

## STATE-OF-THE-ART PAPER

# Hemoxygenase-1 in Cardiovascular Disease

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Hemoxygenase (HO)-1 is an inducible isoform of the first and rate-controlling enzyme of the degradation of heme into iron, carbon monoxide, and biliverdin, the latter being subsequently converted into bilirubin. Several positive biological effects exerted by this enzyme have gained attention, as anti-inflammatory, anti-apoptotic, angiogenic, and cytoprotective functions are attributable to carbon monoxide and/or bilirubin. Thus, the physiological induction of HO-1 may be an adaptive and beneficial response to several possibly noxious stimuli, including heme itself, suggesting a potentially autoprotective and autodefensive role in several pathophysiological states including acute coronary syndromes and stroke. This review article provides a comprehensive overview of the biochemistry, physiology, and pathophysiology of HO-1 in relation to cardiovascular disease (CVD). Furthermore, we present some of the emerging evidence in support of the view that the induction of the HO-1 gene may be a new opportunity to target the pathophysiology of CVD, with therapeutic implications for management. (J Am Coll Cardiol 2008;52:971-8) © 2008 by the American College of Cardiology Foundation

Hemoxygenase (HO)-1, which is inducible by factors that include heavy metals and reactive oxygen species, catalyzes the first and rate-controlling step of the degradation of heme into ferrous ( $\text{Fe}^{2+}$ ) iron, carbon monoxide (CO), and biliverdin, the latter being subsequently converted into bilirubin (1). These products may have physiological and pathological functions, such as in protection from oxidative stress, a process that may be important in the pathophysiology of several cardiovascular diseases (2,3). The growing interest in HO-1 is now such that the induction of the HO-1 gene has been proposed as a new therapy (4). In this review article, we provide a comprehensive overview of enzymatic and biological aspects of HO-1, as well as its reaction products (chiefly, CO and bilirubin) that illustrate the significance of this molecule in the pathophysiology of cardiovascular disease (CVD). In addition, we present some of the salient lines of evidence in support of the therapeutic and clinical implications of the modification of the activity of this enzyme.

## Search Strategy

We performed a literature search (1966 to May 2008) on “hemoxygenase-1” and “heme-oxygenase-1” using Medline, EMBASE, PubMed, and Cochrane electronic biblio-

graphic databases, as well as scanned relevant reference lists from included articles.

## Biochemistry and Genetics of HO-1

Hemoxygenase, originally identified by Tenhunen et al. (5), has 3 isoforms. The first, HO-1, is a 32-kDa protein, inducible by numerous stimuli, that catalyzes the first and rate-limiting step in the degradation of the protoporphyrin ring of tetrapyrrole heme from effete red blood cells, yielding equimolar quantities of biliverdin IXa, CO, and iron (5,6). Biliverdin (through the action of biliverdin reductase) is converted to bilirubin, and iron is sequestered into ferritin. Interestingly, HO-1 utilizes heme as both a prosthetic group and a substrate (1). The second isoform of hemoxygenase, HO-2, a constitutively synthesized 36-kDa protein, is generally unresponsive to any of the inducers of HO-1. The third isoform, HO-3, also catalyzes heme degradation, but much less so when compared with HO-2 (7,8). Although heme is the typical HO-1 inducer, others include endotoxin, heavy metals, oxidants, and hypoxia (Table 1). A common feature of several of these inducers is their ability to generate reactive oxygen species, suggesting that HO-1 provides potent cytoprotective effects (9-16) (Table 1).

**Bilirubin.** Several lines of evidence suggest that biliverdin and bilirubin may be part of a cell defense strategy in response to oxidative stress. Both molecules are natural antioxidants, and high-normal serum levels of bilirubin are inversely related to the atherogenic risk, possibly by inhibitory effects against low-density lipoprotein oxidation and the scavenging of oxygen radicals (17,18). Additionally, an

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**Abbreviations and Acronyms**

- CAD** = coronary artery disease
- cGMP** = cyclic guanine monophosphate
- CO** = carbon monoxide
- CVD** = cardiovascular disease
- GT** = glutathione thymidine dinucleotide
- HO** = hemoxygenase
- NO** = nitric oxide
- VEGF** = vascular endothelial growth factor

inhibitory effect on protein kinase C and protein phosphorylation activity has been shown, both of which lead to inhibition of proatherogenic factors (19, 20). Bilirubin also provides cardioprotection against reperfusion injury, such as by the suppression of the oxidation of lipid membranes (21,22). The hypothesis that the protection provided by bilirubin in the ischemic myocardium could be clinically significant is supported by an inverse correlation between plasma bilirubin and the risk of coronary artery disease (CAD) and de-

creased antioxidant activity of bilirubin in atherosclerotic lesions (23-25). The increased intracellular bilirubin as a consequence of HO-1 induction also implies that CO production may be enhanced. Indeed, Morita et al. (17) have suggested that stimuli that amplify the generation of bilirubin also act in an adaptive reaction against oxidative insults, so that the action of the HO-1 pathway in raising endogenous bilirubin levels may represent an additional option in frustrating oxidative stress that may, eventually, have relevant clinical impact.

**Carbon monoxide.** Morita et al. (17) also summarized the evidence that CO has a physiological role in the regulation of vascular tone similar to that of nitric oxide (NO), one mechanism for which may be increased intracellular cyclic guanine monophosphate (cGMP) (26). However, the precise physiological significance of CO in relation to NO as a vasodilator is contentious. For example, the increase in cGMP is induced in vitro by perhaps 130-fold by NO, whereas it is only induced 4-fold by CO. Rodent models suggest that overproduction of CO might impair NO-elicited generation of soluble guanylate cyclase, resulting in inhibition of the cGMP increase in the aortas of transgenic mice that over-expressed HO-1, suggesting that CO may

result in protection from acute hypertension (27,28). Carbon monoxide may also limit the development of vascular diseases because of an effect on smooth muscle cell proliferation and death, whereas lack of HO-1 in (-/-) KO mice leads to pulmonary hypertension (29-31).

