

# Hyperkalemia: An adaptive response in chronic renal insufficiency

F. JOHN GENNARI and ALAN S. SEGAL

University of Vermont College of Medicine, Burlington, Vermont, USA

## Hyperkalemia: An adaptive response in chronic renal insufficiency.

**Background.** Hyperkalemia is a common feature of chronic renal insufficiency, usually ascribed to impaired  $K^+$  homeostasis. However, various experimental observations suggest that the increase in extracellular  $[K^+]$  actually functions in a homeostatic fashion, directly stimulating renal  $K^+$  excretion through an effect that is independent of, and additive to, aldosterone.

**Methods.** We have reviewed relevant studies in experimental animals and in human subjects that have examined the regulation of  $K^+$  excretion and its relation to plasma  $[K^+]$ .

**Results.** Studies indicate that (1) extracellular  $[K^+]$  in patients with renal insufficiency correlates directly with intracellular  $K^+$  content, and (2) hyperkalemia directly promotes  $K^+$  secretion in the principal cells of the collecting duct by increasing apical and basolateral membrane conductances. The effect of hyperkalemia differs from that of aldosterone in that  $K^+$  conductances are increased as the primary event. The changes in principal cell function and structure induced by hyperkalemia are indistinguishable from the effects seen in adaptation to a high  $K^+$  diet.

**Conclusions.** We propose that hyperkalemia plays a pivotal role in  $K^+$  homeostasis in renal insufficiency by stimulating  $K^+$  excretion. In patients with chronic renal insufficiency, a new steady state develops in which extracellular  $[K^+]$  rises to the level needed to stimulate  $K^+$  excretion so that it again matches intake. When this new steady state is achieved, plasma  $[K^+]$  remains stable unless dietary intake increases, glomerular filtration rate falls, or drugs are given that disrupt the new balance.

Hyperkalemia in chronic renal insufficiency is generally viewed as a sign of impaired potassium homeostasis, usually ascribed to toxic effects on cell membrane  $Na^+$ ,  $K^+$ -ATPase pump activity or to hyporeninemic hypoaldosteronism [1–4]. In this review, we propose that hyperkalemia is not due to a failure in homeostasis, but rather is a physiological success story. Evidence is presented indicating that hyperkalemia is associated with an in-

crease rather than a maldistribution in body  $K^+$  stores, and that it promotes  $K^+$  excretion by directly stimulating  $K^+$  secretion in the principal cells of the collecting duct of the kidney. Based on such evidence, we postulate that hyperkalemia in renal insufficiency is an adaptive change that serves to restore  $K^+$  balance in the face of a reduction in functioning nephrons. This “ $K^+$  adaptation” is the key to establishing a new homeostatic set point at which  $K^+$  excretion again matches intake. In addition, it may provide protection against surges in plasma  $[K^+]$  that might otherwise occur in chronic renal insufficiency.

## INCIDENCE OF HYPERKALEMIA IN RENAL INSUFFICIENCY

The incidence of hyperkalemia in patients with chronic renal insufficiency is difficult to assess because of the almost universal use of drugs that influence plasma  $[K^+]$ . In one study carried out before angiotensin-converting enzyme (ACE) inhibitors were available and which excluded patients receiving diuretics, plasma  $[K^+]$  was significantly elevated (average value, 4.9 mmol/L) even in patients with only mild-moderate renal dysfunction (serum creatinine 2 to 4 mg/dL) [5]. Hyperkalemia appears to occur more frequently in patients with tubulointerstitial disease or diabetes mellitus [6, 7], but is clearly not confined to these disorders [6]. In a random sample of 300 patients with serum creatinine levels of 1.5 to 6.0 mg/dL taken from our clinic, excluding individuals with diabetes and those receiving either diuretics or drugs that impair angiotensin II synthesis or effect, we found an incidence of hyperkalemia (plasma  $[K^+]$  5.0 mmol/L or higher) of 55%. In the small group of patients in this sample meeting our criteria for inclusion ( $N = 18$ , mean serum creatinine, 3.1 mg/dL), hyperkalemia was as likely to occur with glomerular diseases as with tubulointerstitial disease. Based on these limited observations, the prevalence of hyperkalemia in patients with renal insufficiency may exceed 50%.

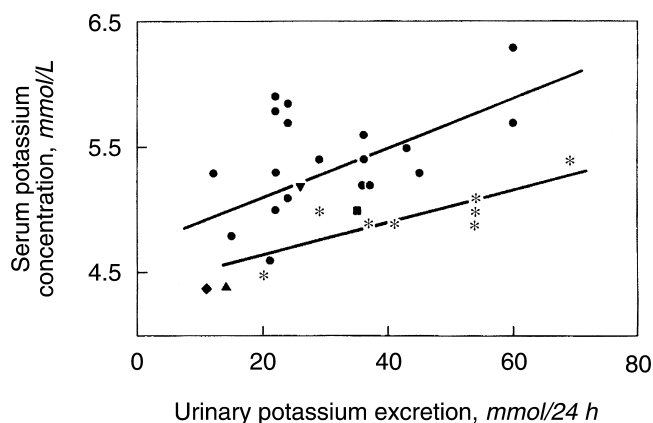
**Key words:** hyperkalemia, potassium, potassium adaptation, kidney, renal failure, ROMK.

Received for publication September 7, 2001

and in revised form December 4, 2001

Accepted for publication December 26, 2001

© 2002 by the International Society of Nephrology



**Fig. 1. Relationship between plasma  $[K^+]$  and urine  $K^+$  excretion in patients with chronic renal insufficiency.** The solid symbols and upper regression line are data obtained in patients with low renin and aldosterone levels; the asterisks and lower regression line are data obtained in patients with normal renin and aldosterone levels. Reprinted with permission from the International Society of Nephrology [7].

## POTASSIUM HOMEOSTASIS IN RENAL INSUFFICIENCY

Regardless of whether hyperkalemia is present in the steady state, patients with renal insufficiency have a diminished ability to acutely excrete a  $K^+$  load and, therefore, have more severe and prolonged hyperkalemia when challenged [8, 9]. Because hyperkalemia stimulates aldosterone secretion, one might expect aldosterone levels to be higher in patients with renal insufficiency than in individuals with normal renal function. However, aldosterone measurements have shown widely varying patterns [3, 7, 10–12], and its importance as a factor in maintaining  $K^+$  balance in renal insufficiency is uncertain. In patients with renal insufficiency, plasma  $[K^+]$  has been shown to correlate directly with daily urinary  $K^+$  excretion regardless of the circulating aldosterone level [7]. In this carefully studied group of patients, those with the highest plasma  $[K^+]$  had the greatest urinary  $K^+$  excretion (Fig. 1). Assuming these patients are in  $K^+$  balance, those with the highest excretion must also have had the highest dietary  $K^+$  intake.

In a related study,  $K^+$  intake was deliberately increased in patients with stable chronic renal insufficiency (mean creatinine clearance, 30 mL/min) by giving them a  $K^+$  supplement of 50 mmol/day [13]. The patients with renal insufficiency increased  $K^+$  excretion to the same degree as control subjects given the same supplement but, in contrast to the control subjects, achieved this goal by increasing plasma  $[K^+]$  by 0.7 mmol/L, to a mean value of 5.4 mmol/L. These observations suggest a causal linkage between the increase in plasma  $[K^+]$  and stimulation of renal  $K^+$  excretion. The respective roles of extracellular  $[K^+]$ , aldosterone, and other factors in maintaining  $K^+$  balance in renal insufficiency are discussed in detail below.

