



A high-resolution cat radiation hybrid and integrated FISH mapping resource for phylogenomic studies across Felidae

Brian W. Davis^a, Terje Raudsepp^a, Alison J. Pearks Wilkerson^a, Richa Agarwala^b, Alejandro A. Schäffer^c, Marlys Houck^d, Bhanu P. Chowdhary^a, William J. Murphy^{a,*}

^a Department of Veterinary Integrative Biosciences, College of Veterinary Medicine and Biomedical Sciences, Texas A&M University, Mail Stop 4458, College Station, TX 77843-4458, USA

^b IEB/NCBI/NLM, National Institutes of Health, Department of Health & Human Services, Bethesda, MD 20894, USA

^c CBB/NCBI/NLM, National Institutes of Health, Department of Health & Human Services, Bethesda, MD 20894, USA

^d San Diego Zoo Conservation Research, San Diego Zoological Park, Escondido, CA 92025, USA

ARTICLE INFO

Article history:

Received 5 August 2008

Accepted 23 September 2008

Available online 5 November 2008

Keywords:

Domestic cat

Radiation hybrid map

Comparative mapping

FISH mapping

Synteny

Chromosome rearrangement

Felidae

ABSTRACT

We describe the construction of a high-resolution radiation hybrid (RH) map of the domestic cat genome, which includes 2662 markers, translating to an estimated average intermarker distance of 939 kilobases (kb). Targeted marker selection utilized the recent feline 1.9x genome assembly, concentrating on regions of low marker density on feline autosomes and the X chromosome, in addition to regions flanking interspecies chromosomal breakpoints. Average gap (breakpoint) size between cat–human ordered conserved segments is less than 900 kb. The map was used for a fine-scale comparison of conserved syntenic blocks with the human and canine genomes. Corroborative fluorescence *in situ* hybridization (FISH) data were generated using 129 domestic cat BAC clones as probes, providing independent confirmation of the long-range correctness of the map. Cross-species hybridization of BAC probes on divergent felids from the genera *Profelis* (serval) and *Panthera* (snow leopard) provides further evidence for karyotypic conservation within felids, and demonstrates the utility of such probes for future studies of chromosome evolution within the cat family and in related carnivores. The integrated map constitutes a comprehensive framework for identifying genes controlling feline phenotypes of interest, and to aid in assembly of a higher coverage feline genome sequence.

© 2008 Elsevier Inc. All rights reserved.

Introduction

During the past decade, increasingly detailed genetic and physical maps of the domestic cat genome have provided tools for mapping and discerning the hereditary basis of morphological variation and genetic diseases in cats, which model phenotypes or pathologies observed in other mammals [1–4]. These include mutations in functional candidate genes that control coat morphology [5–7], as well as those that are causative for monogenic diseases such as polycystic kidney disease [8], and hypertrophic cardiomyopathy [9]. The maps were also used in genome scans to identify disease genes not previously implicated in human studies at the time of the scan [10,11], suggesting the value of multiple animal models in understanding the pathogenesis of human disease. The ability to narrow candidate regions and identify causative mutations in the feline model is currently limited by appropriate animal cohorts and by the quality of feline–human comparative gene maps.

Therefore, we have continued to improve the quality and density of domestic cat radiation hybrid maps to define the evolutionary rearrangements that distinguish the cat from other sequenced mammalian genomes, and facilitate positional reasoning in gene and mutation hunting. These maps can be a tool for both the long-range precision and quality control of genome assemblies [12], as well as studies of the dynamics of mammalian chromosome evolution [13]. For example, the feline 1.9x sequence [14] was assembled based on conserved ordered segments defined by the RH-based feline–human and feline–dog comparative maps [3]. However, a large fraction of the genome remains unassembled due primarily to the low coverage of the sequence traces [14] and secondarily to the incomplete (85%) coverage of the previous cat–human comparative maps [3]. The new map reported here provides enhanced coverage of the feline genome (96%), and has been independently validated using FISH data, and by comparison to a new genetic linkage map [15]. This tool will aid the chromosomal assignment and ordering of scaffolds for the draft genome assembly, facilitate identification of genes controlling feline phenotypes, and provide insight into the details of chromosomal evolution that have occurred since the divergence of the cat and dog genomes from the ancestral carnivore karyotype.

* Corresponding author. Fax: +1 979 845 9972.

E-mail address: wmurphy@cvm.tamu.edu (W.J. Murphy).

Results and discussion

A fine-scale cat radiation hybrid gene map

We analyzed a final set of 2674 markers that were evaluated by formal linkage analysis. Twelve of these markers were later dropped while computing the final map in which linkage groups were computed using a two-point LOD score threshold of 9.0. At this threshold, all but five chromosomes comprised complete linkage groups. Chromosomes A1, C2, and D2 were divided into two linkage groups with gaps at the centromeres; these chromosome arms were merged in a single map. Chromosome E1 formed three RH linkage groups with two gaps: the first caused by the nucleolar organizer region (NOR), and the second because of the severe changes in retention frequency associated with the RH selectable marker (*TK1*) that is found on this chromosome. The three RH groups on chromosome E1 were oriented using evidence from the feline linkage maps and FISH data. On chromosome X, the pseudoautosomal region (PAR) comprises a separate linkage group due to increased retention resulting from co-amplification of Y chromosome-bearing fragments in the PAR [3].

