

A High-Resolution Spatiotemporal Atlas of Gene Expression of the Developing Mouse Brain

Carol L. Thompson,^{1,6} Lydia Ng,^{1,6} Vilas Menon,¹ Salvador Martinez,^{4,5} Chang-Kyu Lee,¹ Katie Glattfelder,¹ Susan M. Sunkin,¹ Alex Henry,¹ Christopher Lau,¹ Chinh Dang,¹ Raquel Garcia-Lopez,⁴ Almudena Martinez-Ferre,⁴ Ana Pombero,⁴ John L.R. Rubenstein,² Wayne B. Wakeman,¹ John Hohmann,¹ Nick Dee,¹ Andrew J. Sodt,¹ Rob Young,¹ Kimberly Smith,¹ Thuc-Nghi Nguyen,¹ Jolene Kidney,¹ Leonard Kuan,¹ Andreas Jeromin,¹ Ajamete Kaykas,¹ Jeremy Miller,¹ Damon Page,¹ Geri Orta,¹ Amy Bernard,¹ Zackery Riley,¹ Simon Smith,¹ Paul Wohnoutka,¹ Michael J. Hawrylycz,^{1,*} Luis Puelles,³ and Allan R. Jones¹

¹Allen Institute for Brain Science, Seattle, WA 98103, USA

²Department of Psychiatry, Rock Hall, University of California at San Francisco, San Francisco, CA 94158, USA

³Department of Human Anatomy and Psychobiology, University of Murcia, E30071 Murcia, Spain

⁴Instituto de Neurociencias UMH-CSIC, A03550 Alicante, Spain

⁵Centro de Investigacion Biomedica en Red de Salud Mental (CIBERSAM) and IMIB-Arrixaca of Instituto de Salud Carlos III, 30120 Murcia, Spain

⁶Co-first author

*Correspondence: mikeh@alleninstitute.org

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SUMMARY

To provide a temporal framework for the genoarchitecture of brain development, we generated *in situ* hybridization data for embryonic and postnatal mouse brain at seven developmental stages for ~2,100 genes, which were processed with an automated informatics pipeline and manually annotated. This resource comprises 434,946 images, seven reference atlases, an ontogenetic ontology, and tools to explore coexpression of genes across neurodevelopment. Gene sets coinciding with developmental phenomena were identified. A temporal shift in the principles governing the molecular organization of the brain was detected, with transient neuromeric, plate-based organization of the brain present at E11.5 and E13.5. Finally, these data provided a transcription factor code that discriminates brain structures and identifies the developmental age of a tissue, providing a foundation for eventual genetic manipulation or tracking of specific brain structures over development. The resource is available as the Allen Developing Mouse Brain Atlas (<http://developingmouse.brain-map.org>).

INTRODUCTION

The diversity of cell types in the brain presents an immense challenge toward understanding cellular organization, connectivity, and function of this organ. The objective definition of cell type remains elusive but should integrate molecular, anatomic, morphological, and physiological parameters. At both a large and small scale, neuroscientists have flocked to genetic strategies that depend upon known molecular markers to label adult cell types for the purpose of isolating or manipulating specific

populations (Siegert et al., 2012; Sugino et al., 2006). However, achieving a fine resolution of cell subtypes will probably require combinatory or intersectional strategies due to the lack of absolute specificity of any single gene marker for a given cell type. Developmental neurobiologists have used careful descriptive analyses and genetic fate mapping for over a decade to specify the developmental origin of cell types, typically utilizing an intersectional strategy to map the fate of cells produced at a specified time from a particular anatomic domain (Joyner and Zervas, 2006). In the retina, a transcription factor (TF) code has been deduced for each branch of the retinal cell lineage (Agathocleous and Harris, 2009; Livesey and Cepko, 2001) and this code is evident even in the adult differentiated neurons (Siegert et al., 2012). The success of creating meaningful definitions of cell types may ultimately rely on a combination of classification metrics that include both terminal molecular characteristics as well as their topological developmental origin.

Morphogenesis and functional development of the mammalian CNS occur via mechanisms regulated by the interaction of genes expressed at specific times and locations during development (Rubenstein and Rakic, 2013; Sanes et al., 2012). Understanding this temporal and regional complexity of gene expression over brain development will be critical to provide a framework to define neuroanatomical subdivisions and the component cell types. To this end, we have generated an extensive data set and resource that provides spatial and temporal profiling of ~2,100 genes across mouse C57Bl/6J embryonic and postnatal development with cellular-level resolution (<http://developingmouse.brain-map.org>). Genes were surveyed by high-throughput *in situ* hybridization (ISH) across seven embryonic and postnatal ages (embryonic day 11.5 [E11.5], E13.5, E15.5, E18.5, postnatal day 4 [P4], P14, and P28), in addition to P56 data available in the Allen Mouse Brain Atlas. This developmental survey comprises 18,358 sagittal and 1,913 coronal ISH experiments, displayed online at 10× resolution and are downloadable via XML. From a neuroanatomical perspective, the Allen Developing Mouse Brain Atlas defines a number of

CNS subdivisions (described in 2D atlas plates and 3D structural models) based on an updated version of the prosomeric model of the vertebrate brain (Puelles et al., 2012; Puelles and Rubenstein, 2003). Furthermore, a novel informatics framework enables navigation of expression data within and across time points. In addition to stage-specific novel reference atlases, the resource provides an innovative ontogenetic ontology of the full brain with over 2,500 hierarchically organized names and definitions, and 434,946 sections of high-resolution spatially and temporally linked ISH data, offering rapid access and a range of visualization and analysis tools.

The chosen stages were intended to survey diverse developmental mechanisms, including regional specification, proliferation, neurogenesis, gliogenesis, migration, axon pathfinding, synaptogenesis, cortical plasticity, and puberty. The genes selected include: (1) ~800 TFs representing 40% of total TFs, with nearly complete coverage of homeobox, basic helix-loop-helix, forkhead, nuclear receptor, high mobility group, and POU domain genes; (2) neurotransmitters and their receptors, with extensive coverage of genes related to dopaminergic, serotonergic, glutamatergic, and GABA-ergic signaling, as well as of neuropeptides and their receptors; (3) neuroanatomical marker genes delineating regions or cell types throughout development; (4) genes associated with signaling pathways relevant to brain development including axon guidance (~80% coverage), receptor tyrosine kinases and their ligands, and Wnt and Notch signaling pathways; (5) a category of highly studied genes coding for common drug targets, ion channels (~37% coverage), G protein-coupled receptors (GPCRs; ~7% coverage), cell adhesion genes (~32% coverage), and genes involved neurodevelopmental diseases, which were expected to be expressed in the adult brain or during development (Table S1 available online). A smaller set of genes was surveyed in “old age” (18–33 months).

Analyses of these data identified molecular signatures associated with key developmental events with precise spatiotemporal regulation. These signatures revealed a shift in the organizing principles governing the molecular profiles of brain regions over development, with the coexistence of both dorsoventral (DV) or longitudinal plate-based, and anteroposterior (AP) or neuromere-based organization strongest at E13.5 leading to areal (progenitor domain)-based organization. Finally, by focusing on TFs, unique combinatorial codes were found that precisely define most brain structures and even pinpoint developmental age, a potential starting point to investigate both how regions are specified as well as how they acquire unique functional properties.

RESULTS

New Developmental Reference Atlases

To provide a consistent anatomical context for analysis of ISH data, we created seven reference atlases spanning E11.5 to P56 (161 annotated plates and 1,898 supporting reference images; Figure 1). The reference atlases used an ontological approach that classifies brain structures based upon their orthogonal areal neuroepithelial origin in the wall of the neural tube (intersection of fundamental neuromeric and longitudinal zonal units), employing a topological and ontogenetic viewpoint

to register the emergence of both transient and definitive nuclear or cortical cell populations in the mantle layer, the final location of postmitotic, terminally differentiated neurons (Puelles and Ferran, 2012; Puelles et al., 2012). Tangentially migrated structures (e.g., pontine nuclei) are classified by postmigratory position. Thus, the reference atlas drawn for the adult contains developmental and morphological concepts that make it distinct, in terms of nomenclature and classification, from that drawn for the Allen Mouse Brain Atlas (Dong, 2008).

The present ontogenetic ontology (Puelles et al., 2013) has 13 levels of anatomic classification. Early levels 1–3 include definition of protosegments (e.g., forebrain), followed by neuromeric AP subdivisions (e.g., prosomeres). Basic DV subdivisions are defined next, including the alar-basal boundary, plus roof, and floor plates (levels 4 and 5). Levels 6–8 cover finer areal regionalization into realistic progenitor domains with known differential fates. Stratification refers first to the distinction between ventricular and mantle zones (level 9) and second to superficial, intermediate, and periventricular transient strata (level 10) of the mantle zone. Adult brain nuclei and other associated structures (tracts, commissures, circumventricular organs, and glands) largely reside in the mantle zone and are represented at levels 11–13.

Automated and Manual Annotation of the Data Facilitates Navigation of Spatial and Temporal Information

For ISH experiments, the sampling density of tissue sections was scaled by specimen size and age, ranging from 80 μm to 200 μm (Supplemental Experimental Procedures). Most genes (63%) were expressed at all ages and 10% were not expressed in the brain at any stage. The remaining genes (27%) were temporally specific, with 19% exhibiting delayed activation across this time course, potentially associated with terminally differentiated cellular phenotypes (e.g., GPCRs and ion channels; Figures S1E and S1F) and 4% of the genes expressed only at early stages (Figures S1A and S1B; Table S2).

