Contents lists available at SciVerse ScienceDirect



Journal of Oral Biosciences



journal homepage: www.elsevier.com/locate/job

Review Recent topics in fibrodysplasia ossificans progressiva

Takenobu Katagiri^{a,b,*}

^a Division of Pathophysiology, Research Center for Genomic Medicine, Saitama Medical University, 1397-1 Yamane, Hidaka-shi, Saitama 350-1241, Japan ^b Project of Clinical and Basic Research for FOP at Saitama Medical University, 1397-1 Yamane, Hidaka-shi, Saitama 350-1241, Japan

ARTICLE INFO

Article history: Received 27 February 2012 Received in revised form 20 March 2012 Accepted 21 March 2012 Available online 17 August 2012

Keywords: Bone morphogenetic protein Receptor Genetic disorder Treatment

ABSTRACT

Fibrodysplasia ossificans progressiva (FOP) is a rare autosomal dominant disorder characterized by progressive heterotopic bone formation in skeletal muscle tissue. Patients with FOP show malformed digits, osteochondroma, and other skeletal abnormalities due to abnormal patterning during development. Heterozygous mutations in the Activin A receptor type I (*ACVR1*) gene, which encodes the bone morphogenetic protein (BMP) type I receptor ALK2, have been identified in not only typical FOP patients but also patients with unusually mild or severe clinical features. The serine/threonine kinase activity of ALK2 may be constitutively activated by mutations in the GS domain or the kinase domain. Based on these findings, selective small chemical inhibitors and allele-specific RNAi approaches for mutant ALK2 have been developed for preventing heterotopic bone formation in FOP. Other novel treatments have also been reported to block heterotopic bone formation in patients with FOP. These findings open the door to the next step in FOP treatment and related research.

© 2012 Japanese Association for Oral Biology. Published by Elsevier B.V. All rights reserved.

Contents

1.	Introduction	119
2.	Typical features of FOP	120
3.	Variations in FOP	120
4.	Development of treatments for preventing heterotopic bone formation	121
5.	Treatments preventing heterotopic bone formation in FOP	121
	Conclusions	
	Conflict of interest	122
	Acknowledgments	122
	References	122

1. Introduction

Fibrodysplasia ossificans progressiva (FOP) is a rare autosomal dominant disorder that causes progressive heterotopic bone formation in skeletal muscle tissue [1–3]. A heterozygous substitution of "G" with "A" in the Activin A receptor type I (ACVR1) gene at position 617 (c.617 G > A) has been identified in both familial and sporadic cases of FOP [4]. The ACVR1 gene encodes ALK2, a type I receptor for bone morphogenetic proteins (BMPs), which are potent bone growth-inducing factors in mammals [3]. BMPs, but not the

related transforming growth factor- β (TGF- β) or activin, are able to induce heterotopic bone formation in skeletal muscle tissue [3,5]. The differences in the biological activities of BMPs and other members of the TGF- β family are explained by the specific intracellular signaling pathways that are activated through cell surface BMP receptors.

The members of the TGF- β family bind to type I and type II receptors on their target cells [5] (Fig. 1). Both types of receptors are transmembrane serine/threonine kinases. Type I and type II receptors recognize different types of substrates and are classified on the basis of their structures. Type I receptors have a conserved "GS" domain, which is a glycine- and serine-rich domain in the intracellular region. These receptors are inactive kinases until bound to a ligand. Type II receptors lack the GS domain and are constitutive active kinases that phosphorylate the GS domain of

^{*} Correspondence address: Division of Pathophysiology, Research Center for Genomic Medicine, Saitama Medical University, 1397-1 Yamane, Hidaka-shi, Saitama 350-1241, Japan. Tel./fax: +81 42 984 0443.

