# The spectrum of podocytopathies: A unifying view of glomerular diseases

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Glomerular diseases encompass a broad array of clinicopathologically defined syndromes which together account for 90% of end-stage kidney disease costing \$20 billion per annum to treat in the United States alone. Recent insights have defined the central role of the podocyte as both the regulator of glomerular development as well as the determinant of progression to glomerulosclerosis. We can now place all glomerular diseases within this spectrum of podocytopathies with predictable outcomes based on podocyte biology impacted by temporal, genetic, and environmental cues. This simplified construct is particularly useful to rationalize clinical effort toward podocyte preservation and prevention of progression as well as to focus basic research effort on understanding podocyte biology and for clinical research toward development of practical monitoring strategies for podocyte injury, dysfunction, and loss.

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KEYWORDS: podocyte; glomerulus; glomerulonephritis; glomerulonephropathy; glomerulosclerosis; nephrotic syndrome Glomerular diseases account for 90% of end-stage kidney disease (ESKD) at a cost of \$20 billion per year in the US.<sup>1</sup> The traditional glomerular disease classification encompasses a bewildering array of descriptive pathologic entities and their clinical counterparts. And yet clinical trials and experience, guided by the hyperfiltration hypothesis of Brenner *et al.*,<sup>2</sup> tell us that effective blood pressure control, angiotensin II inhibition, and reduction in proteinuria are almost uniformly effective in reducing the rate of progression. This implies that there are common mechanisms driving disease progression.

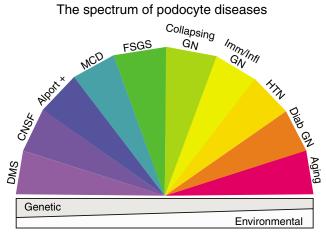
Recent identification of mutations causing glomerular phenotypes, clinical biopsy reports, and experimental model systems reveal a simplified concept of glomerular diseases in which podocyte dysfunction, injury, or loss is a common and determining factor. Toxic, genetic, immune, infectious, oxidant, metabolic, hemodynamic, and other mechanisms can all target the podocyte. Depending on the stage of glomerular development and associated environmental factors, podocyte dysfunction, injury, or loss can result in a broad spectrum of clinical syndromes (Figure 1). Considering glomerular diseases as being related by these common podocyte-dependent mechanisms rather than as separate pathobiological entities facilitates progress towards prevention and treatment. Any national plan aimed at reducing the cost of health care will need to focus basic scientific effort on understanding podocyte biology and clinical research on learning how to prevent and monitor podocyte injury and depletion as major targets for intervention.

The development and maintenance of normal glomerular structure and function requires successful signaling and coordination between all glomerular cells including mesangial, endothelial, parietal epithelial, and visceral epithelial cells (podocytes). This point is emphasized by the critical requirement for vascular endothelial growth factor-A production by podocytes for normal endothelial and mesangial cell development and function.<sup>3,4</sup> However, the podocyte is the key organizer of glomerular development and maintenance.<sup>5,6</sup> This discussion will therefore be focused on the podocyte.

Figure 2 emphasizes, on the one hand, the key role of the podocyte in governing glomerular development culminating in the normal mature glomerulus. On the other, Figure 2 also depicts the impact of podocyte injury and depletion leading

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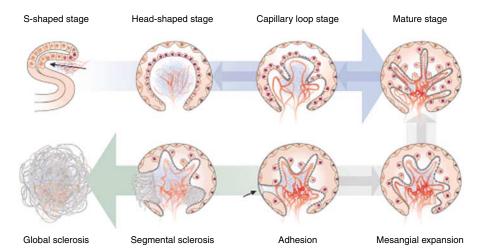
**Figure 1** | **The spectrum of podocyte diseases.** Glomerular diseases can be considered as a consequence of podocyte dysfunction caused by genetic and/or environmental factors. Depending on when, during glomerular development and progression they occur and how significantly and rapidly they impact the glomerulus the clinical phenotype will vary. The clinical syndromes associated with podocyte dysfunction of various types include the following: diffuse mesangial sclerosis (DMS), congenital nephrotic syndrome of the Finnish type (CNSF), Alport's syndrome and variants (Alport + ), MCD, FSGS, collapsing glomerulonephropathy (Collapsing GN), immune and inflammatory glomerulonephropathies (Imm/Inf GN), hypertensive nephropathy (HTN), diabetic glomerulonephropathy (Aging). Together these conditions account for 90% of ESKD.

ultimately to global glomerulosclerosis. Variations in this basic pathway are specialized to particular clinical settings and result in a modified pathologic appearance of the glomerulus (e.g., diabetic glomerulosclerosis). Nevertheless, the major sequence of events can conveniently be considered as a continuum that plays out over time to a greater or lesser extent in an individual person depending on factors contributed by genetic and environmental milieus.

#### **GLOMERULAR DEVELOPMENT**

Podocytes become recognizable by their expression of key proteins in the S-shaped stage of nephron development (Figure 2).<sup>5,6</sup> They surround the in-growing blood vessels and mesenchyme to form a 'halo' or 'head-shaped' structure. By this time, podocalyxin is present on the apical surface of podocytes and tracks the intercellular junctions as they migrate down the lateral cell surface of the podocyte towards the developing glomerular basement membrane (GBM).<sup>7</sup>

During the capillary loop stage of glomerular development, there is a major expansion of the GBM requiring bulk protein synthesis of type IV collagens, laminins, and glycosaminoglycans.<sup>8</sup> At the same time, podocytes begin forming the interdigitating foot processes that markedly increases the space between podocytes necessary for efficient filtration. These foot processes connect the cell to the underlying GBM via an intergrin-linked adhesion mechanism.<sup>9</sup> The specialized intercellular junctions (slit diaphragms) that bridge between foot processes are made up of special proteins



**Figure 2** | **Stages of glomerular development and progression with emphasis on the role of the podocyte.** The S-shaped stage. Developing podocytes acquire podocyte markers during the S-shaped stage of nephron development at which time blood vessels and mesenchyme invade (arrow). The podocytes (round nuclei) separate from the parietal epithelial cells (triangular nuclei) forming what will become Bowman's space. The head-shaped stage: at this stage of glomerular development, the glomerulus consists of a ball of cell surrounded by developing podocytes. The capillary loop stage: this stage of glomerular development includes the infolding of the surface layer in order to enlarge the area available for filtration as well as the development of foot processes that interdigitate between podocytes and abut the underlying GBM which is being synthesized as a collaboration between the podocyte and underlying endothelial and mesangial cells. The mature glomerulus has maximized filtration surface area by developing intertwining finger-like projections coated by fenestrated endothelial cells on the inside, a specialized strong thin GBM in the middle, and interdigitating podocyte foot processes connected by slit diaphragms on the outer surface. Mesangial expansion: loss of some podocytes (20%) is associated with mesangial expansion possibly as an attempt to reduce the filtration surface area. Adhesion formation: loss of podocytes resulting in appearance of bare areas of filtration surface results in adhesion of the bare surface to Bowman's capsule (synechia). Segmental sclerosis: loss of podocytes beyond a critical level results in a fibrotic glomerular response in that part of the glomerulus (segmental sclerosis).

required for the filtration, sensing, and signaling functions of the podocyte. They include nephrin, podocin, CD2AP, neph1, fyn, yes, and others.<sup>10–15</sup> As the filtration surface of the mature glomerulus develops in association with an expanded GBM area, each podocyte has to communicate with and support its multiple and distant foot processes by outreach through major and intermediate cell processes that collectively give rise to the octopus-like shape of the mature podocyte. The GBM itself is constructed as a thin yet strong and efficient filtration surface through contributions by the podocyte, endothelial, and mesangial cells including the specialized  $\alpha$ 3,  $\alpha$ 4, and  $\alpha$ 5 type IV collagen chains.<sup>16</sup>

The key organizational signaling systems required to create and maintain the complex glomerular structure are not well understood. Master regulatory genes including the transcription factor *WT1* that control programs of downstream gene expression are necessary.<sup>17–19</sup> As noted above, coordination between glomerular cells also plays a key role as demonstrated by the critical requirement for vascular endothelial growth factor production by podocytes for normal endothelial and mesangial cell development.<sup>3,4</sup> Ultimately, the process of nephrogenesis results in the production of two kidneys, each containing approximately 1 million nephrons, which collectively filters 100 ml of blood plasma per minute with high efficiency.