Events that shape the development of the brain from an undifferentiated neuroepithelium populated by neural precursors to a mature, functioning organ occur at different times in different regions (Rubenstein and Rakic, 2013; Sanes et al., 2012), and the ability to parse out specific spatial and temporal patterns of gene expression is highly desirable. The standardized generation of ISH data supported the development of a systematic and automated informatics-based data processing pipeline (Ng et al., 2007) for navigation and analysis of this large and complex data set, shown in Figures 2A–2C. Tissue sections from each ISH experiment were aligned to age-matched 3D brain models assembled from 2D reference atlas annotation, and ISH signal was quantified across a voxel grid whose dimensions corresponded to the sampling density of the ISH. The ISH data for each gene can be analyzed in a 3D context as a pure voxel grid or can be contextualized with the neuroanatomic reference atlas. In the online application, this processing supports expression summary statistics, anatomic and temporal-based search, and other advanced search options, all freely available. Using Anatomic Search, for a given age, a user can identify genes enriched within a selected brain structure. Results are rank ordered based upon their selectivity for that structure by comparing

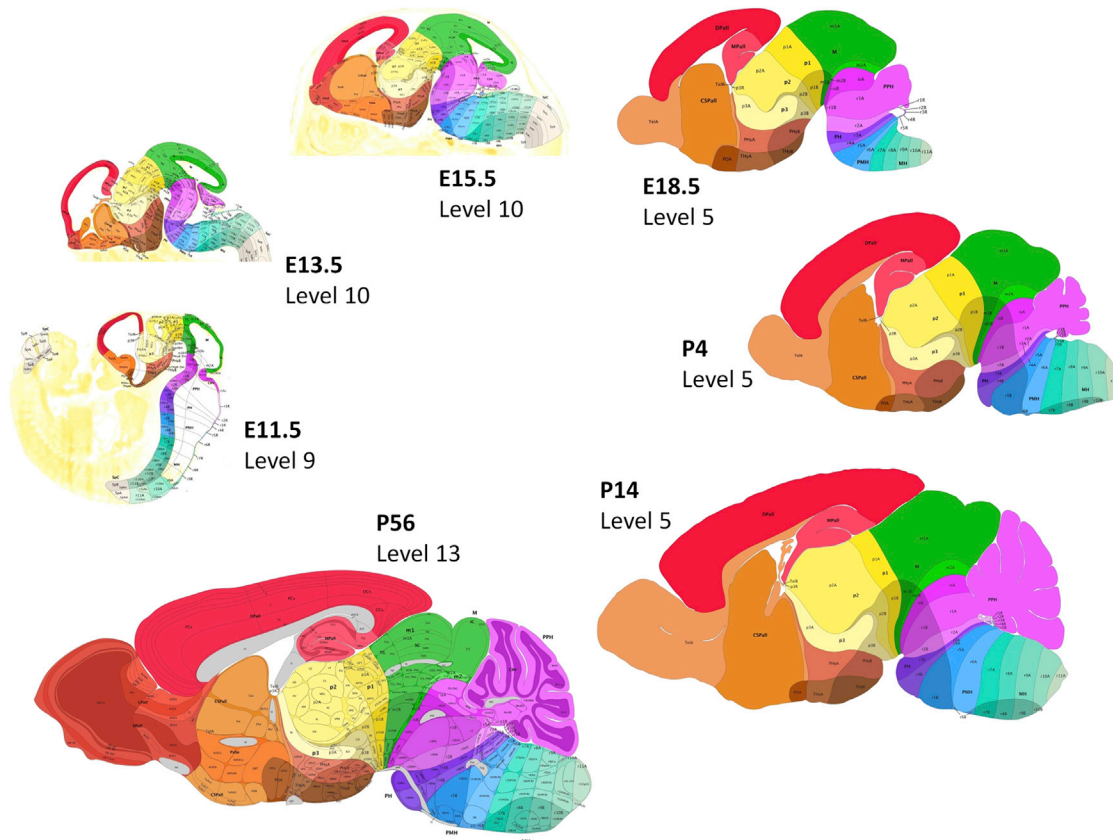


Figure 1. Reference Framework for the Allen Developing Mouse Brain Atlas

Representative reference atlas plates from seven developmental ages surveyed in the project are shown. Because P28 and P56 time points are indistinguishable from a neuroanatomic standpoint, the P56 Nissl images used in the reference atlas for the Allen Mouse Brain Atlas were also annotated using the developmental ontology and are supplied as a reference for both P28 and P56 ISH data.

expression in the target brain structure to expression in adjacent brain structures. With Temporal Search, users find genes showing temporal enrichment for a given structure. In this case, results are rank ordered based upon their selectivity for expression at a given age in comparison to all other ages. These two search options are orthogonal in that Anatomic Search ignores temporal enrichment and Temporal Search ignores anatomic enrichment.

Gene coexpression can suggest shared gene function (Hughes et al., 2000; Nayak et al., 2009), protein interactions (Jansen et al., 2002), and common regulatory pathways (Allocco et al., 2004; Segal et al., 2003). We have previously demonstrated that gene-to-gene spatial correlations in adult mouse brain can identify genes belonging to specific functional classes (Hawrylycz et al., 2011) and cell types, such as astrocytes or oligodendrocytes (Lein et al., 2007). An online tool (NeuroBlast) allows identification of genes whose spatial expression patterns are correlated to that of a given gene of interest. The expression pattern of each gene is summarized by a voxel grid encompassing the brain. Pearson's correlation coefficient is calculated between pairs of genes, over the corresponding voxel sets across the neural primordium. Correlation can also be restricted to a pre-defined anatomic structure. For example, *Wnt3a*, a ligand in the

Wnt signaling pathway, is selectively expressed in the E13.5 cortical hem, a transiently identifiable brain structure that regulates hippocampal development; *Wnt3a* mutant mice fail to generate a recognizable hippocampus (Lee et al., 2000). Pearson's correlation coefficient between *Wnt3a* and the entire gene set across all voxels in the telencephalic vesicle was used to identify genes with spatially comparable expression. The top search returns include eight Wnt signaling genes: *Wnt3a*, *Wnt2b*, *Dkk3*, *Axin2*, *Rspo1*, *Rspo3*, *Nkd1*, and *Rspo2* (DAVID [Huang et al., 2009a, 2009b]). Other highly correlated genes, *Jam3*, *Dmrt3*, *Lmx1a*, *Foxj1*, and *Id3*, could represent candidates for interactions with Wnt signaling or pallial patterning (Figure 2D).

The NeuroBlast tool can also identify coexpression relationships between a TF and potential downstream targets. In the simplest scenario, a TF would be activated in a cell type and then collaborate with other TFs to activate given enhancer/repressor DNA sequences of its target genes. Positively regulated target genes should be expressed shortly after, and over time, the spatial expression of the TF should partly match the expression of its downstream targets. We identified a set of 22 genes highly correlated with the TF *Pou4f1*, which is expressed in the habenula (Figure S2). Seven of the top genes are presumed to be downstream of *Pou4f1*, as shown by altered expression

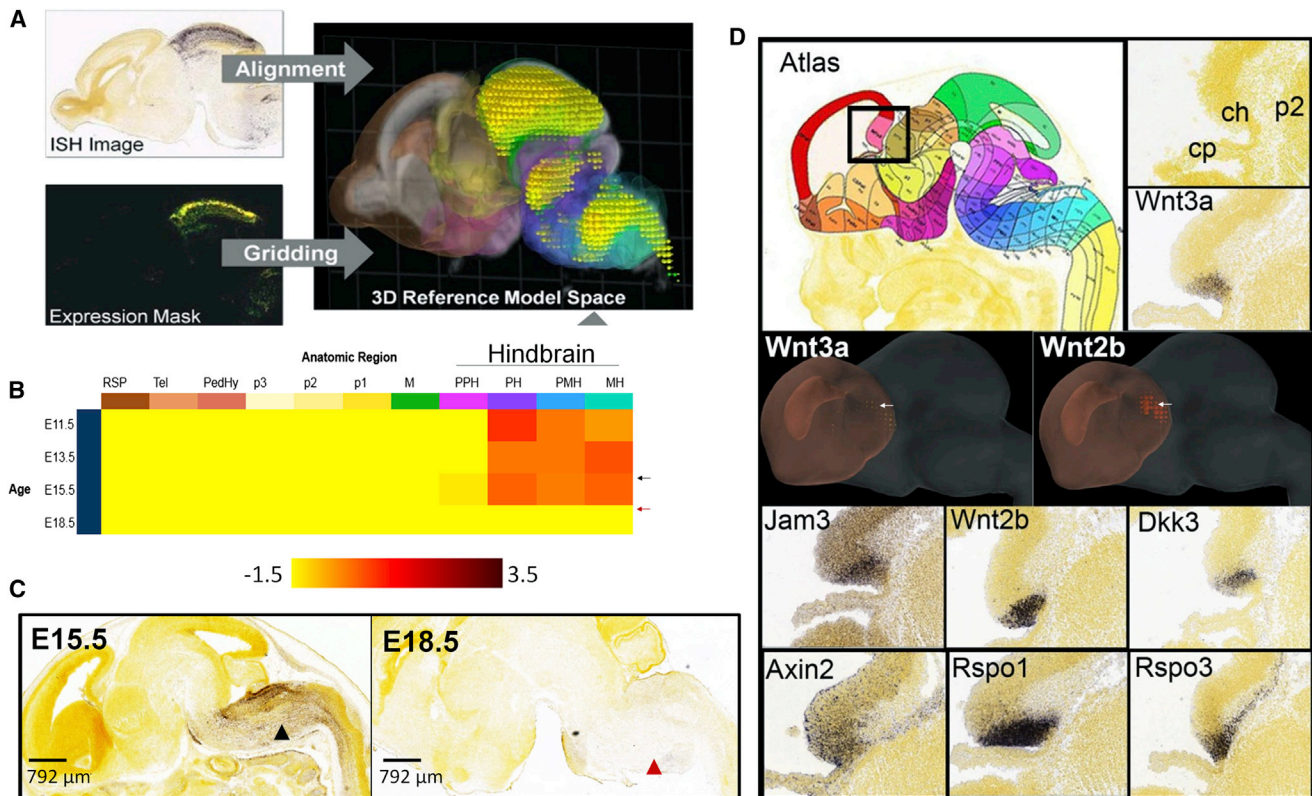


Figure 2. Automated Informatics-Based Pipeline for ISH Image Analysis

(A) Image preprocessing, alignment, signal quantification, and summary are provided by a suite of automated modules. An “Alignment” module registers ISH images to the common coordinates of a 3D reference space (Supplemental Experimental Procedures). The “Gridding” module produces an expression summary in 3D for computational expression analysis. The “Unionize” module generates anatomic structure-based statistics by combining grid voxels with the same 3D structural label. ISH for *Tcfap2b* is shown at E18.5 with its expression mask and 3D expression summary.