E-mail address: katagiri@saitama-med.ac.jp

^{1349-0079/\$-}see front matter © 2012 Japanese Association for Oral Biology. Published by Elsevier B.V. All rights reserved. http://dx.doi.org/10.1016/j.job.2012.03.004

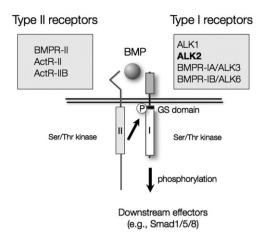


Fig. 1. BMP and its receptors. BMPs bind to type I and type II receptors, which are transmembrane serine/threonine kinases that are expressed on the cell membrane. The type II receptors (BMPR-II, ActR-II, and ActR-IIB) phosphorylate the GS domain of the type I receptors (ALK1, ALK2, BMPR-IA/ALK3, and BMPR-IB/ALK6). The activated type I receptors subsequently phosphorylate downstream effectors, such as Smad1/5/8, in the cytoplasm.

the type I receptors in the ternary complex that is formed on ligand binding. BMP signaling is transduced by 4 types of type I receptors (ALK1, ALK2, BMPR-IA/ALK3, and BMPR-IB/ALK6) and 3 types of type II receptors (BMPR-II, ActR-II, and ActR-IIB) (Fig. 1). The phosphorylation of the GS domain activates the kinase activity of type I receptors by changing their threedimensional structures. Thus, the GS domain is considered the molecular switch of the type I receptor.

The c.617 G > A mutation in the *ACVR1* gene, which was first identified in patients with FOP, causes an arginine-to-histidine substitution at position 206 (p.R206H) in ALK2 [4]. The p.R206H mutation is located in the GS domain and is expected to change the kinase activity of ALK2 in patients with FOP [4]. The over-expression of the mutant ALK2 p.R206H induces a set of BMP signaling cascades in cultured cells [6,7]. Therefore, FOP is considered the first example of a genetic condition caused by the gain of function of BMP receptors in mammals. Recently, genetic diagnoses have identified variants of FOP that have different clinical features and are caused by novel mutations at different positions in ALK2. In addition, some novel potential treatments have been found to block or reduce the heterotopic bone formation induced by the mutant ALK2 in a mouse model of FOP and in patients with FOP.

2. Typical features of FOP

The p.R206H mutation in ALK2 was the first identified substitution in FOP. This typical mutation is found not only in sporadic cases but also in familial cases of FOP in several different populations, including African-American, European American, European (UK), Korean, native Brazilian, and Japanese patients [4,7,8]. FOP is transmitted in an autosomal dominant fashion. In early studies, mutations in a 36 cM region at chromosome 4q27– 31 and in the *NOG* gene (encoding the BMP antagonist noggin) were reported as potentially responsible for FOP, but the *ACVR1* gene was mapped to chromosome 2q23–24 [4,9,10]. Causative mutations in the *NOG* gene have since been excluded by linkage and mutational analyses [11]. The overexpression of BMP-4 in lymphocytes has been reported in FOP patients, suggesting that BMP-4 is one of the downstream target genes of BMP signaling in a positive feedback loop [12]. Most patients with the p.R206H mutation exhibit a malformation of the big toes (hallux), i.e., hallux valgus, as one of the typical phenotypes that can be recognized at birth before heterotopic bone formation begins. In 16 Japanese patients, 29 out of 31 feet (93.5%) showed hallux deformities at various degrees [13]. Patients with the p.R206H mutation also showed osteochondroma of the proximal tibia and other skeletal malformations due to disorganized patterning during development [1].

Heterotopic bone formation in FOP patients can be induced by local skeletal muscle injury, including intramuscular immunization, biopsy, and surgical operation [1]. Influenza-like illnesses are also known to induce heterotopic bone formation in FOP patients. However, heterotopic bone formation is not observed in the diaphragm, tongue, extra-ocular muscles, cardiac muscle, or smooth muscles of patients with FOP [1–3]. The molecular-level differences between the bone-inducible and bone-uninducible muscles are still unknown.

FKBP12 binds to the immunosuppressive drug FK506 and has been suggested to be involved in the suppression of intracellular signaling via type I TGF- β and BMP family receptors [14,15]. It binds to the GS domain of unbound receptors and is released through the phosphorylation of the GS domain by type II receptors in response to ligand binding [15]. The p.R206H mutation has been shown to reduce the affinity of ALK2 for FKBP12, in the absence of ligand binding, in a pH-dependent manner [16].

