

Free Fatty Acids and Fatty Acids of Triacylglycerols in Normal and Hyperkeratotic Human Stratum Corneum*

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The content of the free fatty acids and the fatty acids of triacylglycerols has been measured in human plantar stratum corneum from normal and hyperkeratotic subjects with palmoplantar keratoderma. Fatty acids of triacylglycerols in normal tissues showed a characteristic pattern with a relative abundance of short-chain length and unsaturated fatty acids. Free fatty acid fraction was characterized by the predominance of saturated compounds. The relative amount

of short-chain and monoene fatty acids in the hyperkeratotic stratum corneum was increased. These results seem to show a defect in the maturation of fatty acids in the living epidermis and present new evidence that the abnormality of lipid metabolism can influence the process of desquamation in stratum corneum. *J Invest Dermatol* 87:68-71, 1986

Important progress has been made in the study of the lipid composition of epidermis, particularly stratum corneum. Evidence for the involvement of lipids in stratum corneum cohesion and in a number of scaling skin disorders is well documented [1,2], particularly regarding the accumulation of cholesterol sulfate [3] in x-linked ichthyosis which is associated with a defect of steroid sulfatase [4]. Some of us have recently reported normal values of steroid-sulfatase activity in sole skin epidermis from postmenopausal women with palmoplantar keratoderma (PPK) [5]. It was concluded that abnormal desquamation in PPK was not related to modification of sterolsulfate hydrolysis.

Preliminary studies (Laurent, Nicollier, unpublished data) of epidermis from patients with various forms of PPK have frequently shown the presence of intracellular vacuoles. Such vacuoles with excess of neutral lipids, particularly triacylglycerols (TAG) have been reported in numerous defects of keratinization including psoriasis [6], lamellar ichthyosis [7], harlequin ichthyosis [8], and Refsum's disease [9]. The formation and accumulation of these droplets might be associated with a defect of fatty acid metabolism [1]. The literature is sparse as regards human stratum corneum free fatty acids (FFA) although they account for the major lipids of stratum corneum membrane complexes with cholesterol and ceramides [10,11]. Coon et al [12] have reported that the barrier zone lipids of epidermis include significant amounts of long-chain FFA but they attributed their

findings to contaminants. Later on, the importance of long-chain fatty acids was reported in stratum corneum [13] and particularly in the plantar stratum corneum where C₁₈ and C₂₄ predominate [14].

In order to provide quantitative analytical information about the fatty acids of the human plantar stratum corneum and to further understand the pathogenesis of palmoplantar hyperkeratosis, the fatty acid composition of TAG and FFA was studied both in specimens of normal plantar stratum corneum and specimens from patients with PPK.

MATERIALS AND METHODS

Tissues Normal tissues were obtained from cadavers (7 males and 7 females) within 4 h following death. Hyperkeratotic tissues were obtained from 13 patients with PPK. The diagnoses of all patients in this study were established by clinical and histologic criteria [15]. All the biopsies were performed with a Davol dermatome. Stratum corneum was isolated by heating the skin at 56°C for 3 min, and trypsinization: trypsin (DIFCO, 1:250) 0.3%, 45 min.

Lipid Extraction and Thin-Layer Chromatography Minced stratum corneum was homogenized in chloroform:methanol (2:1) (v:v) and lipids were extracted according to Bowyer and Janice [16]. The combined lipid extracts were washed with 0.2 vol of 0.1 M KCl and 0.2 vol of upper phase solvent containing KCl according to Folch et al [17]. The lower phase was evaporated in a rotary evaporator to dryness under vacuum at 35°C. The lipids were weighed and then redissolved in benzene and stored at -20°C.

The FFA and TAG were separated by thin-layer chromatography (TLC) according to the procedure of Nieminen et al [18]. Lipid classes were identified by cochromatography with the following standards: cholesterol, glycerol trioleate, and oleic acid (Sigma, St. Louis, Missouri). Standards were visualized by spraying with 50% H₂SO₄:ethanol (v:v) and heating at 110°C for 10 min. FFA and TAG were recovered from the silica gel 60 plates (Schleicher et Schull) by extraction with diethylether (2 × 3 ml).