(B and C) Expression summary (B) and ISH (C) for *Hoxa2*. PH, pontine hindbrain; PMH, pontomedullary hindbrain; and MH, medullary hindbrain (last three columns in Expression Summary in B).

(D) *Wnt3a* was used as a seed gene in NeuroBlast to find other genes in the cortical hem at E13.5. The E13.5 reference atlas is shown; the black box indicates the areas shown in the histology images. The area containing the cortical hem is labeled in a reference HP Yellow stained image (ch, cortical hem; p2, prosomere 2; cp, choroid plexus). 3D images of the atlas structures overlaid with gene expression are shown using the Brain Explorer 2 3D viewer, where gray represents the entire brain, and orange represents the telencephalic vesicle (Tel) that was used to constrain the search. Voxels found to have gene expression are highlighted, appearing as “bubbles.” Arrows point to the cortical hem. ISH for genes identified by NeuroBlast are shown (sagittal plane; see also Figures S2, S3, and Table S1).

levels in a knockout model of *Pou4f1* (*Efcfb2*, *Etv1*, *Chrna3*, *Nr4a2*, *Dcc*, *Sncg*, *Wif1* [Quina et al., 2009]). These methods can be used to identify and establish a temporal hierarchy of expression of genes activated downstream of any TF.

Although sophisticated image-processing tools were developed to annotate ISH expression data, the small size of some brain structures relative to adjacent large empty ventricles as occurs in the E11.5 brain presents challenges for automated tissue registration and analysis. Therefore, expert-guided manual annotation of the ISH data was performed on the four prenatal ages (E11.5 through E18.5) to accurately assign gene expression calls and metrics to specific atlas-defined brain structures and is available online (Figure S3).

Mapping Gene Expression to Developmental Phenomena

Analyzing temporal peaks of gene expression over development could identify major developmental phenomena associated with

a specific brain structure or age. RNA expression levels were investigated for seven functional gene categories across 13 brain regions at 6 ages (E13.5–P28; Figure 3A). These categories relate to key developmental events such as regional patterning, neurogenesis, differentiation, migration, axogenesis, and synaptogenesis, in which developmental timing may vary throughout the CNS (Rubenstein and Rakic, 2013; Sanes et al., 2012). For TFs, two primary peaks are evident, with one peak in the E13.5 midbrain. The Temporal Search tool identified five bHLH genes (*Tal2*, *Mxd3*, *Tcfe2a*, *Nhlh2*, and *Neurog1*), and four homeobox genes (*Pou3f3*, *Lhx1*, *Pou2f2*, and *Pou4f3*) within the top 20 returns for genes enriched at E13.5 in the midbrain. Most of these bHLH genes were expressed specifically in the ventricular or periventricular strata of the midbrain wall (Figure 3B), coincident with the timing of peak neurogenesis (Clancy et al., 2001), suggesting a role in growth and generation of neurons in this region. *Tcfe2a*, for example, maintains stem cells in an undifferentiated state (Nguyen et al., 2006) and is essential for midbrain development

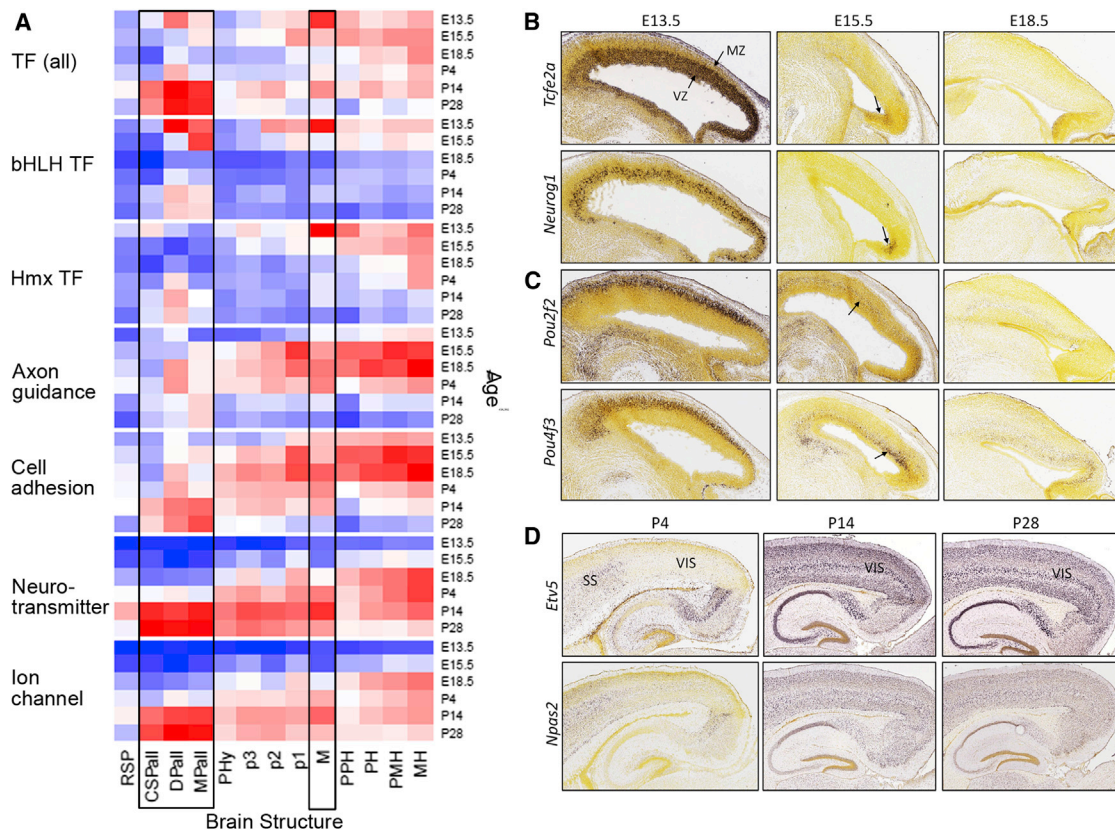


Figure 3. Anatomic and Temporal Expression by Gene Class

(A) Normalized average expression level for gene classes by age and anatomic region. Expression level is calculated as in [Experimental Procedures](#) and normalized across gene class with higher expression levels in red, lower in blue. Abbreviations are the following: genes: bHLH, basic helix loop helix; Hmx, homeobox; structures: RSP, rostral secondary prosencephalon; CSPall, central subpallium; DPall, dorsal pallium/isocortex; MPall, medial pallium; PHy, peduncular hypothalamus; p3, prosomere 3 (prethalamus and prethalamic tegmentum); p2, prosomere 2 (thalamus and thalamic tegmentum); p1, prosomere 1 (pretectum and pretectal tegmentum); M, midbrain; PPH, preoptone hindbrain; PH, pontine hindbrain; PMH, pontomedullary hindbrain; MH, medullary hindbrain. (B–D) Genes identified using online Temporal Search feature. (B and C) Temporal Search for genes enriched in E13.5 midbrain identified bHLH genes expressed in ventricular (VZ) and periventricular zones (B), and homeobox genes in mantle zone (MZ) (C). (D) Temporal Search for genes enriched at P28 in the telencephalic vesicle. Although these genes are expressed in the P4 somatosensory cortex (SS), they exhibit striking lack of expression in visual cortex (VIS). These genes are expressed throughout neocortex after eye opening (P14 and P28; see also [Figure S1](#) and [Table S2](#)).

in zebrafish ([Kim et al., 2000](#)). *Neurog1* marks initiation of neurogenesis and promotes cell cycle exit ([Bertrand et al., 2002](#)), consistent with its expression in the periventricular zone, where postmitotic neurons exit into the mantle zone. In contrast, homeobox genes that peak in the E13.5 midbrain were primarily enriched in mantle zone, which contains postmitotic maturing neurons, suggesting a role for these genes in differentiation or layering ([Figure 3C](#)). Consistent with this hypothesis, *Pou2f2* (*Oct2*) is known to induce neuronal differentiation ([Theodorou et al., 2009](#)), and a close family member of *Pou4f3* (*Brn3c*) regulates the transition from neurogenesis to terminal differentiation ([Lanier et al., 2009](#)). The distinct stratification of bHLH and homeobox genes suggest that these TF classes are utilized in the same manner as in the retina, in which the bHLH activators regulate layer specificity of retinal cell types but not neuronal fate, but the homeobox genes regulate neuronal subtype specification ([Hatakeyama and Kageyama, 2004](#)). Later, the midbrain shows peak expression of axon guidance and cell adhesion genes

around birth, followed by expression of neurotransmitter-related genes and ion channels in postnatal ages, consistent with the expected progression of neural development.

