

data, a detailed mechanistic model of the allosteric conformational changes during DNA mismatch recognition by MutS is proposed.

1756-Pos Board B600

Working Mechanism of the Human Bloom's Syndrome Helicase

Máté Gyimesi, Kata Sarlós, Mihály Kovács.

Eötvös University, Budapest, Hungary.

Genome integrity is indispensable for unperturbed cell functioning. RecQ helicases play essential roles in genome maintenance. Mutations in three of the human RecQ isoforms (BLM, WRN or RECQL4) lead to severe diseases as the Bloom's, the Werner's and the Rothmund-Thomson syndromes, respectively, characterized by increased cancer predisposition and premature aging. Behind the serious genetic disorders stands the lack of repair mechanisms. BLM plays a crucial role in HR-based pathways by dissolving double Holliday-junctions and D-loops. The detailed working mechanism by which these "roadblock remover" functions are achieved is still unclear. We performed extensive kinetic, fluorescence spectroscopic and electrophoretic analyses to investigate the enzymatic cycle of BLM. In these studies wild-type and single tryptophan-containing BLM mutants were used. We demonstrate that BLM randomly and structure specifically binds DNA in the absence of nucleotide. ATP binds to DNA-bound BLM and induces a conformational change. ATP binding, hydrolysis and phosphate release occur rapidly and are followed by the rate limiting step of the cycle. This step is possibly a conformational change induced by DNA during translocation. BLM performs multiple ATPase cycles without dissociating from the DNA track. This results in the processive translocation activity of BLM. In contrast to other helicases (e.g. PcrA), BLM dissociates from the DNA strand at its 5'-end, thereby avoiding futile ATPase cycling. Our results emphasize the importance of investigating the basic working mechanism of different DNA helicases because these mechanisms may differ significantly. Moreover, understanding the basic working mechanism will greatly aid in understanding the complex functions of RecQ helicases.

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Direct Simulation Of Electron Transfer Reactions In DNA Radical Cations

Thomas B. Steinbrecher¹, Thorsten Koslowski², David A. Case¹.

¹Rutgers University, Piscataway, NJ, USA, ²Universitaet Freiburg, Freiburg, Germany.

The electron transfer properties of DNA radical cations are important in DNA damage and repair processes. Fast long-range charge transfer has been demonstrated experimentally, but the subtle influences that experimental conditions as well as DNA sequences and geometries have on the details of electron transfer parameters are still poorly understood.

In this work, we employ an atomistic QM/MM approach, based on a one-electron tight binding Hamiltonian and a classical molecular mechanics forcefield, to conduct nanosecond length MD simulations of electron holes in DNA oligomers. Multiple spontaneous electron transfer events were observed in 100 ns simulations with neighbouring adenine or guanine bases. Marcus parameters of charge transfer could be extracted directly from the simulations. The reorganisation energy lambda for hopping between neighbouring bases was found to be ca. 25 kcal/mol and charge transfer rates of $4.1 \times 10^{-9} \text{ s}^{-1}$ for AA hopping and $1.3 \times 10^{-9} \text{ s}^{-1}$ for GG hopping were obtained.

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Studies of the Translocation Mechanism of Hepatitis C Virus NS3 Helicase with Computationally Mutant Constructs

Yunde D. Shi, Jung-Chi Liao.

Columbia University, New York, NY, USA.

Hepatitis C virus (HCV) NS3 helicase unwinds double-stranded polynucleotide for HCV genome replication. Biochemical and single molecule studies have examined its enzymatic activity in depth, while the detailed translocation mechanism is still unclear. Our previous work has identified a list of hot-spot residues for its dynamic couplings and translocation by using an elastic network model (ENM). To further pinpoint key residues important for the polynucleotide movement, we used molecular dynamic (MD) simulation to study the conformational dynamics of NS3 helicase with computationally mutant constructs H293A, T324A, V432A and R461A. These mutations have been shown critical to the function of NS3 helicase by both experimental studies and ENM. We also simulated mutant constructs, T448A and P230A, which have only been predicted by ENM without experimental tests. Our results were consistent with experimental observations and suggested other important residues for polynucleotide translocation. Moreover, we have identified key hydrogen bond interactions between NS3 helicase and the polynucleotide for future experimental verification.