Gas Liquid Chromatography (GLC) Methyl esters of FFA were prepared by the diazomethane method [19]. The TAG were

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Abbreviations:

- FFA: free fatty acid(s)
- GLC: gas liquid chromatography
- PPK: palmoplantar keratoderma
- TAG: triacylglycerol(s)
- TLC: thin-layer chromatography

Table I. Clinical Details of Patients with Palmoplantar Keratoderma (PPK)

| Classes | Patient Number | Age (years) | Sex | Diagnosis | Age at Onset | Localization |
|-------------|----------------|-------------|-----|--------------------------------|--------------|--|
| Idiopathic | 1 | 51 | F | Keratoderma climactericum | 50 | |
| | 2 | 49 | F | Keratoderma climactericum | 48 | |
| | 3 | 52 | F | Keratoderma climactericum | 51 | |
| | 4 | 56 | M | Striate PPK (Brunauer-Fuhs) | 20 | Soles (circumscribed) Palms (bandlike) |
| | 5 | 50 | M | Idiopathic PPK | 20 | Soles and palms circumscribed |
| | 6 | 39 | M | Mechanical painful callosities | 32 | Soles |
| Symptomatic | 7 | 70 | F | Pityriasis rubra pilaris | 69 | Soles and palms; erythroderma |
| | 8 | 58 | F | PPK with eczema | 57 | Soles |
| | 9 | 52 | F | PPK with eczema | 51 | Soles and palms |
| | 10 | 27 | M | PPK with eczema | 22 | Soles and palms |
| Inherited | 11 | 23 | F | Meleda disease | 3 months | Diffuse PPK of the soles and palms Extension of the lesions on the dorsal areas, ankles, and wrist |
| | 12 | 31 | M | Thost-Unna disease | Childhood | Diffuse PPK sharply limited to the soles and palms |
| | 13 | 32 | M | Epidermolytic PPK (Voerher) | Childhood | Diffuse PPK of the soles and palms |

deacylated with 0.5 N KOH-methanol and fatty acids were esterified by 20% boron trifluoride in methanol using the procedure of Van Wijngaarden [20]. The methyl esters were chromatographed using a Girdel 30 gas chromatograph (Suresnes, France) with a flame ionization detector equipped with a SE 52 open glass capillary column (25 m × 0.2 mm) (Carlo Erba, Milano, Italy), using temperature programming: 100°–290°C at 2°C/min. Peaks were identified by comparison of retention time values to known standards (Applied Sciences, State College, Pennsylvania) and by GLC-mass spectrometry. In all cases, the gas-chromatograms were recorded and the peak areas were measured with a ICR-1B computing integrator (Intersmat, Lyon, France).

Mass Spectrometry The gas chromatographic-mass spectrometric analyses were performed with a Ribier 10-10 B spectrometer (Rueil-Malmaison, France) on line to a Digital pdp 8/m data system (Westminster, Massachusetts). The SE 52 glass capillary column was coupled directly to the ion source of the mass spectrometer without any separator.

Statistical Analysis The data of all fatty acid quantifications in duplicate were statistically analyzed. Significant differences among groups were determined with the Mann Whitney-Wilcoxon test and the analysis of variance with an IBM 4341/2 computer system (Portsmouth, England).

RESULTS

The fatty acid content of plantar stratum corneum was measured in skin specimens from normal and abnormal subjects. A total of 27 skin specimens was studied including 7 specimens obtained from normal women and 7 specimens from normal men which constituted control specimens. Thirteen abnormal skin specimens were as shown in Table I: 6 from patients with idiopathic hyperkeratosis, 4 from patients with symptomatic hyperkeratosis, and 3 from patients with inherited hyperkeratosis. In all cases FFA and fatty acids of TAG were identified by their mass spectra after GLC-mass spectrometry.

In order to point out significant differences in the fatty acid composition between normal and pathologic stratum corneum, the relative abundance of each of them (in percentage of the total identified fatty acids) was compared (Tables II, III). It may be noticed that no hydroxy-fatty acid was identified in all tissues studied. The multivariate variance analysis showed no significant difference according to the sex, either in normal or pathologic tissues. Consequently, the variations were studied irrespective of this parameter. In normal stratum corneum, the relative abundance of C₁₄–C₁₈ fatty acids was lower and that of C₂₀–C₂₈ fatty

acids was higher in the free fraction than in the TAG fraction. Regardless of their chain length, unsaturated (monoene) fatty acids were more abundant in the TAG than in the free fraction. The 2 most abundant monoene fatty acids of the TAG fraction were C₁₈:1 and C₂₂:1. The latter was not found in the free fraction. The major diene fatty acid was C₁₈:2 and no significant difference was observed between the free fraction and TAG fraction.