A second expression peak for TFs was identified in dorsal pallium (isocortex), medial pallium (hippocampus), and central subpallium (striatum/pallidum) at P14 and P28, the period when activity-dependent processes are sculpting the brain's wiring diagrams. A Temporal Search for genes enriched at P28 in the telencephalic vesicle (inclusive of these regions) reveals enrichment for immediate-early genes (*Fos*, *Egr1*, *Homer1*, *Arc*, *Ets2*, *Dusp14*, *Hlf*, *Bcl6*, *Etv5*, and *Per1*), many of which are TFs. Immediate-early genes are rapidly induced following stimuli, believed to reflect neuronal activation. A subset of these genes is induced in the visual cortex and striatum by sleep deprivation ([Thompson et al., 2010](#)), presumably due to increased visual stimulation during sleep deprivation in the light phase. Many immediate-early genes appear to be strongly enriched in visual cortex starting at P14. For instance, expression of *Etv5* and *Npas2* is not detected

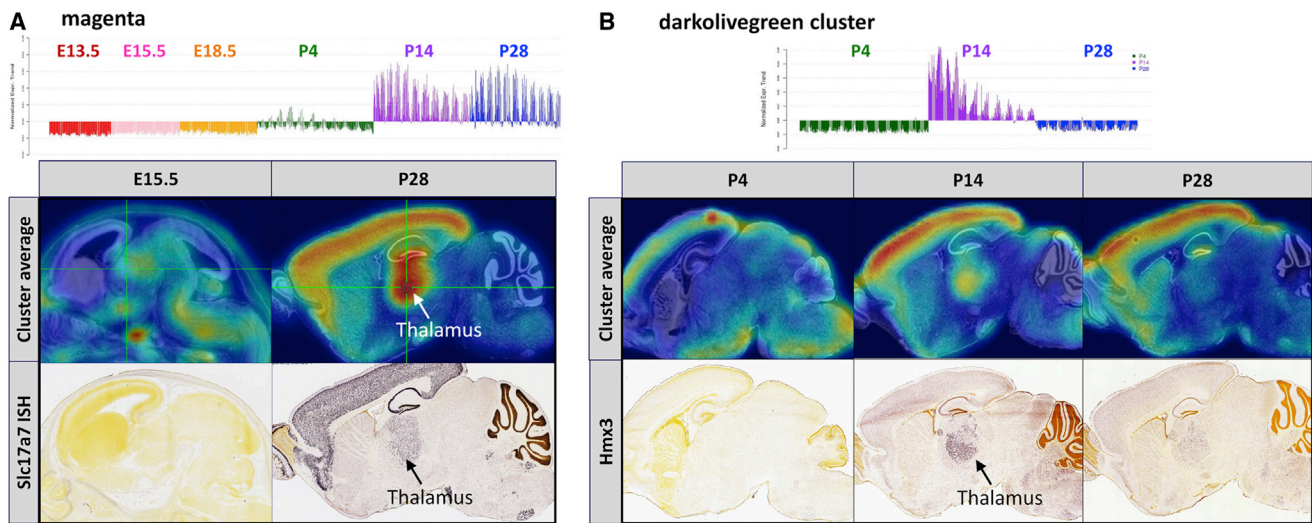


Figure 4. Temporal Expression Patterns in the Diencephalon Identified by WGCNA

(A) Voxalized expression data from six ages were used to cluster genes by WGCNA; the magenta cluster is a temporally regulated cluster. The plot (top) shows the eigengene for the cluster across individual voxels at each age. Underneath, the top panels illustrate average expression levels at the indicated stages. The ISH for a gene example is shown in the bottom panels.

(B) Voxalized expression data from postnatal ages were used to cluster genes by WGCNA. The dark olive green cluster shows strong upregulation at P14 (see also Figures S4, S5, and S6 and Tables S3, S4, and S5).

in the P4 visual cortex (note expression in surrounding cortical regions in Figure 3D), whereas by P14 and P28 visual cortex expression is prominent. Thus, *Etv5* and *Npas2* expression may reflect activity-induced transcription resulting from accrued visual input to the visual cortex after eye opening.

For other gene categories, brainstem exhibits peak expression at mid to late embryonic stages, whereas telencephalic regions exhibit late postnatal peak expression. This trend is observed for the axon guidance, cell adhesion, neurotransmitter, and ion channel gene categories and parallels the timing of maturation of these regions. Neurotransmitter and ion channel classes represent late differentiation variables of the neuronal phenotypes; genes in these categories exhibit very low expression at the earliest age, E13.5, across all brain regions.

To illustrate clusters of genes with temporal coexpression patterns, we focused on the diencephalon. We clustered genes based on their coexpression patterns in voxels annotated for the diencephalon. However, when genes were clustered at each age, no significant coherence was observed across the entire time period (data not shown), although clear differences were observed between embryonic and postnatal ages. This observation led us to group the time points into three periods, for independent analysis of coexpression: “embryonic” (E13.5, E15.5, and E18.5), “postnatal” (P4, P14, and P28), and “all” (E13.5, E15.5, E18.5, P4, P14, and P28). In order to extract expression trends over these time periods, weighted gene coexpression network analysis (WGCNA) was used to group genes into clusters with coexpression patterns across the data set (Zhang and Horvath, 2005). The eigengene of each cluster, a measure of the average expression of all the genes within a cluster, represents an expression trend over time observed for the diencephalon (Horvath, 2011).

In most cases, the clusters were comprised of genes delineating particular spatially discrete, contiguous sets of voxels. Example modules are shown for “embryonic” period clustering (Figure S4). The temporal pattern of expression in the diencephalon is plotted ordering the module eigengenes from E13.5 to E18.5 (Figures S4B–S4K). Because the expression data are comprised of voxels with known anatomic location, the average expression pattern of the cluster can also be plotted back into a 3D model to determine the spatial expression pattern of each cluster. Clustering results are available for “embryonic,” “postnatal,” and “all” (Figures S4–S6) and gene ontology results for a subset of modules (Tables S3, S4, and S5, respectively). The two most frequent anatomic expression patterns in the diencephalon identified by WGCNA clustering across any time frame were expression in the thalamus (Figures S4B–S4D and S5B–S5F) and subsets of diencephalic regions that specifically exclude the thalamus (Figures S4E, S4F, and S5G). The thalamus clusters were enriched in metabotropic glutamate receptor group I pathways, ion transport, and synaptic transmission genes. In some cases, specific nuclei of the thalamus were identified, e.g., the parafascicular nucleus or the posterior ventromedial nucleus (Figures S4C, S4D, and S6E).

Temporal expression patterns can also be identified using the WGCNA approach. When examining the “all” period that spans from E13.5 to P28, two clusters are identified in which genes have strong upregulation of expression in the diencephalon at P14 and P28 (Figure S6). In the magenta cluster (Figure 4A), GO analysis identifies enrichment of genes in the metabotropic glutamate receptor group III pathway ($p = 0.028$; e.g., *Slc17a7*, *Grm4*, *Slc17a6*, *Grin2b*, *Grin2c*, *Grm1*, *Slc1a1*). Examining the postnatal (P4, P14, and P28) cluster identified a set of genes (*Pip1*, *Cnp*, *Mbp*, *Mog*, *Mobp*, and *Olig1*) strongly upregulated

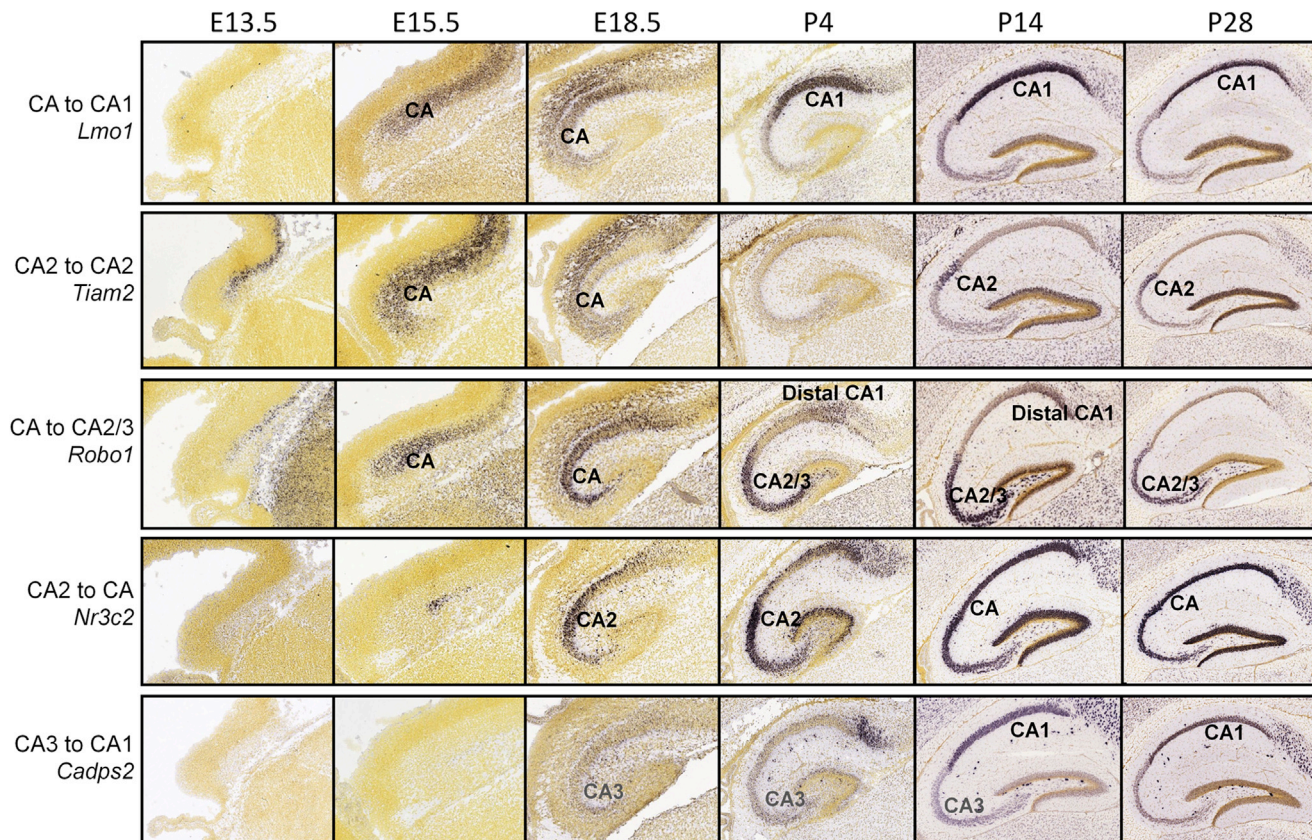


Figure 5. Changes in Specificity of Gene Markers for Hippocampal Fields

The top three genes are expressed initially in the entire CA pyramidal layer in the embryo and eventually display specificity in only one CA field by P28. *Nr3c2* is expressed in a subset of cells at E15.5, with enrichment in CA2 around birth, but is expressed throughout CA by the adult. Finally, *Cadps2* exhibits transient weak expression in CA3 prior to strong CA1 staining in the adult.

at P14 and P28, including genes heavily enriched in oligodendrocytes (Figure S4, cluster grey60). Although oligodendrocytes are produced as early as E18.5 (Hardy and Friedrich, 1996), these data show that several well-known oligodendrocyte genes do not exhibit widespread distribution in the diencephalon until P14. A particularly intriguing temporal expression pattern is the occurrence of strong, thalamus-specific expression of predominantly TF genes at P14 (Figure 4B), a phenomenon that is either weak or undetectable at P4 and P28. The timing may coincide with eye opening and the initial reception of visual stimulation by the thalamus, occurring around P12–P13, or with other delayed synaptogenesis-related developmental events. Note that thalamic nuclei corresponding to visual, somatosensitive, somatomotor, and auditory systems are represented in this cluster.