The computed RH map contains 2662 markers that cover all 18 feline autosomes and the X chromosome, with marker density being fairly uniform across chromosomes (Table 1). The uniform marker density was achieved in part because we computed an intermediate map after 2550 markers had been genotyped (map not shown here) and the final batch of markers to be developed were targeted to gaps in coverage of that intermediate map. We reused some marker input data from earlier maps, but we did not assume that markers were in the same order as in previous maps. Of the 2662 markers, 733 are associated with feline annotated genes or ESTs, 783 are microsatellites, and the remainder are derived from published STS markers or the domestic cat 1.9x genome sequence. The final map has 1602 markers on the maximum-likelihood (MLE)-consensus framework map, 779 markers placed at centiRay (cR) positions relative to the framework map, and 281 markers assigned to larger bins with respect to the framework map. The 1602 markers on the framework map are assigned to 1445 distinct positions; among the 2381 markers assigned to cR positions, there are 2184 distinct positions. Therefore, assuming a feline euchromatic genome size of 2.5 gigabases (Gb) [14], all markers comprising the current map result in one marker every 939 kb, 82% of which reside in unique positions. The binned markers are not ordered or assigned a cR position, thus one marker is assigned a cR position every 1050 kb. Fig. 1 depicts the

integrated RH, FISH and human comparative maps for feline chromosome A3. The complete RH comparative maps and cytogenetic maps are available in supplemental material (Supplemental Table 1, Supplemental Figs. 1 and 2).

Map order confirmation using FISH and linkage map comparisons

The accuracy of the long-range marker order of the RH map was verified using two independent approaches. The first approach used FISH mapping of domestic cat bacterial artificial clones (BAC) isolated from the RPCI-86 10x library. BAC end sequencing of 480 random clones was used to identify, by computational comparison to the dog, human and cat assemblies, a set of probes distributed relatively evenly across all autosomes and the X chromosome. The positions of 129 feline BAC clones were obtained by FISH (Fig. 1, Supplemental Fig. 2, Supplemental Table 2). When combined with previously published cytogenetic mapping data (see Supplemental Fig. 2), the average spacing of FISH markers is approximately one marker every 18 Mb. The FISH based marker order was identical to the RH-based comparative marker order, confirming the long-range assembly of the RH maps.

A second analysis compared the RH map order with marker order derived from a domestic cat linkage map [15]. A comparison of markers mapped on both maps showed the two maps to be 94% consistent, with only 18 of 319 marker order discrepancies between the two maps (Supplemental Table 3). The vast majority of marker order differences involved flips of closely spaced, adjacent markers (Avg. distance=6.8 cM). Only 3 of 17 discrepantly ordered marker pairs were spaced greater than 10 cM apart, in addition to a single marker, FCA1028, assigned to different chromosomes that could not be resolved (Supplemental Table 3). Overall, the three combined mapping approaches validate the long-range marker order across cat chromosomes, and provide anchor points for each RH map to its respective feline chromosome.

Comparative synteny analysis

We identified orthologous positions for 99% of the total 2662 mapped markers in the latest dog and human genome assemblies (Supplemental Table 1). AutoGRAPH [16] was used to visualize chromosome rearrangements between cat, dog and human genomes (Fig. 1, Supplemental Fig. 1). Using conservative criteria to define blocks of conserved marker order (see Materials and methods), we

Table 1
Summary statistics for the domestic cat radiation hybrid map

Domestic cat chromosome	Total markers on map	No. of MLE-consensus framework markers	Avg. marker density (Mb)	RH length (cR5000)	Approximate physical length ^a (Mb)	cR/Mb	Human CSOs	Dog CSOs
A1	246	129	1.0	1964.4	246	8.0	12	13
A2	205	127	0.9	1785.2	180	9.9	15	10
A3	155	97	0.9	1278.6	143	9.0	7	4
B1	190	104	1.0	1466.2	198	7.4	13	19
B2	140	90	1.1	1235.3	148	8.4	2	6
B3	144	96	1.0	950.1	143	6.7	4	4
B4	148	88	0.9	1040.2	138	7.6	4	8
C1	197	125	1.1	1626.7	220	7.4	3	12
C2	158	98	0.9	1302.0	148	8.8	10	8
D1	133	83	0.9	1089.7	123	8.9	5	6
D2	105	58	1.0	813.0	103	7.9	8	3
D3	114	80	0.9	1091.0	103	10.6	10	5
D4	109	69	0.9	992.4	95	10.4	6	3
E1	109	62	0.9	1200.9	95	12.6	6	4
E2	82	52	0.9	552.4	78	7.1	2	4
E3	69	50	0.9	637.3	60	10.6	7	2
F1	102	56	0.7	556.5	75	7.4	10	5
F2	84	47	0.9	608.8	75	8.1	1	3
X	172	91	0.7	960.3	128	7.5	1	1
Total	2662	1602	0.9	21151.0	2493	8.5	126	121

^a Assumes a 2.5 Mbp euchromatic genome and the total cytogenetic fraction estimated for each chromosome in the domestic cat genome [14], excluding the Y chromosome.