3. Variations in FOP

After the identification of the p.R206H mutation in ALK2 in 2006, additional genetic mutations have been found in the coding region of *ACVR1* in patients who show heterotopic bone formation and other skeletal abnormalities (Fig. 2). In the GS domain of ALK2, 4 mutations have been found in addition to p.R206H: p.L196P, p.P198-F199del_insL, p.R202I, and p.Q207E [17–19]. Moreover, several mutations have been found in the serine/threonine kinase domain of ALK2, such as p.R258S, p.G325A, p.G328E/R/W, p.G356D, and p.R375P [18–23]. Some of the mutations in the kinase domain have been suggested to be exposed at the interface with the GS domain and to be involved in the interaction with FKBP12 [19,20].

The clinical features in patients with these mutations in ALK2, except p.R258S, are different from those in patients with the

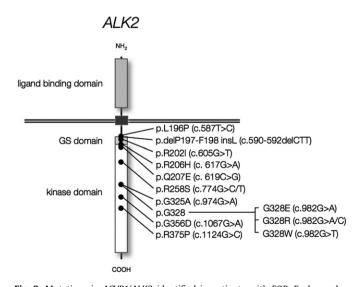


Fig. 2. Mutations in ACVR1/ALK2 identified in patients with FOP. Each number indicates the position of a mutation detected in the cDNA of the ACVR1 gene (c) or in the ALK2 protein (p). Note that all of the mutations are localized to the intracellular domain of ALK2.

p.R206H mutation. Four independent patients with the p.G356D mutation exhibited shortening/truncation malformations of the thumbs and halluces [18,23]. Severe malformations in the thumbs and halluces were observed in other patients who had the p.G328W and p.G328E mutations [18,22]. The tight link between heterotopic bone formation, the mutations in ALK2, and the malformations of the digits suggests that the fine-tuning of BMP signaling is required for the normal development of the skeletal muscle, thumbs, and halluces. Indeed, it was reported that the transient activation of BMP signaling is essential for normal muscle regeneration in adult mice [24]. Moreover, deletion of the *Bmp-4* and/or *Bmp-7* gene causes skeletal abnormalities, including polydactyly, in newborn mice [25].

In contrast to the severe digital phenotypes seen in FOP patients, some patients who have other mutations in ALK2 have milder phenotypes, especially with respect to heterotopic bone formation in the muscle. One patient did not have hallux malformations at birth but developed trismus and heterotopic bone formation in the thigh muscle after a motorbike accident at the age of 21 years [17]. The p.L196P mutation was identified in this patient by sequencing [17]. Modeling of p.L196P in silico revealed a steric clash with the kinase domain that is predicted to weaken its interactions with FKBP12 [17]. The overexpression of the p.L196P mutant in C2C12 myoblasts induced BMP signaling equivalent to that of the p.R206H mutant, at least in vitro, suggesting that the p.L196P mutation activates ALK2. However, ALK2 activity may have been suppressed in vivo (i.e., in the patient) by some unknown molecular mechanism prior to the motorbike accident [26].

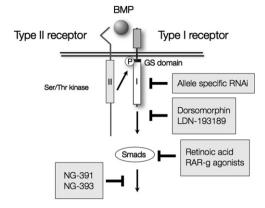
Recently, another novel mutation in ALK2 was identified in a patient with congenital hallux valgus. In this patient, heterotopic bone formation was induced by a viral illness at age of 47 years [27]. Analysis of the *ACVR1* gene revealed heterozygosity of a c974G > C mutation that causes a p.G325A substitution in ALK2 [27]. Another FOP patient, who had normal toes and showed slow progression of heterotopic bone formation, had a c.1124 G > C mutation that caused a p.R375P substitution [18]. Together with the p.L196P case, these FOP cases with late-onset heterotopic bone formation suggest the importance of the genetic diagnosis of FOP even in patients who do not show typical clinical features.

4. Development of treatments for preventing heterotopic bone formation

Although there is no effective treatment for preventing heterotopic bone formation in FOP, several trials of new treatments have been initiated based on the ALK2 findings (Fig. 3).

Because most of the ALK2 mutations found in FOP patients represent activated forms of the BMP receptor, a specific inhibitor of this receptor would prevent the intracellular signaling induced by the mutant ALK2. LDN-193189 was developed on the basis of structure of dorsomorphin, which inhibits the phosphorylation of Smad1/5/8 but not p38 by BMP type I receptors. Treatment with LDN-193189 reduced heterotopic bone formation in mice carrying a p.Q207D mutation in ALK2 [28]; it should be noted that this mutation has not been identified in FOP patients. Recently, two small compounds produced by fungi, NG-391 and NG-393, were shown to inhibit BMP signaling by mutant ALK2 (p.R206H) *in vitro* [29].