Molecular Cohesion of Anatomic Regions over Development

An obvious application of this data set is to find gene markers selective for specific structures over time, to assess the earliest appearance of that structure in the embryo as well as to characterize how sets of developmentally important genes may change over time. Numerous markers were identified from the Allen Mouse Brain Atlas that subdivide the hippocampus CA region

into fields CA1, CA2, and CA3. These genes exhibited complex spatiotemporal expression. First, many markers were not apparent before P14 (e.g., CA2 markers *Sostdc1*, *Stard5*, and *Fgf5*; CA1 markers *Plekhhg1*, *Sstr4*, *Htr1a*, and *Igf1bp4*; available online) and may relate to terminal differential functions of these CA fields rather than to developmental identity. Other markers are expressed in the full CA pyramidal layer at earlier ages, becoming regionalized at later stages (Figure 5), or are regionally restricted at E18.5/P4 and become widely expressed across the CA by P28 (e.g., *Nr3c2*; Figure 5). Other genes show changing specificity, such as *Cadps2*, which is expressed in CA3 at age P4, in both CA1 and CA3 at P14, and is CA1 specific at P28. Within a brain region, a variety of events may drive dynamic or transient gene expression and could provide intriguing clues about the process of development within a given region. In order to provide users with another mode of navigating spatiotemporal gene expression, we created a new version of Anatomic Gene Expression Atlas (AGEA) that incorporates developmental age.

Gene expression profiling has been invaluable for refining our understanding of neuroanatomy and development, insofar as gene expression correlations can recapitulate known functional divisions of the brain, provide a hint of their embryological origin (Ng et al., 2009; Zapala et al., 2005), and serve as fiducials to

compare particular brain structures across species and time (Puelles et al., 2000). The original AGEA released as part of the Allen Mouse Brain Atlas was a powerful tool to identify correlated voxels at age P56 and find corresponding genes. In the Allen Developing Mouse Brain Atlas, AGEA has undergone a significant advance to allow users to explore spatiotemporal genetic relationships and identify voxels in the brain that show highly correlated gene expression across different ages. Thus, the molecular signatures of brain regions (Puelles and Ferran, 2012) can be used to follow the progressive development of anatomic domains as a surrogate for actual fate-mapping experiments.

A correlation map for each fixed age (Ng et al., 2009) is generated by evaluating each seed voxel against every other target voxel in the 3D reference model. The values obtained across the voxels of each map represent the Pearson correlation coefficients between the seed voxel and every other location over the set of 2,000 genes. Correlations are also calculated between each seed voxel and target voxels of adjacent ages, resulting in a combined total of 265,621 online 3D browsable maps. These correlation maps allow visualization of voxels that share a correlated transcriptome profile and typically identify adjacent voxels that reflect local neuroanatomy.

Furthermore, the user can view correlation maps that “walk” across the different ages. In this technique, the highest correlates of a chosen voxel are identified at adjacent ages, thereby enabling a type of anatomic “virtual molecular fate map.” By selecting an initial seed voxel at P28, the user can navigate across time to find the highest correlated voxel at P14, subsequently P4, then E18.5, and so on to provide a “reverse molecular fate map.” A “forward molecular fate map” is similarly constructed by selecting an initial seed voxel at E13.5 and moving forward in time. The thalamus, the olfactory bulb, and cortex each exhibit coherent and identifiable anatomic precursors as shown in reverse correlation maps traced from P28 to E13.5, highlighting the molecularly consistent anatomic origin of these structures (Figure 6A). Once such spatiotemporal correlations are established, the AGEA application lists the most significant correlated genes.

To illustrate a virtual forward molecular fate map, we selected an initial seed voxel in the E13.5 ganglionic eminences. The highest correlated voxel at the next oldest age was calculated and automatically selected in stepwise fashion. The lateral ganglionic eminence (LGE) is a source of striatal projection neurons, and the medial ganglionic eminence (MGE) is a source of pallidal, diagonal, and preoptic projection neurons, as well as of striatal and cortical interneurons; the latter migrate tangentially from the MGE to the cortex and intersperse across the cortical layers among the glutamatergic neurons. In the forward map, a seed point chosen in the subventricular zone (SVZ) of the MGE at E13.5 correlates highly to the P4 cortical SVZ and rostral migratory stream, both of which undergo late neurogenesis and tangential migration; they likely share part of their transcriptomic profiles with the neurogenic subpallial SVZ. A seed point in the E13.5 LGE results in a set of highly correlated voxels in the striatum by P4, consistent with current knowledge about the origin and local radial layering of these neurons. These techniques provide a method for understanding the molecularly defined precursor domains and the development of anatomic structures; its results also serve as tests of the structural interpretations

introduced in the reference atlases. These “virtual fate maps” are based on the assimilation of data from over 2,000 genes and are geared to identify the best temporal match for the correlates of any structure recognizable at the given magnification. While this works easily for broad definitions of structures (e.g., olfactory bulb), it does not necessarily work for finer subdivisions of the brain. In practice, a limit is imposed by the level of neuroanatomical knowledge of the user (aided by the reference atlases). Anatomically expert users may guess where new interesting seed voxels can be found. Exploring true cell-fate specification would require two things: (1) analysis with cellular-level resolution to discriminate diverse cell types present in the brain, and (2) using genes or methods to consistently label cell types over time rather than rely on transiently expressed genes. However, the use of many genes at once provides a measure of relatedness that can inform novel insights about the development of the brain.

Molecular Principles of Brain Organization

TFs are key regulators for the specification of cell fate during neural development and thus the profiling of ~800 TFs with a relatively fine spatiotemporal sampling may reveal organizing principles of the brain. Multidimensional scaling (MDS) was applied to the binarized (on-off) manual annotation expression data. The MDS visualization allows for qualitative comparison of the relationship of gene expression and structural development. Points represent anatomic structures at a given developmental age, and the distance between them represents proximity on the basis of gene expression. A progressive change was observed in how TF expression correlates with progressive brain regionalization from E11.5 to E18.5 (Figure 7). At E11.5, brain structures clustered primarily by their longitudinal zonal origin within the major DV columns or “plates” (roof, alar, basal, and floor), and secondarily by neuromeric location along the AP axis, jointly defining a checkerboard pattern of primary histogenetic areas (Figure 7). This implies that gene functions shared along the longitudinal dimension of the whole neural tube—underpinning subsequent segmental serial similarity, known as metamery (Puelles and Rubenstein, 2003)—are activated earlier and more distinctly than differential AP molecular patterning of the neuromeric domains. In Figures 7B and 7C, gene expression patterns were overlaid to demonstrate the clear plate-based (DV) and neuromeric (AP) organization. Between E11.5 and E18.5, a gradual shift occurs in the molecular organization of the brain, resulting in the emergence of a secondary organization with mixed DV and AP features, appearing areally by E18.5 (as shown by the stronger AP organization). By E18.5, structures derived from alar and basal plates are no longer demarcated easily on the sole basis of their TF expression, possibly the result of DV tangential migrations (data not shown). The same is true for floor and roof plate-derived structures, although a distinction remains between alar-basal and roof-floor. Therefore, by late prenatal stages, brain regional identity is defined areally, instead of by plate-of-origin or neuromere; this switch occurs between E15.5 and E18.5, as shown by TF expression.

In general, alar-derived structures in the forebrain and midbrain show the largest variation over time, followed by basal-derived structures in the same two brain parts. The roof