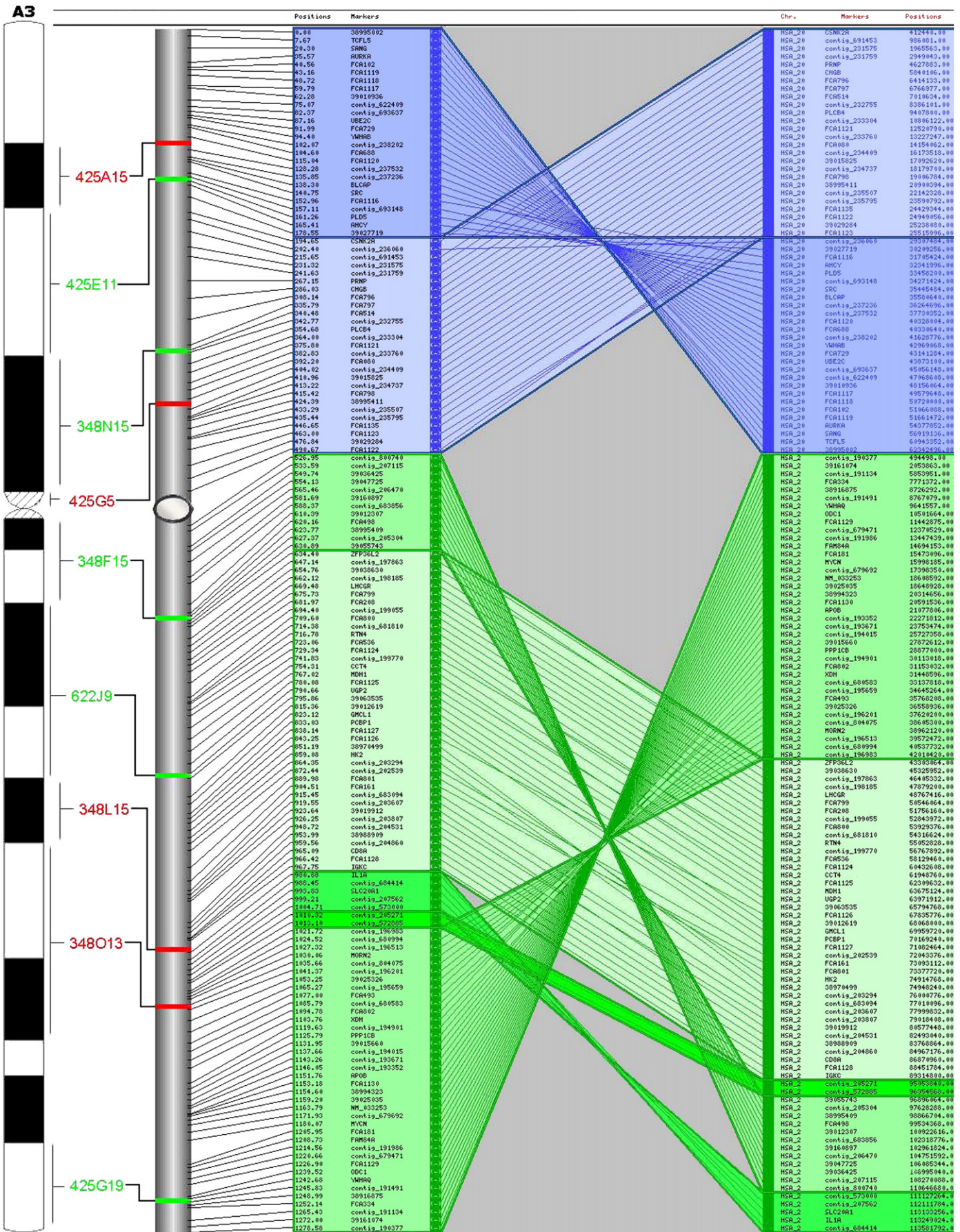


Fig. 1. Integrated RH, FISH, and comparative maps for feline chromosome A3. CSOs for human chromosome 20 and 2 are shown to the right of each cat chromosome map (only the map scale is shown). The inferred centromere position is shown as a gray oval. The comparative map represents the output from an AutoGRAPH [16] based analysis.

identified 126 conserved segments ordered (CSOs, or homologous syntenic blocks—HSBs [13]) between the cat and human genomes, and 121 between the cat and dog genomes (Supplemental Table 1, Supplemental Fig. 1). Though these figures include singletons, we only considered a singleton to represent a CSO if it was present in one comparison species (i.e. human or dog) and part of a multimarker stretch of conserved gene order in the other species. We interpret these singletons to represent evidence of lineage-specific rearrangements.

