

## HIGH MOLECULAR MASS AMINO ACYL-tRNA SYNTHETASE COMPLEXES IN EUKARYOTES

Chi V. DANG, Deborah L. JOHNSON<sup>†</sup> and David C. H. YANG\*

*The Johns Hopkins University School of Medicine, Baltimore, MD 21205, <sup>†</sup>Department of Molecular Biophysics and Biochemistry, Yale University, New Haven, CT 06520 and \*Department of Chemistry, Georgetown University, Washington, DC 20057, USA*

Received 4 February 1982

### 1. Introduction

Aminoacyl-tRNA synthetases (AARS) are enzymes which play an indispensable role in protein biosynthesis by catalyzing the formation of aminoacyl-tRNA from amino acid, the cognate tRNA, and ATP by highly selective intermolecular interactions [57]. Joachimiak and Barciszewski [41] have provided an extensive compilation of the properties of the aminoacyl-tRNA synthetases; however, information on the eukaryotic high  $M_r$  ( $HM_r$ ) complexes of aminoacyl-tRNA synthetases was lacking. Here, we intend to fill this void by providing a summary of the properties of the eukaryotic aminoacyl-tRNA synthetase complexes.

Eukaryotic aminoacyl-tRNA synthetases may occur as complexes with  $M_r$ -values of  $>10^6$  in contrast to the prokaryotic counterparts which have  $M_r$ -values of  $\leq 250\,000$ . These eukaryotic  $HM_r$ -AARS complexes appear ubiquitous in a wide spectrum of cell types from yeast to human placenta as shown in table 1. Although not all 20 aminoacyl-tRNA synthetases were examined in each case shown in table 1, it appears that the AARS commonly associated with  $M_r$  complexes are those specific for Arg, Gln, Glu, Ile, Leu, Lys and Met. The properties of these  $HM_r$ -AARS complexes are most consistent with multienzyme complexes of aminoacyl-tRNA synthetases [19,20,43, 46]. The physicochemical properties, composition, and stoichiometry of the more rigorously characterized complexes are shown in table 2.

The mechanism(s) of intermolecular interaction between the aminoacyl-tRNA synthetases is not known, but the putative interactions of aminoacyl-tRNA synthetases with a variety of biomolecules have been suggested to play a role in complex formation as shown in table 3. Our present knowledge of the func-

tional significance of  $HM_r$ -AARS is profoundly lacking; however, interactions of the aminoacyl-tRNA synthetases with other components of the protein biosynthetic machinery and other enzymes suggest the intriguing possibility of higher organization of eukaryotic protein biosynthesis. Table 4 is a summary of the possible interactions of the aminoacyl-tRNA synthetases with subcellular components and other enzymes.

This presentation is a brief summary of the properties of the high molecular weight eukaryotic aminoacyl-tRNA synthetase complexes. We hope that this compilation will complement that presented in [41] and will provide useful information for workers in this and other related fields.

### References

- [1] Agris, P. F., Woolverton, D. K. and Setzer, D. (1976) Proc. Natl. Acad. Sci. USA 73, 3857–3861.
- [2] Agris, P. F., Setzer, D. and Gehrk, C. W. (1977) Nucleic Acids Res. 4, 3803–3818.
- [3] Alzhanova, A. T., Fedorov, A. N., Ovchinnikov, L. P. and Spirin, A. S. (1980) FEBS Lett. 120, 225–229.
- [4] Arbeeny, C. M., Briden, K. L. and Stirewalt, W. S. (1979) Biochim. Biophys. Acta 564, 191–201.
- [5] Bandyopadhyay, A. K. and Deutscher, M. P. (1971) J. Mol. Biol. 60, 113–122.
- [6] Bandyopadhyay, A. K. and Deutscher, M. P. (1973) J. Mol. Biol. 74, 257–261.
- [7] Berg, B. H. (1975) Biochim. Biophys. Acta 395, 164–172.
- [8] Berg, B. H. (1975) Biochim. Biophys. Acta 395, 173–178.
- [9] Berg, B. H. (1975) Biochim. Biophys. Acta 414, 93–98.
- [10] Berg, B. H. (1977) Biochim. Biophys. Acta 479, 153–171.
- [11] Berg, B. H. (1978) Biochim. Biophys. Acta 521, 274–287.
- [12] Bont, W. S., Geels, J. and Rezelman, G. (1976) Mol. Biol. Rep. 2, 379–384.