The comparison between normal and pathologic tissues showed that C₁₄–C₁₈ FFA were more abundant in hyperkeratotic stratum corneum than in normal tissues whereas C₂₀–C₂₈ were less abundant. The most striking features in the TAG fraction were the significantly higher values found for C₁₈:1, C₁₈:2 fatty acids and the lower values for C₂₂:1 in hyperkeratotic stratum corneum than in normal stratum corneum.

The differences observed between pathologic and normal tissues are summarized in Table III. The 2 most important findings were: (1) a relative increase of the short chain length fatty acids (C₁₄–C₁₈) and unsaturated (monoene) fatty acids in hyperkeratotic stratum corneum; these modifications of fatty acid composition are particularly obvious in the free fraction; and (2) no significant difference among the different groups of pathologic tissue except for symptomatic keratoderma; in this group, there was a significant increase of di- and triene unsaturated fatty acids.

DISCUSSION

The study of FFA and fatty acids of TAG in the plantar stratum corneum from normal and hyperkeratotic subjects, after identification by their mass spectra, emphasizes some findings. In normal tissues, fatty acids of TAG showed a characteristic pattern: a relative abundance of unsaturated fatty acids with short-chain length, particularly C₁₈:1 and C₂₁:1. This predominance of unsaturated acids is in good agreement with the results reported by Lampe et al [14]. The differences observed for FFA between our results and those of Lampe et al [14] are due to the fact that these authors have not reported the values for odd-chain fatty acids which represent nearly 20% of the free fraction and 11.4% of the total fatty acids according to Ansari et al [13]. However, it is possible to characterize FFA by the abundance of saturated compounds.

The striking feature in hyperkeratotic stratum corneum is the relative abundance of short-chain and monoene fatty acids. The stratum corneum fatty acids initially derive from the content of lamellar bodies which are synthesized primarily within the spinous cells. A comparison of the fatty acid composition of lipids from the living layers with that found in the stratum corneum

Table II. Fatty Acid Composition of Normal and Hyperkeratotic Stratum Corneum Lipids

| | Percent Free Fatty Acids (means) ^a | | | Percent Fatty Acids of Triacylglycerols (means) ^a | | |
|--------------------|---|---|-----------------------|--|---|-----------------------|
| | Normal Stratum Corneum (n = 14) | Hyperkeratotic Stratum Corneum (n = 13) | <i>p</i> ^b | Normal Stratum Corneum (n = 13) | Hyperkeratotic Stratum Corneum (n = 13) | <i>p</i> ^b |
| C ₁₄ :0 | 0.7 | 2.0 | 0.001 | 3.9 | 2.2 | 0.044 |
| C ₁₅ :0 | 0.8 | 2.2 | 0.000 | 2.9 | 2.7 | NS |
| C ₁₆ :0 | 9.2 | 17.5 | 0.001 | 18.2 | 14.9 | NS |
| C ₁₆ :1 | 0.2 | 0.4 | 0.001 | 2.8 | 2.6 | NS |
| C ₁₆ :1 | 0.5 | 2.1 | 0.001 | 0.8 | 2.7 | NS |
| C ₁₇ :0 | 1.7 | 2.6 | 0.041 | 2.6 | 2.5 | NS |
| C ₁₇ :1 | — | 0.8 | — | 0.6 | 1.3 | NS |
| C ₁₈ :0 | 11.9 | 16.8 | 0.002 | 14.8 | 12.2 | NS |
| C ₁₈ :1 | 1.2 | 3.3 | 0.002 | 3.1 | 5.7 | 0.000 |
| C ₁₈ :1 | 9.6 | 20.2 | 0.000 | 17.6 | 25.1 | 0.003 |
| C ₁₈ :2 | 7.2 | 6.7 | NS | 7.9 | 11.6 | NS |
| C ₁₉ :0 | 1.9 | 1.4 | 0.012 | 1.3 | 0.9 | NS |
| C ₁₉ :1 | — | — | — | — | 0.2 | — |
| C ₂₀ :0 | 5.1 | 2.9 | 0.000 | 2.2 | 1.4 | NS |
| C ₂₀ :1 | — | 0.8 | — | 0.3 | 0.9 | NS |
| C ₂₀ :1 | — | 0.9 | — | 1.0 | 2.3 | NS |
| C ₂₀ :2 | — | 0.4 | — | 0.2 | 1.1 | 0.008 |
| C ₂₀ :3 | — | 0.1 | — | — | 0.8 | — |
| C ₂₁ :0 | 2.6 | 0.9 | 0.000 | 1.1 | 0.4 | 0.008 |
| C ₂₂ :0 | 11.0 | 3.9 | 0.000 | 2.6 | 1.3 | 0.011 |
| C ₂₂ :1 | — | 0.3 | — | 0.2 | 0.5 | 0.011 |
| C ₂₂ :1 | — | 1.2 | — | 8.5 | 3.5 | 0.003 |
| C ₂₃ :0 | 7.1 | 1.9 | 0.000 | 1.3 | 0.4 | 0.009 |
| C ₂₄ :0 | 18.0 | 6.2 | 0.000 | 3.3 | 1.4 | 0.003 |
| C ₂₄ :1 | — | 1.6 | — | 0.2 | 0.9 | 0.003 |
| C ₂₅ :0 | 4.2 | 1.1 | 0.000 | 0.9 | 0.5 | 0.004 |
| C ₂₆ :0 | 4.5 | 1.5 | 0.000 | 0.8 | 0.3 | 0.008 |
| C ₂₇ :0 | 1.0 | 0.1 | 0.001 | 0.2 | — | — |
| C ₂₈ :0 | 1.5 | 0.4 | 0.001 | 0.6 | 0.1 | 0.008 |