Comparative coverage of cat CSOs on the human genome assembly was estimated as previously described [17]. Specifically, comparative coverage is defined as the sum of the physical span of human CSOs in cat, divided by the size of the human genome after excluding centromere, telomere, and heterochromatic regions, or regions lacking any cross-species homology in multi-species alignments. Estimated comparative coverage with the human genome is 96%, an 11% increase over the previous cat–human comparative map [3]. The mean gap size is 867 kb (range=0.12–2.3 Mb), with 68% of the gaps being less than 1 Mb, and 97% less than 2 Mb.

An examination of chromosome rearrangement rates between cat–human and cat–dog, as well as the lineage-specific chromosomal breakpoints, is presented in Fig. 2. A similar number of CSOs distinguish cat and human and cat and dog genomes (126 versus 121 respectively), despite the fact that dog and cat share a much more recent (55 million years ago [Mya] versus 95 Mya) common ancestry (Fig. 2). This is accounted for by the well-documented accelerated rate of canine chromosome evolution [18–20].

In comparison to human and cat, the dog genome has been punctuated by a very high proportion of interchromosomal rearrangements (Fig. 2), while very few blocks of cat–dog conserved synteny are further disrupted by inversions (Fig. 1 and Supplementary Fig. 1). Specifically, 86% of the rearrangements between cat and dog genomes involve interchromosomal breakpoints, compared to only 19% of human–cat rearrangements, which are predominately inversions (Fig. 2). Further, while the relative proportion of interchromosomal to intrachromosomal rearrangements is similar in the two carnivore lineages (roughly twice as many interchromosomal rearrangements), the overall rate of breakage is approximately twice as high in the canid lineage versus the feline lineage. The refined definition of cat–dog–human chromosome breakpoints in this study will allow for a detailed

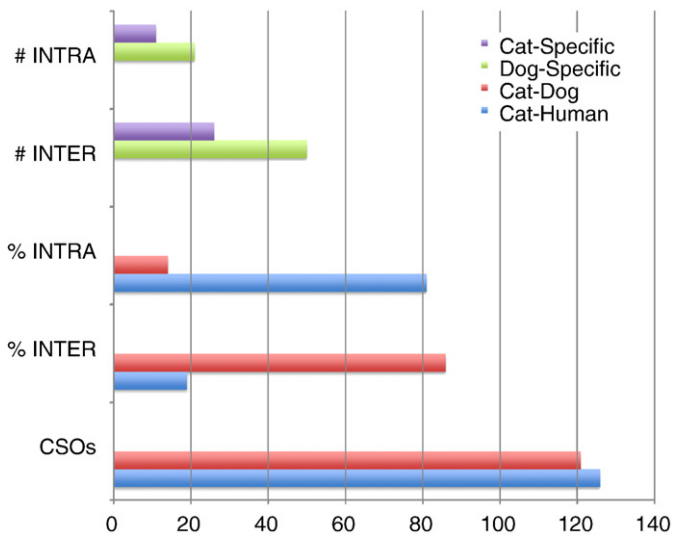


Fig. 2. Lineage-specific rearrangement totals for the domestic dog and domestic cat and rearrangement frequencies between the domestic cat, domestic dog, and human genomes. The lineage-specific totals are #INTRA=the number of intrachromosomal and #INTER=the number of interchromosomal breakpoints. %INTRA=the percentage of rearrangements between two genomes that are the result of intrachromosomal breakpoints. %INTER=the percentage of rearrangements between two genomes that are the result of interchromosomal breakpoints.