## TRANSCELLULAR POTASSIUM HOMEOSTASIS IN RENAL INSUFFICIENCY

Before reviewing the regulation of  $K^+$  excretion in patients with chronic renal insufficiency, it is important to address the issue of whether the hyperkalemia seen in these patients is due to impaired entry of  $K^+$  into cells or reflects an increase in intracellular  $K^+$  stores. Two general approaches—measurement of total body or striated muscle  $K^+$  stores, and assessment of cell membrane  $Na^+$ ,  $K^+$ -ATPase activity or function—have been used to address this issue. Both approaches have limitations and have provided conflicting results. The key studies are discussed below.

### Body potassium stores

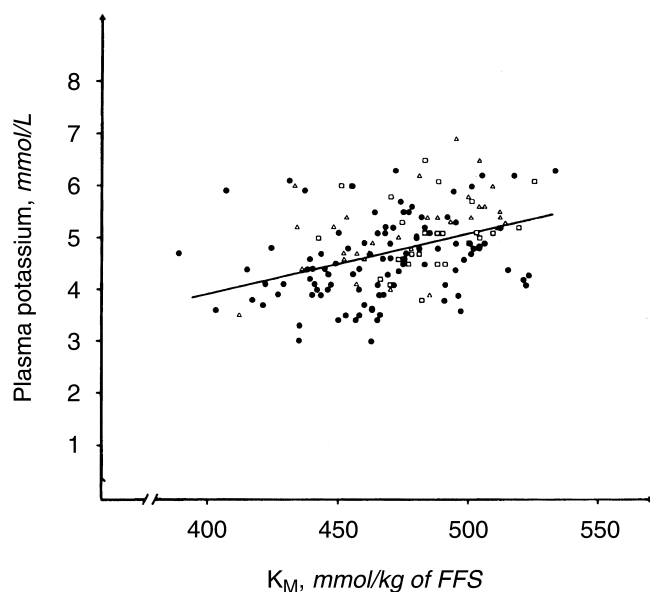
Measurements of total body  $K^+$  content in patients with renal insufficiency show low or normal values, a finding advanced to support the view that  $K^+$  entry into cells is impaired [14–22]. It should be emphasized, however, that these measurements give little information about intracellular  $K^+$  content [14, 22]. Total body stores will be reduced compared to normal individuals, for example, if there is a loss of muscle mass (the main reservoir of intracellular  $K^+$ ) due to uremia, even though intracellular  $K^+$  in the remaining muscle cells may be normal or increased [16, 23]. A more direct assessment of intracellular  $K^+$  can be obtained by analyzing the  $K^+$  content of striated muscle obtained in biopsy samples. Such measurements have shown, in most instances, that the  $K^+$  content in muscle tissue in patients with even advanced renal insufficiency is either indistinguishable from individuals with normal renal function or increased [14, 19, 20].

### Correlation between extracellular and intracellular $[K^+]$

The key issue of whether extracellular  $[K^+]$  correlates directly with intracellular  $[K^+]$  has been addressed in a systematic fashion in only one study [19]. In this study, in which the  $K^+$  content of muscle samples was assessed in patients with renal insufficiency (serum creatinine range, 3.6 to 19.5 mg/dL), plasma  $[K^+]$  correlated directly with muscle  $K^+$  content (Fig. 2). The authors found an even stronger correlation between plasma and intracellular  $[K^+]$  when they estimated the latter from the ratio of  $K^+$  content to water content. Therefore, hyperkalemia is associated both with a higher muscle  $K^+$  content and a higher intracellular  $K^+/H_2O$  ratio.

### Assessment of ATPase activity and function

In red cells, white cells, and myocytes taken from patients and experimental animals with renal insufficiency,  $Na^+$ ,  $K^+$ -ATPase activity is consistently decreased compared to normal controls [1, 2, 4, 24–26]. However, when  $Na^+$ ,  $K^+$ -ATPase function is tested in animals and hu-



**Fig. 2. Relationship between plasma  $[K^+]$  and muscle  $K^+$  content in 103 patients with renal insufficiency.** Muscle  $K^+$  content ( $K_M$ ) is given in mmol/kg of fat-free solids (FFS). The closed symbols are samples taken from patients not receiving dialysis therapy, and the open symbols are from patients receiving either peritoneal or hemodialysis therapy. Reprinted with permission from the International Society of Nephrology [19].

mans with renal insufficiency, no abnormality is detected. Stimulation of  $Na^+,K^+$ -ATPase activity by either insulin or  $K^+$  administration, assessed by  $K^+$  entry into cells, is not impaired [6, 8, 9, 26, 27]. In one study,  $K^+$  entry into cells in response to insulin was actually greater in uremic rats than in normal controls [26]. The reason for the decrease in  $Na^+,K^+$ -ATPase activity in ex vivo samples remains unexplained.

### Effects of aldosterone

The activity of  $Na^+,K^+$ -ATPase is stimulated by aldosterone, and therefore hypoaldosteronism has been implicated to cause hyperkalemia by impairing  $K^+$  entry into cells [1, 3, 4]. However, the role of aldosterone in transcellular  $K^+$  equilibrium remains controversial [28]. Although many studies in experimental animals and human subjects appear to demonstrate a major role for sustained changes in aldosterone in transcellular  $K^+$  homeostasis [29–32], most have not been controlled for changes in body  $K^+$  stores or for the effect of catecholamines [28, 31, 33]. Blockade of the normal surge in aldosterone after acute  $K^+$  administration has no effect on the transcellular disposition of administered  $K^+$  [34, 35]. An additional problem with positing an important effect of aldosterone on transcellular  $K^+$  movement is that striated muscle cells, the major intracellular reservoir of  $K^+$ , lack mineralocorticoid receptors [36].

