm: a proteomics approach. Proc. Biol. Sci. 272, 2117–2126.

Bluthé, R.M., Castanon, N., Pousset, F., Bristow, A., Ball, C., Lestage, J., Michaud, B., Kelley, K.W., and Dantzer, R. (1999). Central injection of IL-10 antagonizes the behavioural effects of lipopolysaccharide in rats. Psychoneuroendocrinology *24*, 301–311.

Bluthé, R.M., Kelley, K.W., and Dantzer, R. (2006). Effects of insulin-like growth factor-I on cytokine-induced sickness behavior in mice. Brain Behav. Immun. 20, 57–63.

Borre, Y.E., O'Keeffe, G.W., Clarke, G., Stanton, C., Dinan, T.G., and Cryan, J.F. (2014). Microbiota and neurodevelopmental windows: implications for brain disorders. Trends Mol. Med. *20*, 509–518.

Braniste, V., Al-Asmakh, M., Kowal, C., Anuar, F., Abbaspour, A., Toth, M., Korecka, A., Bakocevic, N., Guan, N.L., Kundu, P., et al. (2014). The gut microbiota influences blood-brain barrier permeability in mice. Sci. Transl. Med. 6, 263ra158.

Bravo, J.A., Forsythe, P., Chew, M.V., Escaravage, E., Savignac, H.M., Dinan, T.G., Bienenstock, J., and Cryan, J.F. (2011). Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc. Natl. Acad. Sci. USA *108*, 16050–16055.

Breder, C.D., Hazuka, C., Ghayur, T., Klug, C., Huginin, M., Yasuda, K., Teng, M., and Saper, C.B. (1994). Regional induction of tumor necrosis factor alpha expression in the mouse brain after systemic lipopolysaccharide administration. Proc. Natl. Acad. Sci. USA *91*, 11393–11397.

Cai, W., Khaoustov, V.I., Xie, Q., Pan, T., Le, W., and Yoffe, B. (2005). Interferon-alpha-induced modulation of glucocorticoid and serotonin receptors as a mechanism of depression. J. Hepatol. *42*, 880–887.

Cani, P.D., Lecourt, E., Dewulf, E.M., Sohet, F.M., Pachikian, B.D., Naslain, D., De Backer, F., Neyrinck, A.M., and Delzenne, N.M. (2009). Gut microbiota fermentation of prebiotics increases satietogenic and incretin gut peptide production with consequences for appetite sensation and glucose response after a meal. Am. J. Clin. Nutr. 90, 1236–1243.

Clarke, G., Grenham, S., Scully, P., Fitzgerald, P., Moloney, R.D., Shanahan, F., Dinan, T.G., and Cryan, J.F. (2013). The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. Mol. Psychiatry *18*, 666–673.

Conn, A.R., Fell, D.I., and Steele, R.D. (1983). Characterization of alpha-keto acid transport across blood-brain barrier in rats. Am. J. Physiol. 245, E253–E260.

Dantzer, R., Konsman, J.P., Bluthe, R.M., and Kelley, K.W. (2000). Neural and humoral pathways of communication from the immune system to the brain: parallel or convergent? Auton. Neurosci. *85*, 60–65.

Dantzer, R., O'Connor, J.C., Freund, G.G., Johnson, R.W., and Kelley, K.W. (2008). From inflammation to sickness and depression: when the immune system subjugates the brain. Nat. Rev. Neurosci. 9, 46–56.

de Theije, C.G., Wopereis, H., Ramadan, M., van Eijndthoven, T., Lambert, J., Knol, J., Garssen, J., Kraneveld, A.D., and Oozeer, R. (2014). Altered gut microbiota and activity in a murine model of autism spectrum disorders. Brain Behav. Immun. 37, 197–206.

Desbonnet, L., Garrett, L., Clarke, G., Kiely, B., Cryan, J.F., and Dinan, T.G. (2010). Effects of the probiotic Bifidobacterium infantis in the maternal separation model of depression. Neuroscience *170*, 1179–1188.