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Elucidation of the Mechanism of an Epigenetic Switch by Single-molecule Assays

Laura Finzi, Chiara Zurla, Carlo Manzo, David Dunlap.

Emory University, Atlanta, GA, USA.

The lambda bacteriophage epigenetic switch determines the growth lifestyle of the virus after infection of its host (*E. coli*). It is now clear that the switch consists of a ~2.3 kbp-long DNA loop mediated by the lambda repressor protein. Using tethered particle microscopy (TPM), magnetic tweezers and AFM, our laboratory has novel, direct evidence of loop formation and breakdown by the repressor, the first characterization of the thermodynamics and kinetics of the looping reaction and its dependence on DNA supercoiling and repressor non-specific binding. These *in vitro* data provide insight into the different possible nucleoprotein complexes and into the lambda repressor-mediated looping mechanism which leads to predictions for that *in vivo*. The significance of this work consists not only of the new insight into a paradigmatic epigenetic switch that governs lysogeny vs. lysis, but also the detailed mechanics of regulatory DNA loops mediated by proteins bound to multipartite operators and capable of different levels of oligomerization.

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Elasticity of Sub-micron DNA Molecules Studied with Axial Optical Tweezers

Yih-Fan Chen, David P. Wilson, Krishnan Raghunathan,

Jens-Christian Meiners.

University of Michigan, Ann Arbor, MI, USA.

Understanding the elasticity of sub-micron DNA molecules is important because many crucial biological structures and processes occur on this length scale. Using optical tweezers to manipulate DNA molecules, however, is difficult when molecules are shorter than about one micron. The reason is that conventional optical tweezers stretch molecules laterally in the focal plane of the microscope objective, a mode in which steric hindrances from the coverslip and other surface effects are substantial. To overcome the problem, we developed and calibrated an axial optical tweezers that makes this length scale accessible by stretching the molecule in the axial direction of the laser beam. By varying the laser intensity, different stretching forces were applied to the DNA molecule, and the axial position of the tethered microsphere was obtained from its diffraction pattern.

We measured the force-extension relationships of four short ds-DNA molecules, which are 1298 bp-, 662 bp-, 390 bp-, and 247 bp-long, using the axial optical tweezers. Using a modified worm-like chain (WLC) model for the extended DNA molecule that incorporates excluded-volume entropic effects from the coverslip and microsphere are taken into account, we obtained effective persistence lengths and excluded-volume forces for these molecules. The fitted values for the persistence length decrease with the contour length of the DNA, which is qualitatively consistent with observations by Seol et al. on longer, micron- and sub-micron sized constructs (Seol 2007). Moreover, the excluded-volume forces are close to the theoretical predictions by Segall et al (Segall 2006).

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Structure Conversion Of Human Telomeric Sequence Studied By Single-molecule Tethered Particle Motion

Jen-Fei Chu^{1,2}, Ta-Chau Chang^{1,3}, Hung-Wen Li³.

¹Institute of Atomic and Molecular Sciences, Academia Sinica, Taipei, Taiwan, ²Department of Chemistry, National Taiwan Normal University, Taipei, Taiwan, ³Department of Chemistry, National Taiwan University, Taipei, Taiwan.

Telomeres contain G-rich tandem repeats of single-stranded DNA sequences at 3' tail. The G-rich sequences can be folded into a secondary structure named G-quadruplexes by Hoogsteen base pairing in the presence of monovalent cations (such as Na⁺, K⁺). The folding of telomeric DNA into the G-quadruplexes may inhibit telomerase activity for the proliferation of cancer cells. Moreover, the change of a quadruplex conformation may play an important role in biological effect. Thus, understanding structure conversion between the folded and unfolded G-quadruplex structures, and how the structure conversion is mediated by ions, its anti-sense sequence and its stabilizers are important to telomere biology. Here, we have directly monitored the conversion between the folded and unfolded structures in human telomeric AGGG(TTAGGG)₃ sequence by the single-molecule tethered particle motion (TPM) method. TPM method monitors the DNA length change caused by the G-quadruplex formation, and allows us to monitor the conversion mechanism