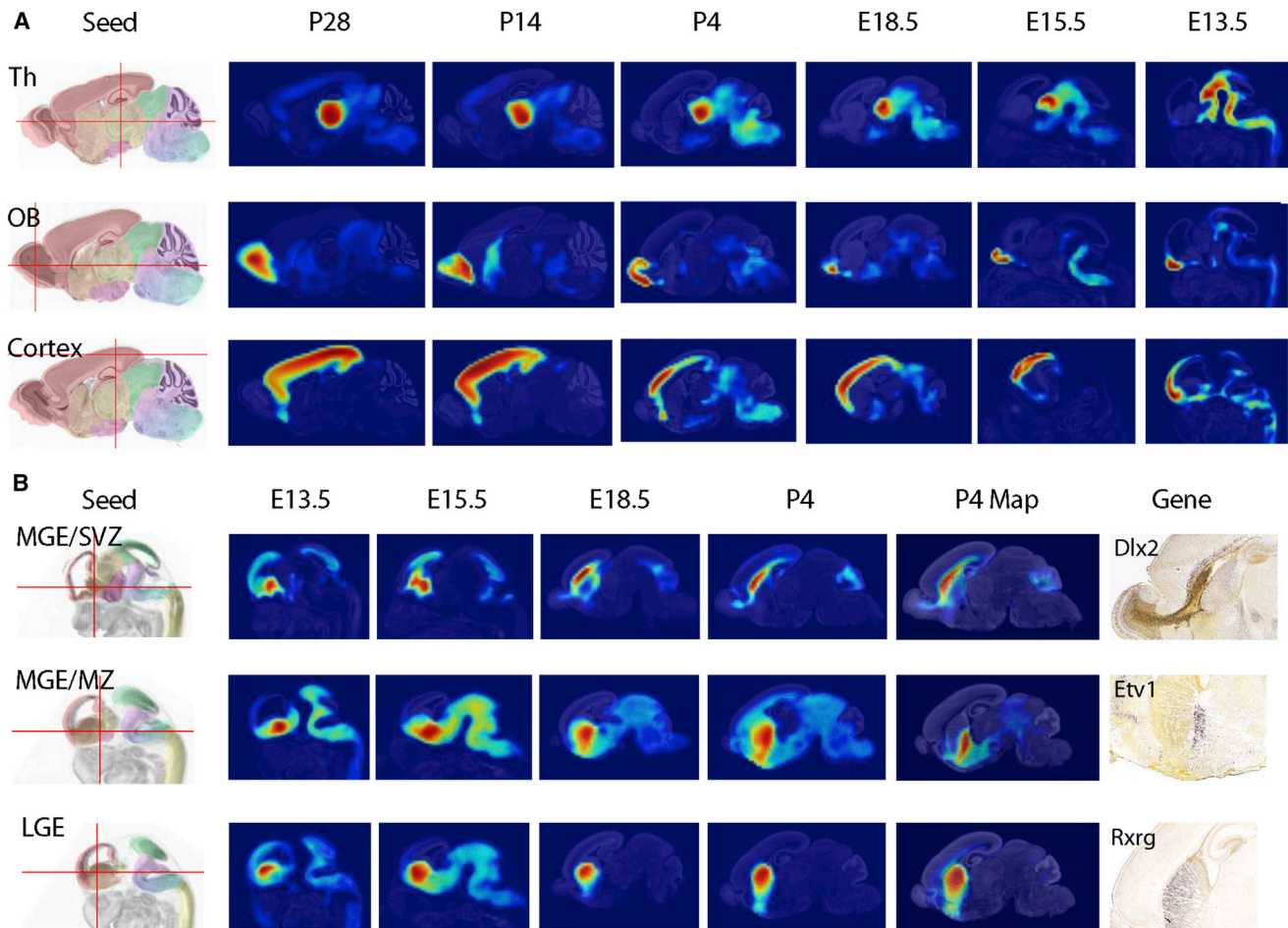


Figure 6. Virtual Fate Maps Using AGEA

(A) Virtual (reverse) fate mapping is constructed starting with an initial seed voxel selected at P28. The highest correlated voxel at the next youngest age is calculated in stepwise fashion iteratively until E13.5, and a correlation map is generated at each age. Method is shown for thalamus (Th), olfactory bulb (OB), and cortex.

(B) Virtual (forward) fate map of the ganglionic eminences. The initial seed voxel was selected manually at E13.5, and the highest correlated voxel at the next oldest age was automatically selected in stepwise fashion until P28. ISH data at P4 for a supporting gene are shown for each example: *Dlx2* for MGE/SVZ; *Etv1* for MGE/MZ; and *Rxrg* for LGE.

and floor plate-derived structures brain-wide, as well as the alar- and basal-derived hindbrain structures, show the least variable expression over time. Some subregions of the alar telencephalon, including neocortex, hippocampus, and olfactory bulb (red samples in left MDS plots), follow a unique trajectory separate from other alar plate-derived structures. These samples reasonably cluster with other alar structures at E11.5, when the plate-of-origin dominates region identity, but as they differentiate they become increasingly distinct from all other brain regions. One caveat is that TF expression is not necessarily linked to mechanisms of anatomic regionalization (boundary building), since other functions exist (e.g., control of proliferation and neurogenesis). These analyses are intended to assess the most evident principles of organization based upon a broad sampling of genes, acknowledging that selected functionally relevant markers can be used for more precise investigation of longitudinal or transverse boundaries.

The change from plate and neuromeric organization to largely areal organization reflects an acquisition of mature properties and a loss of early patterning cues. Finer subdivisions emerge as distinct structures over this period of embryonic development, lending to the dominance of areal and even strata-related identity by E18.5. We used the binarized TF data to assess the emergence of complexity over this time period, defined as the number of distinct binarized spatial expression patterns exhibited by the TFs within a given brain structure. For example, there are 12 distinct level 5 structures in the diencephalon in the reference atlas; a given gene can be “on” (detected) or “off” (undetected) in each structure, resulting in one of 4,096 (2^{12}) possible combinations. Taking all the TFs into account, the complexity of a region is the total number of distinct spatial expression patterns observed within that region. Based upon independent analysis of four brain regions (secondary prosencephalon, diencephalon, midbrain, and hindbrain), the number

A Dorsoventral plate of origin Anteroposterior/neuromeric location

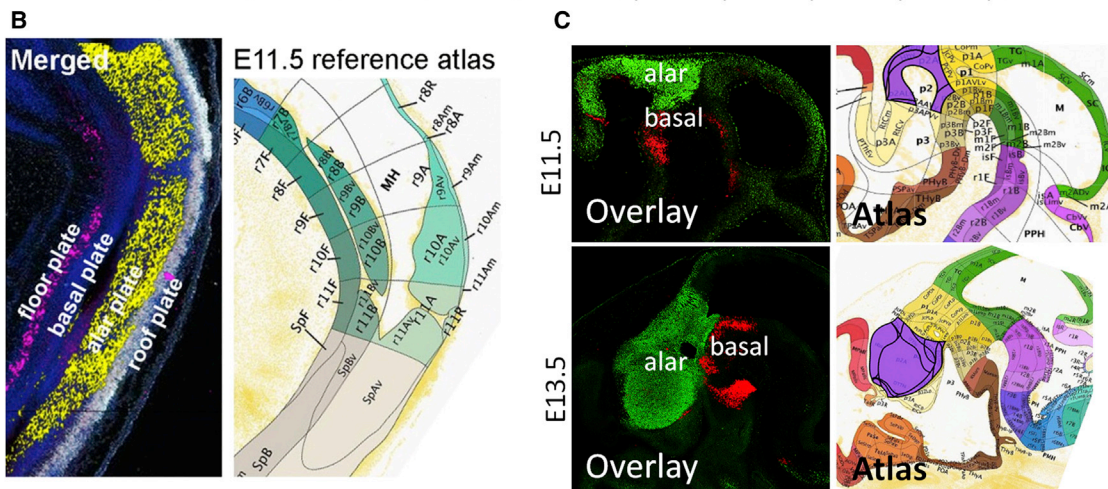
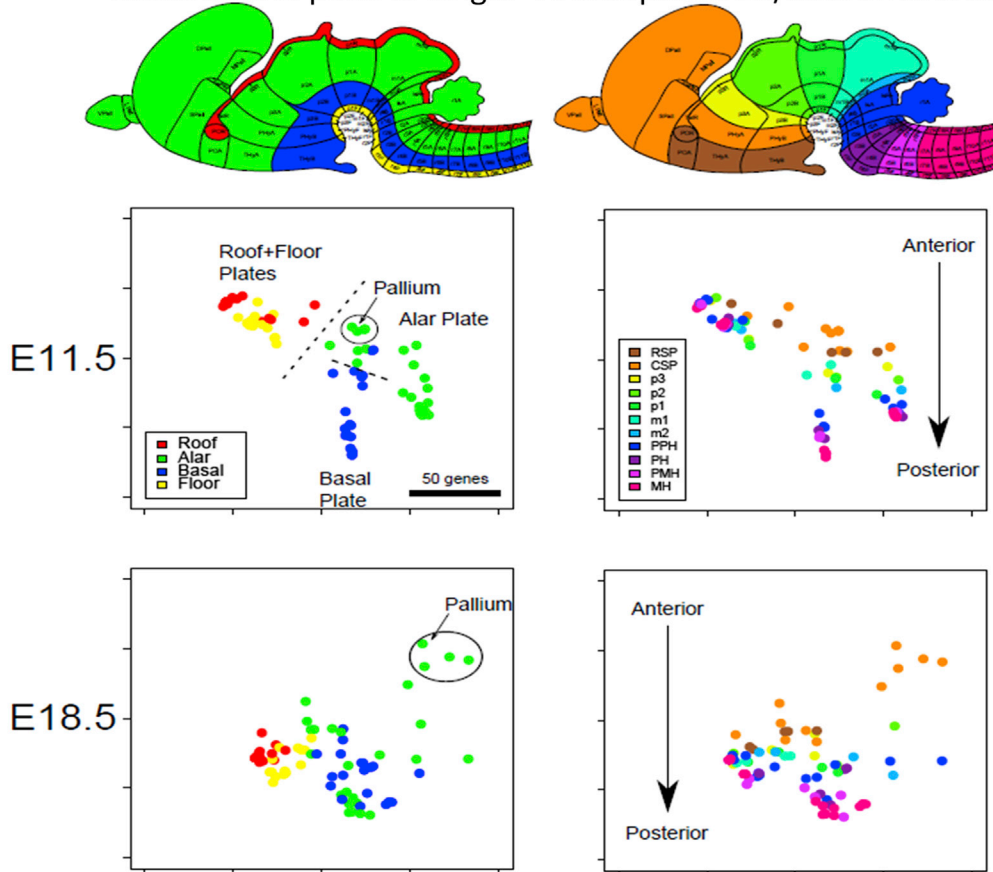


Figure 7. Multidimensional Scaling Shows a Shift from Dorsoventral- or Plate-Based to Anteroposterior Neuromere-Based Organization of the Embryonic Brain

(A) Two-dimensional visualization of regions characterized by differences in TF expression, using standard MDS for two embryonic ages. The brain schematic on the top shows brain structures color coded by DV plates or AP/neuromeric position. The distance between any two regions (dots) represents the number of genes that are differentially expressed between them, as determined by “expressed” versus “undetected” calls in the manual annotation. Left: structures are colored by DV location (roof, red; alar, green; basal, blue; yellow, floor); right: regions are colored by AP location, divided into the following gross categories: rostral secondary prosencephalon (RSP), caudal secondary prosencephalon (CSP), prosomeres 1–3 (p1, p2, and p3), mesomeres 1–2 (m1 and m2), prepontine hindbrain (PPH), pontine hindbrain (PH), pontomedullary hindbrain (PMH), and medullary hindbrain (MH).