Desbonnet, L., Clarke, G., Shanahan, F., Dinan, T.G., and Cryan, J.F. (2014). Microbiota is essential for social development in the mouse. Mol. Psychiatry 19, 146–148. Deshmukh, H.S., Liu, Y., Menkiti, O.R., Mei, J., Dai, N., O'Leary, C.E., Oliver, P.M., Kolls, J.K., Weiser, J.N., and Worthen, G.S. (2014). The microbiota regulates neutrophil homeostasis and host resistance to Escherichia coli K1 sepsis in neonatal mice. Nat. Med. *20*, 524–530.

Diaz Heijtz, R., Wang, S., Anuar, F., Qian, Y., Björkholm, B., Samuelsson, A., Hibberd, M.L., Forssberg, H., and Pettersson, S. (2011). Normal gut microbiota modulates brain development and behavior. Proc. Natl. Acad. Sci. USA *108*, 3047–3052.

Driver, C. (2014). Rabies: risk, prognosis and prevention. Nurs. Times 110, 16-18.

Forsythe, P., Bienenstock, J., and Kunze, W.A. (2014). Vagal pathways for microbiome-brain-gut axis communication. Adv. Exp. Med. Biol. 817, 115–133.

Foster, J.A., and McVey Neufeld, K.A. (2013). Gut-brain axis: how the microbiome influences anxiety and depression. Trends Neurosci. 36, 305–312.

Frost, G., Sleeth, M.L., Sahuri-Arisoylu, M., Lizarbe, B., Cerdan, S., Brody, L., Anastasovska, J., Ghourab, S., Hankir, M., Zhang, S., et al. (2014). The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. Nat. Commun. 5, http://dx.doi.org/10.1038/ncomms4611.

Fukumoto, S., Tatewaki, M., Yamada, T., Fujimiya, M., Mantyh, C., Voss, M., Eubanks, S., Harris, M., Pappas, T.N., and Takahashi, T. (2003). Short-chain fatty acids stimulate colonic transit via intraluminal 5-HT release in rats. Am. J. Physiol. Regul. Integr. Comp. Physiol. 284, R1269–R1276.

Funkhouser, L.J., and Bordenstein, S.R. (2013). Mom knows best: the universality of maternal microbial transmission. PLoS Biol. *11*, e1001631.

Gershon, M.D. (2013). 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. Curr. Opin. Endocrinol. Diabetes Obes. *20*, 14–21.

Goehler, L.E., Gaykema, R.P., Opitz, N., Reddaway, R., Badr, N., and Lyte, M. (2005). Activation in vagal afferents and central autonomic pathways: early responses to intestinal infection with Campylobacter jejuni. Brain Behav. Immun. *19*, 334–344.

Gordon, J.I., Dewey, K.G., Mills, D.A., and Medzhitov, R.M. (2012). The human gut microbiota and undernutrition. Sci. Transl. Med. *4*, 137ps112.

Hooper, L.V., Littman, D.R., and Macpherson, A.J. (2012). Interactions between the microbiota and the immune system. Science 336, 1268–1273.

House, P.K., Vyas, A., and Sapolsky, R. (2011). Predator cat odors activate sexual arousal pathways in brains of Toxoplasma gondii infected rats. PLoS ONE 6, e23277.

Hsiao, E.Y. (2013). Immune dysregulation in autism spectrum disorder. Int. Rev. Neurobiol. *113*, 269–302.

Hsiao, E.Y., McBride, S.W., Chow, J., Mazmanian, S.K., and Patterson, P.H. (2012). Modeling an autism risk factor in mice leads to permanent immune dysregulation. Proc. Natl. Acad. Sci. USA *109*, 12776–12781.

Hsiao, E.Y., McBride, S.W., Hsien, S., Sharon, G., Hyde, E.R., McCue, T., Codelli, J.A., Chow, J., Reisman, S.E., Petrosino, J.F., et al. (2013). Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. Cell *155*, 1451–1463.

Hughes, D.P. (2005). Parasitic manipulation: a social context. Behav. Processes 68, 263–266.

Hyland, N.P., and Cryan, J.F. (2010). A Gut Feeling about GABA: Focus on GA-BA(B) Receptors. Front. Pharmacol. *1*, 124.

