



Feeding behaviour after injection of α -adrenergic receptor agonists into the median raphe nucleus of food-deprived rats

Anderson Savaris Ribas*, Rafael Appel Flores, Aparecida Marcelino de Nazareth, Moacir Serralvo Faria, Mariana Graciela Terenzi, José Marino-Neto, Marta Aparecida Paschoalini

Department of Physiological Sciences, Centre of Biological Sciences-CCB, Federal University of Santa Catarina (UFSC), 88040-970, Florianópolis, SC, Brazil

ARTICLE INFO

Article history:

Received 15 April 2011

Received in revised form 29 July 2011

Accepted 25 August 2011

Keywords:

Adrenergic receptors

Food intake

Median raphe nucleus

Phenylephrine

ABSTRACT

This study investigated the participation of median raphe nucleus (MnR) α 1-adrenergic receptors in the control of feeding behaviour. The α 1-adrenergic agonist phenylephrine (PHE) and α 2-adrenergic agonist clonidine (CLON) (at equimolar doses of 0, 6 and 20 nmol) were injected into the MnR of: a) rats submitted to overnight fasting (18 h); or b) rats maintained with 15 g of lab chow/day for 7 days. Immediately after the drug injections, the animals were placed in the feeding chamber and feeding and non-ingestive behaviours such as grooming, rearing, resting, sniffing and locomotion were recorded for 30 min. The results showed that both doses of PHE injected into the MnR of overnight fasted animals decreased food intake accompanied by an increase in the latency to start feeding. A reduction in feeding duration was observed only after treatment of the MnR with the 20 nmol dose of PHE. Both locomotion duration and sniffing frequency increased after injection with the highest dose PHE into the MnR. Feeding frequency and the other non-ingestive behaviours remained unchanged after PHE treatment in the MnR. Both doses of PHE injected into the MnR of food-restricted rats decreased food intake. This hypophagic response was accompanied by a decrease in feeding duration only after treatment of the MnR with the highest dose of PHE. The latency to start feeding and feeding frequency were not affected by injection of either dose of PHE into the MnR. While both doses of PHE increased sniffing duration, the highest dose of PHE increased resting duration and resting frequency. Treatment with CLON into the MnR did not affect feeding behaviour in either of the food deprivation conditions. The present results indicate the inhibitory functional role of α 1-adrenergic receptors within the MnR on feeding behaviour.

© 2011 Elsevier Inc. Open access under the [Elsevier OA license](http://www.elsevier.com/locate/elsevier).

1. Introduction

The raphe nuclei are constituted of serotonergic neurons grouped into eight nuclei located in the midline of the brainstem. These nuclei are considered to be the main source of prosencephalic serotonin (5-hydroxytryptamine) (5-HT) [1–4]. The dorsal raphe nucleus and the median raphe nucleus (MnR) also contain distinct subpopulations of non-serotonergic neurons that occur in equal or greater numbers compared to the serotonergic neurons [5–9]. In the MnR, GABAergic neurons are located in both midline and lateral regions across the rostro-caudal extent of the MnR. 5-HT neurons are found just lateral and adjacent to the population of GABAergic neurons at the midline with little overlap or co-localisation between the two populations [10]. Non-serotonergic neurotransmitters are co-localised with serotonergic neurotransmitters [10–14], and include gamma-amino butyric acid (GABA), glutamate and corticotropin-releasing factor (CRF)

[15–17]. Thus, both 5-HT and non-5-HT neurotransmitters may be co-released within the raphe nuclei as well as in projection areas [18,19]. It is estimated that 20% of the 5-HT innervations originating in the MnR predominantly reach the dorsal hippocampus, medial septum and hypothalamus [5,20].

A great density of 5-HT_{1A} receptors is found in the MnR [21–24] and they function as autoreceptors that regulate the synthesis and release of 5-HT in their projection areas [21,25]. Agonists of 5-HT_{1A} receptors, such as 8-hydroxy-2-(di-n-propylamino) tetralin (8-OH-DPAT), stimulate food intake when injected into the MnR [26–29]. Besides the high density of 5-HT receptors in the MnR, a great density of α 1- and α 2-adrenergic receptors in this nucleus has been reported [30–32]. MnR neurons receive noradrenergic innervation from the locus coeruleus/subcoeruleus, lateral tegmental area, and projections from the adrenaline (AD) (C1 + C2) medullary nuclei and A1/A2 cell groups [33–37]. The noradrenergic inputs to MnR exert tonic facilitatory control of 5-HT release through α 1-adrenergic receptors and inhibitory control by α 2-adrenergic receptors [21,30].

Previous studies in our laboratory revealed AD but not noradrenaline (NA) injected into the MnR decreased food intake and shortened

* Corresponding author. Tel.: +55 48 37219352; fax: +55 48 37219672.
E-mail address: andersonsavarisribas@hotmail.com (A.S. Ribas).

meal duration in food-restricted rats [38]. On the other hand, injection of AD into the MnR of free-feeding rats increased food intake, feeding frequency and decreased the latency to start feeding [39]. Since the inhibitory action of 5-HT on feeding behaviour has been extensively reported [25,40–43] and due to the serotonergic innervation of the hypothalamic paraventricular nucleus, an important region where 5-HT has been implicated in feeding regulation arises from MnR serotonergic projections [41,44,45]. We have suggested that in food-restricted rats, AD-induced hypophagia may be due to the activation of α 1-adrenergic receptors on 5-HT MnR neurons resulting in 5-HT release and an indirect inhibitory action of AD on feeding [38]. In free-feeding rats, the food intake effects evoked by AD injections may be attributed to the activation of α 2-adrenergic receptors on 5-HT MnR neurons that could act to suppress the release of 5-HT and its inhibitory action on feeding behaviour [39].