**Iron.** Ferrous iron, a possible electron donor with cytotoxic potential in generating reactive oxygen species is an additional regulator of HO-1 (32). Thus ferritin, primarily a liver-derived, iron-binding protein, may act as an indirect antioxidant by sequestering iron. The augmentation of intracellular ferritin through HO-1 reduces the cytotoxic effects of heme and hydrogen peroxide in vascular endothelial cells (33), and may protect against ischemia-reperfusion injury through its cytoprotective effects on the endothelium and its ability to keep labile iron pools low and thereby reduce oxidant-induced lipid peroxidation (34). However, one hesitates to leap to the conclusion that these relationships are causal. Nevertheless, the weight of published data suggests that, of the 3 products of HO-1, the potentially protective effects of bilirubin and/or CO seem likely to exceed the potentially harmful effects of iron.

**Pathophysiological Processes of HO-1**

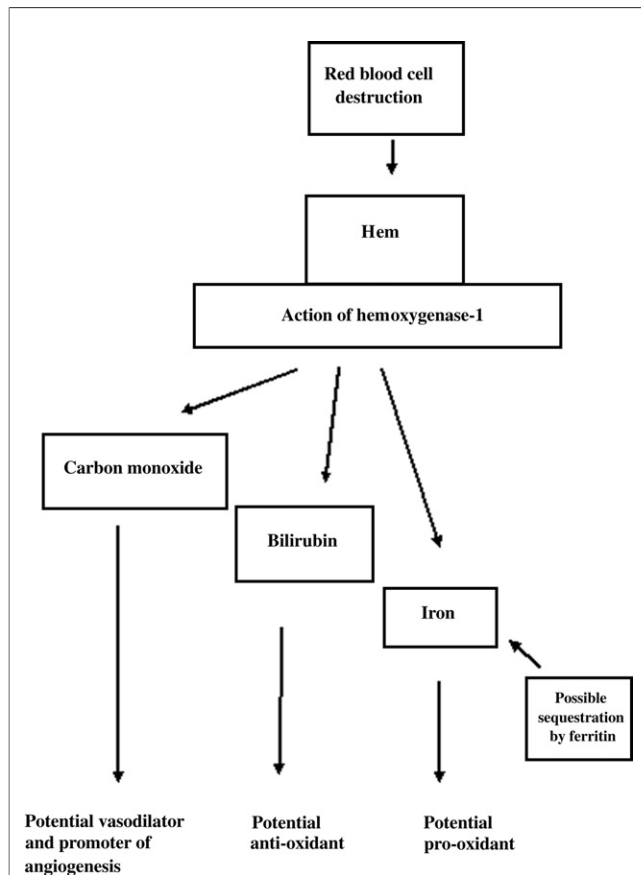
The previous section provided evidence to suggest that HO-1 can (through its products CO and bilirubin) function as a potentially important cytoprotective molecule. Many workers assume that the up-regulation of HO-1 by stress-causing agents could mediate cytoprotection against subsequent noxious stimuli and that this can be an important physiological process. However, although physiologically low concentrations of heme are cytoprotective as they induce the rapid up-regulation of HO-1, excess pathological amounts of heme out-strip the ability of HO-1 to metabolize it so that residual heme (liberating free iron) may act deleteriously on tissue by pro-oxidative and proinflammatory effects (35-37). Mechanisms by which HO-1 provides protection against cardiopathology include antioxidant activity of bilirubin (22-25), sequestration of iron by ferritin (33), and an antifibrinolytic and vasodilative effect of CO (26-31,38) (Fig. 1).

**Inflammation and antioxidant function.** The mechanisms by which HO-1 is anti-inflammatory generate considerable research activity but remain unclear, although animal model clues exist, such as a relationship between HO-1 and cytokines (39-41). A rat model of hepatic ischemic and reperfusion injury (which activates toll-like receptor-4 signaling) has been used as evidence of a novel mechanism by which HO-1 exerts adaptive cytoprotective and anti-inflammatory functions (41,42). In the latter, cobalt-protoporphyrin-induced HO-1 over-expression reduced liver damage and down-regulated activation of signal transducers and activator of transcription 1 by the type-1

**Table 1 Inducers/Stimulators of HO-1 Activity**

Cytokines (IL-10, IL-13, IL-18)
Endotoxin*
Growth factors (TGF-beta, PDGF, VEGF)
Heme
Heavy metals (e.g., sodium arsenite, cadmium, tin, lead)
Hypoxia* and hyperoxia*
Nitric oxide
Oxidants (e.g., hydrogen peroxide,* peroxytrile*)
Oxidized LDL
Thiol scavengers
Ultraviolet radiation*

Pooled from references 9-17, 70, 71, and elsewhere. \*Also promoters of oxidative stress. HO = hemoxygenase; IL = interleukin; LDL = low-density lipoprotein; PDGF = platelet-derived growth factor; TGF = transforming growth factor; VEGF = vascular endothelial growth factor.



**Figure 1** Schematic of Hemoxygenase-1-Catalyzed Biochemical Reactions

Hem released from red blood cells is enzymatically converted into carbon monoxide, bilirubin, and iron. The former is a vasodilator and promoter of angiogenesis. Bilirubin has potential antioxidant activity. Free ferrous iron has potential pro-oxidant activity, although this may be limited by its sequestration by ferritin.

interferon pathway downstream of toll-like receptor-4, which in turn decreased CXCL-10 production. The anti-oxidant activity of bilirubin may feed through to antiatherogenic properties—possibly by protecting low density lipoprotein cholesterol from oxidation (22,43,44).