Jiménez, E., Marín, M.L., Martín, R., Odriozola, J.M., Olivares, M., Xaus, J., Fernández, L., and Rodríguez, J.M. (2008). Is meconium from healthy newborns actually sterile? Res. Microbiol. *159*, 187–193.

Kang, D.W., Park, J.G., Ilhan, Z.E., Wallstrom, G., Labaer, J., Adams, J.B., and Krajmalnik-Brown, R. (2013). Reduced incidence of Prevotella and other fermenters in intestinal microflora of autistic children. PLoS ONE *8*, e68322.

Li, W., Dowd, S.E., Scurlock, B., Acosta-Martinez, V., and Lyte, M. (2009). Memory and learning behavior in mice is temporally associated with dietinduced alterations in gut bacteria. Physiol. Behav. *96*, 557–567.

Li, Y., Xiang, Y.Y., Lu, W.Y., Liu, C., and Li, J. (2012). A novel role of intestine epithelial GABAergic signaling in regulating intestinal fluid secretion. Am. J. Physiol. Gastrointest. Liver Physiol. 303, G453–G460.

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Lu, B., Nagappan, G., Guan, X., Nathan, P.J., and Wren, P. (2013). BDNFbased synaptic repair as a disease-modifying strategy for neurodegenerative diseases. Nat. Rev. Neurosci. *14*, 401–416.

Lucas, P., Landete, J., Coton, M., Coton, E., and Lonvaud-Funel, A. (2003). The tyrosine decarboxylase operon of Lactobacillus brevis IOEB 9809: characterization and conservation in tyramine-producing bacteria. FEMS Microbiol. Lett. *229*, 65–71.

Lyte, M. (2013). Microbial endocrinology in the microbiome-gut-brain axis: how bacterial production and utilization of neurochemicals influence behavior. PLoS Pathog. 9, e1003726.

Lyte, M. (2014). Microbial endocrinology and the microbiota-gut-brain axis. Adv. Exp. Med. Biol. *817*, 3–24.

Macnab, V., and Barber, I. (2012). Some (worms) like it hot: fish parasites grow faster in warmer water, and alter host thermal preferences. Glob. Change Biol. *18*, 1540–1548.

Maitland, D.P. (1994). A parasitic fungus infecting yellow dungflies manipulates host perching behavior. P. Roy. Soc. B.-Biol. Sci. 258, 187–193.

Marcobal, A., Kashyap, P.C., Nelson, T.A., Aronov, P.A., Donia, M.S., Spormann, A., Fischbach, M.A., and Sonnenburg, J.L. (2013). A metabolomic view of how the human gut microbiota impacts the host metabolome using humanized and gnotobiotic mice. ISME J. 7, 1933–1943.

Matsumoto, M., Kibe, R., Ooga, T., Aiba, Y., Kurihara, S., Sawaki, E., Koga, Y., and Benno, Y. (2012). Impact of intestinal microbiota on intestinal luminal metabolome. Sci. Rep. 2, http://dx.doi.org/10.1038/srep00233.

Mayer, E.A., Knight, R., Mazmanian, S.K., Cryan, J.F., and Tillisch, K. (2014). Gut microbes and the brain: paradigm shift in neuroscience. J. Neurosci. *34*, 15490–15496.

Mayer, E.A., Tillisch, K., and Gupta, A. (2015). Gut/brain axis and the microbiota. J. Clin. Invest. 125, 926–938.

Mazmanian, S.K., Round, J.L., and Kasper, D.L. (2008). A microbial symbiosis factor prevents intestinal inflammatory disease. Nature 453, 620–625.

McNulty, N.P., Yatsunenko, T., Hsiao, A., Faith, J.J., Muegge, B.D., Goodman, A.L., Henrissat, B., Oozeer, R., Cools-Portier, S., Gobert, G., et al. (2011). The impact of a consortium of fermented milk strains on the gut microbiome of gnotobiotic mice and monozygotic twins. Sci. Transl. Med. *3*, http://dx.doi.org/10.1126/scitranslmed.3002701.

Mead, J., and Ashwood, P. (2015). Evidence supporting an altered immune response in ASD. Immunol. Lett. *163*, 49–55.