Recently, retinoic acid and related chemical compounds were found to be potent inhibitors of BMP signaling induced heterotopic bone formation in the skeletal muscle [30]. Because BMPs induce heterotopic bone formation via endochondral ossification, retinoic acid was examined as an inhibitor of chondrogenesis *in vitro* and *in vivo*. The inhibitory activity of retinoic acid is



Heterotopic bone formation

Fig. 3. Treatments for preventing heterotopic bone formation in FOP. Treatments developed for preventing heterotopic bone formation are indicated in this schematic of BMP signal transduction. The details are explained in the text.

mediated via RAR- γ but not RAR- α or RAR- β [30]. A synthetic selective agonist of RAR- γ inhibited heterotopic bone formation induced by BMP-2 in skeletal muscle [30]. Other types of RAR- γ agonists also had similar effects on heterotopic bone formation. Among them, CD1530 blocked FOP-like heterotopic bone formation in mutant mice carrying the ALK2 p.Q207D mutation [30]. CD1530 has been suggested to inhibit BMP signaling by reducing SMAD protein levels [30].

Chemical inhibitors of BMP receptors may suppress the intracellular signaling induced by both wild-type and mutant ALK2. To develop a specific inhibitor of mutant ALK2, allele-specific RNAi has been tested. Double-stranded small RNAs with simple mutations targeting the p.R206H-coding transcript were not allele specific and inhibited both the mutant and wild-type *ACVR1* transcripts *in vitro* [31]. However, the introduction of a single additional nucleotide mutation in the RNA generated an allelespecific dsRNA that suppressed the expression of only the mutant ALK2 [31]. This allele-specific inhibition by RNAi was confirmed to suppress expression of both the p.R206H and p.G356D mutant proteins [31]. Thus, allele-specific RNAi may represent a novel type of treatment for FOP.

5. Treatments preventing heterotopic bone formation in FOP

There has been one case report of FOP that described the prevention of heterotopic bone formation for 14 years [32]. The patient, who had a c.617 G > A mutation, received bone marrow transplantation for the treatment of intercurrent aplastic anemia. No heterotopic bone formation occurred during the 14 years during which the patient received immunosuppressive drug treatment (prednisone, cyclosporine, and methotrexate) after the bone marrow transplantation [32]. However, heterotopic bone formation resumed when the patient discontinued these medications. This resumption of heterotopic bone formation indicates that bone marrow transplantation is not an effective treatment for FOP in spite of the fact that BMP-4 overexpression in lymphocytes has been reported previously to underlie heterotopic bone formation. This case also suggests that the immune system may be involved in the induction of bone formation in FOP patients.

Recently, it was reported that a combination of rosiglitazone and low doses of prednisone effectively prevented heterotopic bone formation in a patient with FOP [33]. The suggested molecular mechanism of this action is that rosiglitazone induces the activation of PPAR- γ , which promotes the differentiation of bone marrow mesenchymal stem cells into adipocytes rather than into osteoblasts. These two case reports suggest that treatments that do not directly block BMP signaling may still effectively prevent heterotopic bone formation in FOP patients.

A recent finding describing a role for vascular cells suggests an alternative treatment strategy [34]. Endothelial cells express some cell surface-specific markers, such as Von Willebrand factor (vWF) and TIE-2. Immunohistochemical staining of the heterotopic bones formed in FOP patients showed that both chondrocytes and osteoblasts, which were identified by staining with anti-Sox9 and anti-osteocalcin respectively, were co-stained with vWF and Tie-2. In normal bone tissues, these endothelial markers were not detected in either chondrocytes or osteoblasts, suggesting that endothelial cells transdifferentiate into mesenchymal cells in FOP patients. Indeed, the over-expression of ALK2 p.R206H, but not wild-type ALK2, induced the mesenchymal transdifferentiation of cultured endothelial cells into osteoblasts, chondrocytes, and adipocytes [34]. Moreover, the treatment of endothelial cells with BMP-4 and TGF-β2 did not induce heterotopic bone formation in vivo but did induce the same changes in vitro [34]. These findings suggest that the conversion of endothelial cells into mesenchymal cells is a critical molecular mechanism underlying heterotopic bone formation in FOP patients.