**Apoptosis.** Hemoxygenase-1 appears to have a role in reducing the proapoptotic effects of tumor necrosis factor (TNF), hyperglycemia, and iron, some of which could involve CO (45-48). At the intracellular level, this may

involve expression of p38 mitogen-activated protein kinase enzymes and possibly the activation of nuclear factor kappa B (49,50). Adenovirus-mediated transfection of the HO-1 gene into rat hearts resulted in a reduction in infarct size that was accompanied by decreases in lipid peroxidation and in proapoptotic Bax and proinflammatory interleukin-1-beta protein abundance, with a parallel increase in antiapoptotic Bcl-2 protein level (51). Various models of transplantation, hyperglycemia, and tissue culture suggest HO-1 inhibits apoptosis by suppressing cytotoxic, inflammatory, and signaling cytokines (52-55).

**Hypoxia and ischemia/reperfusion injury.** Hypoxia is thought to be a key determinant in clinical pathology, and thus several lines of research have linked it (albeit indirectly) with HO-1. For example, the HO-1-bilirubin pathway can defend cells from reoxygenation injury, and restricting vascular smooth muscle cell growth by increasing the release of CO may represent a route to limiting pulmonary hypertension (56,57). Interestingly, in a rat model, monotherapy with either CO or biliverdin did not alter the survival of heart grafts, and dual treatment increased survival from 0% to 80%, with a significant decrease of myocardial injury and improved cardiac function (58). This provides tantalizing data that may conceivably translate into a human therapy, as is implied by various reviewers, for example, Chen et al. (59) and Immenschuh and Schroder (60).

**Angiogenesis.** A link between HO-1 and angiogenesis is relatively recent. Transfection of rabbit cells with the human HO-1 gene resulted in a 2-fold increase in blood vessel formation (61); other investigators used a transfection model to show increased blood flow and, crucially, a relationship with the angiogenic stimulant vascular endothelial growth factor (VEGF) (62,63). Jozkowicz et al. (64) showed that CO could drive VEGF expression, and Busolati et al. (65) linked inflammation with angiogenesis by proposing a dual action of HO-1 in an anti-inflammatory action and in the promotion of VEGF-driven angiogenesis. The role of HO-1 in angiogenesis has been recently reviewed (66,67).

Thus, interest in HO-1 in cardiology may be justified by aspects such as inflammation, antioxidant functions, apoptosis, hypoxia, and ischemia/reperfusion injury, and angiogenesis (Fig. 1).

**Table 2** Studies Reporting the Association Between HO-1 and CAD in Humans

Author (Ref. #)	Year	n	Disease	Key Findings
Chen et al. (80)	2005	135	AMI, UAP, SAP	HO-1 expression in patients with CAD significantly higher than in patients without CAD.
Gulesserian et al. (76)	2005	199	CAD	Low HO-1 level inducibility, may represent an independent prognostic marker for restenosis after angioplasty.
Kaneda et al. (75)	2002	577	CAD	Patients with shorter GT (<25 repeats) less likely to have CAD than patients with long GT (>29 repeats).
Li et al. (79)	2006	110	CAD	Significantly higher HO-1 protein leukocyte expression in patients with CAD than in patients without CAD.

AMI = acute myocardial infarction; CAD = coronary artery disease; GT = glutathione thymidine dinucleotide; HO = hemoxygenase; SAP = stable angina pectoris; UAP = unstable angina pectoris.

**Table 3 Studies Reporting the Association of HO-1 With PVDs and DM**

Author (Ref. #)	Year	n	Disease	Key Findings
Chen et al. (74)	2002	474	DM	Greater expression of long GT repeats ( $\geq 32$ ), thus might have higher oxidative stress and increased risk for CAD.
Schillinger et al. (70)	2002	271	CAD, AAA, PAD	Significant differences of HO-1 expression among 3 groups of patients: group with AAA had lower risk than other groups; thus up-regulation of HO-1 may be a protective anti-inflammatory factor against development of AAA.
Schillinger et al. (69)	2004	381	PVD	Significant short (<25 GT) repeats in HO-1 gene expression confer a reduced risk for restenosis after balloon angioplasty.
Dick et al. (73)	2005	472	PVD	Significant short (<25 GT) repeats in HO-1 gene expression confer a reduced risk for MI.

AAA = abdominal aortic aneurysms; DM = diabetes mellitus; GT = glutathione thymidine dinucleotide; MI = myocardial infarction; PAD = peripheral artery disease; PVD = peripheral vascular disease; other abbreviations as in Table 2.

### Clinical Aspects of HO-1 in CVDs

The evidence for the protective role of HO-1 in clinical CVD is not only supported by experimental findings in cell culture and animal models (as discussed) but also by clinical studies in humans (Tables 2 and 3).

**Genetics of HO-1.** Exner et al. (68) reported that the number of glutathione thymidine dinucleotide (GT) repeats in the promoter region of the HO-1 gene modulates the level of gene transcription. The presence/absence of short/long GT repeats had a bearing on 6-month restenosis after femoropopliteal balloon dilatation, possibly associated with differences in levels of inflammatory marker C-reactive protein (69). They also reported that this polymorphism may be important in abdominal aortic aneurysm and renal (but not cardiac) allografting (70-72). A 21-month follow-up of 472 patients with peripheral artery disease indicated that the HO-1 genotype is potentially protective against adverse coronary events (73). Others (74,75) have also looked at this polymorphism in CAD and/or diabetes mellitus, speculating that diabetic persons carrying longer (GT) repeats might have higher oxidative stress and increased susceptibility to the development of CAD (i.e., the patients with fewer GT repeats were less likely to have CAD). The long (>29 repeats) polymorphic allele of the HO-1 gene promoter, which leads to low HO-1 inducibility, may be an independent prognostic marker for restenosis after percutaneous coronary intervention and stent implantation (76). Another polymorphism [the T(-413)A (AA/TA+TT) variant] of the HO-1 gene is associated with an increased incidence of hypertension in women (77). Other polymorphisms and a microsatellite marker seem to have no significant role on outcome of kidney transplantation (78).