#### REPLACEMENT OF PODOCYTES AND COMPENSATORY PODOCYTE HYPERTROPHY

The mature podocyte has limited capacity to divide in situ.<sup>20,21</sup> To what extent podocytes can be replaced at all during adult life, and if so, how and at what rate, has not yet been established. One potential mechanism for podocyte replacement is stem cell immigration from the bone marrow via blood as has been demonstrated in model systems.<sup>22-24</sup> A second possibility is the migration of cells from the parietal epithelial cell layer along Bowman's capsule onto the glomerular tuft together with a switch to the podocyte phenotype. In the first case, the stem cell has to traverse the GBM to home to its destination, whereas in the second case this is not necessary. A related question is the extent to which podocytes can hypertrophy to replace lost neighbors or to compensate for glomerular enlargement. Both human biopsy analysis as well as animal models show that podocytes can undergo significant hypertrophy but to a limited extent.<sup>25-27</sup> In a model system, as podocytes continue to enlarge they express different proteins reflecting an 'adaptation' stage and subsequently a 'decompensation' stage before becoming lost from glomeruli.<sup>27</sup> Whatever the mechanisms involved in podocyte hypertrophy and replacement it is possible that they occur slowly so that podocyte injury leading to podocytopenia can become a major threat to the glomerulus.

### **PROGRESSION TO GLOMERULOSCLEROSIS**

The sequence of anatomic events resulting from podocyte injury has been comprehensively described in multiple animal model systems by Kriz.<sup>20</sup> This includes the appearance

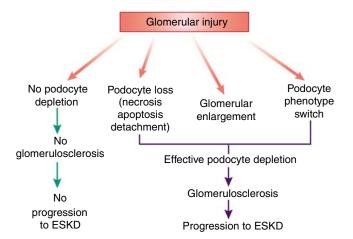
of denuded areas of GBM, adhesion to Bowman's capsule (synechia formation), focal segmental glomerulosclerosis, and global glomerulosclerosis associated with misdirected filtration into the interstitial compartment contributing to interstitial injury and fibrosis. The consequences of different degrees of podocyte loss is exemplified using rat models of regulated podocyte depletion.<sup>28,29</sup> In these models, loss < 20% of podocytes causes mesangial expansion alone. Loss of more then 20% of podocytes leads to the appearance of denuded areas of GBM resulting in adhesion of the glomerular capillary loop to Bowman's capsule (synechia formation). With 20-40% podocyte loss, selected glomerular capillary loops become devoid of podocytes and result in a scarring response limited to that particular loop (segmental sclerosis). With more than 40% podocyte loss, the glomerulus becomes progressively more sclerotic until at >60% podocyte loss glomeruli become globally sclerotic and nonfiltering. These stages of podocyte depletion are accompanied by corresponding degrees of proteinuria and, as an increasing proportion of glomeruli become involved by measurable reduction in the clearance function of the kidney.

Glomerular enlargement is also associated with development of glomerulosclerosis.<sup>30,31</sup> This is true for reduction in renal mass (nephron number) as well as endocrine conditions such as acromegaly and obesity in man and in experimental models of overexpression of growth factors, partial nephrectomy, and high calorie intake. The sequence of events flowing from glomerular enlargement as outlined above is determined by the limited extent to which podocytes can hypertrophy to accommodate the increased GBM area to be served.<sup>20,27</sup> Exceeding this capacity appears to lead to glomerulosclerosis via similar mechanisms to those outlined for direct podocyte depletion.

### THE PODOCYTE DEPLETION HYPOTHESIS

This hypothesis states that whatever the initial insult to the glomerulus (immune, toxic, infectious, ischemic, etc), the outcome depends on whether or not the complement of normal mature podocytes becomes depleted (Figure 3). On the one hand, if as a result of glomerular injury podocytes are not depleted, then the glomerulus has the capacity to remodel and recover essentially normal structure and function. On the other hand, if significant podocyte depletion occurs, then the glomerulus (or that part of the glomerulus) will not recover its normal structure and function. If progressive podocyte depletion is allowed to occur over time, then this will be associated with progressive glomerulosclerosis leading to progressive loss of renal function culminating in ESKD. Podocyte depletion can occur as a result of (a) direct podocyte loss because of necrosis, apoptosis, or detachment; (b) by glomerular enlargement leading to relative podocyte depletion; or (c) by a switch of the podocyte phenotype to one which cannot maintain normal glomerular structure and function.

As outlined above, many potential mechanisms may trigger podocyte injury and depletion. The loss of some



**Figure 3** | **The podocyte depletion hypothesis.** This hypothesis states that the outcome for an individual glomerulus will depend on whether glomerular injury results in significant podocyte depletion or not. Podocyte 'depletion' is considered to be depletion of the complement of mature podocytes necessary for maintenance of the normal mature glomerulus. Podocyte 'depletion' can therefore occur by podocyte loss itself, by glomerular enlargement leading to expansion of GBM area to be served, or by alteration of the podocyte phenotype such that the podocyte-derived cell is no longer able to fulfill its normal mature functions, such as occurs in collapsing glomerulopathy and crescentic nephritis.

podocytes for any reason may trigger a vicious cycle leading to the loss of more podocytes.<sup>32</sup> Angiotensin II may itself directly amplify this effect,<sup>33</sup> thus providing a potential explanation for the well-known protective effect of angiotensin II inhibition in slowing and preventing progression in glomerular diseases.

## THE SPECTRUM OF SYNDROMES ASSOCIATED WITH ALTERED PODOCYTE BIOLOGY

As indicated by Figure 1 and outlined in sections below, the different clinical syndromes of glomerular diseases can each be directly related to various aspects of altered podocyte biology. These include podocyte developmental arrest, defective podocyte products, phenotypic switch and podocyte loss, dedifferentiation, and proliferation.

#### Arrested glomerular development

Glomerular developmental arrest as a cause of diffuse mesangial sclerosis. Mutations that result in arrested glomerular developmental at the 'head-shaped' or early capillary loop stage of glomerular development (Figure 2) cause the clinical syndrome of diffuse mesangial sclerosis (DMS). Such arrested glomeruli can still filter to some extent but they leak protein into the filtrate (nephrotic syndrome) and rapidly progress to ESKD. The clinical syndrome of DMS includes proteinuria from birth (congenital nephrotic syndrome) and progression to ESKD within 6–12 months of age.<sup>34,35</sup> The pathologic appearance of DMS glomeruli includes a halo-shaped distribution of podocytes surrounding a matrix-containing glomerular center ('DMS'), absent foot processes, and lack of normal glomerular capillary loops.