Messaoudi, M., Lalonde, R., Violle, N., Javelot, H., Desor, D., Nejdi, A., Bisson, J.F., Rougeot, C., Pichelin, M., Cazaubiel, M., and Cazaubiel, J.M. (2011). Assessment of psychotropic-like properties of a probiotic formulation (Lactobacillus helveticus R0052 and Bifidobacterium longum R0175) in rats and human subjects. Br. J. Nutr. *105*, 755–764.

Meuer, K., Pitzer, C., Teismann, P., Krüger, C., Göricke, B., Laage, R., Lingor, P., Peters, K., Schlachetzki, J.C., Kobayashi, K., et al. (2006). Granulocyte-colony stimulating factor is neuroprotective in a model of Parkinson's disease. J. Neurochem. 97, 675–686.

Miller, T.L., and Wolin, M.J. (1979). Fermentations by saccharolytic intestinal bacteria. Am. J. Clin. Nutr. 32, 164–172.

Minuk, G.Y. (1986). Gamma-aminobutyric acid (GABA) production by eight common bacterial pathogens. Scand. J. Infect. Dis. *18*, 465–467.

Mitchell, R.W., On, N.H., Del Bigio, M.R., Miller, D.W., and Hatch, G.M. (2011). Fatty acid transport protein expression in human brain and potential role in fatty acid transport across human brain microvessel endothelial cells. J. Neurochem. *117*, 735–746.

Montiel-Castro, Augusto, J., Báez-Yáñez, Mario, G., and Pacheco-López, G. (2014). Social neuroeconomics: the influence of microbiota in partner-choice and sociality. Curr. Pharm. Des. 20, 4774–4783.

Mulle, J.G., Sharp, W.G., and Cubells, J.F. (2013). The gut microbiome: a new frontier in autism research. Curr. Psychiatry Rep. 15, 337.

Neufeld, K.M., Kang, N., Bienenstock, J., and Foster, J.A. (2011). Reduced anxiety-like behavior and central neurochemical change in germ-free mice. Neurogastroenterol. Motil. *23*, 255–264.

O'Mahony, S.M., Clarke, G., Borre, Y.E., Dinan, T.G., and Cryan, J.F. (2015). Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. Behav. Brain Res. 277, 32–48.

Ochoa-Repáraz, J., Mielcarz, D.W., Ditrio, L.E., Burroughs, A.R., Begum-Haque, S., Dasgupta, S., Kasper, D.L., and Kasper, L.H. (2010). Central nervous system demyelinating disease protection by the human commensal Bacteroides fragilis depends on polysaccharide A expression. J. Immunol. *185*, 4101–4108.

Ozogul, F., Kuley, E., Ozogul, Y., and Ozogul, I. (2012). The Function of Lactic Acid Bacteria on Biogenic Amines Production by Food-Borne Pathogens in Arginine Decarboxylase Broth. Food Sci. Technol. Res. *18*, 795–804.

Paredes, R.G., and Agmo, A. (1992). GABA and behavior: the role of receptor subtypes. Neurosci. Biobehav. Rev. *16*, 145–170.

PrabhuDas, M., Bonney, E., Caron, K., Dey, S., Erlebacher, A., Fazleabas, A., Fisher, S., Golos, T., Matzuk, M., McCune, J.M., et al. (2015). Immune mechanisms at the maternal-fetal interface: perspectives and challenges. Nat. Immunol. *16*, 328–334.

Prakash, A., Medhi, B., and Chopra, K. (2013). Granulocyte colony stimulating factor (GCSF) improves memory and neurobehavior in an amyloid- β induced experimental model of Alzheimer's disease. Pharmacol. Biochem. Behav. *110*, 46–57.

Pyndt Jørgensen, B., Hansen, J.T., Krych, L., Larsen, C., Klein, A.B., Nielsen, D.S., Josefsen, K., Hansen, A.K., and Sørensen, D.B. (2014). A possible link between food and mood: dietary impact on gut microbiota and behavior in BALB/c mice. PLoS ONE 9, e103398.

Raison, C.L., Capuron, L., and Miller, A.H. (2006). Cytokines sing the blues: inflammation and the pathogenesis of depression. Trends Immunol. 27, 24–31.