\* To whom correspondences should be addressed

- [13] Brevet, A., Kellermann, O., Tonetti, H. and Waller, J.-P. (1979) *Eur. J. Biochem.* **99**, 551–558.
- [14] Carias, J. R., Mouricout, M., Quintard, B., Thomas, J. C. and Julien, R. (1978) *Eur. J. Biochem.* **87**, 583–590.
- [15] Charezinski, M. and Borkowski, T. (1981) *Arch. Biochem. Biophys.* **207**, 241–247.
- [16] Dang, C. V., Grothusen, J. R., Zimmerman, J. K. and Hilderman, R. H. (1981) Miami Winter Symposium. abst., in press.
- [17] Dang, C. V., Glinski, R. L., Gainey, P. C. and Hilderman, R. H. (1982) *Biochemistry*, in press.
- [18] Dang, C. V. and Yang, D. C. H. (1978) in: *Biomolecular Structure and Function* (Agris, P. F. et al. eds) pp. 575–580, Academic Press, New York.
- [19] Dang, C. V. and Yang, D. C. H. (1978) *Biochem. Biophys. Res. Commun.* **80**, 709–714.
- [20] Dang, C. V. and Yang, D. C. H. (1979) *J. Biol. Chem.* **254**, 5350–5356.
- [21] Denney, R. M. (1977) *Arch. Biochem. Biophys.* **183**, 157–167.
- [22] Deutscher, M. P. (1974) *Methods Enzymol.* **29**, 577–583.
- [23] Diatewa, M. and Stahl, A. J. C. (1981) *Nucleic Acids Res.* **9**, 6293–6304.
- [24] Dignam, J. D., Rhodes, D. G. and Deutscher, M. P. (1980) *Biochemistry* **19**, 4978–4984.
- [25] Dimitrijevic, L. and Godefroy-Colburn, Th. (1974) *FEBS Lett.* **45**, 194–201.
- [26] Dimitrijevic, L. (1977) *FEBS Lett.* **79**, 37–41.
- [27] Enger, M. D., Ritter, P. O. and Hampel, A. E. (1978) *Biochemistry* **17**, 2435–2438.
- [28] Geels, J., Bont, W. S. and Rezelman, G. (1971) *Arch. Biochem. Biophys.* **144**, 773–774.
- [29] Glinski, R. L., Gainey, P. C., Mawhinney, T. P. and Hilderman, R. H. (1979) *Biochem. Biophys. Res. Commun.* **88**, 1052–1061.
- [30] Goto, T. and Schweiger, A. (1973) *Hoppe-Seyler's Z. Physiol. Chem.* **354**, 1027–1033.
- [31] Graf, H. (1976) *Biochim. Biophys. Acta* **425**, 175–181.
- [32] Hampel, A. E. and Enger, M. D. (1973) *J. Mol. Biol.* **79**, 285–293.
- [33] Hampel, A. E., Ritter, P. O. and Enger, M. D. (1978) *Nature* **276**, 844–845.
- [34] Harris, C. L., Marin, K. and Stewart, D. (1977) *Biochem. Biophys. Res. Commun.* **79**, 657–662.
- [35] Hele, P. and Hebert, L. (1977) *Biochim. Biophys. Acta* **479**, 311–321.
- [36] Hradec, J. and Dusek, Z. (1969) *Biochem. J.* **115**, 873–880.
- [37] Hradec, J., Dusek, Z., Bermek, E. and Matthaei, H. (1971) *Biochem. J.* **123**, 959–966.
- [38] Hradec, J. and Dusek, Z. (1980) *Mol. Biol. Rep.* **6**, 245–248.
- [39] Irvin, J. D. and Hardesty, B. (1972) *Biochemistry* **11**, 1915–1920.
- [40] Jakubowski, H. (1979) *FEBS Lett.* **103**, 71–76.
- [41] Joachimiak, A. and Barciszewski, J. (1980) *FEBS Lett.* **119**, 201–211.
- [42] Johnson, D. L., Dang, C. V. and Yang, D. C. H. (1980) *J. Biol. Chem.* **255**, 4362–4366.
- [43] Johnson, D. L. and Yang, D. C. H. (1981) *Proc. Natl. Acad. Sci. USA* **78**, 4059–4062.
- [44] Kellermann, O., Brevet, A., Tonetti, H. and Waller, J.-P. (1978) *Eur. J. Biochem.* **88**, 205–210.
- [45] Kellermann, O., Viel, C. and Waller, J.-P. (1978) *Eur. J. Biochem.* **88**, 197–204.
- [46] Kellermann, O., Brevet, A., Tonetti, H. and Waller, J.-P. (1979) *Eur. J. Biochem.* **99**, 541–550.
- [47] Lamkin, A. F., Smith, D. W. and Hurlbert, R. B. (1973) *Biochemistry* **12**, 4137–4145.
- [48] Moline, G., Hampel, A. and Enger, M. D. (1974) *Biochem. J.* **143**, 191–195.
- [49] Norton, S. J., Key, M. D. and Scholes, S. W. (1965) *Arch. Biochem. Biophys.* **109**, 7–12.
- [50] Quintard, B., Mouricout, J. R., Carias, J. R. and Julien, R. (1978) *Biochem. Biophys. Res. Commun.* **85**, 999–1006.
- [51] Rapaport, E., Zamecnik, P. C. and Baril, E. F. (1981) *Proc. Natl. Acad. Sci. USA* **78**, 838–842.
- [52] Ritter, P. O., Enger, M. D. and Hampel, A. (1976) in: *Onco-developmental Gene Expression* (Fishman, W. H. and Sells, S. eds) pp. 47–56, Academic Press, New York.
- [53] Ritter, P. O., Enger, M. D. and Hampel, A. (1979) *Biochim. Biophys. Acta* **562**, 377–385.
- [54] Roberts, W. K. and Coleman, W. H. (1972) *Biochem. Biophys. Res. Commun.* **46**, 206–214.
- [55] Roberts, W. K. and Olsen, M. L. (1976) *Biochim. Biophys. Acta* **454**, 480–492.
- [56] Saxholm, H. J. K. and Pitot, H. C. (1979) *Biochim. Biophys. Acta* **562**, 386–399.
- [57] Schimmel, P. R. and Söll, D. (1979) *Annu. Rev. Biochem.* **48**, 601–648.
- [58] Shafer, S. J., Olexa, S. and Hillman, R. (1976) *Insect Biochem.* **6**, 405–411.
- [59] Smith, D. W. E., Silbert, P. E. and McNamara, A. L. (1979) *Biochim. Biophys. Acta* **562**, 453–461.
- [60] Smulson, M., Lin, C. S. and Chirikjian, J. G. (1975) *Arch. Biochem. Biophys.* **167**, 458–468.
- [61] Som, K. and Hardesty, B. (1975) *Arch. Biochem. Biophys.* **166**, 507–517.
- [62] Tanaka, W. K., Som, K. and Hardesty, B. (1976) *Arch. Biochem. Biophys.* **172**, 252–260.
- [63] Tscherne, J. S., Weinstein, I. B., Lanks, K. W., Gersten, N. B. and Cantor, C. R. (1973) *Biochemistry* **12**, 3859–3865.
- [64] Ussery, M. A., Tanaka, W. K. and Hardesty, B. (1977) *Eur. J. Biochem.* **72**, 491–500.
- [65] Vadeboncoeur, C. and Lapointe, J. (1980) *Brain Res.* **188**, 129–133.
- [66] Vadeboncoeur, C. and Lapointe, J. (1980) *Eur. J. Biochem.* **109**, 581–587.
- [67] Vellekamp, G. J. and Kull, F. J. (1981) *Eur. J. Biochem.* **118**, 261–269.
- [68] Vennegoor, C. and Bloemendaal, H. (1970) *Eur. J. Biochem.* **15**, 161–170.
- [69] Vennegoor, C. and Bloemendaal, H. (1972) *Eur. J. Biochem.* **26**, 462–473.
- [70] Vennegoor, C. J. G. M., Stols, A. L. H. and Bloemendaal, H. (1972) *J. Mol. Biol.* **65**, 375–378.
- [71] Vennegoor, C. and Bloemendaal, H. (1974) *Methods Enzymol.* **29**, 585–600.