The recent identification of phospholipase C epsilon (PLCE1) truncating mutations as a cause of DMS is helpful for understanding glomerular biology.<sup>36</sup> As PLCE1 is expressed by the podocyte in the developing glomerulus, the fact that mutations in PLCE1 gene prevent normal glomerular development confirms the central role of the podocyte in this process. Similarly, WT1 is a major podocyte transcription factor present early in podocyte development, which regulates expression of key proteins by the podocyte including nephrin and podocalyxin.<sup>37,38</sup> Mutations in the WT1 gene also cause DMS.<sup>17-19,39</sup> Furthermore, mutations in the  $\beta$ 2laminin (*LAMB2*) gene cause DMS presumably as a result of defective GBM expansion and assembly required for normal glomerular development.<sup>40,41</sup> The fact that mutations in the above noted genes expressed by podocytes all cause a similar glomerular phenotype provides a glimpse of the part of this regulatory machinery necessary for early stages of glomerular development.

Glomerular developmental arrest as a cause of the congenital nephrotic syndrome of the Finnish type. Glomerular developmental arrest at the late capillary loop stage of glomerular development causes massive life-threatening proteinuria as congenital nephrotic syndrome of the Finnish type. Having constructed the enormous filtration surface area during the capillary loop stage of glomerular development, it becomes necessary that the filtration machine efficiently prevent loss of blood protein into the filtrate. This goal is achieved in part through the interdigitating podocyte foot processes and specialized intercellular junctions (slit diaphragms). By these mechanisms the podocyte provides physical support to the GBM to counteract the hydrostatic pressure of blood as well as supplies special molecules required by the GBM for its normal structure and function. The foot processes and slit diaphragms also serve as information mechanisms that signal outside-in to the podocyte about space between foot processes, pressure, and similar information required to maintain efficient filtration. The proteins that make up the slit diaphragm and its signaling system include nephrin, podocin, neph1, and their signaling partners fyn, yes, nck, CD2AP, and others.<sup>10–15,32–46</sup>

Failure to construct normal foot processes and intercellular junctions during development results in massive proteinuria before and immediately after birth, giving rise to the syndrome of congenital nephrotic syndrome of the Finnish type. The major causes of congenital nephrotic syndrome of the Finnish type in man are mutations in the *NPHS1* gene coding for the slit diaphragm protein nephrin and in the *NPHS2* gene coding for the foot process membrane protein podocin.<sup>42,43</sup> These mutations result in grossly normal capillary loop development with the key exception that foot processes and slit diaphragms fail to form normally. The result is a leaky glomerular filter and consequent life-threatening loss of protein into the urine.

### Defective GBM construction and maintenance (Alport's syndrome and variants)

The requirement of the GBM to be strong and rugged yet thin and discriminating is achieved in part by special type IV

collagen trimers comprised an  $\alpha$ 3,  $\alpha$ 4, and  $\alpha$ 5 chain of type IV collagen.<sup>16</sup> These type IV collagens are produced by the podocyte and inserted into the GBM during development and adult life as required for GBM maintenance. Mutations in the  $\alpha$ 3 and  $\alpha$ 4 chains (COL4A3 and COL4A4) can result in a thin GBM ('thin GBM disease'). Thin GBM is liable to rupture causing hematuria, but does not usually cause progressive loss of renal function or significant proteinuria.<sup>47</sup> Rarely, mutations in  $\alpha 3$  and  $\alpha 4$  type IV collagen genes (COL4A3 and COL4A4) are associated with progressive loss of kidney function.<sup>48</sup> Mutations in  $\alpha 5$  type IV collagen COL4A5 cause Alport's syndrome resulting in defects in the glomerular GBM, the lens of the eye, and the cochlear membrane. The associated clinical syndrome includes renal failure, deafness, and visual defects.<sup>49</sup> Because the COL4A5 gene is on the X chromosome, Alport's syndrome is a sex-linked condition primarily affecting males.

#### Podocyte phenotypic switch and minimal change disease

In minimal change disease (MCD) podocytes undergo a phenotypic switch from a cell with foot processes protruding from its basal surface to a cell without basal processes but which has acquired microvillus protrusions on its apical cell surface ('microvillus transformation') similar to those present in renal tubular cells. This remarkable alteration in podocyte structure is not associated with reduction in expression of key podocyte proteins such as nephrin or podocin, such as is seen in forms of focal segmental glomerulosclerosis (FSGS),<sup>50,51</sup> although podocyte  $\alpha$ -dystroglycan, responsible in part for adherence of the podocyte to the underlying GBM is reduced in MCD.<sup>52</sup> MCD can therefore be considered as a podocyte phenotypic switch. It is typically triggered by allergic and immune events including lymphomas implying that circulating factors such as cytokines may trigger the switch, possibly in genetically susceptible individuals.<sup>53</sup> This concept is consistent with the ability of glucocorticoids to rapidly induce a switch reversal back to the normal podocyte phenotype, analogous to glucocorticoid being used to promote maturation (and surfactant production) by alveolar epithelial cells in the immature lung. The clinical consequence of the 'switched' podocyte in MCD is decreased efficiency of the filtration mechanism with loss of large amounts of albumin into the glomerular filtrate causing nephrotic syndrome. Podocytes are not lost into the urine in MCD with maintenance of the normal podocytes numbers compatible with the excellent long-term prognosis for renal function.54

# Podocyte depletion from the glomerulus (podocytopenia) and FSGS

FSGS is a heterogeneous condition that has been recognized as being associated with podocyte injury for more than 30 years.<sup>55,56</sup> The sequence of events by which podocyte injury results in denudation of GBM, adhesions to Bowman's capsule, and progression through segmental to global sclerosis has been rigorously mapped by Kriz<sup>20</sup> and in numerous experimental reports. Direct quantitative demonstration of podocyte loss as a cause of FSGS has been demonstrated in rat models.<sup>28,29</sup> The concept that podocyte depletion causes FSGS in man is also supported by glomerular podocin mRNA being relatively depleted in FSGS compared with normal glomeruli<sup>57</sup> and finding that podocytes are lost into the urine in FSGS.<sup>58</sup> These data are compatible with the hypothesis that FSGS can be caused by podocyte depletion in man, but whether or not all FSGS in man is caused by podocyte depletion is not yet established.

For podocytes to serve their function, they must cover the filtration surface area with foot processes. Glomerular enlargement is known to be directly associated with development of FSGS, and is found in association with obesity, endocrine conditions such as acromegaly and diabetes, loss of kidney mass through nephrectomy, chronic glomerulosclerosis, and also in glomerulopenia resulting from premature birth and other causes.<sup>30,31,59,60</sup> Quantitative morphologic data show that FSGS is associated with increased GBM length per podocyte as further support for the hypothesis that relative podocyte depletion resulting from glomerular enlargement can contribute to FSGS.25 Furthermore, in model systems of glomerular enlargement in rats the podocyte number did not increase as glomerular volume increased.<sup>27</sup> Rather, compensatory podocyte hypertrophy occurred up to a threshold point before FSGS supervened, as would be predicted if relative podocyte depletion was taking place in the enlarging glomerulus.