(B and C) Examples of genes showing DV organization at E11.5 in the hindbrain (B) and in the diencephalon (C). Genes in (B) are floor plate, *Arx*; alar plate, *Asc1*; roof plate, *Msx1*. Genes in (C) are alar plate, *Tcf7l2* and basal plate, *Foxa1*.

of distinct expression patterns increased from E11.5 to E13.5, with a 2-fold increase in secondary prosencephalon. We detected no significant increase in the diversity of patterns after E13.5 (Figure S7). Based on the spatial patterns shared by the largest number of genes, it appears that the common expression modes at E11.5 were defined by expression throughout a large DV/AP region (e.g., Hox genes in the hindbrain and spinal cord) or by genes restricted to a longitudinal plate (e.g., *Shh* and *Pax7*). In the older embryo, however, the most frequent spatial expression patterns were restricted to individual brain structures (e.g., pallium or olfactory bulb). The peak in expression patterns at E13.5 could be due to the temporary coexistence of both DV (plate)-based and neuromeric/AP-based patterning.

The TFs were further analyzed to determine whether brain regions (defined as atlas ontology level 7 for pallium and level 5 for other brain structures) can be distinguished by a binary pattern of TF expression at each age across embryonic development (E11.5–E18.5); basically, we sought unique combinatorial expression patterns to define each age by brain structure combination. In order to identify putative genes that are involved in structural identity, we used a criterion that a gene must be expressed in all descendants of a given atlas structure down to level 10, the deepest level of the ontology short of individual nuclei or layers, in order to be called “widely expressed” for that level 5/7 brain structure (as opposed to “locally expressed” or “not expressed”).

To find a binarized TF code, we identified for each structure a unique set of widely expressed and not expressed genes. Several pairs of regions cannot be distinguished based on this criterion; these pairs of regions show widespread expression of the same genes and not expression of the same genes. Although differences noted in locally expressed genes imply that the expression patterns in such brain structures are not identical, they cannot be definitively distinguished with any combination of TFs. In addition, the “locally expressed” characterization means that transitivity of distinction between gene pairs is not preserved: if regions A and B cannot be distinguished, and regions B and C cannot be distinguished, it does not necessarily follow that regions A and C cannot be distinguished, because there might be a gene with widespread expression in A that shows local expression in B and no expression in C.

We identified a minimal set of ~80 TFs that provide a unique signature for every “distinguishable” region over four ages; 830 out of 13,944 total possible structure and region-pairs cannot be distinguished, the vast majority of which are pairs of regions at E18.5 (Figure S8B). For the remaining regions with distinct signatures, Figure S8A shows a spatiotemporal TF code at key prenatal stages in development. These genes include known region-specific markers such as *Foxg1*, a marker of telencephalic development, or the set of Hox genes, known to be involved in hindbrain and spinal cord patterning. The list also includes genes involved in reprogramming to stem cells, or in vitro transdifferentiation. However, some of the selected genes have not been as widely studied and are thus potentially interesting candidates for further analysis for their role in structural identity along both the spatial and temporal axes.

This minimal TF code is not unique, and alternative or complementary codes could exist. Indeed, the full set of ~800 TFs itself

forms a comprehensive code, although it provides no more information than the minimal 83 gene set. The majority of genes in the minimal TF code presented here are necessary (i.e., some pairs of anatomic structures are distinguished by a single gene). The remaining genes may still be biologically relevant, in that they distinguish particular subregions from each other. Overall, this analysis shows that a reduced set of less than 100 TFs is sufficient to generate a unique spatiotemporal code for all distinguishable primary/secondary brain structures at a medium-scale partitioning of the developing mouse brain wall. A simpler example of how the TF code can distinguish six structures at four ages is shown (Figure 8A).

To demonstrate the utility of this TF code for cross-platform comparisons of developmental time and region, we used published microarray data sets for mouse embryonic hypothalamus and preoptic area sampled from E11 to E15, and midbrain floor plate from E10. Cross-platform comparison is compounded by the underappreciated challenges of converting a scale of microarray expression values into a thresholded, binarized expression call comparable to our manual annotation data; thus, a perfect match was not anticipated. A mismatch score was calculated between the microarray set and the age \times anatomic structure-specific TF code, and using this score, the appropriate age and anatomic structure for each microarray sample could be identified based upon the best match of each sample to the TF code (Figure 8B).

DISCUSSION

The Allen Developing Mouse Brain Atlas uses histological and molecular profiling to provide a window into the temporal dynamics of over 2,100 genes over neural development in the mouse. Due to compromises of scale, a number of key genes surely are not represented in this Atlas. The gene set was selected to survey key functional classes and categories based on known pathways important for development. Ninety percent of these genes were detected in brain at some stage of development, as compared to the Allen Mouse Brain Atlas encompassing more than 20,000 genes in the C57Bl/6J P56 mouse, of which 78.8% are expressed at some level in the adult murine brain (Lein et al., 2007). It is notable that even using a preselected set of roughly 2,000 genes, representing 10% of the genome, the analyses of the resulting data set provided great insight into the organization of the brain, underpinning significantly our ontology (Puelles et al., 2013) and reference atlases.

While neuroanatomists have long used expression of key genes to guide their understanding of brain architecture, only more recently have integrated studies over genome-scale data sets been possible (Bota et al., 2003; Diez-Roux et al., 2011; Dong et al., 2009; Hawrylycz et al., 2010; Lein et al., 2007; Ng et al., 2009, 2010; Puelles and Ferran, 2012; Swanson, 2003; Thompson et al., 2008). In this resource, we provided a temporal framework to understand the genoarchitecture of brain development and new tools for the community to access these data. The manual annotation data that interpret expression patterns based on the ontology and the seven reference atlases provide support for users unfamiliar with neuroanatomy, aiding them to assign

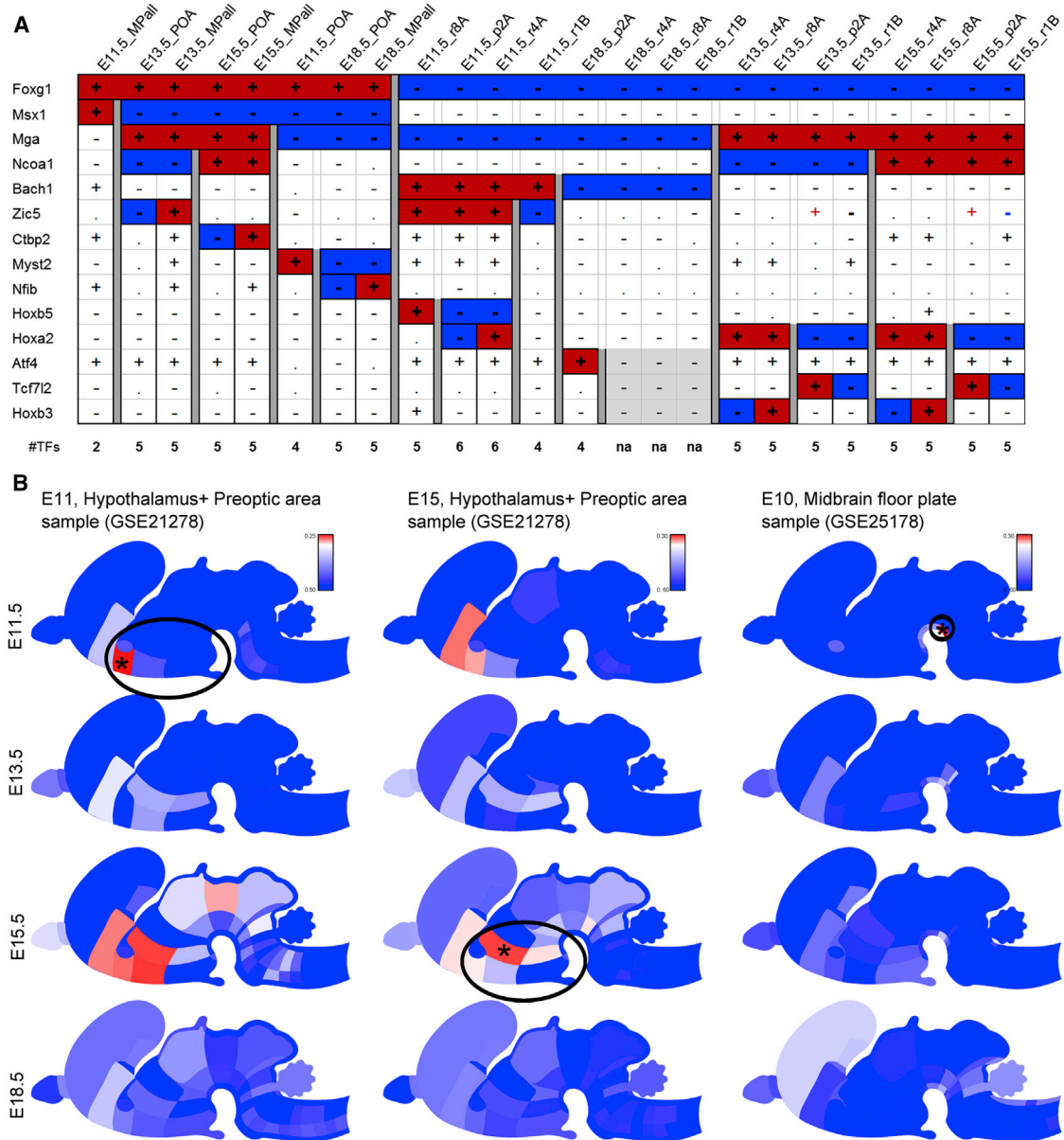


Figure 8. A Transcription Factor Code Can Uniquely Identify the Developmental Age and Anatomic Structure in a Sample Profiled by Microarray

(A) Fourteen genes can distinguish six brain structures at four ages; in this example, three atlas structures at E18.5 (gray shade) remain indistinguishable with this code.

(B) Identifying the anatomic region and biological age of a microarray sample based upon the TF code. For each sample, the GEO ID is given; the best match to a given age × region combination in the ADMB is color coded (red, high correlation; blue, low correlation; asterisk, best match). In each case, the TF code accurately identifies the closest age × brain structure. Note the anatomic criteria used for obtaining the microarray samples may have differed in part with our criteria, leading to the dispersion of the correlative results (see also Figures S7 and S8).

observed ISH signal to atlas structures; discovery tools such as NeuroBlast and AGEA enable users to achieve explicit identification of new genes of interest. Furthermore, in the three youngest embryonic ages, ISH, 3D models, and AGEA tools are available for the entire embryo, encompassing not only spinal cord and peripheral nervous system but also organs such as lung, heart, and kidney.