6. Conclusions

All patients with FOP exhibit mutations in the GS domain or kinase domain of ALK2, a type I BMP receptor. Although most FOP patients share many common clinical features, several novel phenotypic variations that characterize FOP have also been found. The genetic diagnosis of FOP through the sequencing of the *ACVR1* gene is useful. The development of specific inhibitors of ALK2 and other types of treatment will be useful to prevent heterotopic bone formation in FOP patients.

Conflict of interest

Takenobu Katagiri received a grant-in-aid from the Takeda Science Foundation; a grant-in-aid from Daiichi-Sankyo Co., Ltd. Takenobu Katagiri was awarded the Lion Dental Research Award from the Japanese Association for Dental Biosciences (JAOB) supported by Lion Corporation.

Acknowledgments

I thank the members of the Division of Pathophysiology, Research Center for Genomic Medicine, Saitama Medical University, members of the Project of Clinical and Basic Research for FOP at Saitama Medical University, and members of the Research Committee on FOP of the Japanese Ministry of Health, Labour, and Welfare for their valuable comments and discussion. This work was supported in part by the Health and Labor Sciences Research Grants for Research on Measures for Intractable Research from the Ministry of Health, Labour, and Welfare of Japan; a grant-in-aid from the Ministry of Education, Culture, Sports, Science, and Technology of Japan; a grant-in-aid from the Support Project for the Formation of a Strategic Center in a Private University from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

References

 Kitterman JA, Kantanie S, Rocke DM, Kaplan FS. latrogenic harm caused by diagnostic errors in fibrodysplasia ossificans progressiva. Pediatrics 2005;116: e654–61.