Mutations in genes coding for key podocyte proteins also cause FSGS. Autosomal recessive mutations such as NPHS2 (podocin) and PLCE1 (PLC epsilon non-truncating mutation) cause FSGS in children and young adults.<sup>14,36</sup> Autosomal dominant gain of function mutations in genes coding for α-actinin 4 and TRPC6 cause FSGS in adults.<sup>61-65</sup> We do not yet know whether these mutations cause FSGS through promoting podocyte loss in man or through some other mechanism. Similarly, experimental deletions of genes coding for proteins that play a role in podocyte structure and function causes FSGS in model systems under normal aging or stressed conditions (reviewed in Mundel et al.,<sup>10</sup> Pavenstadt et al.,<sup>11</sup> Asanuma and Mundel,<sup>12</sup> Ly et al.,<sup>13</sup> Antignac,<sup>14</sup> and Huber and Benzing<sup>15</sup>). Transforming growth factor $\beta$  overexpression is an example of a model where podocyte number has been shown to be reduced in association with development of FSGS.<sup>66</sup> Circulating factors,<sup>67</sup> toxic factors including drugs, oxidants, and deposition of glycosphingolipids in Fabry's disease<sup>68</sup> and crystallization of myeloma protein in podocytes<sup>69</sup> all cause podocyte injury leading to FSGS and account in part for the heterogeneity of this pathologic entity.<sup>56</sup>

### Podocyte dedifferentiation causes 'collapsing glomerulopathy'

HIV-associated nephropathy and pamidronate toxicity are associated with a glomerular phenotype described as 'collapsing glomerulopathy' in which the glomerulus loses its normal structure including its capillary loops.<sup>56,70–73</sup> In this condition, mature podocyte markers including WT1, nephrin, podocalyxin, GLEPP1, the cyclin-inhibitors p21, and p27 and foot processes are lost. In contrast, markers of immature dividing podocytes are acquired and the podocyte becomes capable of proliferating. Expression of the HIV protein nef specifically by podocytes in a transgenic model system is sufficient to cause this phenotype, thereby confirming that this is a podocyte disease.<sup>70</sup> In the presence of such dedifferentiated podocytes (absence of normal mature podocytes), the whole glomerulus can be viewed as itself dedifferentiating backwards towards a phenotype resembling the head- or halo-shaped early capillary loop stage of glomerular development where podocytes are arranged on the peripheral surface of the glomerular tuft before capillary loop formation and which is also present in DMS (Figure 2). Collapsing glomerulopathy is therefore mechanistically and structurally distinct from other forms of FSGS. Accumulating data suggest that eradication of the HIV infection allows podocytes to differentiate back to the mature phenotype with improvement in the filter characteristics and preservation of renal function.74

# Podocyte injury, loss, and proliferation in immune and inflammatory glomerulopathies

The outcome in immune and inflammatory glomerulonephropathy is strikingly variable even within descriptive pathologic types. According to the podocyte depletion hypothesis, whatever the underlying form of immune injury, those individuals in whom immune injury causes podocyte loss would develop proteinuria, glomerulosclerosis, and progressive loss of kidney function, whereas those where similar injury did not result in podocyte loss would not develop glomerulosclerosis. For example, in immunoglobulin A nephropathy biopsy analysis shows a correlation between podocyte depletion and development of glomerulosclerosis.<sup>75</sup> People with active immunoglobulin A nephropathy, systemic lupus erithematosus, and membranoproliferative glomerulonephritis have increased numbers of podocytes in their urine,<sup>76–79</sup> which can be eliminated by effective treatment.<sup>80</sup> Membranous nephropathy is well known to be associated with podocyte injury owing to insertion of complement C5b-9 complex into podocyte plasma membranes.<sup>81-83</sup> To what extent the difference between progressive and nonprogressive membranous nephropathy is determined by whether or not podocytes are lost from glomeruli is not yet known.

*The special case of crescentic nephritis.* In crescentic nephritis, dedifferentiated podocytes in the process of losing their mature markers can be identified as present in the glomerular crescent filling Bowman's space, possibly contributing to podocytopenia via this mechanism.<sup>84,85</sup> Quaggin and co-workers have demonstrated using a transgene model of Von Hippel–Lindau gene deletion from podocytes that podocytes themselves can drive the crescentic process in part via expression of the hypoxia-inducible factor target gene

Cxcr4.<sup>86</sup> They also demonstrate that this signaling system is present in crescentic human glomeruli from patients with systemic vasculitis. Thus, under certain conditions podocytes can change their phenotype, proliferate to form at least a part of the glomerular crescent, and themselves drive the crescentic process towards a sclerotic outcome.

### Hypertensive glomerulosclerosis and the podocyte

There is no doubt that systemic hypertension superimposed on glomerular disease of any kind leads to more rapid loss of renal function.<sup>87</sup> Blood pressure control and use of angiotensin-converting enzyme-inhibitors in glomerular diseases reduces loss of podocytes in the urine.<sup>75–77</sup> African Americans are particularly susceptible to developing ESKD in association with hypertension and serve as an important example of hypertension-related kidney failure.<sup>88</sup> Low glomerular number associated with low birth weight cannot account for this susceptibility, at least in African Americans.<sup>89</sup> Biopsy material from adult hypertensive African Americans shows that about 13% have typical FSGS lesions.<sup>90</sup> A large proportion of the remainder have a focal and global solidification of the glomerulus described as 'decompensated benign nephrosclerosis?<sup>91</sup> Kimmelstiel and Wilson<sup>92</sup> and Helmchen and Wenzel93 noted that the earliest glomerular changes in this lesion, which they called 'alterative glomerulitis', was 'swelling of the epithelial cells of the glomerular loops' and that 'early fibrinous adhesions to the parietal layer of Bowman's capsule may be present'. The recent observation that mutations in WT1 are associated with FSGS in the African-American population fits in with the general concept that a podocyte abnormality could play a role.<sup>94</sup> African Americans, like the general hypertensive population, are protected from progression by angiotensin-converting enzyme inhibition and effective blood-pressure control.95 To what extent progression to ESKD in different hypertensive groups can be attributed primarily or secondarily to podocyte dysfunction/depletion has not yet been determined.

# Diabetic glomerulosclerosis as a consequence of glomerular enlargement and podocyte loss

The concept that podocyte injury is central to progression in diabetic nephropathy is now well established.<sup>96–98</sup> Pathologic biopsy reports in type I diabetic glomerulosclerosis and type II diabetic glomerulosclerosis directly link reduced glomerular podocyte number to increased proteinuria99-102 and show that glomerular podocyte number is the best predictor of glomerular outcome in type II diabetes in Pima Indians.<sup>103</sup> This biopsy data are supported by urine podocyte number data in man where progressive diabetic glomerulopathy is associated with increased podocyturia and data showing that some diabetic treatment strategies reduce podocyturia.<sup>104-107</sup> Furthermore, there is extensive experimental data showing that podocytes injury is an early detectable event after onset of diabetes as well as reports linking progressive loss of podocytes to progression of glomerular disease in model systems.<sup>108</sup> Thus, the concept of podocyte loss as a driver of the glomerulosclerotic process in diabetes is well documented. The underlying mechanisms are likely to be in part related to oxidant injury of the podocyte combined with glomerular enlargement resulting from insulin and other growth factors.