### Renal regulation of $K^+$ homeostasis

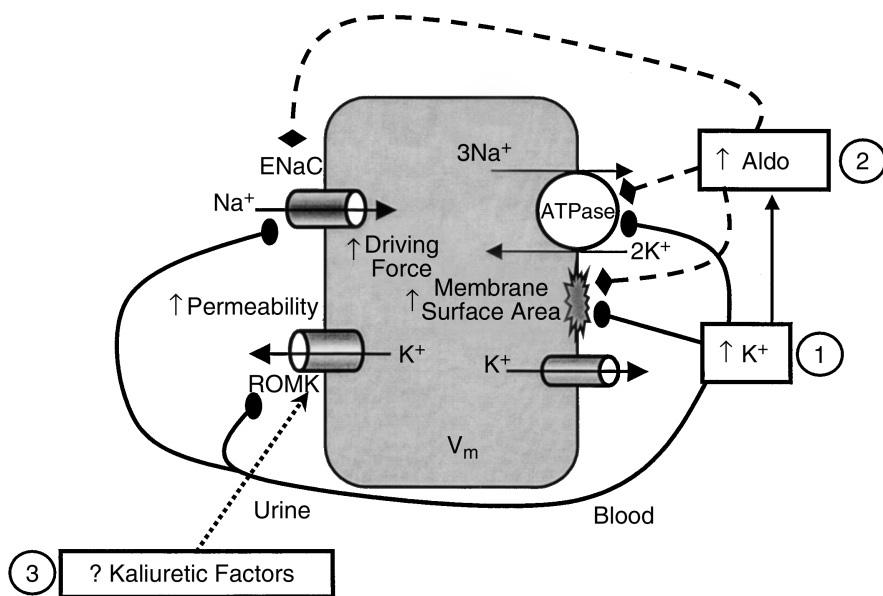
Regardless of whether  $K^+$  distribution in the body is normal or abnormal in patients with chronic renal insufficiency, excretion of  $K^+$  must eventually match intake or lethal hyperkalemia will supervene within days. Potassium excretion by the kidney is tightly regulated and is determined primarily by events beyond the early distal tubule where either reabsorption or secretion of  $K^+$  can occur [37–40]. Potassium excretion almost always exceeds the amount delivered to the early distal tubule (except under conditions of sustained  $K^+$  depletion), indicating that the rate of secretion is the key determinant of  $K^+$  excretion. Secretion of  $K^+$  occurs in the collecting duct and is mediated primarily by apical membrane  $K^+$  (ROMK) channels in the principal cells [39–41], although it may be augmented by “maxi- $K$ ” channels in the apical membrane of intercalated cells under conditions of high urine flow rates [42].

When renal function is normal, aldosterone and tubular flow rate to the distal nephron appear to be the key factors regulating  $K^+$  excretion [43], and plasma  $[K^+]$  remains within normal levels over a wide range of dietary  $K^+$  intake. However, as discussed below, the specific role of aldosterone remains somewhat controversial and other regulatory influences, perhaps even extracellular  $[K^+]$  itself, may play a role [44]. In adrenalectomized dogs on fixed aldosterone replacement, for example, an increase in plasma  $[K^+]$  increases urinary  $K^+$  excretion without any change in  $Na^+$  excretion [45]. In individuals with renal insufficiency, flow rate to the remaining functioning tubules is already maximized so that other factors must come into play to regulate  $K^+$  excretion. We will examine the evidence that plasma  $[K^+]$  itself is a key player, which interacts with aldosterone and other factors to regulate  $K^+$  excretion in patients with renal insufficiency.

### Principal cell $K^+$ secretion

Potassium secretion through principal cells of the collecting duct is a two-step process involving (1) cellular  $K^+$  entry across the basolateral membrane via the  $Na^+,K^+$ -ATPase pump, and (2)  $K^+$  exit across the apical membrane via  $K^+$  channels that open to allow secretion into an electronegative lumen (Fig. 3). The electronegativity of the lumen is largely due to  $Na^+$  reabsorption through the epithelial  $Na^+$  channel, ENaC (Fig. 4A). Potassium secretion through apical  $K^+$  channels constitutes the rate-limiting step of  $K^+$  secretion, and occurs through the ROMK potassium channel in the principal cells [46, 47].

*Effect of hyperkalemia on  $K^+$  secretion.* The specific effects of peritubular  $[K^+]$  on distal tubular  $K^+$  secretion and renal  $K^+$  excretion were first elucidated in a series of in situ microperfusion studies in rats [48]. In these experiments, acute hyperkalemia was induced by intra-



**Fig. 3. Major factors that regulate  $K^+$  secretion in principal cells.** Sodium is reabsorbed across the luminal membrane through ENaC  $Na^+$  channels, with the resultant cellular depolarization increasing the electrical driving force for  $K^+$  secretion through ROMK  $K^+$  channels. (1) Elevation of peritubular  $[K^+]$  (circular arrowheads) increases the density of luminal ENaC and ROMK channels, which both promote  $K^+$  secretion by increasing the electrical driving force and  $K^+$  permeability, respectively. Increases in peritubular  $[K^+]$  also activate the  $Na^+,K^+$ -ATPase pump in the BLM and stimulate aldosterone release. (2) Aldosterone (diamond arrowheads) increases the density of ENaC (but not ROMK) channels and activates the  $Na^+,K^+$ -ATPase pump, both of which increase the driving force for  $K^+$  secretion. The surface area of the BLM containing the  $Na^+,K^+$ -ATPase pump undergoes amplification during prolonged exposure to either increased peritubular  $[K^+]$  or aldosterone. (3) Kaliuretic factors, including  $K^+$  itself, have been proposed that somehow directly increase  $K^+$  secretion. For example, high luminal  $[K^+]$  may directly increase the activity of ROMK channels.

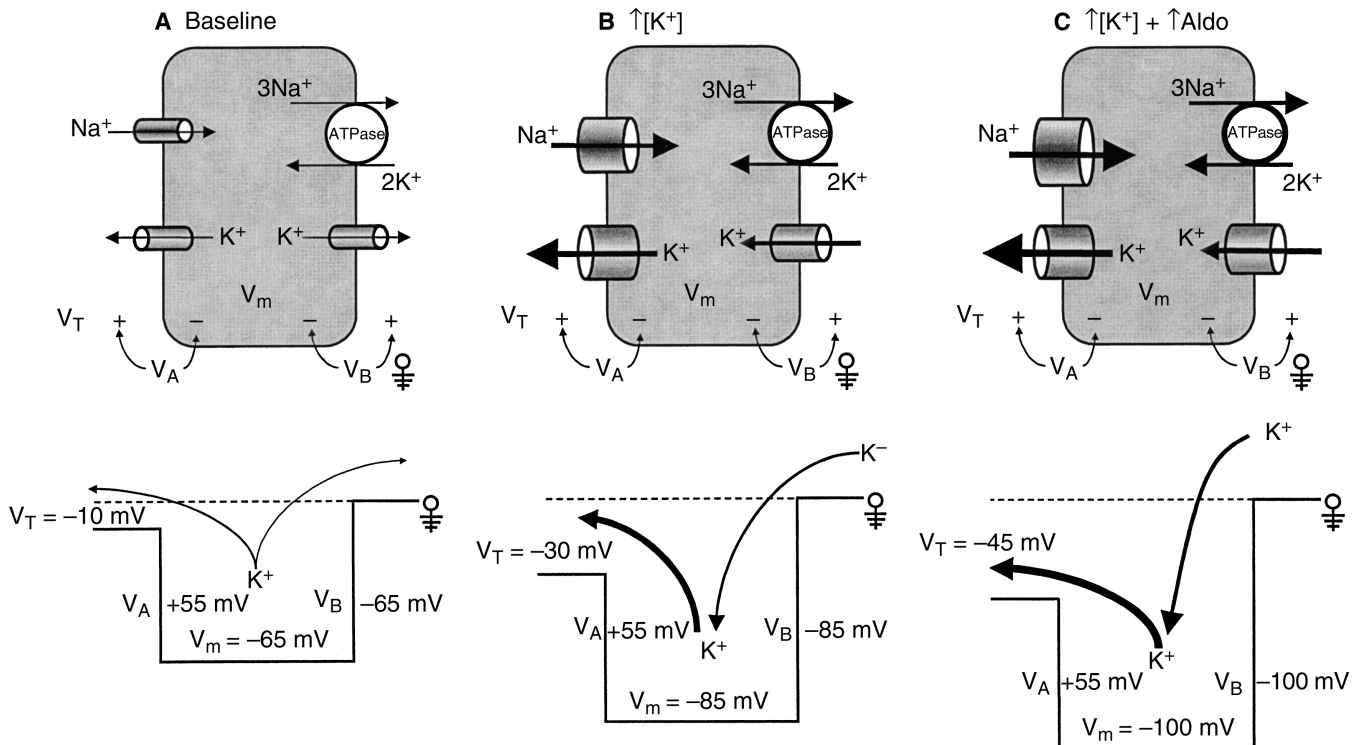
venous  $K^+$  administration into adrenalectomized rats maintained at a fixed aldosterone level. Hyperkalemia more than doubled  $K^+$  secretion in perfused surface distal tubules and acutely increased urinary  $K^+$  excretion. The increase in  $K^+$  secretion induced by hyperkalemia was the same whether aldosterone secretion rate was fixed at low or high levels, indicating independence of this effect of hyperkalemia from aldosterone. Moreover,  $Na^+$  delivery, pH, tubular flow rate, and transepithelial potential difference were all controlled, indicating a specific effect of peritubular  $[K^+]$  on  $K^+$  secretion [48].