- [2] Shore EM, Kaplan FS. Insights from a rare genetic disorder of extra-skeletal bone formation, fibrodysplasia ossificans progressiva (FOP). Bone 2008;43: 427–33.
- [3] Katagiri T. Heterotopic bone formation induced by bone morphogenetic protein signaling: fibrodysplasia ossificans progressiva. J Oral Biosci 2010;52:33-41.
- [4] Shore EM, Xu M, Feldman GJ, Fenstermacher DA, Cho TJ, Choi IH, Connor JM, Delai P, Glaser DL, LeMerrer M, Morhart R, Rogers JG, Smith R, Triffitt JT, Urtizberea JA, Zasloff M, Brown MA, Kaplan FS. A recurrent mutation in the BMP type I receptor ACVR1 causes inherited and sporadic fibrodysplasia ossificans progressiva. Nat Genet 2006;38:525–7.
- [5] Katagiri T, Suda T, Miyazono K. The bone morphogenetic proteins. In: Miyazono K, Derynck R, editors. The TGF-beta family. New York: Cold Spring Harbor; 2008. p. 121–49.
- [6] Fukuda T, Kanomata K, Nojima J, Kokabu S, Akita M, Ikebuchi K, Jimi E, Komori T, Maruki Y, Matsuoka M, Miyazono K, Nakayama K, Nanba A, Tomoda H, Okazaki Y, Ohtake A, Oda H, Owan I, Yoda T, Haga N, Furuya H, Katagiri T. A unique mutation of ALK2, G356D, found in a patient with fibrodysplasia ossificans progressiva is a moderately activated BMP type I receptor. Biochem Biophys Res Commun 2008;377:905–9.
- [7] Fukuda T, Kohda M, Kanomata K, Nojima J, Nakamura A, Kamizono J, Noguchi Y, Iwakiri K, Kondo T, Kurose J, Endo K, Awakura T, Fukushi J, Nakashima Y, Chiyonobu T, Kawara A, Nishida Y, Wada I, Akita M, Komori T, Nakayama K, Nanba A, Maruki Y, Yoda T, Tomoda H, Yu PB, Shore EM, Kaplan FS, Miyazono K, Matsuoka M, Ikebuchi K, Ohtake A, Oda H, Jimi E, Owan I, Okazaki Y, Katagiri T. Constitutively activated ALK2 and increased SMAD1/5 cooperatively induce bone morphogenetic protein signaling in fibrodysplasia ossificans progressiva. J Biol Chem 2009;284:7149–56.
- [8] Nakajima M, Haga N, Takikawa K, Manabe N, Nishimura G, Ikegawa S. The ACVR1 617 G > A mutation is also recurrent in three Japanese patients with fibrodysplasia ossificans progressiva. J Hum Genet 2007;52:473–5.
- [9] Feldman G, Li M, Martin S, Urbanek M, Urtizberea JA, Fardeau M, LeMerrer M, Connor JM, Triffitt J, Smith R, Muenke M, Kaplan FS, Shore EM. Fibrodysplasia ossificans progressiva, a heritable disorder of severe heterotopic ossification, maps to human chromosome 4q27-31. Am J Hum Genet 2000;66:128–35.
- [10] Lucotte G, Semonin O, Lutz P. A de novo heterozygous deletion of 42 basepairs in the noggin gene of a fibrodysplasia ossificans progressiva patient. Clin Genet 1999;56:469–70.
- [11] Xu MQ, Feldman G, Le Merrer M, Shugart YY, Glaser DL, Urtizberea JA, Fardeau M, Connor JM, Triffitt J, Smith R, Shore EM, Kaplan FS. Linkage exclusion and mutational analysis of the noggin gene in patients with fibrodysplasia ossificans progressiva (FOP). Clin Genet 2000;58:291–8.
- [12] Feldman GJ, Billings PC, Patel RV, Caron RJ, Guenther C, Kingsley DM, Kaplan FS, Shore EM. Over-expression of BMP4 and BMP5 in a child with axial skeletal malformations and heterotopic ossification: a new syndrome. Am J Med Genet A 2007;143:699–706.
- [13] Nakashima Y, Haga N, Kitoh H, Kamizono J, Tozawa K, Katagiri T, Susami T, Fukushi J, Iwamoto Y. Deformity of the great toe in fibrodysplasia ossificans progressiva. J Orthop Sci 2010;15:804–9.
- [14] Kugimiya F, Yano F, Ohba S, Igawa K, Nakamura K, Kawaguchi H, Chung UI. Mechanism of osteogenic induction by FK506 via BMP/Smad pathways. Biochem Biophys Res Commun 2005;338:872–9.
- [15] Nishanian TG, Waldman T. Interaction of the BMPR-IA tumor suppressor with a developmentally relevant splicing factor. Biochem Biophys Res Commun 2004;323:91–7.
- [16] Groppe JC, Wu J, Shore EM, Kaplan FS. In vitro analyses of the dysregulated R206H ALK2 kinase-FKBP12 interaction associated with heterotopic ossification in FOP. Cells Tissues Organs 2011;194:291–5.
- [17] Gregson CL, Hollingworth P, Williams M, Petrie KA, Bullock AN, Brown MA, Tobias JH, Triffitt JT. A novel ACVR1 mutation in the glycine/serine-rich domain found in the most benign case of a fibrodysplasia ossificans progressiva variant reported to date. Bone 2011;48:654–8.
- [18] Kaplan FS, Xu M, Seemann P, Connor JM, Glaser DL, Carroll L, Delai P, Fastnacht-Urban E, Forman SJ, Gillessen-Kaesbach G, Hoover-Fong J, Koster B, Pauli RM, Reardon W, Zaidi SA, Zasloff M, Morhart R, Mundlos S, Groppe J, Shore EM. Classic and atypical fibrodysplasia ossificans progressiva (FOP) phenotypes are caused by mutations in the bone morphogenetic protein (BMP) type I receptor ACVR1. Hum Mutat 2009;30:379–90.
- [19] Petrie KA, Lee WH, Bullock AN, Pointon JJ, Smith R, Russell RG, Brown MA, Wordsworth BP, Triffitt JT. Novel mutations in ACVR1 result in atypical features in two fibrodysplasia ossificans progressiva patients. PloS One 2009;4:e5005.
- [20] Bocciardi R, Bordo D, Di Duca M, Di Rocco M, Ravazzolo R. Mutational analysis of the ACVR1 gene in Italian patients affected with fibrodysplasia ossificans progressiva: confirmations and advancements. Eur J Hum Genet 2009;17:311–8.
- [21] Ratbi I, Borcciadi R, Regragui A, Ravazzolo R, Sefiani A. Rarely occurring mutation of ACVR1 gene in Moroccan patient with fibrodysplasia ossificans progressiva. Clin Rheumatol 2012;29:119–121.
- [22] Carvalho DR, Navarro MM, Martins BJ, Coelho KE, Mello WD, Takata RI, Speck-Martins CE. Mutational screening of ACVR1 gene in Brazilian fibrodysplasia ossificans progressiva patients. Clin Genet 2010;77:171–6.
- [23] Furuya H, Ikezoe K, Wang L, Ohyagi Y, Motomura K, Fujii N, Kira J, Fukumaki Y. A unique case of fibrodysplasia ossificans progressiva with an ACVR1 mutation, G356D, other than the common mutation (R206H). Am J Med Genet A 2008;146A:459–63.