**CAD.** Considerable animal data justify interest in HO-1 in human CAD, where the clear implication is that increased activity of the HO-1 gene (and therefore its products) is beneficial. Expression of the HO-1 protein was assessed in monocytes and lymphocytes from patients with acute myocardial infarction, patients with unstable angina pectoris, and patients with stable angina pectoris (79,80). There were significant differences of HO-1 expression—highest for the group with acute myocardial infarction, followed by the group with unstable angina pectoris, and finally by the group with stable angina pectoris. Within the patients with

angiographically-defined CAD, HO-1 was highest in those with a greater disease burden. One interpretation of these data is that higher HO-1 expression is a consequence of the disease process and so may be a defense (self-limiting) mechanism. Morsi et al. (81) provided insightful data by obtaining endothelial cells from patients with advanced or early lesions and from coronary arteries free of disease. The HO-1 expression and (crucially) its biological activity (in terms of bilirubin release per mg of protein) were only present in cells from advanced atherosclerotic lesions. Interestingly, there also is a strong correlation between HO-1 and VEGF, although one hesitates before leaping to the conclusion that this raised VEGF may have been driven by HO-1 and/or its products.

**Diabetes mellitus.** In humans, the (GT)<sub>n</sub> HO-1 gene promoter polymorphism may influence clinical outcome, a putative mechanism being resistance/susceptibility to oxidative stress (74-76). De Silva et al. (82), examining retinal pigment epithelium, found significantly decreased levels of HO-1 messenger ribonucleic acid (mRNA), namely, 340 to 450 HO-1 mRNA copies/ng of total ribonucleic acid, in tissue from diabetic patients as compared with 425 to 8,000 HO-1 mRNA copies/ng of total ribonucleic acid in retinal pigment epithelium from normal donors and 460 to 7,605 copies/ng in hypertensive donor eyes. Increased monocyte HO-1 gene expression in diabetic patients falls upon metabolic improvement, possibly related to oxidative stress, although others found lower HO-1 skeletal muscle cell expression (83,84). Leukocyte HO-1 gene expression is significantly lower in patients with and without diabetic microangiopathy compared with control subjects, correlates negatively with a marker of oxidative stress, glycosylated hemoglobin, and diabetes duration, and normalization of blood glucose results in a reduction in HO-1 antigen in the cytoplasm of mononuclear leukocytes (85,86). Arredondo et al. (87) assessed the length of (GT)<sub>n</sub> repeats in the HO-1 gene promoter and also HO-1 enzymatic activity in mononuclear cells from diabetic patients. Although patients had significantly greater iron stores and HO activity than did control subjects, with a positive association between serum iron and HO activity in the diabetic patients, allelic frequency did not differ significantly between diabetic patients and control subjects.

**Cerebrovascular disease.** Beschoner *et al.* (88) demonstrated increased accumulation of HO-1+ microglia/macrophages at hemorrhagic lesions as early as 6 h after traumatic brain injury trauma that was still pronounced after 6 months. In contrast, after focal cerebral infarctions, HO-1+ microglia/macrophages accumulated within focal hemorrhages only and were absent in nonhemorrhagic regions. They speculated that prolonged expression of HO-1 in glial cells in human brains after traumatic brain injury and cerebral infarction helps in the recovery of neuronal tissue after these insults. Morgan *et al.* (89) assessed the role of GT repeats in the HO-1 promoter in 69 patients with cerebral aneurysms and 230 age-matched control subjects, and found that patients were more likely to have more than 36 repeats than were control subjects. The authors speculate that facilitated up-regulation of HO-1 may be protective against the development of intracranial aneurysms. They wisely point out, however, that because of the relatively small sample size and modest statistical significance, the data must be interpreted with caution and the association needs to be confirmed in large studies. For example, follow-up of 472 patients with advanced peripheral artery disease found that the status of the short/long GT genotype failed to identify 40 patients who had a cerebrovascular event but did instead associate with 48 patients who had a coronary event (73).

### A Role for HO-1 in CVD?

Naturally, a caveat for the association(s) between greater activity of the HO-1 gene, its enzyme product, and the products of the enzyme relate to “cause or effect” phenomenon. It could be argued that raised levels and activity simply reflect more serious disease and an attempt by the body to limit the disease. Although data point to the likelihood that the activity of HO-1 leads to an active protection against the disease process, that may be one speculation too far. Perhaps the disease process effectively swamps the ability of HO-1 to limit cell damage that could lead to clinical disease (Tables 2 and 3). Possible mechanisms by which HO-1 contributes to pathogenesis are summarized in Figure 1.

Unsurprisingly, the combination of tissue protective and smooth muscle relaxing properties makes HO-1 an interesting objective for the drug treatment of CVD (4). Immenschuh and Ramadori (90) speculated that the therapeutic approaches intended at moderately increasing HO-1 expression in tissue might be beneficial in a number of disease states that probably relate to vascular disorders. Leaving aside the potential of gene therapy as being distant from the clinic (91), some current pharmacological agents act to induce HO-1. Some statins seem able to increase endothelial HO-1 mRNA levels in a concentration- and time-dependent fashion, although, whereas atorvastatin enhances the expression of endothelial nitric oxide synthase, HO-1 is not significantly affected (92-94). However, other

studies show that simvastatin activates HO-1 in vascular smooth muscle cells *in vitro* and *in vivo* (95). Aspirin (30 to 300  $\mu$ M) increased human umbilical vein endothelial cell HO-1 protein levels in a concentration-dependent fashion up to 5-fold over basal levels (96), and more recent evidence points to a possible role for a peroxisome proliferators-activated receptors system (97).