In a related study, stimulation of distal tubular  $K^+$  secretion by increases in plasma  $[K^+]$  was shown to occur primarily when  $[K^+]$  was increased from 4 to 6 mmol/L, with no further effect at higher concentrations [49]. Therefore, the effect is prompt, occurs over a range of plasma  $[K^+]$  not associated with adverse events in humans, and acts in a homeostatic fashion to increase urinary  $K^+$  excretion. Based on these studies, one can estimate that more than half of the stimulation of  $K^+$  secretion in the distal tubule is induced by hyperkalemia, with the remainder stimulated by aldosterone [48]. In support of these results in intact rats, several studies in isolated perfused rabbit collecting ducts have shown that  $K^+$  secretion is stimulated by an increase in peritubular  $[K^+]$  [50–53].

The nature of this stimulation has been explored using electrophysiological techniques in isolated rabbit collecting tubules by Muto et al (Fig. 4) [53]. In their study, an increase in peritubular  $[K^+]$  initiated the following sequence of events: (1) immediate activation of the  $Na^+,K^+$ -ATPase pump, (2) an increase in the  $K^+$  conduc-

tance in both the apical and basolateral membranes, and (3) an increase in the apical  $Na^+$  conductance. The increase in apical  $K^+$  conductance is due to an increase in the ROMK  $K^+$  conductance, and the increase in apical  $Na^+$  conductance is due to an increase in the ENaC  $Na^+$  conductance. The latter plays a key role because inhibition of ENaC by amiloride attenuates activation of the  $Na^+,K^+$ -ATPase pump, which decreases cellular  $K^+$  entry and therefore  $K^+$  secretion. Conversely, these effects are amplified by treatment with mineralocorticoids, which stimulates ENaC [54–56]. Careful microperfusion studies also have shown that an acute elevation of peritubular  $[K^+]$  directly activates the basolateral  $Na^+,K^+$ -ATPase pump [49, 57], which secondarily increases the apical  $Na^+$  conductance [52, 53]. Both of these changes elevate the driving force for  $K^+$  secretion through apical ROMK channels (Fig. 4B). However, the signaling mechanisms underlying the “crosstalk” between the luminal and peritubular membranes remain unknown [58].

*Interaction between  $K^+$  and aldosterone.* The interaction between extracellular  $[K^+]$  and  $K^+$  excretion involves the adrenal cortex as well as the kidney (Figs. 3 and 4). An increase in extracellular  $[K^+]$  is well known to stimulate aldosterone secretion by the adrenal gland, an effect facilitated by simultaneous stimulation of angiotensin II secretion [34, 59–61]. This effect is apparent in patients with renal insufficiency as well; aldosterone correlates directly with plasma  $[K^+]$  in these patients [7]. Aldosterone increases the density and activity of  $Na^+,K^+$ -ATPase pumps in the basolateral membrane (BLM) and  $Na^+$  channels in the apical membrane [52,



**Fig. 4. Electrophysiology of  $K^+$  secretion and  $K^+$  adaptation in cortical collecting duct (CCD) principal cells.** The luminal membrane has ENaC  $Na^+$  channels and ROMK  $K^+$  channels, and the peritubular membrane has  $K^+$  channels and  $Na^+,K^+$ -ATPase pumps. The thickness of the arrow through the channels is proportional to the driving force and the size of the channel is proportional to the conductance. Stimulation of the pump is denoted by increased thickness. (A) At baseline,  $K^+$  entering across the BLM via the  $Na^+,K^+$ -ATPase pump is subsequently secreted across the apical membrane into an electronegative lumen while some  $K^+$  recycles back into the blood. (B) An increase in peritubular  $[K^+]$  activates the pump and leads to an increase in the  $K^+$  conductance in the BLM, resulting in hyperpolarization of  $V_B$ . When  $V_B$  becomes more negative than  $E_K^{BLM}$ , the direction of  $K^+$  current reverses, and  $K^+$  enters the cell. An elevation of peritubular  $[K^+]$  also increases the density of apical membrane conductances; more  $Na^+$  current via ENaC increases lumen electronegativity ( $V_T$ ) and therefore the driving force for  $K^+$  secretion through a higher density of ROMK channels. (C) The combination of increased peritubular  $[K^+]$  and mineralocorticoids further augments the changes described in panel B. Note that aldosterone leads to an increase in the driving force for  $K^+$  secretion through ROMK channels, but does not increase the density of these channels. The **Appendix** contains a more detailed description of the electrophysiological profile under each condition. (Adapted from [56]; used with permission from the American Physiological Society).

53, 58, 62, 63], but does not increase the density of apical  $K^+$  channels [64]. These effects of aldosterone, coupled with the increase in the density of apical  $K^+$  channels induced by peritubular hyperkalemia, result in both an increase in  $K^+$  permeability and driving force, which is an extremely favorable setting for  $K^+$  secretion [54, 55, 62, 63, 65].

Additional gain is built into the system because an increase in extracellular  $[K^+]$  also stimulates aldosterone secretion, which enhances the effect of increased peritubular  $[K^+]$  on  $K^+$  secretion. Moreover, if stimulation of secretion is prolonged, the surface area of the basolateral membrane undergoes amplification [66]. Such a chain of events explains why  $K^+$  adaptation can be rapid [67] and robust [68]. In addition to stimulating  $K^+$  secretion in the kidney, aldosterone stimulates  $K^+$  secretion by the colonic epithelium, promoting stool  $K^+$  loss [69–71]. Indirect evidence suggests that hyperkalemia also stimulates colonic  $K^+$  secretion independent of aldosterone [69, 70].