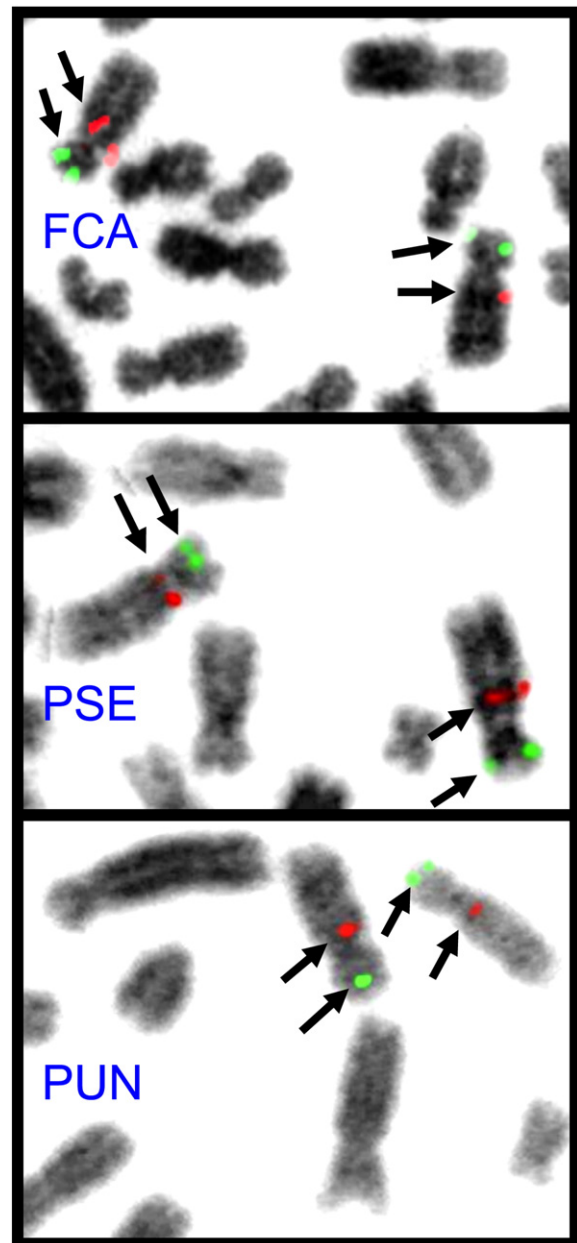


Fig. 3. FISH results for two feline BAC clone probes (green: 348A7; red: 348J11) hybridized to B4 orthologues on domestic cat (Fca), serval (Pse) and snow leopard (Pun) metaphase chromosomes. Note the same relative positioning of each probe in all three species. A summary of all FISH results relative to domestic cat chromosomes can be found in Supplementary Fig. 2.

comparative analysis of their sequence properties once high-coverage cat genome sequence becomes available.

Utility of FISH probes in other species of Felidae

We examined the applicability of the domestic cat FISH probes to hybridize to chromosomes of other divergent members of the cat family: serval (*Profelis serval*), and snow leopard (*Panthera uncia*). These two species are representatives of felid lineages that last shared a common ancestor with the domestic cat approximately 8 and 10 Mya, respectively, and span the earliest nodes of the felid phylogeny [21]. We tested all probes from two domestic cat chromosomes, B4 and F1, that had differences in G-banding patterns documented between at least two of the three species [22]. It has been suggested that the B4 chromosome of species of the genus *Panthera*

(and several additional felid species) is distinguished from members of the domestic cat lineage and the serval by a pericentric inversion. In addition, domestic cat chromosome F1 is orthologous to chromosomes F3 of serval and snow leopard, but differs in its G-banding pattern, suggesting possible gene order differences between the two chromosomes.

100% of domestic cat BAC probes tested ($n=22$ independent hybridizations) on serval and snow leopard chromosome preparations produced successful hybridization results (Fig. 3). Furthermore, hybridization results suggested the marker order (as shown in Supplemental Fig. 2) is conserved across all three felid species on these two chromosomes (Fig. 3). Using our probes, we found no evidence of a pericentric inversion distinguishing snow leopard chromosome B4 from domestic cat or serval. Specifically, a BAC probe that hybridizes just below the centromere on domestic cat B4 localizes to the same location of the long arm, rather than the short arm (in the case of a pericentric inversion), of snow leopard chromosome B4 (Fig. 3). Furthermore, the marker order was identical across all three cat species for chromosome F1 and its homologue F3. Therefore the observed G-banding differences across the felids studied may not be due to large scale structural changes (inversions), but either to smaller paracentric inversions inside of the markers we applied here, or due to compositional sequence changes between species that do not affect gene order.

Conclusions and future directions

We report a 939-kilobase resolution, RH-based, physical map of the feline genome. The most important result of this study is the notable improvement in the comparative genome alignments between the cat–human and cat–dog. The comparative coverage of the human genome is 96%, closing more than two-thirds of the coverage deficiency of the most recent map [3], with evolutionary breakpoints (gaps) resolved to less than 900 kilobase resolution on average. The enhanced map resolution allowed for the identification of several novel small conserved segments generated by intrachromosomal rearrangements not observed in previous maps. The high marker density of this map makes it vital to the construction of the forthcoming assembly and annotation of a higher coverage (than the currently available 1.9x) feline genome sequence. These resources will stimulate and facilitate the identification of feline genes of interest using genetic linkage and selective sweep mapping approaches in domestic and natural populations of felid species.

The high success rate of the domestic cat BAC probes on other felid species suggests they will be a reliable resource for cross-species synteny analysis in all felids. Because results were unable to detect any rearrangement that would explain the observed cytological differences between the orthologous domestic cat F1 and *Profelis* and *Panthera* F3 chromosomes, these data suggest karyotypic conservatism may be more extreme across Felidae than previously appreciated, and that further application of these, and additional, probes may help resolve other documented karyotypic differences within the cat family. Human BAC probes are routinely applied in other primate lineages that span 40–50 million year divergence times [23]. Our results suggest that a large proportion of domestic cat BAC probes will successfully hybridize to chromosomes of other feliform carnivores (i.e. linsangs, hyenids, herpestids, viverrids), a group that radiated in a similar evolutionary timeframe [24].

Materials and methods

Marker selection

Pairs of primers were designed from sequence contigs from the 1.9x feline genome assembly [14] that had reciprocal best hits to one or both orthologous chromosomes of dog (CanFam2) and human (Hs36) genome assemblies. Repetitive sequences were masked using Repeat-

masker, and primers were designed with Primer3 [25]. Each primer pair was tested by PCR in cat DNA, hamster DNA, and a 10:1 hamster:cat mixture of DNA. Only those markers showing robust amplification in cat and the 10:1 DNAs were chosen for further genotyping in the RH panel.