### Conclusions

Several of the numerous pathophysiological processes in atherosclerosis are, in theory, amenable to the action of 2 of the products of HO-1 (i.e., CO and bilirubin), whereas the third (i.e., iron) may be toxic. However, iron may be sequestered by ferritin, and the vast weight of published reports focuses on possible benefits of CO and bilirubin. For example, oxidative injury (such as to low-density lipoprotein cholesterol), which is thought to be a common feature of many pathophysiological processes, may be attenuated by HO-1. Thus, CO and bilirubin may play an important beneficial role in conditions such as hypertension, acute renal injury, and lung injury (17,37), and may well operate through the up-regulation of HO-1 in endothelial cells by various stimuli (such as hypoxia).

In addition, HO-1 is induced by some of the well-established cardiovascular risk factors, and appears to have a protective role in the vascular wall against atherogenesis through several pathways. However, in contrast to the implication of intracellular and pericellular activity of HO-1, little is known about plasma levels of this enzyme and, in particular, whether raised or lowered levels are present in CVD, and if such levels correlate with bilirubin and other plasma biomarkers. For example, if there is raised plasma HO-1 in CVD, is this indicative of potential or present protection (possibly driven by the pathology) or would it simply reflect leakage from damaged cells?

A number of therapeutic agents that are able to manipulate HO-1 gene expression have been recognized, suggesting that manipulation of the HO-1 gene might be a new avenue in the prevention and/or treatment of CVD (5,90). However, the more direct gene therapy approach, proven in animals, remains an intriguing opportunity to treat cardiovascular (and other) diseases. Nevertheless, whether these preliminary but promising reports come to fruition in the clinical setting is unknown, and a continuing weakness in the study of HO-1 is the lack of good clinical data. One example is a fascinating case report of a 6-year-old boy with severe HO-1 deficiency (98). Consistent with the cell biology and animal models described above (Fig. 1), he exhibited hematuria, proteinuria, a microcytic hemolytic anemia, increased iron-binding capacity, ferritin, and iron deposition alongside raised von Willebrand factor (marking endothelial cell damage). Crucially, serum bilirubin was constantly low whereas serum heme was extremely high. Undoubtedly,

more attention to HO-1 biology may provide novel insights into the pathophysiology of CVD.