*$K^+$  sensors,  $K^+$  channels, and kaliuresis.* The transient hyperkalemia induced by  $K^+$  ingestion and absorption from the gut is almost certainly the proximate signal for the acute increase in  $K^+$  excretion by the kidney [72–75]. This renal response begins within 30 minutes of receiving a  $K^+$  load [6, 72, 75], and probably prevents any detectable change in systemic  $[K^+]$  [73, 74]. Although aldosterone secretion also increases within minutes in response to  $K^+$  loading [34, 75], its role in the rapid increase in  $K^+$  excretion that follows an oral  $K^+$  load remains controversial [47, 74, 75]. Some investigators have postulated the existence of a neuroenteric kaliuretic reflex involving a splanchnic  $K^+$  sensor that senses dietary  $K^+$  and somehow stimulates renal tubular  $K^+$  secretion [74, 76]. Another possibility is that peritubular  $[K^+]$  itself is the most important kaliuretic factor.

#### ADAPTIVE MECHANISMS OF $K^+$ EXCRETION

Sustained hyperkalemia does not occur in individuals with normal renal function despite marked increases in

potassium intake, because an adaptive change in distal tubular  $K^+$  secretion occurs such that intake is matched by rapid and equivalent increases in  $K^+$  excretion [31, 49, 77, 78]. This response of the kidney to high dietary  $K^+$  intake, first demonstrated in rats [79], is termed potassium adaptation. The distal tubular events induced by a high  $K^+$  diet are identical to those observed when extracellular  $[K^+]$  is increased, suggesting that such an increase may be the signal for  $K^+$  adaptation. In experimental animals, a high  $K^+$  diet induces an increase in ROMK channels, ENaC channels, and  $Na^+,K^+$ -ATPase pump activity, in the absence of an increase in aldosterone secretion. By contrast, when rats are given exogenous aldosterone or maintained on a low  $Na^+$  diet to increase endogenous aldosterone levels, the density of apical ENaC channels increases, but there is no change in the number of ROMK channels [47]. Although sustained hyperkalemia does not occur when renal function is normal in experimental animals or humans ingesting a high  $K^+$  diet, transient hyperkalemia has been observed in the postprandial period. In human subjects with normal renal function ingesting a liquid formula diet containing large amounts of  $K^+$  (5 mmol/kg body weight) for a five-day period, serum  $K^+$  rose transiently to between 5 and 6 mmol/L after meals (R. Tannen, personal communication). Although the factors signaling the initiation of renal  $K^+$  adaptation remain to be defined, one factor could be a repetitive postprandial increase in extracellular  $[K^+]$ .

Whatever the underlying mechanisms,  $K^+$  adaptation is characterized by changes in apical  $K^+$  and  $Na^+$  conductances and in basolateral  $Na^+,K^+$ -ATPase pump activity that can occur in the absence of aldosterone. In addition to an increase in plasma  $[K^+]$ , urinary  $[K^+]$  may be part of the signaling mechanism, as a recent study has demonstrated that an elevation of luminal  $[K^+]$  also increases the density of ROMK channels [80], an effect that may contribute to  $K^+$  adaptation.

### **$K^+$ adaptation in renal insufficiency**

How might hyperkalemia induce  $K^+$  adaptation in the setting of an overall reduction in functioning nephron mass, when each remaining nephron faces an increased workload? As discussed earlier, the effects of hyperkalemia on the collecting duct are virtually identical to those seen during  $K^+$  adaptation. They include (1) an amplification of basolateral membrane area [81] and  $Na^+,K^+$ -ATPase pump activity [82, 83], (2) an increase in apical  $Na^+$  delivery and reabsorption [84], and (3) an increase in  $K^+$  excretion per nephron in order to match  $K^+$  intake [85]. Based on these observations, the following cascade of events may occur in renal insufficiency (Fig. 3). As ingested  $K^+$  is absorbed and retained due an initial impairment in  $K^+$  excretion, hyperkalemia develops, amplifying the normal signal to excrete  $K^+$  (aldosterone and

other putative kaliuretic factors). This augmented signal will lead not only to activation of the  $Na^+,K^+$ -ATPase pump and an increase in apical  $Na^+$  channel density, but also to an increase in apical  $K^+$  channel density [58], producing an extremely favorable milieu for  $K^+$  secretion. Because of the decrease in functioning nephrons in renal insufficiency, the development of sustained hyperkalemia may actually be required for full  $K^+$  adaptation in many of these patients.

### **HYPERKALEMIA: SIGNAL AND EFFECTS**

Based on the evidence presented in this review, we propose the following scheme. Either a reduction in GFR or an increase in dietary  $K^+$  intake causes a transient disequilibrium resulting in an increase in body  $K^+$  stores, reflected by an increase in plasma  $[K^+]$  when aldosterone or other kaliuretic factors fail to correct the imbalance. In the absence of a rapid ability to increase renal  $K^+$  excretion and correct this disequilibrium, extracellular  $[K^+]$  rises until it reaches a level sufficient to produce a sustained augmentation of renal  $K^+$  excretion, so that excretion again matches intake. In this new steady state, plasma  $[K^+]$  will remain stable at an elevated level until either dietary  $K^+$  intake, aldosterone secretion, or renal function changes. In patients with low aldosterone levels, dietary  $K^+$  and renal function become the primary determinants of plasma  $[K^+]$ . If dietary intake falls or renal function improves, plasma  $[K^+]$  will fall. If intake increases or renal function deteriorates, plasma  $[K^+]$  will rise until balance is again achieved.

### **Fragility of the new steady state**

Although a new steady state may be achieved in which plasma  $[K^+]$  is elevated but stable in patients with renal insufficiency, this delicate balance can easily be disrupted. It is apparent from Figure 1 that simply increasing dietary  $K^+$  intake can raise extracellular  $[K^+]$  to potentially dangerous levels. Potassium balance is more commonly disrupted by physician intervention. Indeed, the most common cause of severe hyperkalemia in patients with renal insufficiency is physician-prescribed  $K^+$  supplements [86, 87]. In addition to  $K^+$  itself, drugs that impair  $K^+$  homeostasis are often responsible for disrupting the new steady state in these patients [88]. Aside from drug effects, a wide range of factors may influence transcellular  $K^+$  distribution and exacerbate hyperkalemia in patients with decreased renal function, including metabolic acidosis, reduced  $\beta$ -adrenergic activity, and perhaps toxic effects on cell membrane  $Na^+,K^+$ -ATPase [4].

### **Therapeutic implications**

Recognition that mild to moderate hyperkalemia is an adaptive response should lead to tolerance of a steady-state serum  $[K^+]$  of 5.0 to 5.5 mmol/L in patients with

chronic renal insufficiency. In patients with higher serum  $[K^+]$  levels, intervention should be directed at uncovering factors such as inadvertent potassium intake (for example, “salt substitutes”) or drugs that impair  $K^+$  excretion or transcellular distribution. Given the relationship shown in Figure 1, important intervention in all patients with chronic renal insufficiency and hyperkalemia should be dietary counseling to assure that foods high in  $K^+$  content are not ingested in excessive amounts. Reduction in dietary  $K^+$  intake to 2 to 3 g/day may even be sufficient to allow continuation of angiotensin inhibiting drugs that can prolong renal survival. Chronic Kayexalate therapy should be reserved for patients who are unable to regulate their dietary intake, and acute intervention should be reserved for those patients with extremely high serum  $[K^+]$  levels ( $>6.5$  mmol/L).