RH genotyping

RH genotyping was performed on the 5000-rad feline whole genome radiation hybrid panel using previously described methods [4]. Markers were dropped before map computation for one of the following reasons: weak amplification, high hamster background amplification, or excessively high retention frequency ($>70\%$ and not predicted to reside on the selectable locus chromosome or near a centromere) or excessively low retention frequency compared to other markers on the same chromosome. These new genotypes were merged with vectors from previous maps for map computation.

Map construction

Two-point linkage groups were computed at a LOD score of 9.0. Markers within each chromosome or linkage group were ordered using a reduction from the problem of RH mapping to the traveling salesman problem (TSP), as implemented in the software *rh_tsp_map* [26,27]. The computations to construct the map were done using programs from the software package *rh_tsp_map* (ftp://ftp.ncbi.nih.gov/pub/agarwala/rhmapping/rh_tsp_map.tar.gz) and using the package CONCORDE (<http://www.isye.gatech.edu/~wcook/rh>) [28] linked with QSOPT (<http://www.isye.gatech.edu/~wcook/qsopt>) to solve the TSP instances to guaranteed optimality. Details of the mapping procedures are described in the tutorial accompanying *rh_tsp_map*. The three linkage groups on chromosome E1, and the PAR and X-specific region on the X chromosome maps were oriented and ordered using data from linkage maps and FISH.

Before computing the final map shown here, we used similar procedures to compute an intermediate map when approximately 2550 markers had been developed, amplified, and scored on the 5000 rad panel. The intermediate map was used for two different purposes that both led to a much-improved final map. First, for any clone and triplet of (apparently) consecutive markers in which the retention patterns showed two obligate breaks, the problematic marker/clone pairs were reevaluated using the strategy described in [17]. This substantial effort in local quality control explains why over 99% of the markers at the start of final map computation could be included in the final map, despite rigorous flips tests of marker order implemented in *rh_tsp_map*.

Second, the intermediate map was used to identify holes in cat–human and cat–dog map comparisons, so that the last batch of markers to be developed could be targeted to many of these holes. This substantial effort in global quality control explains why the coverage of the new map (96%) is so much higher than the coverage of the previous cat RH map (85%; [3]).

FISH mapping

480 clones from three randomly selected plates of the domestic cat RPCI-86 BAC library were end-sequenced (SP6 end only) using described protocols [29]. These sequences were repeatmasked and queried against the feline 1.9x genome assembly and human and dog genome assemblies using BLAT to identify best hits and positions in each assembly (Supplemental Table 2). A collection of BAC clones spaced across all cat autosomes and the X chromosome were selected, grown, and DNA extracted using the Qjagen large construct procedure. BAC clone DNA was labeled with biotin and/or digoxigenin (Bio-Nick and Dig-Nick kits; Roche Molecular Biochemicals) and hybridized to domestic cat metaphase chromosomes. Images for a minimum of 30 metaphase spreads were captured and analyzed with a Zeiss Axioplan2 fluorescent microscope equipped with Cytovision/Genus

V. 2.7 (Applied Imaging). Identification of cat chromosomes and assignment of markers to specific chromosome bands followed the nomenclature of O'Brien and Nash [30]. A subset of these probes ($n=11$) were also hybridized to metaphase spreads from individuals representing two divergent felid species: *Profelis serval* (serval) and *Panthera uncia* (snow leopard).

Comparative analysis

For each domestic cat locus, physical positions for orthologous genes were obtained by using either BLAT [31] or discontinuous MegaBLAST (<http://www.ncbi.nlm.nih.gov/blast/>; [32]) searches to the feline, human and canine reference genome assemblies (felCat3, hg18 and canFam2, respectively). In a small number of cases, BLAT hits to one species (either human or dog) genome were not covered in the top alignment net in the second genome. In these cases, we identified the nearest corresponding stretch of orthology using the dog and human alignment nets of the UCSC Genome Browser. Conserved segments ordered (homologous synteny blocks) were defined by searching for runs of two or more uninterrupted markers on the same chromosome between two species. Inverted segments were defined by runs of three or more markers, each separated by 1 Mbp. Some out of place markers were expected due to mapping/genotyping errors or limitations of RH mapping resolution. These were assigned to the closest conserved segment if the intervening markers did not span more than a few Mb. Markers that were binned or placed with a LOD score <0.5 were not used in determining marker order, though they could be used to determine the extent of comparative coverage. The program AutoGRAPH [16] was used to visualize ordered and unordered conserved segments between cat–human and cat–dog comparisons.

Acknowledgments

We thank Keith Durkin, Julie Fronczek, Jan Janecka, William Nash, and Ashley Seabury for helpful advice and technical support. We thank Cynthia King for the serval blood sample. Jim Mullikin provided a list of candidate contigs for primer design. Marilyn Menotti-Raymond shared the marker positions on a new genetic map computed by AAS. This work was supported by funds from the Morris Animal Foundation (Grant D06FE-063) to WJM and the Winn Feline Foundation (Grant 06-026) to WJM, TR, and BPC. This research was supported in part by the intramural research program of the NIH, NLM (RA, AAS).

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.ygeno.2008.09.010](https://doi.org/10.1016/j.ygeno.2008.09.010).