#### High calorie intake, obesity, and glomerulosclerosis

Several experimental models demonstrate that high-calorie diet results in glomerular enlargement and glomerulosclerosis.<sup>109–111</sup> This process may be podocyte-dependent as suggested by analysis of the mechanism for the protective effect of dietary restriction in rats.<sup>27</sup> Obese humans also develop proteinuria and FSGS,<sup>26,56,112</sup> and obesity is an independent risk factor for progression to ESKD.<sup>113,114</sup> The mechanism by which obesity and high-calorie intake target the podocyte is not known, but like diabetes is likely to include glomerular enlargement under the influence of growth factors and oxidant injury to podocytes.

#### Aging-associated glomerulopathy

Glomerulosclerosis is closely associated with aging, so that by over 40 years most individuals have sclerotic glomeruli and the proportion increases with further aging.<sup>115-119</sup> ESKD is also directly related to aging with a peak incidence of treated ESKD at 64 years.<sup>1</sup> The observed reduction in treated ESKD above 65 years of age is probably the result of a selection not to treat ESKD or a failure to recognize ESKD in old age, particularly in women with small muscle mass and therefore lower serum creatinine levels. Ninety percent of ESKD is because of glomerular disease.<sup>1</sup> In older age, most of this ESKD is currently assigned to the category of hypertension and/or diabetes as a causative mechanism although the validation of this assignment is not well established, leaving open the possibility that the aging process itself may contribute significantly age-associated ESKD by as yet unrecognized and potentially treatable mechanisms.

In rats, development of glomerulosclerosis occurs in relation to aging itself.<sup>118,119</sup> This process can be accelerated by high-calorie intake and retarded by calorie restriction in the absence of either hypertension or diabetes.<sup>27</sup> This tendency to age-related glomerulosclerosis in rats is strain-dependant, thereby demonstrating that genetic background also plays a role in age-associated glomerulopathy.

Glomerulosclerosis in the aging rat is a 'podocyte disease' as was first pointed out by Floege *et al.*<sup>118</sup> and supported by recent data.<sup>27</sup> The underlying mechanisms will possibly include alterations in DNA methylation and/or histone modifications (epigenetic mechanisms) that cause miss-expression of key genes in the podocyte as well oxidant injury to mitochondrial DNA and other macromolecules.<sup>120,121</sup> This would be analogous to other age-associated diseases that impact key cells which, like podocytes, are highly differentiated post-mitotic cells with limited capacity to divide. Examples include neurons in Alzheimer's and Parkinson's disease, pancreatic  $\beta$  cells in diabetes, photoreceptors in the eye, hair cells in the ear, and conducting cells

in the heart. Age-associated glomerulopathy can therefore be seen as a senescent process, which sets up the podocyte, and thereby the whole glomerulus, for failure. This aging process can be accelerated by a combination of factors including genetic background, diet, and superimposed conditions such as hypertension and/or overt diabetes.

### PREVENTION AND TREATMENT IMPLICATIONS OF THE PODOCYTE INJURY/DEPLETION HYPOTHESIS OF GLOMERULOSCLEROSIS AND PROGRESSION

Podocyte depletion (absolute or relative to glomerular enlargement) can be considered as the mechanistic explanation underlying the hyperfiltration hypothesis of Brenner et al.<sup>2</sup> that has proved so useful in preventing progression in the clinic. According to this hypothesis, the primary clinical focus should be on minimizing factors that tend to promote injury, podocyte detachment, and depletion. This includes control of systemic and intraglomerular hypertension and particularly the use of angiotensin II blockade as a central element of the prevention strategy. In this sense, reducing podocyte injury and depletion and reducing proteinuria are facets of the same strategy. Additional clinical strategies such as reducing glomerular enlargement and oxidant injury through calorie restriction, weight control, reduction in circulating insulin, and other growth factor levels will probably be confirmed to be clinically important by future studies. The permissive role of mutations and polymorphisms of genes that play a role in the propensity to lose podocytes in individual patients will need to be defined and targeted for therapeutic intervention. Measurement of podocyte products in the urine as a potential mechanism for monitoring accelerated podocyte loss non-invasively holds good potential for clinical application. Podocyte replacement by stem cells may prove to be a useful strategy.

#### CONCLUSION

A unifying concept, which puts podocytes at the center of the glomerular disease spectrum, is conceptually simple and makes intuitive sense to the practicing clinician. It also reinforces rational clinical and scientific approaches towards preventing progression.

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#### REFERENCES

- US Renal Data System USRDS 2003 Annual Data Report: Atlas of End-Stage Renal Disease in the United States. Bethesda, MD: National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases, 2003.
- 2. Brenner BM, Lawler EV, Mackenzie HS. The hyperfiltration theory: a paradigm shift in nephrology. *Kidney Int* 1996; **49**: 1774–1777.
- Eremina V, Sood M, Haigh J et al. Glomerular-specific alterations of VEGF-A expression lead to distinct congenital and acquired renal diseases. J Clin Invest 2003; 111: 707–716.
- Eremina V, Cui S, Gerber H et al. Vascular endothelial growth factor a signaling in the podocyte-endothelial compartment is required for

mesangial cell migration and survival. *J Am Soc Nephrol* 2006; **17**: 724–735.