## SUMMARY AND FUTURE DIRECTIONS

We propose that hyperkalemia plays a pivotal role in  $K^+$  homeostasis. It stimulates  $K^+$  excretion and restores balance when  $K^+$  intake outstrips the kidney’s ability to respond in patients with renal insufficiency. Hyperkalemia promotes  $K^+$  excretion by directly stimulating  $K^+$  secretion in collecting duct principal cells. The cellular mechanisms by which  $K^+$  secretion is increased by hyperkalemia remain to be determined, but the effect is independent of, and additive to, the effects of aldosterone. In patients with renal insufficiency, extracellular  $[K^+]$  rises to the level needed to stimulate collecting duct  $K^+$  secretion so that renal  $K^+$  excretion again matches intake. When this new steady state is achieved, extracellular  $[K^+]$  remains stable unless dietary intake increases further, GFR falls, or drugs are given that disrupt the new balance. Management of hyperkalemia in patients with renal insufficiency should focus on preventing disruption of this new steady state by reviewing drug therapy and dietary counseling.

Our proposal that the elevation in plasma  $[K^+]$  seen in many patients with chronic renal insufficiency is an adaptive signal for promoting kaliuresis and that hyperkalemia is necessary for homeostasis requires further basic and clinical investigation. For example, the physiological studies done by Muto et al on rabbits with normal renal function could be repeated using healthy tubules from animals with renal insufficiency [52, 53]. Our thesis would predict that these tubules would show an exaggerated kaliuretic response compared to those from animals with normal renal function. To examine whether the initial kaliuretic signal comes from sensing dietary  $K^+$  or peritubular  $[K^+]$ , it might be possible to somehow “low  $[K^+]$ -clamp” one renal artery of adrenalectomized animals fed a high  $K^+$  diet, and then compare the response of tubules from each kidney. A reasonable starting point for clinical experiments would be to place individuals

with normal renal function on a high potassium diet and make frequent postprandial measurements of plasma  $[K^+]$  to see whether transient hyperkalemia occurs. If so, one could then examine the relationship between dietary  $K^+$  and the increase in renal  $K^+$  excretion.

Reprint requests to F. John Gennari, M.D., 2308 Rehab, UHC campus, Fletcher Allen Health Care, Burlington, Vermont 05401, USA.  
E-mail: fgennari@zoo.uvm.edu

## APPENDIX

Refer to Figure 4. By convention, the luminal barrier voltage  $V_A$  is positive, the basolateral barrier voltage  $V_B$  is negative, and the peritubular side is ground. Therefore, the transepithelial voltage,  $V_T = V_A + V_B$ , is negative (that is, electronegative lumen). The Nernst potential (in mV) for  $K^+$  across the luminal membrane is:

$$E_K^{\text{apical}} = (25.4) \ln([K^+]_{\text{lumen}}/[K^+]_{\text{cell}})$$

and that across the peritubular membrane is:

$$E_K^{\text{BLM}} = (25.4) \ln([K^+]_{\text{blood}}/[K^+]_{\text{cell}})$$

The equations for the  $K^+$  current across each membrane are:

*Apical  $K^+$  current:*

$$\begin{aligned} I_K^{\text{Apical}} &= G_K^{\text{Apical}} * [(V_T - V_B) - E_K^{\text{Apical}}] \\ &= G_K^{\text{Apical}} * [V_A - E_K^{\text{Apical}}] \end{aligned}$$

where  $G_K^{\text{Apical}}$  is the sum of all luminal  $K^+$  conductances.

- $V_T$  hyperpolarizes if  $V_B$  hyperpolarizes and/or if the lumen becomes more negative (for example, with mineralocorticoid treatment and activation of ENaC).
- $V_T$  depolarizes if  $V_B$  depolarizes and/or if the lumen becomes less negative (such as, amiloride blockade of ENaC).
- Note that since  $V_A$  is always positive and  $E_K^{\text{Apical}}$  is always negative, the apical  $K^+$  current is always directed outward (cell  $\rightarrow$  lumen):

*Basolateral  $K^+$  current:*

$$I_K^{\text{BLM}} = G_K^{\text{BLM}} * [V_B - E_K^{\text{BLM}}]$$

where  $G_K^{\text{BLM}}$  is the sum of all peritubular  $K^+$  conductances.

- $K^+$  current across the BLM is directed outward (cell  $\rightarrow$  blood) under control conditions because  $V_B$  is less negative than  $E_K^{\text{BLM}}$ . However, high peritubular  $[K^+]$  and/or treatment with mineralocorticoids lowers  $V_A$  (by increasing inward current through ENaC) and can cause  $V_B$  (and therefore  $V_T$ ) to hyperpolarize (both by increasing BLM  $G_K$  and by activating the pump) more negative than  $E_K^{\text{BLM}}$ , leading to  $K^+$  entry across the BLM (blood  $\rightarrow$  cell), promoting more  $K^+$  secretion through apical ROMK channels.

## REFERENCES

1. VAN YPERSELE DE STRIHOUC: Potassium homeostasis in renal failure. *Kidney Int* 11:491–504, 1977
2. DRUML W, KELLY RA, MAY RC, MITCH WE: Abnormal cation transport in uremia. Mechanisms in adipocytes and skeletal muscle from uremic rats. *J Clin Invest* 81:1197–1203, 1988
3. DEFONZO RA: Hyperkalemia and hyporeninemic hypoaldosteronism. *Kidney Int* 17:118–134, 1980
4. SALEM M, ROSA RM, BATLLE DC: Extrarenal potassium tolerance in chronic renal failure: Implications for the treatment of acute hyperkalemia. *Am J Kidney Dis* 18:421–440, 1991
5. WIDMER B, GERHARDT RE, HARRINGTON JT, COHEN JJ: Serum electrolyte and acid base composition. The influence of graded degrees of chronic renal failure. *Arch Intern Med* 139:1099–1102, 1979
6. GONICK HC, KLEEMAN CR, RUBINI ME, MAXWELL MH: Functional