References

- Menotti-Raymond, V.A. David, R. Agarwala, A.A. Schäffer, R. Stephens, S.J. O'Brien, W.J. Murphy, Radiation hybrid mapping of 304 novel microsatellites in the domestic cat genome, *Cytogenet. Genome Res.* 102 (2003) 272–276.
- Menotti-Raymond, V.A. David, M.E. Roelke, Z.Q. Chen, K.A. Menotti, S. Sun, A.A. Schäffer, J.F. Tomlin, R. Agarwala, S.J. O'Brien, W.J. Murphy, Second-generation integrated genetic linkage/radiation hybrid maps of the domestic cat (*Felis catus*), *J. Heredity* 94 (2003) 95–106.
- W.J. Murphy, B. Davis, V.A. David, R. Agarwala, A.A. Schäffer, A.J. Pearks Wilkerson, B. Neelam, S.J. O'Brien, M. Menotti-Raymond, A 1.5-Mb-resolution radiation hybrid map of the cat genome and comparative analysis with the canine and human genomes, *Genomics* 89 (2007) 189–196.
- W.J. Murphy, S. Sun, Z. Chen, N. Yuhki, D. Hirschmann, M. Menotti-Raymond, S.J. O'Brien, A radiation hybrid map of the cat genome: implications for comparative mapping, *Genome Res.* 10 (2000) 691–702.
- J.S. Kehler, V.A. David, A.A. Schäffer, K. Bajema, E. Eizirik, D.K. Ryugo, S.S. Hannah, S.J. O'Brien, M. Menotti-Raymond, Four independent mutations in the feline fibroblast growth factor 5 gene determine the long-haired phenotype in domestic cats, *J. Heredity* 98 (2007) 555–566.
- L.A. Lyons, D.L. Imes, H.C. Rah, R.A. Grahn, Tyrosinase mutations associated with Siamese and Burmese patterns in the domestic cat (*Felis catus*), *Anim. Genet.* 36 (2005) 119–126.
- A. Schmidt-Küntzel, E. Eizirik, S.J. O'Brien, M. Menotti-Raymond, Tyrosinase and tyrosinase related protein 1 alleles specify domestic cat coat color phenotypes of the albino and brown loci, *J. Heredity* 96 (2005) 289–301.
- L.A. Lyons, D.S. Biller, C.A. Erdman, M.J. Lipinski, A.E. Young, B.A. Roe, B. Qin, R.A. Grahn, Feline polycystic kidney disease mutation identified in PKD1, *J. Am. Soc. Nephrol.* 15 (2004) 2548–2555.
- K.M. Meurs, X. Sanchez, R.M. David, N.E. Bowles, J.A. Towbin, P.J. Reiser, J.A. Kittleson, M.J. Munro, K. Dryburgh, K.A. Macdonald, M.D. Kittleson, A cardiac myosin binding protein C mutation in the Maine Coon cat with familial hypertrophic cardiomyopathy, *Hum. Mol. Genet.* 14 (2005) 3587–3593.
- J.C. Fyfe, M. Menotti-Raymond, V.A. David, L. Brichta, A.A. Schäffer, R. Agarwala, W.J. Murphy, W.J. Wedemeyer, B.L. Gregory, B.G. Buzzell, M.C. Drummond, B. Wirth, S.J. O'Brien, An ~140-kb deletion associated with feline spinal muscular atrophy implies an essential LIX1 function for motor neuron survival, *Genome Res.* 16 (2006) 1084–1090.
- M. Menotti-Raymond, V.A. David, A.A. Schäffer, R. Stephens, D. Wells, R. Kumar-Singh, S.J. O'Brien, K. Narfström, Mutation in CEP290 discovered for cat model of human retinal degeneration, *J. Heredity* 98 (2007) 211–220.
- C. Hitte, J. Madeoy, E.F. Kirkness, C. Priat, T.D. Lorentzen, F. Senger, D. Thomas, T. Derrien, C. Ramirez, C. Scott, G. Evanno, B. Pullar, E. Cadieu, V. Oza, K. Lourgang, D.B. Jaffe, S. Tacher, S. Dreano, N. Berkova, C. André, P. Deloukas, C. Fraser, K. Lindblad-Toh, E.A. Ostrander, F. Galibert, Facilitating genome navigation: survey sequencing and dense radiation-hybrid gene mapping, *Nat. Rev. Genet.* 6 (2005) 643–648.
- W.J. Murphy, D.M. Larkin, A. Everts-van der Wind, G. Bourque, G. Tesler, L. Auviel, J. E. Beaver, B.P. Chowdhary, F. Galibert, L. Gatzke, C. Hitte, S.N. Meyers, D. Milan, E.A. Ostrander, G. Pape, H.G. Parker, T. Raudsepp, M.B. Rogatcheva, L.B. Schouk, L.C. Skow, M. Welge, J.E. Womack, J. O'Brien, S. P.A. Pevzner, H.A. Lewin, Dynamics of mammalian chromosome evolution inferred from multispecies comparative maps, *Science* 309 (2005) 613–617.