The temporal resolution of these data provided several major findings. First, gene expression exhibits complex dynamics over development; a set of marker genes at one stage may not necessarily define the same brain structure at a distant stage of development. However, by integrating the data of ~2,000 genes, large brain areas (i.e., at the level of thalamus, cortex, or striatum) and relatively smaller subregions can be tracked in

a stepwise fashion from embryonic to postnatal ages, demonstrating their molecular coherence across development, irrespective of emergent changes. Over the course of embryonic development, we observed that the organizing principles for the brain shift from a largely DV or longitudinal, plate-based organization of the brain (classic columns) to an AP, neuromeric, or transversally delimited organization of the brain that eventually transforms by orthogonal intersection of DV and AP units into the areal organization of individual histogenetic or progenitor domains, key for understanding the production of differential cell types. This order (AP to plate to areal) is consistent with the purposeful ordering of the reference atlas ontology, reflecting the order of key stages of developmental patterning. Although the hallmarks of AP patterning remain through the time course (previously observed in gene expression from adult tissues [Zapala et al., 2005]), the molecular signature of the major longitudinal plates appears to be transient; alar or basal plate signatures become indistinguishable by E18.5, and discrete late neuronal populations or complexes more closely identify with their final areal position/context.

Due to the complexity of developmental gene expression, it would be useful to have a molecular signature or “barcode” that identifies a particular brain structure at a given stage of brain development. This barcode could enable the development of intersectional strategies to target and manipulate cells at a precise stage of development and could also help identify the developmental age of cells generated from pluripotent stem cells by directed differentiation *in vitro*. The developmental phenomena that underlie brain development in tetrapods and possibly in all vertebrates have striking similarities in the types of genes and networks activated to govern the precise development of each brain region, though the timing of individual regions may vary; several neuroanatomists are developing panmammalian ontologies that assume a common developmental progression underlies this process in humans, nonhuman primates, and mice, hopefully without undermining the future panvertebrate developmental and adult brain ontology predicted by evolutionary theory and genomics. Thus, the identification of a perfected TF code that could potentially align homologous structures along comparable developmental stages across different vertebrate species is highly possible, irrespective of predictable variations. The TF code introduced in this paper is a humble beginning to the deduction of a molecular signature that describes brain regionalization in its entirety, as an extension of the codes previously deduced for simpler systems such as retinal development (Hatakeyama and Kageyama, 2004), and should illuminate hypercomplex systems such as human brain development. However, in our approach a shared code may not necessarily consist of, or contain, all factors that are causative for the specification of cell types. The TF code presented includes genes known to be key factors in direct reprogramming to specific cell types in culture (e.g., *Ascl1*, *Pou3f2*, *Sox2*, *Gata2*, *Nr2f1*, and *Foxg1*), and it probably also includes TFs that are involved in more downstream developmental differentiation processes such as axogenesis or dendritic maturation, providing a hallmark of the developmental age of the region. Future efforts could be targeted to refine the TF code to find causative genes and panmammalian or panvertebrate genes.

EXPERIMENTAL PROCEDURES

ISH

A high-throughput ISH platform described previously (Lein et al., 2007) generated ISH data for ~2,000 genes across seven ages including four embryonic (days postconception: E11.5, E13.5, E15.5, and E18.5) and three postnatal ages (P4, P14, and P28 days after birth, where day of birth is P0), with the addition of a yellow nuclear counterstain and modified protocols optimized for each age. Full methodological details are supplied in the [Supplemental Experimental Procedures](#).

Reference Atlases

For each reference atlas, tissue sections were stained by Nissl/cresyl violet or a nuclear HP Yellow stain to aid identification of anatomic structures for expert delineation (done by L.P. on the basis of an ontogenetic ontology based upon the prosomeric model [Puelles et al., 2012]). High-resolution images of tissue sections were obtained from automated microphotographic digitalizing systems and processed through our standard image pipeline, then exported to Adobe Illustrator CS graphics software for delineation of brain structures. Line drawings were converted to polygons corresponding to individual structures that were named systematically according to the ontology in the Illustrator file, converted to scalable vector graphics (SVG), databased, and lofted into 3D for use in the informatics pipeline.

Informatics Processing and Data Analysis

Full methodological details on the pipeline including development of NeuroBlast and AGEA are supplied in the [Supplemental Experimental Procedures](#). Pearson correlation was used to compare expression profiles. The statistical package R (<http://www.r-project.org/>) was used for data analysis and visualization. Expression clusters were visualized by projecting voxel expression data into a plane of section. Using expression values for the voxels of the diencephalon, we created coexpression gene networks using WGCNA (Zhang and Horvath, 2005). Gene ontology analysis was performed using DAVID (Huang et al., 2009a, 2009b). For the MDS analysis, all manually annotated data were binarized to expressed (= 1) and not expressed (= 0) calls for each anatomic structure at level 5 of the ontology (level 7 for pallial substructures). The distance between each pair of structures was calculated as the number of genes expressed in one structure that were not expressed in the other; this is equivalent to the Manhattan distance between the structures' expression vectors. This distance matrix was then projected onto three dimensions using the classical MDS package *cmdscale* in R. For visualization in two dimensions, the first two coordinates were chosen to plot the structure labels. In all plots shown, the eigenvalue-based goodness of fit measure as reported by the *cmdscale* package was at least 0.65.

Manual Annotation

ISH experiments were annotated by expert developmental neuroanatomists. Complete sets of image series of E11.5, E13.5, E15.5, and E18.5 experiments were manually annotated. Three metrics were used: intensity, density, and pattern. These metrics were scored for each brain structure according to a standard scheme (Figure S3) and entered into the hierarchically organized ontology of anatomical structures. At each developmental stage, annotation was performed for anatomic structures belonging to the most detailed level of the ontology (down to Level 10) that were identifiable as exhibiting differential expression. For example, if the pallium exhibited a homogeneous pattern but the subpallium exhibited a different pattern, annotation would be recorded for each of these structures individually. When a given brain structure was annotated, that annotation data was intended to represent the complete set of “child” or “descendent” structures of this level in the hierarchical tree (corresponding to an anatomical region), such that the expression call for pallium would then apply to its children: medial pallium, lateral pallium, ventral pallium, and dorsal pallium.

Manual annotation was not performed for every structure at every level of the ontology, which amounts to over 1,500 brain structures. Instead, the annotation strategy ensured that every “branch” of the ontological tree was annotated. For example, for the four major parts of the brain: forebrain, midbrain, hindbrain, and spinal cord, a given gene may have expression in only

diencephalon. Therefore, midbrain, hindbrain, and spinal cord would be annotated as “undetected,” and the forebrain expression may be addressed by providing the actual expression information for diencephalon, while producing an “undetected” call for the sibling structure, secondary prosencephalon.

Transcription Factor Code

For every pair of anatomic structures, TFs were identified that show widespread expression in one structure and no expression in the other, generating a combinatorial set of structure pairs, each linked to a set of TFs. Widespread expression was defined as expression in all children of the structure to level 10, the deepest level of the ontology. Next, we identified all pairs of brain structures that could be distinguished only by a single gene. All of these genes were included in the final set. For the remaining structure pairs, identifying a minimal set of TFs to distinguish each brain region is equivalent to the set cover problem (an NP-hard problem). We used a heuristic pruning approach to approximate a minimal set: starting with the full set of unselected genes, we randomly removed one and reexamined the remaining data to identify structure pairs that now had a single gene distinguishing the pair members. These genes were added to the final list, and the pruning process continued until all remaining genes were crucial to distinguish at least one structure pair. An exhaustive search over every possible selection path was not feasible, so this process was repeated 100 times and the gene set with the fewest members was selected.

The TF code was applied to three Affymetrix mouse genome microarray data sets from the Gene Expression Omnibus (GEO) with IDs GSE21278 and GSE25178. Because the tissues profiled in these data sets do not correspond exactly to specific anatomic structures defined in the atlas ontology described here, we compared the thresholded expression profile from the GEO data sets to the full TF code for every time point and structure in the ontology and ranked the matches using the following metric:

$$\text{Match score} = \frac{4 * FP + FN}{FP + FN + TP + TN}$$

where FN = number of genes called “present” in the GEO set but “undetected” in our code, FP = number of genes called “absent” in the GEO set but called “widely expressed” in our code, TN = number of genes called “absent” in the GEO set and “undetected” in our code, and TP = number of genes called “present” in the GEO set and “widely expressed” in our code. For each GEO sample data set, we ranked all the structures by how well they scored according to this match score. The brain structure with the best match score for each of the GEO data sets is starred (Figure 8).

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, eight figures, and five tables and can be found with this article online at <http://dx.doi.org/10.1016/j.neuron.2014.05.033>.

AUTHOR CONTRIBUTIONS

C.L.T. and L.N. spearheaded the creation of the resource with guidance from J.L.R.R., A.R.J., and L.P. N.D., K.S., J.K., A.J., G.O., Z.R., and S.S. contributed to optimization of methods specific for embryonic and postnatal *in situ* hybridization process. L.N., C.L., C.D., W.B.W., A.J.S., R.Y., and L.K. developed analysis algorithms, informatics pipeline, visualization software, and database infrastructure specific for the ADMBA resource. L.N., V.M., C.K.L., T.N.N., C.L.T., S.M.S., K.G., A.H., J.M., D.P., S.M., J.L.R.R., A.K., D.P., and M.J.H. performed data analysis and interpretation. S.M., R.G.L., A.M.F., and A.P. created the manual annotation data set by reviewing each individual embryonic ISH experiment. C.L.T., M.J.H., V.M., L.P., and J.L.R.R. wrote the paper. L.P. created the reference atlases and ontology for seven ages of mouse brain development with assistance from C.L.T., L.N., and K.G. Finally, A.B., J.H., and P.W. oversaw laboratory pipeline activities.

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