- [24] Ono Y, Calhabeu F, Morgan JE, Katagiri T, Amthor H, Zammit PS. BMP signalling permits population expansion by preventing premature myogenic differentiation in muscle satellite cells. Cell Death Differ 2011;18:222–34.
- [25] Katagiri T, Boorla S, Frendo JL, Hogan BL, Karsenty G. Skeletal abnormalities in doubly heterozygous Bmp4 and Bmp7 mice. Dev Genet 1998;22:340–8.
- [26] Ohte S, Shin M, Sasanuma H, Yoneyama K, Akita M, Ikebuchi K, Jimi E, Maruki Y, Matsuoka M, Namba A, Tomoda H, Okazaki Y, Ohtake A, Oda H, Owan I, Yoda T, Furuya H, Kamizono J, Kitoh H, Nakashima Y, Susami T, Haga N, Komori T, Katagiri T. A novel mutation of ALK2, L196P, found in the most benign case of fibrodysplasia ossificans progressiva activates BMP-specific intracellular signaling equivalent to a typical mutation, R206H. Biochem Biophys Res Commun 2011;407:213–8.
- [27] Whyte MP, Wenkert D, Demertzis JL, Dicarlo EF, Westenberg E, Mumm S. Fibrodysplasia ossificans progressiva: Middle-age onset of heterotopic ossification from a unique missense mutation (c.974G > C, p.G325A) in ACVR1. J Bone Miner Res. 2012;27:729–37.
- [28] Yu PB, Deng DY, Lai CS, Hong CC, Cuny GD, Bouxsein ML, Hong DW, McManus PM, Katagiri T, Sachidanandan C, Kamiya N, Fukuda T, Mishina Y, Peterson RT, Bloch KD. BMP type I receptor inhibition reduces heterotopic [corrected] ossification. Nat Med 2008;14:1363–9.

- [29] Fukuda T, Uchida R, Inoue H, Ohte S, Yamazaki H, Matsuda D, Katagiri T, Tomoda H. Fungal pyrrolidine-containing metabolites inhibit alkaline phosphatase activity in bone morphogenetic protein-stimulated myoblastoma cells. Acta Pharm Sin B. 2012;2:23–7.
- [30] Shimono K, Tung WE, Macolino C, Chi AH, Didizian JH, Mundy C, Chandraratna RA, Mishina Y, Enomoto-Iwamoto M, Pacifici M, Iwamoto M. Potent inhibition of heterotopic ossification by nuclear retinoic acid receptorgamma agonists. Nat Med 2011;17:454–60.
- [31] Takahashi M, Katagiri T, Furuya H, Hohjoh H. Disease-causing allele-specific silencing against the ALK2 mutants, R206H and G356D, in fibrodysplasia ossificans progressiva. Gene Ther. 2012;19:781–5.
- [32] Kaplan FS, Glaser DL, Shore EM, Pignolo RJ, Xu M, Zhang Y, Senitzer D, Forman SJ, Emerson SG. Hematopoietic stem-cell contribution to ectopic skeletogenesis. J Bone Joint Surg Am 2007;89:347–57.
- [33] Gatti D, Viapiana O, Rossini M, Silvano A. Rosiglitazone therapy is associated with major clinical improvements in a patient with fibrodysplasia ossificans progressiva. J Bone Miner Res 2011;25:1460–2.
- [34] Medici D, Shore EM, Lounev VY, Kaplan FS, Kalluri R, Olsen BR. Conversion of vascular endothelial cells into multipotent stem-like cells. Nat Med 2010;16:1400–6.