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## REFERENCES

1. Ponka P. Cell biology of heme. *Am J Med Sci* 1999;318:241-56.
2. Maxwell SR, Lip GYH. Free radicals and antioxidants in cardiovascular disease. *Br J Clin Pharmacol* 1997;44:307-17.
3. Victor VM, Rocha M. Targeting antioxidants to mitochondria: a potential new therapeutic strategy for cardiovascular diseases. *Curr Pharm* 2007;13:845-63.
4. Stocker R, Perrella MA. Heme oxygenase-1: a novel drug target for atherosclerotic diseases? *Circulation* 2006;114:2178-89.
5. Tenhunen R, Marver HS, Schmid R. The enzymatic conversion of heme to bilirubin by microsomal hemoxygenase. *Proc Natl Acad Sci USA* 1968;61:748-55.
6. Ortis de Montellano PR. The mechanism of hemoxygenase. *Curr Opin Chem Biol* 2000;4:221-6.
7. Hayashi S, Omata Y, Sakamoto H, et al. Characterization of rat heme oxygenase-3 gene, implication of processed pseudogenes derived from heme oxygenase-2 gene. *Gene* 2004;336:241-50.
8. McCoubrey WK, Huang TJ, Maines MD. Isolation and characterization of a cDNA from the rat brain that encodes hemoprotein heme oxygenase-3. *Eur J Biochem* 1997;247:725-32.
9. Abraham NG, Kappa A. Heme oxygenase and cardiovascular system. *Free Radic Biol Med* 2005;39:1-25.
10. Otterbein LE, Choi AMK. Heme oxygenase: colors of defense against cellular stress. *Am J Physiol* 2000;279:1029-103.
11. Keyse SM, Tyrrell RM. Heme oxygenase is the major 32-kDa stress protein induced in human skin fibroblasts by UVA radiation, hydrogen peroxide, and sodium arsenite. *Proc Natl Acad Sci USA* 1989;86:99-103.
12. Maeshima H, Sato M, Ishikawa K, Katagata Y, Yoshida T. Participation of altered upstream stimulatory factor in the induction of rat heme oxygenase-1 by cadmium. *Nucleic Acids Res* 1996;24:2959-65.
13. Tomaro ML, Batle AM. Bilirubin: its role in cytoprotection against oxidative stress. *Int J Biochem Cell Biol* 2002;34:216-20.
14. Rizzardini M, Terao M, Falciani F, Cantoni L. Cytokine induction of haem oxygenase mRNA in mouse liver: interleukin 1 transcriptionally activates the haem oxygenase gene. *Biochem J* 1993;290:343-7.
15. Tyrell RM, Applegate LA, Tromvoukis Y. The proximal promoter region of the human hemoxygenase contains elements involved in stimulation of transcriptional activity by a variety of agents including oxidants. *Carcinogen* 1993;14:761-75.
16. Doi K, Akaike T, Fujii S, et al. Induction of haemoxygenase-1 nitric oxide and ischaemia in experimental solid tumors and implications for tumor growth. *Br J Cancer* 1999;80:1945-54.
17. Morita T. Hemoxygenase and atherosclerosis. *Arterioscler Thromb Vasc Biol* 2005;25:17861-81.
18. Neuzil J, Stocker R. Free and albumin-bound bilirubin are efficient co-antioxidants for alpha-tocopherol, inhibiting plasma and low density lipoprotein lipid peroxidation. *J Biol Chem* 1994;269:16712-9.
19. Hansen TW, Mathiesen SB, Walaas SI. Bilirubin has widespread inhibitory effects on protein phosphorylation. *Pediatr Res* 1996;39:1072-7.
20. Amit Y, Boneh A. Bilirubin inhibits protein kinase C activity and protein kinase C-mediated phosphorylation of endogenous substrates in human skin fibroblasts. *Clin Chem Acta* 1993;223:103-11.
21. Clark JE, Foresti R, Sarathchandra P, Kaur H, Green CJ, Motterlini R. Heme oxygenase-1 derived bilirubin ameliorates postschismic myocardial dysfunction. *Am J Physiol* 2000;278:643-51.
22. Stocker RY, Yamamoto AF, McDonagh AN, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. *Science* 1987;235:1043-6.
23. Hopkins PN, Wu LL, Hunt SC, James BC, Vincent GM, Williams RR. Higher serum bilirubin is associated with decreased risk for early familial coronary artery disease. *Arterioscler Thromb Vasc Biol* 1996;16:250-5.
24. Schwertner HA, Jackson WG, Tolan G. Association of low serum concentration of bilirubin with risk of coronary artery disease. *Clin Chem* 1994;40:18-23.
25. Kawamura K, Ishikawa K, Wada Y, et al. Bilirubin from heme oxygenase-1 attenuates vascular endothelial activation and dysfunction. *Arterioscler Thromb Vasc Biol* 2005;25:155-60.
26. Furchgott RF, Jothianandan D. Endothelium dependent and independent vasodilatation involving cyclic GMP: relaxation induced by nitric oxide, carbon monoxide and light. *Blood Vessels* 1991;28:52-61.
27. Kajimura M, Shimoyama M, Tsuyama S, et al. Visualization of gaseous monoxide reception by soluble guanylate cyclase in the rat retina. *FASEB J* 2003;17:506-8.
28. Motterlini R, Gonzales A, Foresti R, Clark JE, Green CJ, Winslow RM. Heme oxygenase-1-derived carbon monoxide contributes to the suppression of acute hypertensive response in vivo. *Circ Res* 1998;83:568-77.
29. Morita T, Kourembanas S. Endothelial cell expression of vasoconstrictors and growth factors is regulated by smooth muscle cell-derived carbon monoxide. *J Clin Invest* 1995;96:2676-82.
30. Liu XM, Chapman GB, Peyton KJ, Schafer AI, Durant W. Carbon monoxide inhibits apoptosis in vascular smooth muscle cells. *Cardiovasc Res* 2002;55:396-405.
31. Yet SF, Perrella MA, Layne MD, et al. Hypoxia induces severe right ventricular dilatation and infarction in heme oxygenase-1 null mice. *J Clin Invest* 1999;103: 23-9.
32. Fogg S, Agarwal A, Nick HS, Visner GA. Iron regulates hyperoxia-dependent human heme oxygenase 1 gene expression in pulmonary endothelial cells. *Am J Respir Cell Mol Biol* 1999;20:97-804.
33. Balla J, Vercellotti GM, Jeney V, et al. Heme, heme oxygenase, and ferritin: how the vascular endothelium survives (and dies) in an iron-rich environment. *Antioxid Redox Signal* 2007;9:2119-37.
34. Selmecli L, Antal M, Horkay F, et al. Enhanced accumulation of pericardial fluid ferritin in patients with coronary artery disease. *Cor Artery Dis* 2000;11:53-6.
35. Wagener F, Volk HD, Willis D, et al. Different faces of the heme-heme oxygenase system in inflammation. *Pharmacol Res* 2003; 55:551-71.
36. Nath KA, Balla G, Vercellotti GM, et al. Induction of heme oxygenase is a rapid protective response in rhabdomyolysis in the rat. *J Clin Invest* 1992;90:267-70.
37. Choi AMK. Heme-oxygenase-1 protects the heart. *Circ Res* 2001;89: 105-9.
38. Fujita T, Toda K, Karimova A, et al. Paradoxical rescue from ischemic lung injury by inhaled carbon monoxide driven by derepression of fibrinolysis. *Nat Med* 2001;7:598-604.
39. Ito T, Okada T, Miyashita H, et al. Interleukin-10 expression mediated by an adeno-associated virus vector prevents monocrotaline-induced pulmonary arterial hypertension in rats. *Circ Res* 2007;101: 734-41.
40. Zabalgoitia M, Colston JT, Reddy SV, et al. Carbon monoxide donors or heme oxygenase-1 (HO-1) overexpression blocks interleukin-18-mediated NF-kappaB-PTEN-dependent human cardiac endothelial cell death. *Free Radical Biol Med* 2008;44:284-98.
41. Tsuchihashi S, Zhai Y, Bo Q, Busuttill RW, Kupiec-Weglinski JW. Heme oxygenase-1 mediated cytoprotection against liver ischemia/reperfusion injury: inhibition of type-1 interferon signaling. *Transplantation* 2007;83:1628-34.
42. Shen XD, Ke B, Zhai Y, et al. CD154-CD40 T-cell costimulation pathway is required in the mechanism of hepatic ischemia/reperfusion injury, and its blockade facilitates and depends on heme oxygenase-1 mediated cytoprotection. *Transplantation* 2002;74:315-9.
43. Mayer M. Association of serum bilirubin concentration with risk of coronary artery disease. *Clin Chem* 2000;46:1723-7.
44. Wu TW, Fung KP, Wu J, Yang CC, Weisel RD. Antioxidation of human low density lipoprotein by unconjugated and conjugated bilirubins. *Biochem Pharmacol* 1996;51:859-62.
45. Petrache I, Otterbein LE, Alam J, Wiegand GW, Choi AM. Heme oxygenase-1 inhibits TNF-alpha-induced apoptosis in cultured fibroblasts. *Am J Physiol Lung Cell Mol Physiol* 2000;278:L312-9.
46. Ferris CD, Jaffrey SR, Sawa A, et al. Haem oxygenase-1 prevents cell death by regulating cellular iron. *Nat Cell Biol* 1999;1:152-7.