- Saxen L. Organogenesis of the kidney. In: Barlow PW, Green PB, White CC (eds). *Development and Cell Biology (Series 19)*. Cambridge, UK: Cambridge University Press, 1987, pp 1–171.
- 6. Dressler GR. The cellular basis of kidney development. *Annu Rev Cell Dev Biol* 2006; **22**: 509–529.
- Schnabel E, Dekan G, Miettininen A *et al.* Biogenesis of podocalyxin- the major glomerular sialoprotein – in the newborn rat kidney. *Eur J Cell Biol* 1989; 48: 313–326.
- Abrahamson DR. Structure and development of the glomerular capillary wall and basement membrane. *Am J Physiol* 1987; 253: F783-F794.
- 9. Drenckhahn D, Franke RP. Ultrastructural organization of contractile and cytoskeletal proteins in glomerular podocytes of chicken, rat, and man. *Lab Invest* 1988; **59**: 673–682.
- Mundel P, Shankland SJ. Podocyte biology and response to injury. J Am Soc Nephrol 2002; 13: 3005–3015.
- 11. Pavenstadt H, Kriz W, Kretzler M. Cell biology of the glomerular podocyte. *Physiol Rev* 2003; **83**: 253–307.
- 12. Asanuma K, Mundel P. The role of podocytes in glomerular pathobiology. *Clin Exp Nephrol* 2003; **7**: 255–259.
- 13. Ly J, Alexander M, Quaggin SE. A podocentric view of nephrology. *Curr Opin Nephrol Hypertens* 2004; **13**: 299–305.
- 14. Antignac C. Molecular basis of steroid-resistant nephrotic syndrome. *Nefrologia* 2005; **25**(Suppl 2): 25–28.
- Huber TB, Benzing T. The slit diaphragm: a signaling platform to regulate podocyte function. *Curr Opin Nephrol Hypertens* 2005; 14: 211–216.
- Hudson B, Tryggvason K, Sundamoorthy M et al. Alport's syndrome, Goodpasture's syndrome and type IV collagen. N Eng J Med 2003; 348: 2543–2556.
- 17. Yang Y, Jeanpierre C, Dressler GR *et al.* WT1 and PAX-2 podocyte expression in Denys–Drash syndrome and isolated diffuse mesangial sclerosis. *Am J Pathol* 1999; **154**: 181–192.
- Guo JK, Menke AL, Gubler MC *et al.* WT1 is a key regulator of podocyte function: reduced expression levels cause crescentic glomerulonephritis and mesangial sclerosis. *Hum Mol Genet* 2002; **11**: 651-659.
- 19. Orloff MS, Jyengar SK, Winkler CA *et al.* Variants in the Wilms' tumor gene are associated with focal segmental glomerulosclerosis in the African American population. *Physiol Genomics* 2005; **21**: 212–221.
- Kriz W. Podocyte is the major culprit accounting for the progression of chronic renal disease. *Microsc Res Tech* 2002; 15: 189–195.
- 21. Griffin SV, Petermann AT, Durvasula RV *et al.* Podocyte proliferation and differentiation in glomerular disease: role of cell-cycle regulatory proteins. *Nephrol Dial Transplant* 2003; **18**(Suppl 6): vi8-vi13.
- 22. Vigneau C, Zheng F, Polgar K *et al.* Stem cells and kidney injury. *Curr Opin Nephrol Hypertens* 2006; **15**: 238–244.
- Sugimoto H, Mundel TM, Sund M et al. Bone marrow-derived stem cells repair basement membrane collagen defects and reverse genetic kidney disease. Proc Natl Acad Sci USA 2006; 103: 7321–7326.
- 24. Prodromidi El, Poulsom R, Jeffery R *et al*. Bone marrow-derived cells contribute to podocyte regeneration and amelioration of renal disease in a mouse model of Alport's syndrome. *Stem Cells* 2006; **24**: 2448–2455.
- Bhathena DB. Glomerular basement membrane length to podocyte ratio in human nephronopenia: implications for focal segmental glomerulosclerosis. Am J Kidney Dis 2003; 41: 1179–1188.
- Chen HM, Liu ZH, Zeng CH et al. Podocyte lesions in patients with obesity-related glomerulopathy. Am J Kidney Dis 2006; 48: 772–779.
- Wiggins J, Goyal M, Sanden S *et al.* Podocyte hypertrophy, 'adaptation' and 'decompensation' associated with glomerular enlargement and glomerulosclerosis in the aging rat: prevention by calorie restriction. *J Am Soc Nephrol* 2005; 16: 2953–2966.
- Kim YH, Goyal M, Kurnit D *et al.* Podocyte depletion and glomerulosclerosis have a direct relationship in the PAN-treated rat. *Kidney Int* 2001; **60**: 957–968.
- 29. Wharram B, Goyal M, Wiggins J *et al.* Podocyte depletion causes glomerulosclerosis. Diphtheria toxin-induced podocyte depletion in rats expressing the human DTR transgene. *J Am Soc Nephrol* 2005; **16**: 2941–2952.
- 30. Fogo A, Ichikawa I. Evidence for a pathogenic linkage between glomerular hypertrophy and sclerosis. *Am J Kidney Dis* 1991; **17**: 666–669.
- 31. Fogo AB. Glomerular hypertension, abnormal glomerular growth, and progression of renal diseases. *Kidney Int Suppl* 2000; **75**: S15–S21.

- Ichikawa I, Ma J, Motojima M et al. Podocyte damage damages podocytes: autonomous vicious cycle that drives local spread of glomerular sclerosis. Curr Opin Nephrol Hypertens 2005; 14: 205–210.
- Hoffman S, Podlich D, Hahnel B et al. Angiotensin II type 1 receptor overexpression in podocytes induces glomerulosclerosis in transgenic rats. J Am Soc Nephrol 2004; 15: 1475–1487.
- Habib R, Gubler M, Antignac C *et al*. Diffuse mesangial sclerosis: a congenital glomerulopathy with nephritic syndrome. *Adv Nephrol Necker Hosp* 1993; 22: 44–57.
- Sibley R, Striegel J. Nephrotic syndrome in the fist year of life. In: Tisher CC and Brenner BM (eds). *Renal Pathol* (vol 2) Philadelphia: J.B. Lippencott Company, 1994, pp 1300–1306.
- Hinkes B, Wiggins R, Gbadegesin R et al. Positional cloning uncovers mutations in PLCE1 responsible for a nephrotic syndrome variant that may be reversible. Nat Genet 2006; 38: 1397–1405.
- Wagner N, Wagner KD, Xing Y *et al.* The major podocyte protein nephrin is transcriptionally activated by the Wilms' tumor suppressor WT1. *J Am Soc Nephrol* 2004; **15**: 3044–3051.
- Palmer RE, Kotsianti A, Cadman B et al. WT1 regulates the expression of the major glomerular podocyte membrane protein Podocalyxin. Curr Biol 2001; 11: 1805–1809.
- Mucha B, Ozaltin F, Hinkes BG, *et al.*, Members of the APN Study Group. Mutations in Wilms' tumor 1 gene cause isolated steroid resistant nephritic syndrome and occur in exons 8 and 9. *Pediatr Res* 2006; **59**: 325–331.
- Zenker M, Aigner T, Wendler O et al. Human laminin beta2 deficiency causes congenital nephrosis with mesangial sclerosis and distinct eye abnormalities. Hum Mol Genet 2004; 13: 2625–2632.
- Hasselbacher K, Wiggins R, Zapke V et al. Recessive mutations in LAMB2 expand the clinical spectrum of LAMB2-associated disorders. *Kidney Int* 2006; **70**: 1008–1012.
- Kestila M, Lenkkeri U, Mannikko M *et al.* Positionally cloned gene for a novel glomerular protein – nephrin – is mutated in congenital nephrotic syndrome. *Mol Cell* 1998; 1: 575–582.
- Boute N, Gribouval O, Roselli S *et al.* NPHS2, encoding the glomerular protein podocin, is mutated in autosomal recessive steroid-resistant nephrotic syndrome. *Nat Genet* 2000; 24: 349–354.
- Verma R, Wharram B, Kovari I *et al.* Fyn binds to and phosphorylates the kidney slit diaphragm component Nephrin. *J Biol Chem* 2003; 6: 20716–20723 Erratum in: *J Biol Chem* 2005; 280: 26640.
- Verma R, Kovari I, Soofi A *et al.* Nephrin ectodomain engagement results in Src kinase activation, nephrin phosphorylation, Nck recruitment, and actin polymerization. *J Clin Invest* 2006; **116**: 1346–1359.
- Shih NY, Li J, Cotran R *et al.* CD2AP localizes to the slit diaphragm and binds to nephrin via a novel C-terminal domain. *Am J Pathol* 2001; **159**: 2303–2308.
- 47. Savige J, Rana K, Tonna S *et al.* Thin basement membrane nephropathy. *Kidney Int* 2003; **64**: 1169–1178.
- van der Loop FT, Heidet L, Timmer ED *et al.* Autosomal dominant Alport syndrome caused by a COL4A3 splice site mutation. *Kidney Int* 2000; 58: 1870–1875.
- Jais JP, Knebelmann B, Giatras I *et al*. X-linked Alport syndrome: natural history in 195 families and genotype–phenotype correlations in males. *J Am Soc Nephrol* 2000; **11**: 649–657.
- Sharif K, Goyal M, Kershaw D *et al.* Glomerular epithelial cell (podocyte) phenotypes as defined by expression and distribution of GLEPP1 in the developing glomerulus and in minimal change nephropathy, congenital nephrotic syndrome and focal segmental glomerulosclerosis. *Exp Nephrol* 1998; 6: 234–244.
- Barisoni L, Kriz W, Mundel P *et al.* The dysregulated podocyte phenotype: a novel concept in the pathogenesis of collapsing idiopathic focal segmental glomerulosclerosis and HIV-associated nephropathy. *J Am Soc Nephrol* 1999; **10**: 51-61.
- 52. Regele HM, Fillipovic E, Langer B *et al.* Glomerular expression of dystroglycans is reduced in minimal change nephrosis but not in focal segmental glomerulosclerosis. *J Am Soc Nephrol* 2000; **11**: 403–412.
- 53. Grimbert P, Audard V, Remy P *et al.* Recent approaches to the pathogenesis of minimal-change nephrotic syndrome. *Nephrol Dial Transplant* 2003; **18**: 245–248.
- Nakamura T, Ushiyama C, Suzuki S *et al.* The urinary podocyte as a marker for the differential diagnosis of idiopathic focal glomerulosclerosis and minimal-change nephrotic syndrome. *Am J Nephrol* 2000; 20: 175–179.
- D'Agati VD, Fogo AB, Bruijn JA, Jennette JC. Pathologic classification of focal segmental glomerulosclerosis: a working proposal. *Am J Kidney Dis* 2004; 43: 368-382.