Table I  
Occurrence of high  $M_r$  aminoacyl-tRNA synthetases in eukaryotes<sup>a</sup>

Source	Ala	Arg	Asp	Asn	Cys	Gln	Glu	Gly	His	Ile	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val	References
Mouse liver	+	+	+	+	ND	+	+	+	+	+	+	+	+	+	+	+	ND	+	+	[7-11]	
Mouse liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[64]	
Mouse embryo	+	+	+	ND	ND	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[64]	
Rat liver	+	+	+	ND	ND	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[5,6,22,24]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[12,28]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[18-20,42]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[43]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[56]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[68-71]	
Rat liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[16,17,29,30]	
Rat mammary gland	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[35]	
Rat skeletal muscle	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[4]	
Rabbit reticulocytes	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[61]	
Porcine thyroid gland	+	+	+	+	ND	+	+	+	+	+	+	+	+	+	+	+	ND	+	+	[67]	
Sheep liver	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[13,44-46]	
Bovine brain	+	+	+	+	ND	+	+	+	+	+	+	+	+	+	+	+	ND	+	+	[65,66]	
Calf brain	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[15]	
Human placenta	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	ND	+	+	[21]	
Chick embryo	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[64]	
Friend leukemia cells	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[64]	
Chinese hamster ovary cells	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[27,33,52,53]	
Ehrlich ascites cells <sup>a</sup>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[54,55]	
<i>Drosophila</i> <sup>a</sup>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	[58]	
Wheat germ <sup>a</sup>																			+	[50]	
Yeast <sup>a</sup>																				[25,26]	

<sup>a</sup> Only a few synthetase activities were examined; ND = not determined

Table 2  
Physicochemical properties and composition of eukaryotic aminoacyl-tRNA synthetase complexes <sup>a,b</sup>