47. Brouard S, Otterbein LE, Anrather J, et al. Carbon monoxide generated by heme oxygenase-1 suppresses endothelial cell apoptosis. *J Exp Med* 2000;192:1015-26.
48. Iori E, Pagnin E, Gallo A, et al. Heme oxygenase-1 is an important modulator in limiting glucose-induced apoptosis in human umbilical vein endothelial cells. *Life Sci* 2008;82:383-92.
49. Silva G, Cunha A, Gregoire IP, Seldon MP, Soares MP. The antiapoptotic effect of heme oxygenase-1 in endothelial cells involves the degradation of p38 alpha MAPK isoform. *J Immunol* 2006;177:1894-903.
50. Brouard S, Berberat PO, Tobiasch E, Seldon MP, Bach FH, Soares MP. Heme oxygenase-1-derived carbon monoxide requires the activation of transcription factor NF-kappa B to protect endothelial cells from tumor necrosis factor-alpha-mediated apoptosis. *J Biol Chem* 2002;277:17950-61.
51. Melo LG, Agrawal R, Zhang L, et al. Gene therapy strategy for long-term myocardial protection using adeno-associated virus-mediated delivery of heme oxygenase gene. *Circulation* 2002;105:602-7.
52. Zhen-Wei X, Jian-Le S, Qi Q, Wen-Wei Z, Xue-Hong Z, Zi-Li Z. Heme oxygenase-1 improves the survival of discordant cardiac xenograft through its anti-inflammatory and anti-apoptotic effects. *Pediatr Transplant* 2007;11:850-9.
53. Asija A, Peterson SJ, Stec DE, Abraham NG. Targeting endothelial cells with heme oxygenase-1 gene using VE-cadherin promoter attenuates hyperglycemia-mediated cell injury and apoptosis. *Antioxid Redox Signal* 2007;9:2065-74.
54. Rushworth SA, MacEwan DJ. HO-1 underlies resistance of AML cells to TNF-induced apoptosis. *Blood* 2008;111:3793-801.
55. Sass G, Shembade ND, Tiegs G. Tumour necrosis factor alpha (TNF)-TNF receptor 1-inducible cytoprotective proteins in the mouse liver: relevance of suppressors of cytokine signalling. *Biochem J* 2005;385:537-44.
56. Clark JE, Foresti R, Green CJ, Motterlini R. Dynamics of haem oxygenase-1 expression and bilirubin production in cellular protection against oxidative stress. *Biochem J* 2000;348:615-6.
57. Christou H, Morita T, Hsieh CM, et al. Prevention of hypoxia-induced pulmonary hypertension by enhancement of endogenous heme oxygenase-1 in the rat (erratum in: *Circ Res* 2002;90:e66-a). *Circ Res* 2000;86:1224-9.
58. Nakao A, Neto JS, Kanno S, et al. Protection against ischemia/reperfusion injury in cardiac and renal transplantation with carbon monoxide, biliverdin and both. *Am J Transplant* 2005;5:282-91.
59. Chen YH, Yet SF, Perrella MA. Role of heme oxygenase-1 in the regulation of blood pressure and cardiac function. *Exper Biol Med* 2003;228:447-53.
60. Immenschuh S, Schroder H. Heme oxygenase-1 and cardiovascular disease. *Histol Histopathol* 2006;21:679-85.
61. Deramandt BM, Braunstein S, Remy P, Abraham NG. Gene transfer of human heme oxygenase into coronary endothelial cells potentially promotes angiogenesis. *J Cell Biochem* 1998;68:121-7.
62. Suzuki M, Iso-o N, Takeshita S, et al. Facilitated angiogenesis induced by heme oxygenase-1 gene transfer in a rat model of hindlimb ischemia. *Biochem Biophys Res Commun* 2003;302:138-43.
63. Abdel-Aziz MT, el-Asmar MF, el-Miligy D, et al. Retrovirus-mediated human heme oxygenase-1 (HO-1) gene transfer into rat endothelial cells: the effect of HO-1 inducers on the expression of cytokines. *Int J Biochem Cell Biol* 2003;35:324-32.
64. Jozkowicz A, Huk I, Nigisch A, et al. Hemoxygenase and angiogenic activity of endothelial cells: stimulation by carbon monoxide and inhibition by tin protoporphyrin-IX. *Antioxid Redox Signal* 2003;5:155-62.
65. Bussolati B, Ahmed A, Pemberton H, et al. Bifunctional role for VEGF-induced heme oxygenase-1 in vivo: induction of angiogenesis and inhibition of leukocytic infiltration. *Blood* 2004;103:3:761-6.
66. Bussolati B, Mason JC. Dual role of VEGF-induced heme-oxygenase-1 in angiogenesis. *Antioxid Redox Signal* 2006;8:1153-63.
67. Dulak J, Deshane J, Jozkowicz A, Agarwal A. Heme oxygenase-1 and carbon monoxide in vascular pathobiology: focus on angiogenesis. *Circulation* 2008;117:231-41.
68. Exner M, Schillinger M, Minar E, et al. Heme oxygenase-1 gene promoter microsatellite polymorphism is associated with restenosis after percutaneous transluminal angioplasty. *J Endovasc Ther* 2001;8:433-40.
69. Schillinger M, Exner M, Minar E, et al. Heme oxygenase-1 genotype and restenosis after balloon angioplasty: a novel vascular protective factor. *J Am Coll Cardiol* 2004;43:950-7.
70. Schillinger M, Exner M, Mlekusch W, et al. Heme oxygenase-1 gene promoter polymorphism is associated with abdominal aortic aneurysm. *Thromb Res* 2002;106:131-6.
71. Exner M, Böhmig GA, Schillinger M, et al. Donor heme oxygenase-1 genotype is associated with renal allograft function. *Transplantation* 2004;77:538-42.
72. Ullrich R, Exner M, Schillinger M, et al. Microsatellite polymorphism in the heme oxygenase-1 gene promoter and cardiac allograft vasculopathy. *J Heart Lung Transplant* 2005;10:1600-5.
73. Dick P, Schillinger M, Minar E, et al. Haem oxygenase-1 genotype and cardiovascular adverse events in patients with peripheral artery disease. *Eur J Clin Invest* 2005;35:731-7.
74. Chen YH, Lin SJ, Lin MW, et al. Microsatellite polymorphism in promoter of heme oxygenase-1 gene is associated with susceptibility to coronary artery disease in type 2 diabetic patients. *Hum Genet* 2002;111:1-8.
75. Kaneda H, Ohno M, Taguchi J, et al. Heme oxygenase-1 gene promoter polymorphism is associated with coronary artery disease in Japanese patients with coronary risk factors. *Arterioscler Thromb Vasc Biol* 2002;22:1680-5.
76. Gulesserian T, Wenzel C, Endler G, et al. Clinical restenosis after coronary stent implantation is associated with the heme oxygenase-1 gene promoter polymorphism and the heme oxygenase-1 99G/C variant. *Clin Chem* 2005;51:1661-5.
77. Ono K, Mannami T, Iwai N. Association of a promoter variant of the haeme oxygenase-1 gene with hypertension in women. *J Hypertens* 2003;21:1497-3.
78. Turpeinen H, Kyllönen LE, Parkkinen J, Laine J, Salmela KT, Partanen J. Heme oxygenase 1 gene polymorphisms and outcome of renal transplantation. *Int J Immunogenet* 2007;34:253-7.
79. Li YG, Wang DM, Chen SM, et al. Haem oxygenase-1 expression and coronary heart disease—association between levels of haem oxygenase-1 expression and angiographic morphology as well as the quantity of coronary lesions. *Acta Cardiol* 2006;61:295-300.
80. Chen SM, Li YG, Wang DM. Study on changes of heme oxygenase-1 expression in patients with coronary heart disease. *Clin Cardiol* 2005;28:197-201.
81. Morsi WG, Shaker OG, Ismail EF, et al. HO-1 and VEGF gene expression in human arteries with advanced atherosclerosis. *Clin Biochem* 2006;39:1057-62.
82. da Silva JL, Stoltz RA, Dunn MW, Abraham NG, Shibahara S. Diminished heme oxygenase-1 mRNA expression in RPE cells from diabetic donors as quantitated by competitive RT/PCR. *Curr Eye Res* 1997;16:380-6.
83. Avogaro A, Pagnin E, Calò L. Monocyte NADPH oxidase subunit p22(phox) and inducible hemoxygenase-1 gene expressions are increased in type II diabetic patients: relationship with oxidative stress. *J Clin Endocrinol Metab* 2003;88:1753-9.
84. Bruce CR, Carey AL, Hawley JA, Febbraio MA. Intramuscular heat shock protein 72 and heme oxygenase-1 mRNA are reduced in patients with type 2 diabetes: evidence that insulin resistance is associated with a disturbed antioxidant defense mechanism. *Diabetes* 2003;52:2338-45.
85. Adakalakeswari A, Balasubramanyam M, Rema M, Mohan V. Differential gene expression of NADPH oxidase (p22phox) and hemoxygenase-1 in patients with type 2 diabetes and microangiopathy. *Diabet Med* 2006;23:666-74.
86. Schiekofer S, Galasso G, Andrassy M, Aprahamian T, Schneider J, Rocnik E. Glucose control with insulin results in reduction of NF-kappaB-binding activity in mononuclear blood cells of patients with recently manifested type 1 diabetes. *Diabetes Obes Metab* 2006;8:473-82.
87. Arredondo M, Jorquera D, Carrasco E, Albala C, Hertrampf E. Microsatellite polymorphism in the heme oxygenase-1 gene promoter is associated with iron status in persons with type 2 diabetes mellitus. *Am J Clin Nutr* 2007;86:1347-53.
88. Beschoner R, Adiodah D, Schwab JM, et al. Long-term expression of hemoxygenase-1 (HO-1, HSP-32) following focal cerebral infarctions and traumatic brain injury in humans. *Acta Neuropathol* 2000;100:377-84.