- Grisham E, Churg J. Focal glomerular sclerosis in nephrotic patients: an electron microscopic study of glomerular podocytes. *Kidney Int* 1975; 7: 111–122.
- Schmid H, Henger A, Cohen CD *et al.* Gene expression profiles of podocyte-associated molecules as diagnostic markers in acquired proteinuric diseases. *J Am Soc Nephrol* 2003; 14: 2958–2966.
- Hara M, Yanagihara T, Kihara I. Urinary podocytes in primary focal segmental glomerulosclerosis. *Nephron* 2001; 89: 342–347.
- Cusumano AM, Bodkin NL, Hansen BC et al. Glomerular hypertrophy is associated with hyperinsulinemia and precedes overt diabetes in aging rhesus monkeys. Am J Kidney Dis 2002; 40: 1075–1085.
- Luyckx VA, Brenner BM. Low birth weight, nephron number, and kidney disease. *Kidney Int Suppl* 2005; 97: S68–S77.
- Pollak MR. The genetic basis of FSGS and steroid-resistant nephrosis. Semin Nephrol 2003; 23: 141–146.
- 62. Monteiro EJ, Pereira AC, Pereira AB *et al.* NPHS2 mutations in adult patients with primary focal segmental glomerulosclerosis. *J Nephrol* 2006; **19**: 366–371.
- Kaplan JM, Kim SH, North KN *et al.* Mutations in ACTN4, encoding alpha-actinin-4, cause familial focal segmental glomerulosclerosis. *Nat Genet* 2000; 24: 251–256.
- Winn MP, Conlon PJ, Lynn KL *et al.* A Mutation in the TRPC6 cation channel causes familial focal segmental glomerulosclerosis. *Science* 2005; **308**: 1801–1804.
- 65. Reiser J, Polu KR, Moller CC *et al.* TRPC6 is a glomerular slit diaphragm-associated channel required for normal renal function. *Nat Genet* 2005; **37**: 663–664.
- 66. Wu DT, Bitzer M, Ju W, Mundel P *et al.* TGF-beta concentration specifies differential signaling profiles of growth arrest/differentiation and apoptosis in podocytes. *J Am Soc Nephrol* 2005; **16**: 3211–3221.
- Gohh RY, Yango AF, Morrissey PE et al. Preemptive plasmapheresis and recurrence of FSGS in high-risk renal transplant recipients. Am J Transplant 2005; 5: 2907–2912.
- Fischer EG, Moore MJ, Lager DJ. Fabry disease: a morphologic study of 11 cases. *Mod Pathol* 2006; **19**: 1295–1301.
- 69. Nasr SH, Preddie DC, Markowitz GS *et al.* Multiple myeloma, nephrotic syndrome and crystalloid inclusions in podocytes. *Kidney Int* 2006; **69**: 616–620.
- 70. Shah SN, He CJ, Klotman P. Update on HIV-associated nephropathy. *Curr Opin Nephrol Hypertens* 2006; **15**: 450-455.
- 71. Barisoni L, Kopp JB. Modulation of podocyte phenotype in collapsing glomerulopathies. *Microsc Res Tech* 2002; **57**: 254–262.
- Schwimmer JA, Markowitz GS, Valeri A et al. Collapsing glomerulopathy. Semin Nephrol 2003; 23: 209–218.
- Yang Y, Gubler MC, Beaufils H. Dysregulation of podocyte phenotype in idiopathic collapsing glomerulopathy and HIV-associated nephropathy. *Nephron* 2002; 91: 416–423.
- Atta MG, Gallant JE, Hafizur-Rahman M *et al.* Antiretroviral therapy in the treatment of HIV-associated nephropathy. *Nephrol Dial Transplant* 2006; 21: 2809–2813.
- Lemley KV, Lafayette RA, Safai M et al. Podocytopenia and disease severity in IgA nephropathy. Kidney Int 2002; 61: 1475–1485.
- Hara M, Yanagihara T, Takada T *et al*. Urinary excretion of podocytes reflects disease activity in children with glomerulonephritis. *Am J Nephrol* 1998; **18**: 35-41.
- Nakamura T, Ushiyama C, Suzuki S *et al.* Urinary podocytes for the assessment of disease activity in lupus nephritis. *Am J Med Sci* 2000; **320**: 112–116.
- Vogelmann SU, Nelson WJ, Myers BD *et al*. Urinary excretion of viable podocytes in health and renal disease. *Am J Physiol Renal Physiol* 2003; 285: F40–F48.
- Kanno K, Kawachi H, Uchida Y et al. Urinary sediment podocalyxin in children with glomerular diseases. Nephron Clin Pract 2003; 95: c91–c99.
- Nakamura T, Ushiyama C, Suzuki S *et al.* Effects of angiotensinconverting enzyme inhibitor, angiotensin II receptor antagonist and calcium antagonist on urinary podocytes in patients with IgA nephropathy. *Am J Nephrol* 2000; **20**: 373–379.
- Cybulsky AV, Rennke HG, Feintzeig ID *et al.* Complement-induced glomerular epithelial cell injury. Role of the membrane attack complex in rat membranous nephropathy. *Clin Invest* 1986; **77**: 1096–1107.
- Akano N, Yoshioka K, Aya N *et al.* Immunoelectron microscopic localization of membrane attack complex and hepatitis B e antigen in membranous nephropathy. *Virchows Arch A Pathol Anat Histopathol* 1989; **414**: 325–330.
- Couser WG, Nangaku M. Cellular and molecular biology of membranous nephropathy. J Nephrol 2006; 19: 699–705.