- impairment in chronic renal disease. 3. Studies of potassium excretion. *Am J Med Sci* 261:281–290, 1971
7. SCHAMBELAN M, SEBASTIAN A, BIGLIERI EG: Prevalence, pathogenesis, and functional significance of aldosterone deficiency in hyperkalemic patients with chronic renal insufficiency. *Kidney Int* 17:89–101, 1980
  8. KEITH NM, OSTERBERG AE: The tolerance for potassium in severe renal insufficiency. *J Clin Invest* 26:773–783, 1947
  9. PEREZ GO, PELLETA R, OSTER JR, *et al*: Blunted kaliuresis after an acute potassium load in patients with chronic renal failure. *Kidney Int* 24:656–662, 1983
  10. HENE RJ, BOER P, KOOMANS HA, MEES EJ: Plasma aldosterone concentrations in chronic renal disease. *Kidney Int* 21:98–101, 1982
  11. WEIDMANN P, MAXWELL MH, ROWE P, *et al*: Role of the renin-angiotensin-aldosterone system in the regulation of plasma potassium in chronic renal disease. *Nephron* 15:35–49, 1975
  12. BERL T, KATZ FH, HENRICH WL, *et al*: Role of aldosterone in the control of sodium excretion in patients with advanced chronic renal failure. *Kidney Int* 14:228–235, 1978
  13. KAHN T, KAJI DM, NICOLIS G, *et al*: Factors related to potassium transport in chronic stable renal disease in man. *Clin Sci Mol Med* 54:661–666, 1978
  14. MITCH WE, WILCOX CS: Disorders of body fluids, sodium and potassium in chronic renal failure. *Am J Med* 72:536–550, 1982
  15. BODDY K, KING PC, LINDSAY RM, *et al*: Exchangeable and total body potassium in patients with chronic renal failure. *BMJ* 1:140–142, 1972
  16. LETTERI JM, ASAD SN, CASELNOVA R, *et al*: Creatinine excretion and total body potassium in renal failure. *Clin Nephrol* 4:58–61, 1975
  17. BERLYNE GM, VAN LAETHEM L, BEN ARI J: Exchangeable potassium and renal potassium handling in advanced chronic renal failure in man. *Nephron* 8:264–269, 1971
  18. GRAHAM JA, LAWSON DH, LINTON AL: Muscle biopsy water and electrolyte contents in chronic renal failure. *Clin Sci* 38:583–591, 1970
  19. BERGSTRÖM J, ALVSTRAND A, FURST P, *et al*: Muscle intracellular electrolytes in patients with chronic uremia. *Kidney Int* 24(Suppl 16):S153–S160, 1983
  20. COTTON JR, WOODARD T, CARTER NW, KNOCHEL JP: Resting skeletal muscle membrane potential as an index of uremic toxicity. A proposed new method to assess adequacy of hemodialysis. *J Clin Invest* 63:501–506, 1979
  21. PATRICK J, JONES NF, BRADFORD B, GAUNT J: Leucocyte potassium in uraemia: Comparisons with erythrocyte potassium and total exchangeable potassium. *Clin Sci* 43:669–678, 1972
  22. PATRICK J: Assessment of body potassium stores. *Kidney Int* 11:476–490, 1977
  23. KOPPLE JD, COBURN JW: Metabolic studies of low protein diets in uremia. I. Nitrogen and potassium. *Medicine (Baltimore)* 52:583–595, 1973
  24. CLAUSEN T, EVERTS ME: Regulation of the Na,K-pump in skeletal muscle. *Kidney Int* 35:1–13, 1989
  25. CHENG JT, KAHN T, KAJI DM: Mechanism of alteration of sodium potassium pump of erythrocytes from patients with chronic renal failure. *J Clin Invest* 74:1811–1820, 1984
  26. GOECKE IA, BONILLA S, MARUSIC ET, ALVO M: Enhanced insulin sensitivity in extrarenal potassium handling in uremic rats. *Kidney Int* 39:39–43, 1991
  27. ALVSTRAND A, WAHREN J, SMITH D, DEFONZO RA: Insulin-mediated potassium uptake is normal in uremic and healthy subjects. *Am J Physiol* 246:E174–E180, 1984
  28. SPITAL A, STERNS RH: Extrarenal potassium adaptation: The role of aldosterone. *Clin Sci Colch* 76:213–219, 1989
  29. YOUNG DB, JACKSON TE: Effects of aldosterone on potassium distribution. *Am J Physiol* 243:R526–R530, 1982
  30. SUGARMAN A, BROWN RS: The role of aldosterone in potassium tolerance: Studies in anephric humans. *Kidney Int* 34:397–403, 1988
  31. BIA MJ, DEFONZO RA: Extrarenal potassium homeostasis. *Am J Physiol* 240:F257–F268, 1981
  32. ALEXANDER EA, LEVINSKY NG: An extrarenal mechanism of potassium adaptation. *J Clin Invest* 47:740–748, 1968
  33. COX M, STERNS RH, SINGER I: The defense against hyperkalemia: The roles of insulin and aldosterone. *N Engl J Med* 299:525–532, 1978
  34. PRATT JH: Role of angiotensin II in potassium-mediated stimulation of aldosterone secretion in the dog. *J Clin Invest* 70:667–672, 1982
  35. CLARK BA, BROWN RS, EPSTEIN FH: Effect of atrial natriuretic peptide on potassium-stimulated aldosterone secretion: potential relevance to hypoaldosteronism in man. *J Clin Endocrinol Metab* 75:399–403, 1992
  36. ARRIZA JL, WEINBERGER C, CERELLI G, *et al*: Cloning of human mineralocorticoid receptor complementary DNA: Structural and functional kinship with the glucocorticoid receptor. *Science* 237:268–275, 1987
  37. MALNIC G, MUTO S, GIEBISCH G: Regulation of potassium excretion, in *The Kidney, Physiology, and Pathophysiology*, (3rd ed), edited by SELDIN DW, GIEBISCH G, Philadelphia, Lippincott Williams & Wilkins, 2000, pp 1575–1613
  38. GIEBISCH G, MALNIC G, BERLINER RW: Control of renal potassium excretion, in *The Kidney* (6th ed), edited by BERLINER BM, Philadelphia, Saunders, 2000, pp 417–454
  39. GIEBISCH G, WANG W: Potassium transport: From clearance to channels and pumps. *Kidney Int* 49:1624–1631, 1996
  40. GIEBISCH G: Renal potassium transport: Mechanisms and regulation. *Am J Physiol* 274:F817–F833, 1998
  41. GOOD DW, WRIGHT FS: Luminal influences on potassium secretion: Sodium concentration and fluid flow rate. *Am J Physiol* 236:F192–F205, 1979
  42. WODA CB, BRAGIN A, KLEYMAN TR, SATLIN LM: Flow-dependent K<sup>+</sup> secretion in the cortical collecting duct is mediated by a maxi-K channel. *Am J Physiol (Renal Physiol)* 280:F786–F793, 2001
  43. STANTON BA: Regulation of Na<sup>+</sup> and K<sup>+</sup> transport by mineralocorticoids. *Semin Nephrol* 7:82–90, 1987
  44. YOUNG DB: Quantitative analysis of aldosterone's role in potassium regulation. *Am J Physiol* 255:F811–F822, 1988
  45. YOUNG DB: Relationship between plasma potassium concentration and renal potassium excretion. *Am J Physiol* 242:F599–F604, 1982
  46. GIEBISCH G: Physiological roles of renal potassium channels. *Semin Nephrol* 19:458–471, 1999
  47. PALMER LG: Potassium secretion and the regulation of distal nephron K channels. *Am J Physiol* 277:F821–F825, 1999
  48. FIELD MJ, STANTON BA, GIEBISCH GH: Differential acute effects of aldosterone, dexamethasone, and hyperkalemia on distal tubular potassium secretion in the rat kidney. *J Clin Invest* 74:1792–1802, 1984
  49. STANTON BA, GIEBISCH GH: Potassium transport by the renal distal tubule: Effects of potassium loading. *Am J Physiol* 243:F487–F493, 1982
  50. WINGO CS, SELDIN DW, KOKKO JP, JACOBSON HR: Dietary modulation of active potassium secretion in the cortical collecting tubule of adrenalectomized rabbits. *J Clin Invest* 70:579–586, 1982
  51. MUTO S, SANSOM S, GIEBISCH G: Effects of a high potassium diet on electrical properties of cortical collecting ducts from adrenalectomized rabbits. *J Clin Invest* 81:376–380, 1988
  52. MUTO S, GIEBISCH G, SANSOM S: An acute increase of peritubular K stimulates K transport through cell pathways of CCT. *Am J Physiol* 255:F108–F114, 1988
  53. MUTO S, ASANO Y, SELDIN D, GIEBISCH G: Basolateral Na<sup>+</sup> pump modulates apical Na<sup>+</sup> and K<sup>+</sup> conductances in rabbit cortical collecting ducts. *Am J Physiol* 276:F143–F158, 1999
  54. SANSOM SC, O'NEIL RG: Mineralocorticoid regulation of apical cell membrane Na<sup>+</sup> and K<sup>+</sup> transport of the cortical collecting duct. *Am J Physiol* 248:F858–F868, 1985
  55. SANSOM SC, O'NEIL RG: Effects of mineralocorticoids on transport properties of cortical collecting duct basolateral membrane. *Am J Physiol* 251:F743–F757, 1986
  56. LING BN, KEMENDY AE, KOKKO KE, *et al*: Regulation of the amiloride-blockable sodium channel from epithelial tissue. *Mol Cell Biochem* 99:141–150, 1990
  57. FUJII Y, KATZ AI: Direct Na<sup>+</sup>-K<sup>+</sup> pump stimulation by K<sup>+</sup> in cortical collecting tubules: A mechanism for early renal K<sup>+</sup> adaptation. *Am J Physiol* 257:F595–F601, 1989
  58. MUTO S: Potassium transport in the mammalian collecting duct. *Physiol Rev* 81:85–116, 2001