Reigstad, C.S., Salmonson, C.E., Rainey, J.F., 3rd, Szurszewski, J.H., Linden, D.R., Sonnenburg, J.L., Farrugia, G., and Kashyap, P.C. (2014). Gut microbes promote colonic serotonin production through an effect of short-chain fatty acids on enterochromaffin cells. FASEB J. *29*, 1395–1403.

Rosenberg, E., Sharon, G., Atad, I., and Zilber-Rosenberg, I. (2010). The evolution of animals and plants via symbiosis with microorganisms. Environ. Microbiol. Rep. 2, 500–506.

Rosenfeld, C.S. (2015). Microbiome Disturbances and Autism Spectrum Disorders. Drug Metab. Dispos., in press. http://dx.doi.org/10.1124/dmd.115. 063826.

Round, J.L., and Mazmanian, S.K. (2009). The gut microbiota shapes intestinal immune responses during health and disease. Nat. Rev. Immunol. 9, 313–323.

Round, J.L., and Mazmanian, S.K. (2010). Inducible Foxp3+ regulatory T-cell development by a commensal bacterium of the intestinal microbiota. Proc. Natl. Acad. Sci. USA *107*, 12204–12209.

Savignac, H.M., Kiely, B., Dinan, T.G., and Cryan, J.F. (2014). Bifidobacteria exert strain-specific effects on stress-related behavior and physiology in BALB/c mice. Neurogastroenterol. Motil. *26*, 1615–1627.

Savignac, H.M., Tramullas, M., Kiely, B., Dinan, T.G., and Cryan, J.F. (2015). Bifidobacteria modulate cognitive processes in an anxious mouse strain. Behav. Brain Res. 287, 59–72.

Selkrig, J., Wong, P., Zhang, X., and Pettersson, S. (2014). Metabolic tinkering by the gut microbiome: Implications for brain development and function. Gut Microbes *5*, 369–380.

Sharon, G., Segal, D., Ringo, J.M., Hefetz, A., Zilber-Rosenberg, I., and Rosenberg, E. (2010). Commensal bacteria play a role in mating preference of Drosophila melanogaster. Proc. Natl. Acad. Sci. USA *107*, 20051–20056.

Sharon, G., Garg, N., Debelius, J., Knight, R., Dorrestein, P.C., and Mazmanian, S.K. (2014). Specialized metabolites from the microbiome in health and disease. Cell Metab. *20*, 719–730.

Shyu, W.C., Lin, S.Z., Yang, H.I., Tzeng, Y.S., Pang, C.Y., Yen, P.S., and Li, H. (2004). Functional recovery of stroke rats induced by granulocyte colony-stimulating factor-stimulated stem cells. Circulation *110*, 1847–1854.

Sobrino Crespo, C., Perianes Cachero, A., Puebla Jimenez, L., Barrios, V., and Arilla Ferreiro, E. (2014). Peptides and food intake. Front. Endocrinol. (Lausanne) *5*, 58.

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Steenbergen, L., Sellaro, R., van Hemert, S., Bosch, J.A., and Colzato, L.S. (2015). A randomized controlled trial to test the effect of multispecies probiotics on cognitive reactivity to sad mood. Brain Behav. Immun., in press. http://dx.doi.org/10.1016/j.bbi.2015.04.003.

Stilling, R.M., Bordenstein, S.R., Dinan, T.G., and Cryan, J.F. (2014a). Friends with social benefits: host-microbe interactions as a driver of brain evolution and development? Front. Cell. Infect. Microbiol. *4*, 147.

Stilling, R.M., Dinan, T.G., and Cryan, J.F. (2014b). Microbial genes, brain & behaviour - epigenetic regulation of the gut-brain axis. Genes Brain Behav. *13*, 69–86.

Sudo, N., Chida, Y., Aiba, Y., Sonoda, J., Oyama, N., Yu, X.N., Kubo, C., and Koga, Y. (2004). Postnatal microbial colonization programs the hypothalamicpituitary-adrenal system for stress response in mice. J. Physiol. 558, 263–275.