Source	$M_r (\times 10^{-3})$	$s_{20,w}$	Ala	Arg	Asp	Asn	Cys	Gln	Glu	Gly	His	Ile	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val	References
Chinese hamster ovary	-	30	-	+	-	-	+	-	-	-	+	-	-	+	-	+	-	-	-	-	-	-	[52]
Rat liver	-	18	-	-	-	-	(+)	-	-	-	(1)	2	1	-	-	-	-	-	-	-	-	-	[69,71]
Rat liver <sup>c</sup>	1000	18	-	+	-	-	+	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	[43]
Rat liver	900	24	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	[20]
Rat liver <sup>d</sup>	285	12	-	(2)	-	-	-	-	-	-	-	-	-	(2)	-	-	-	-	-	-	-	-	[117]
Rat mammary gland	-	20-28	-	+	-	-	+	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	[35]
Rabbit reticulocytes	550	16	-	+	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	[61]
Sheep liver	1000	-	-	+	-	-	-	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	[46]
Human placenta	-	17-20	-	+	-	-	-	+	+	-	+	+	-	-	-	-	-	-	-	-	-	-	[21]

<sup>a</sup> Only complexes characterized with published activity profiles are included in table 2.

<sup>b</sup> A plus signifies that the synthetase activity for the indicated amino acid is present in the complexes.

<sup>c</sup> The numbers indicate the stoichiometry determined by active site titration; parentheses signify tentative assignments (details in [43])

<sup>d</sup> The numbers indicate tentative values of the stoichiometry

**Table 3**  
**Biomolecules with putative role in  $HM_f$  aminoacyl-tRNA synthetase complex formation<sup>a</sup>**

Biomolecule	Method of analysis	Enzyme source	References
Carbohydrate	SDS-polyacrylamide gel staining (periodic acid Schiff reagent)	Thr-RS; rat liver	[24]
	Gas chromatography of enzyme hydrolysate	Lys-, Arg-RS; rat liver	[16,29]
Lipid			
Cholesteryl ester, cholesterol	Extraction of enzyme preparation, paper chromatography	Complex; rat liver	[6]
Cholesteryl 14-methylhexadecanoate	Extraction of enzyme preparation	Complex; rat mammary gland	[35]
Ergosterol	Extraction of enzyme preparation	Rat liver	[36,37]
Glycolipid	Paper chromatography of enzyme preparation extract	Lys-RS; yeast	[25]
		Complex; rat liver	[56]
Ribonucleic acid			
tRNA, 4 S RNA	Extraction of enzyme preparation amino acid acceptor activity	Complex; mouse liver	[9]
		Complex; rat liver	[5]
		Complex; rat liver	[56]
$HM_f$ RNA	Analysis of binding by gel filtration and sucrose gradient ultracentrifugation	Ehrlich ascites cells	[55]
	Affinity chromatography	Rabbit reticulocytes	[3]

<sup>a</sup> Studies included in table 3 vary in levels of exactness and enzyme purity

Table 4  
The interaction of aminoacyl-tRNA synthetases with subcellular components and other enzyme activities<sup>a,b</sup>

Organelle or enzyme	Method of analysis	Source	References
Microsome	Cell fractionation	Chinese hamster ovary	[32]
		Rat skeletal muscle	[4]
		Rat liver	[63]
		Chicken embryo	[49]
		Yeast	[26]
		Wheat germ	[50]
Ribosome	Copurification, reconstitution	Rat liver	[63]
	Copurification	Chinese hamster ovary	[48]
	Copurification or reconstitution	Rabbit reticulocyte	[39,59,62,64]
	Enzyme activity stimulation	Rabbit reticulocyte	[31]
	Protection of enzyme activity	Wheat germ	[14]
	Copurification	Yellow lupin seed	[40]
		Friend leukemic cells, chicken embryo, mouse liver, mouse embryo	[64]
Elongation Factors	Copurification	Ehrlich ascites cell	[55]
	Copurification	Rat liver	[60]
		Rabbit reticulocyte	[38]
Peptidyl acetyltransferase	Copurification	Rat liver	[56]
Initiation Factors	Copurification	Rabbit reticulocyte	[38]
tRNA Methyltransferase	Copurification	Human and mouse leukocytes	[1,2]
tRNA Sulfurtransferase	Copurification	Rat liver	[34]
Ribonuclease	Copurification	Porcine thyroid gland	[67]
DNA Polymerase $\alpha$	Copurification	Hela cell	[51]

<sup>a</sup> Mitochondrial enzymes are different from the cytosolic enzymes [23] and are not considered; nucleolar aminoacyl-tRNA synthetase activities have been detected in purified nucleoli [47]

<sup>b</sup> Studies included in table 4 vary in levels of enzyme purity