- Yang D, Goyal M, Sharif K *et al.* Glomerular epithelial protein 1 and podocalyxin-like protein 1 in inflammatory glomerular disease (crescentic nephritis) in rabbit and man. *Lab Invest* 1996; **74**: 571–584.
- Moeller MJ, Soofi A, Hartmann I *et al.* Podocytes populate cellular crescents in a murine model of inflammatory glomerulonephritis. *J Am Soc Nephrol* 2004; 15: 61–67.
- Ding M, Cui S, Li C *et al.* Loss of tumor supressor Vhlh leads to upregulation of Cxcr4 and rapidly progressive glomerulonephritis in mice. *Nature Medicine* 2006; **12**: 1081–1087.
- 87. Marin R, Gorostidi M, Fernandez-Vega F *et al.* Systemic and glomerular hypertension and progression of chronic renal disease: the dilemma of nephrosclerosis. *Kidney Int Suppl* 2005; **99**: S52–S56.
- Toto RD. Proteinuria and hypertensive nephrosclerosis in African Americans. *Kidney Int Suppl* 2004; 92: S102–S104.
- 89. Hughson MD, Bertram JF, Hoy WE *et al.* Hypertension, glomerular number, and birth weight in African Americans and white subjects in the southeastern United States. *Kidney Int* 2006; **69**: 640–642.
- Fogo A, Breyer JA, Smith MC *et al.* Accuracy of the diagnosis of hypertensive nephrosclerosis in African Americans: a report from the African American Study of Kidney Disease (AASK) trial. AASK pilot study investigators. *Kidney Int* 1997; **51**: 244–252.
- Fogo AB. Hypertensive risk factors in kidney disease in African Americans. *Kidney Int Suppl* 2003; 83: S17–S21.
- Kimmelstiel P, Wilson C. Benign and malignant hypertension and nephrosclerosis. A clinical and pathologic study. *Am J Pathol* 1936; 12: 45–81.
- Helmchen U, Wenzel U. Benign and malignant nephrosclerosis and renovascular disease. In: Tisher CC and Brenner BM (eds). *Renal Pathol* (vol 2) Philadelphia: J.B. Lippencott Company, 1994, pp 1201–1236.
- 94. Orloff MS, Iyengar SK, Winkler CA *et al.* Variants in the Wilms' tumor gene are associated with focal segmental glomerulosclerosis in the African American population. *Physiol Genomics* 2005; **21**: 212–221.
- Douglas JG, Agodoa L. ACE inhibition is effective and renoprotective in hypertensive nephrosclerosis: the African American Study of Kidney Disease and Hypertension (AASK) trial. *Kidney Int Suppl* 2003; 83: S74–S76.
- 96. Lemley KV. A basis for accelerated progression of diabetic nephropathy in Pima Indians. *Kidney Int Suppl* 2003; **83**: S38–S42.
- 97. Hayden MR, Whaley-Connell A, Sowers JR. Renal redox stress and remodeling in metabolic syndrome, type 2 diabetes mellitus, and diabetic nephropathy: paying homage to the podocyte. *Am J Nephrol* 2005; **25**: 553–569.
- Wolf G, Chen S, Ziyadeh FN. From the periphery of the glomerular capillary wall toward the center of disease: podocyte injury comes of age in diabetic nephropathy. *Diabetes* 2005; 54: 1626–1634.
- Pagtalunan ME, Miller PL, Jumping-Eagle S et al. Podocyte loss and progressive glomerular injury in type II diabetes. J Clin Invest 1997; 99: 342–348.
- Steffes MW, Schmidt D, McCrery R et al. International Diabetic Nephropathy Study Group: glomerular cell number in normal subjects and in type I diabetic patients. *Kidney Int* 2001; 59: 2104–2113.
- 101. White KE, Bilous RW, Marshall SM *et al.* The European Study for the Prevention of Renal Disease in Type I diabetes (ESPRIT): podocyte number in normotensive type I diabetic patients with albuminuria. *Diabetes* 2002; **51**: 3083–3089.
- Dalla Vestra M, Masiero A, Roiter AM *et al.* Is podocyte injury relevant in diabetic nephropathy? Studies in patients with Type 2 diabetes. *Diabetes* 2003; **52**: 1031–1035.
- Meyer TW, Bennett PH, Nelson RG. Podocyte number predicts long-term urinary albumin excretion in Pima Indians with type II diabetes and microalbuminuria. *Diabetologia* 1999; 42: 1341–1344.
- Nakamura T, Ushiyama C, Suzuki S *et al*. Urinary excretion of podocytes in patients with diabetic nephropathy. *Nephrol Dial Transplant* 2000; 15: 1379–1383.
- Nakamura T, Ushiyama C, Osada S et al. Pioglitazone reduces urinary podocyte excretion in type 2 diabetes patients with microalbuminuria. *Metabolism* 2001; 50: 1193–1196.
- Gross ML, El-Shakmak A, Szabo A *et al.* ACE-inhibitors but not endothelin receptor blockers prevent podocyte loss in early diabetic nephropathy. *Diabetologia* 2003; 46: 856–868.
- Petermann AT, Pippin J, Krofft R *et al.* Viable podocytes detach in experimental diabetic nephropathy: potential mechanism underlying glomerulosclerosis. *Nephron Exp Nephrol* 2004; **98**: e114-e123.
- Gassler N, Elger M, Kranzlin B *et al.* Podocyte injury underlies the progression of focal segmental glomerulosclerosis in the fa/fa Zucker rat. *Kidney Int* 2001; **60**: 106–116.

- Kleinknecht C, Laouari D, Hinglais N *et al.* Role of amount and nature of carbohydrates in the course of experimental renal failure. *Kidney Int* 1986; **30**: 687-693.
- 110. Tapp DC, Wortham WG, Addison JF *et al.* Food restriction retards body growth and prevents end-stage renal pathology in remnant kidneys of rats regardless of protein intake. *Lab Invest* 1989; **60**: 184–195.
- 111. Keenan KP, Coleman JB, McCoy CL *et al.* Chronic nephropathy in *ad libitum* overfed Sprague–Dawley rats and its early attenuation by increasing degrees of dietary (caloric) restriction to control growth. *Toxicol Pathol* 2000; **28**: 788–798.
- 112. Kambham N, Markowitz GS, Valeri A *et al*. Obesity-related glomerulopathy: an emerging epidemic. *Kidney Int* 2001; **59**: 1498–1509.
- 113. Rutkowski P, Klassen A, Sebekova K *et al.* Renal disease in obesity: the need for greater attention. *J Ren Nutr* 2006; **16**: 216–223.
- Iseki K, Ikemiya Y, Kinjo K *et al.* Body mass index and the risk of development of end stage renal disease in a screened cohort. *Kidney Int* 2004; 65: 1870–1876.

- 115. Kaplan C, Pasternack B, Shah H *et al.* Age-related incidence of sclerotic glomeruli in human kidneys. *Am J Pathol* 1975; **80**: 227–234.
- 116. Lindeman RD, Tobin J, Shock NW. Longitudinal studies on the rate of decline in renal function with age. J Am Geriatric Soc 1985; **33**: 278–285.
- 117. Anderson S, Brenner BM. The aging kidney: structure, function, mechanisms and therapeutic implications. *J Am Ger Soc* 1987; **35**: 590–593.
- 118. Floege J, Hackman B, Kliem V *et al.* Age-related glomerulosclerosis and interstitial fibrosis in Milan normotensive rats: a podocyte disease. *Kidney Int* 1997; **51**: 230–243.
- 119. Brandis A, Bianchi G, Reale E *et al.* Age-dependant glomerulosclerosis and proteinuria occurring in rats of the Milan normotensive strain and not in rats of the Milan hypertensive strain. *Lab Invest* 1986; **55**: 234-243.
- Mathers JC. Nutritional modulation of ageing: genomic and epigenetic approaches. *Mech Ageing Dev* 2006; **127**: 584–589.
- 121. Balaban RS, Nemoto S, Finkel T. Mitochondria, oxidants, and aging. *Cell* 2005; **120**: 483–495.