Theis, K.R., Schmidt, T.M., and Holekamp, K.E. (2012). Evidence for a bacterial mechanism for group-specific social odors among hyenas. Sci. Rep. 2, 615.

Theis, K.R., Venkataraman, A., Dycus, J.A., Koonter, K.D., Schmitt-Matzen, E.N., Wagner, A.P., Holekamp, K.E., and Schmidt, T.M. (2013). Symbiotic bacteria appear to mediate hyena social odors. Proc. Natl. Acad. Sci. USA *110*, 19832–19837.

Tillisch, K., Labus, J., Kilpatrick, L., Jiang, Z., Stains, J., Ebrat, B., Guyonnet, D., Legrain-Raspaud, S., Trotin, B., Naliboff, B., et al. (2013). Consumption of fermented milk product with probiotic modulates brain activity. Gastroenterology 144, 1394–1401.

van Dam, A.M., Brouns, M., Louisse, S., and Berkenbosch, F. (1992). Appearance of interleukin-1 in macrophages and in ramified microglia in the brain of endotoxin-treated rats: a pathway for the induction of non-specific symptoms of sickness? Brain Res. 588, 291–296.

Vandvik, P.O., Wilhelmsen, I., Ihlebaek, C., and Farup, P.G. (2004). Comorbidity of irritable bowel syndrome in general practice: a striking feature with clinical implications. Aliment. Pharmacol. Ther. *20*, 1195–1203.

Velagapudi, V.R., Hezaveh, R., Reigstad, C.S., Gopalacharyulu, P., Yetukuri, L., Islam, S., Felin, J., Perkins, R., Borén, J., Oresic, M., and Bäckhed, F.

(2010). The gut microbiota modulates host energy and lipid metabolism in mice. J. Lipid Res. 51, 1101–1112.

Vetulani, J. (2013). Early maternal separation: a rodent model of depression and a prevailing human condition. Pharmacol. Rep. 65, 1451–1461.

Vijay-Kumar, M., Aitken, J.D., Carvalho, F.A., Cullender, T.C., Mwangi, S., Srinivasan, S., Sitaraman, S.V., Knight, R., Ley, R.E., and Gewirtz, A.T. (2010). Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. Science 328, 228–231.

Wall, R., Cryan, J.F., Ross, R.P., Fitzgerald, G.F., Dinan, T.G., and Stanton, C. (2014). Bacterial neuroactive compounds produced by psychobiotics. Adv. Exp. Med. Biol. *817*, 221–239.

Wikoff, W.R., Anfora, A.T., Liu, J., Schultz, P.G., Lesley, S.A., Peters, E.C., and Siuzdak, G. (2009). Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. Proc. Natl. Acad. Sci. USA *106*, 3698–3703.

Williams, B.B., Van Benschoten, A.H., Cimermancic, P., Donia, M.S., Zimmermann, M., Taketani, M., Ishihara, A., Kashyap, P.C., Fraser, J.S., and Fischbach, M.A. (2014). Discovery and characterization of gut microbiota decarboxylases that can produce the neurotransmitter tryptamine. Cell Host Microbe *16*, 495–503.

Yano, J.M., Yu, K., Donaldson, G.P., Shastri, G.G., Ann, P., Ma, L., Nagler, C.R., Ismagilov, R.F., Mazmanian, S.K., and Hsiao, E.Y. (2015). Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. Cell *161*, 264–276.

Zhao, L.R., Navalitloha, Y., Singhal, S., Mehta, J., Piao, C.S., Guo, W.P., Kessler, J.A., and Groothuis, D.R. (2007). Hematopoietic growth factors pass through the blood-brain barrier in intact rats. Exp. Neurol. *204*, 569–573.

Zhu, C.B., Blakely, R.D., and Hewlett, W.A. (2006). The proinflammatory cytokines interleukin-1beta and tumor necrosis factor-alpha activate serotonin transporters. Neuropsychopharmacology *31*, 2121–2131.

Zucchi, R., Chiellini, G., Scanlan, T.S., and Grandy, D.K. (2006). Trace amineassociated receptors and their ligands. Br. J. Pharmacol. *149*, 967–978.