59. MCKENNA TJ, ISLAND DP, NICHOLSON WE, LIDDLE GW: The effects of potassium on early and late steps in aldosterone biosynthesis in cells of the zona glomerulosa. *Endocrinology* 103:1411-1416, 1978
60. HIMATHONGKAM T, DLUHY RG, WILLIAMS GH: Potassium-aldosterone-renin interrelationships. *J Clin Endocrinol Metab* 41:153-159, 1975
61. GANN DS, DELEA CS, GILL JR, *et al*: Control of aldosterone secretion by change in body potassium in normal man. *Am J Physiol* 207:104-108, 1964
62. MUTO S: Action of aldosterone on renal collecting tubule cells. *Curr Opin Nephrol Hypertens* 4:31-40, 1995
63. KOEPPEN B, GIEBISCH G: Cellular electrophysiology of potassium transport in the mammalian cortical collecting tubule. *Pflügers Arch* 405:S143-S146, 1985
64. PALMER LG, ANTONIAN L, FRINDT G: Regulation of apical K and Na channels and Na/K pumps in rat cortical collecting tubule by dietary K. *J Gen Physiol* 104:693-710, 1994
65. SANSOM SC, AGULIAN S, MUTO S, *et al*: K activity of CCD principal cells from normal and DOCA-treated rabbits. *Am J Physiol* 256:F136-F142, 1989
66. KASHGARIAN M, TAYLOR CR, BINDER HJ, HAYSLETT JP: Amplification of cell membrane surface in potassium adaptation. *Lab Invest* 42:581-588, 1980
67. JACKSON CA: Rapid renal potassium adaptation in rats. *Am J Physiol* 263:F1098-F1104, 1992
68. RABELINK TJ, KOOMANS HA, HENE RJ, DORHOUT MEES EJ: Early and late adjustment to potassium loading in humans. *Kidney Int* 38:942-947, 1990
69. FISHER KA, BINDER HJ, HAYSLETT JP: Potassium secretion by colonic mucosal cells after potassium adaptation. *Am J Physiol* 231:987-994, 1976
70. HAYES CP JR, MCLEOD ME, ROBINSON RR: An extrarenal mechanism for the maintenance of potassium balance in severe chronic renal failure. *Trans Assoc Am Physicians* 80:207-216, 1967
71. FRIZZELL RA, SCHULTZ SG: Effect of aldosterone on ion transport by rabbit colon in vitro. *J Membr Biol* 39:1-26, 1978
72. KEITH NM, OSTERBERG AE, BURCHALL HB: Some effects of potassium salts in man. *Ann Intern Med* 16:879-892, 1942
73. RABINOWITZ L, GREEN DM, SARASON RL, YAMAUCHI H: Homeostatic potassium excretion in fed and fasted sheep. *Am J Physiol* 254:R357-R380, 1988
74. RABINOWITZ L: Aldosterone and potassium homeostasis. *Kidney Int* 49:1738-1742, 1996
75. HENE RJ, KOOMANS HA, RABELINK AJ, *et al*: Mineralocorticoid activity and the excretion of an oral potassium load in normal man. *Kidney Int* 34:697-703, 1988
76. RABINOWITZ L: Model of homeostatic regulation of potassium excretion in sheep. *Am J Physiol* 254:R381-R388, 1988
77. WRIGHT FS, STRIEDER N, FOWLER NB, GIEBISCH G: Potassium secretion by distal tubule after potassium adaptation. *Am J Physiol* 221:437-448, 1971
78. SILVA P, BROWN RS, EPSTEIN FH: Adaptation to potassium. *Kidney Int* 11:466-475, 1977
79. MALNIC G, KLOSE R, GIEBISCH G: Micropuncture study of renal potassium excretion in the rat. *Am J Physiol* 206:674-686, 1964
80. SACKIN H, SYN S, PALMER LG, *et al*: Regulation of ROMK by extracellular cations. *Biophys J* 80:683-697, 2001
81. ZALUPS RK, STANTON BA, WADE JB, GIEBISCH G: Structural adaptation in initial collecting tubule following reduction in renal mass. *Kidney Int* 27:636-642, 1985
82. SCHERZER P, WALD H, CZACZKES JW: Na-K-ATPase in isolated rabbit tubules after unilateral nephrectomy and Na<sup>+</sup> loading. *Am J Physiol* 248:F565-F573, 1985
83. MUJAJIS SK, KURTZMAN NA: Regulation of renal Na-K-ATPase in the rat: effect of uninephrectomy. *Am J Physiol* 251:F506-F512, 1986
84. VEHASKARI VM, HERING-SMITH KS, KLAHR S, HAMM LL: Increased sodium transport by cortical collecting tubules from remnant kidneys. *Kidney Int* 36:89-95, 1989
85. FINE LG, YANAGAWA N, SCHULTZE RG, *et al*: Functional profile of the isolated uremic nephron: potassium adaptation in the rabbit cortical collecting tubule. *J Clin Invest* 64:1033-1043, 1979
86. RIMMER JM, HORN JF, GENNARI FJ: Hyperkalemia as a complication of drug therapy. *Arch Intern Med* 147:867-869, 1987
87. PONCE SP, JENNINGS AE, MADIAS NE, HARRINGTON JT: Drug-induced hyperkalemia. *Medicine (Baltimore)* 64:357-370, 1985
88. GENNARI FJ: Disorders of potassium metabolism, in *Suki and Massry's Therapy of Renal Diseases and Related Disorders* (3rd ed), edited by SUKI WN, MASSRY SG, Boston, Kluwer, 1998, pp 54-82