89. Morgan L, Hawe E, Palmen J, Montgomery H, Humphries SE, Kitchen N. Polymorphism of the heme oxygenase-1 gene and cerebral aneurysms. *Br J Neurosurg* 2005;19:317-21.
90. Immenschuh S, Ramadori G. Gene regulation of hemoxygenase-1 as a therapeutic target. *Biochem Pharmacol* 2000;60:1121-8.
91. Liu X, Simpson JA, Brunt KR, et al. Preemptive heme oxygenase-1 gene delivery reveals reduced mortality and preservation of left ventricular function 1 yr after acute myocardial infarction. *Am J Physiol Heart Circ Physiol* 2007;293:48-59.
92. Grosser N, Erdmann K, Hemmerle A, et al. Rosuvastatin up regulates the antioxidant defence protein heme oxygenase-1. *Biochem Biophys Res Commun* 2004;325:871-6.
93. Grosser N, Hemmerle A, Berndt G, et al. The antioxidant defense protein heme oxygenase 1 is a novel target for statins in endothelial cells. *Free Radic Biol Med* 2004;37:2064-71.
94. Dulak J, Loboda A, Jazwa A, et al. Atorvastatin affects several angiogenic mediators in human endothelial cells. *Endothelium* 2005;12:233-41.
95. Lee TS, Chang CC, Zhu Y, Shyy JY. Simvastatin induces heme oxygenase-1: a novel mechanism of vessel protection. *Circulation* 2004;110:1296-302.
96. Grosser N, Abate A, Oberle S, et al. Heme oxygenase-1 induction may explain the antioxidant profile of aspirin. *Biochem Biophys Res Commun* 2003;308:956-60.
97. Krönke G, Kadl A, Ikonomu E, et al. Expression of heme oxygenase-1 in human vascular cells is regulated by peroxisome proliferator-activated receptors. *Arterioscler Thromb Vasc Biol* 2007;27:1276-82.
98. Yachie A, Niida Y, Wada T, et al. Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. *J Clin Invest* 1991;103:129-35.

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**Key Words:** cardiovascular disease ■ hemoxygenase-1 ■ heme ■ bilirubin ■ carbon monoxide.