- J.U. Pontius, J.C. Mullikin, D.R. Smith, K. Lindblad-Toh, S. Gnerre, M. Clamp, J. Chang, R. Stephens, B. Neelam, N. Volkovskiy, A.A. Schäffer, R. Agarwala, K. Narfstrom, W.J. Murphy, U. Giger, A.L. Roca, A. Antunes, M. Menotti-Raymond, N. Yuhki, J. Pecon-Slatery, W.E. Johnson, G. Bourque, G. Tesler, S.J. O'Brien, Initial sequence and comparative analysis of the cat genome, *Genome Res.* 17 (2007) 1675–1689.
- M. Menotti-Raymond, V.A. David, A.A. Schäffer, J.F. Tomlin, E. Eizirik, C. Phillip, D. Wells, J.U. Pontius, S.S. Hannah, S.J. O'Brien, A third-generation autosomal genetic linkage map of the domestic cat, *Felis silvestris catus*, *Genomics* (in press).
- T. Derrien, C. André, F. Galibert, C. Hitte, AutoGRAPH: an interactive web server for automating and visualizing comparative genome maps, *Bioinformatics* 23 (2007) 498–499.
- A. Everts-van der Wind, D.M. Larkin, C.A. Green, J.S. Elliott, C.A. Olmstead, R. Chiu, J.E. Schein, M.A. Marra, J.E. Womack, H.A. Lewin, A high-resolution whole-genome cattle-human comparative map reveals details of mammalian chromosome evolution, *Proc. Natl. Acad. Sci. U. S. A.* 102 (2005) 18526–18531.
- W.G. Nash, J.C. Menninger, J. Wienberg, H.M. Padilla-Nash, S.J. O'Brien, The pattern of phylogenomic evolution of the Canidae, *Cytogenet. Cell Genet.* 95 (2001) 210–224.
- F. Yang, A.S. Graphodatsky, P.C. O'Brien, A. Colabella, N. Solanky, M. Squire, D.R. Sargan, M.A. Ferguson-Smith, Reciprocal chromosome painting illuminates the history of genome evolution of the domestic cat, dog and human, *Chromosome Res.* 8 (2000) 393–404.
- A.S. Graphodatsky, P.L. Perelman, N.V. Sokolovskaya, V.R. Beklemisheva, N.A. Serdukova, G. Dobigny, S.J. O'Brien, M.A. Ferguson-Smith, F. Yang, Phylogenomics of the dog and fox family (Canidae, Carnivora) revealed by chromosome painting, *Chromosome Res.* 16 (2008) 129–143.
- W.E. Johnson, E. Eizirik, J. Pecon-Slatery, W.J. Murphy, A. Antunes, E. Teeling, S.J. O'Brien, The late Miocene radiation of modern Felidae: a genetic assessment, *Science* 311 (2006) 73–77.
- D.H. Wurster-Hill, W.R. Centerwall, The interrelationships of chromosome banding patterns in canids, mustelids, hyena, and felids, *Cytogenet. Cell Genet.* 34 (1982) 178–192.
- R. Stanyon, M. Rocchi, O. Capozzi, R. Roberto, D. Misceo, M. Ventura, M.F. Cardone, F. Bigoni, N. Archidiacono, Primate chromosome evolution: ancestral karyotypes, marker order and neocentromeres, *Chromosome Res.* 16 (2008) 17–39.
- E. Eizirik, W.J. Murphy, *Carnivora*. in: S.B. Hedges, S. Kumar, S., (Ed.), *Assembling the Timetree of Life*, Oxford University Press, Oxford, in press.
- S. Rozen, H. Skaletsky, Primer3 on the WWW for general users and for biologist programmers, *Methods Mol. Biol.* 132 (2000) 365–386.
- R. Agarwala, D.L. Applegate, D. Maglott, G.D. Schuler, A.A. Schäffer, A fast and scalable radiation hybrid map construction and integration strategy, *Genome Res.* 10 (2000) 350–364.
- A.A. Schäffer, E.S. Rice, W. Cook, R. Agarwala, rh_tsp_map 3.0: end-to-end radiation hybrid mapping with improved speed and quality control, *Bioinformatics* 23 (2007) 1156–1158.
- D.L. Applegate, *The Traveling Salesman Problem: A Computational Study*, Princeton University Press, Princeton, 2006.
- D.M. Larkin, A. Everts-van der Wind, M. Rebeiz, P.A. Schweitzer, S. Bachman, C. Green, C.L. Wright, E.J. Campos, L.D. Benson, J. Edwards, L. Liu, K. Osogawa, J.E. Womack, P.J. de Jong, H.A. Lewin, A cattle-human comparative map built with cattle BAC-ends and human genome sequence, *Genome Res.* 13 (2003) 1966–1972.
- S.J. O'Brien, W.G. Nash, Genetic mapping in mammals: chromosome map of domestic cat, *Science* 216 (1982) 257–265.
- W.J. Kent, BLAT—the BLAST-like alignment tool, *Genome Res.* 12 (2002) 656–664.
- Z. Zhang, S. Schwartz, L. Wagner, W. Miller, A greedy algorithm for aligning DNA sequences, *J. Comput. Biol.* 7 (2000) 203–214.