^aCompositions are expressed in weight percentages of identified fatty acid methyl esters.

^bSignificant differences for *p* < 0.05 Mann-Whitney Wilcoxon test. NS = Nonsignificant, *p* > 0.05. — = Not detected.

indicates that the normal stratum corneum lipids are strikingly enriched in long-chain fatty acid, fully saturated acyl groups [2]. We can presume that such maturation of fatty acids in hyperkeratotic skin does not occur. Two clusters of fatty acids in epidermis can be distinguished [13]: one of these clusters ranged from C₁₂–C₁₈ and the other one ranged from C₁₉–C₂₈. The particular fatty acid composition of the hyperkeratotic stratum corneum may be due either to a decreased oxidation of fatty acids of the first cluster which supplies cell energy [13] or to a defect

in chain elongation leading to the fatty acids of the second cluster [21] and to an increase in desaturase activity [13]. The hypothesis of unspecific perturbation in the fatty acid metabolism of the hyperkeratotic stratum corneum is supported by the fact that the shift toward short-chain and monoene fatty acids was constantly found in PPK from various causes. Hence, it is not disease-specific. However, the molecular mechanism(s) of the metabolism perturbation remains unknown. How can these fatty acid modifications influence desquamation in the hyperkeratotic stratum

Table III. Relative Amounts of the Different Fatty Acid Types in Normal and Hyperkeratotic Stratum Corneum and Ratio of Fatty Acids Proportionate to Their Chain Length

| Fatty Acid Type | Normal Stratum Corneum (n = 14) | Hyperkeratotic Stratum Corneum | | | |
|---|---------------------------------|--------------------------------|----------------------------|-----------------------------|------------------------------|
| | | Total (n = 13) | 1 → 6 ^a (n = 6) | 7 → 10 ^a (n = 4) | 11 → 13 ^a (n = 3) |
| Free fatty acid | | | | | |
| Saturated | 81.2 | 62.5 | 59.5 | 57.7 | 69.4 |
| Monoene | 11.5 | 31.0 | 35.7 | 30.0 | 24.7 |
| di and triene | 7.2 | 6.6 | 4.7 | 12.3 | 6.0 |
| C ₁₄ → C ₁₈ above C ₁₈ | 0.8 | 2.7 | 3.2 | 4.2 | 1.5 |
| Triacylglycerol | | | | | |
| Saturated | 56.7 | 41.6 | 41.1 | 39.8 | 42.3 |
| Monoene | 35.1 | 45.6 | 50.3 | 39.4 | 44.5 |
| di and triene | 8.1 | 12.8 | 8.5 | 20.8 | 13.2 |
| C ₁₄ → C ₁₈ above C ₁₈ | 3.0 | 4.9 | 4.8 | 5.6 | 4.9 |

^aPatients as numbered in Table I, were gathered in relation with the pathology.

corneum? We have to consider the role of fatty acids in membrane fluidity. Elias [2] has proposed a heterogeneous, 2-compartment model of the stratum corneum that ascribes a special role to intercellular lipids in the regulation of both barrier function and desquamation. For the latter he concluded that neutral lipids can influence the process of normal desquamation by their mass impact in stratum corneum lipid thermal transition. It is well known that the phase transition of lipid bilayers decreases with increasing unsaturation or decreasing chain length of the fatty acid chains [22]. Consequently, because of the abundance of neutral lipids in stratum corneum [14] the phase transition of the intercellular lipids is lower in the hyperkeratotic stratum corneum than in the normal tissue and can explain its abnormal desquamation.

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