This suggestion was reinforced by data showing that treatment with clonidine (CLON), an α 2-adrenergic agonist, into the MnR of free-feeding rats resulted in a hyperphagic response with an intensity, duration as well as frequency similar to that induced by AD [46]. In this case, an inhibitory influence activated by adrenergic inputs was removed due to α 2-adrenergic stimulation, which in turn decreased the concentration of catecholamines in the synapse. The feeding behaviour effects induced by CLON in the MnR were specifically mediated by α 2-adrenergic activation since previous injection of an α 2-adrenergic antagonist into the MnR blocked the hyperphagic response evoked by this drug [47]. In contrast, phenylephrine (PHE) treatment in the MnR failed to modify feeding behaviour in rats with free access to food. This lack of change in the feeding response after MnR α 1-adrenergic receptor activation was attributed to the presence of elevated adrenergic inputs on MnR neurons which restrain food intake under free-feeding conditions [48]. This suggestion was reinforced by data showing that α 1-adrenergic receptor blockade within the MnR increased food intake in free-feeding rats [47].

In order to strengthen the inhibitory functional role that α 1-adrenergic receptors within the MnR exert on food intake, the present study was designed to evaluate the feeding behaviour effects evoked by injection of α 1-adrenergic receptor agonists into the MnR of food-deprived animals since it has been reported that the inhibitory serotonergic tone is lower in animals submitted to a food restriction regimen than in free-feeding animals [49–51]. In addition, the feeding effects caused by MnR α 2-adrenergic receptor activation were also investigated.

2. Materials and methods

2.1. Animals and surgery

All the experimental procedures described below were conducted in strict adherence to the recommendations found in the “Principles of animal care” and were approved by the local Committee for Ethics in Animal Research (CEUA-UFSC, protocol # PP00367). Male adult Wistar rats ($n=108$) weighing 270–310 g at the time of surgery were used in this study and were group-housed in a room at 21 ± 2 °C under a 12 h lighting cycle (lights on at 07:00 h) with free access to food and water, except when food restriction was applied. The rats were anaesthetised with ketamine hydrochloride (100 mg/kg, i.p.) and xylazine (20 mg/kg, i.p.) and were placed in a stereotaxic apparatus (Insight Instruments, Ribeirão Preto, SP, BRA) for unilateral stainless steel guide cannula (30 G) implantation according to the coordinates ($AP \pm 7.8$, $L \pm 3.0$ and $DV \pm 9.0$) described by Paxinos and Watson [52]. The guide-cannula was inserted at a lateral angle of 20° to avoid the sagittal sinus, and cerebral aqueduct obstruction and was aimed at 2 mm dorsal to the MnR. The cannula was anchored to the skull with jeweller screws and fixed with dental cement; the cannula was maintained patent between experiments by an inner removable stylet.

2.2. Drugs and injections

Phenylephrine hydrochloride (6 and 20 nmol, purchased from Sigma Chemical Co., St. Louis, MO, USA) and clonidine hydrochloride (6 and 20 nmol, purchased from Tocris Bioscience, Ellisville, MO, USA) were freshly dissolved in 0.9% saline, which was used as the vehicle in control experiments. The drug injections were made through an inner cannula (33 G) that extended 2 mm beyond the tip of the guide cannula connected by polyethylene tubing (PE10) to a Hamilton microsyringe (1 μ l) fitted to an injection pump. The injected volumes (0.2 μ l) were administered over a period of 60 s and a further 60 s was allowed for the solution to diffuse from the cannula. The drug doses used in the experiments were derived from our previous studies [38,39,46,47].

2.3. Experimental procedures

For 1 week after surgery, the rats were housed individually with free access to food and water. After this period, the animals were separated into two experimental groups. 1) Food-restricted rats: in this group, the animals were submitted to a food restriction regimen whereby the daily food allotment was limited to 15 g of laboratory chow, delivered at 17:00 h for 7 days. The rats continued to have free accesses to water. After 4 days of food restriction, the body weight declined by approximately 10% and remained stable until the day of the experiment. The experimental session began on the eighth day of food restriction. The rats were habituated, on two occasions before the experiments, to the handling and injection procedures. During the habituation, the rats received mock injections to allow acclimation to the microinfusion procedures. No solutions were delivered on mock injection days by an inner cannula that exhibited the same guide cannula length. The rat chow not consumed at the end of the habituation sessions was offered to the animals at 17:00 h, in order to complete 15 g of chow pellets/day. 2) Overnight fast (18 h): in this group the animals were deprived of food overnight and the morning prior to all experiments (18 h), but continued to have free accesses to water. Each animal received only one drug treatment. All procedures were carried out between 13:00 h and 16:30 h. Immediately after the drug injection into the MnR, the rats were placed in the recording chamber constructed with transparent glass ($49 \times 34 \times 32$ cm), containing food and tap water (in a bottle placed outside the test cage with a spout projected through the wall of the cage). The session was recorded by a webcam perpendicularly located 60 cm above the cage floor for subsequent detailed behavioural analysis through Etholog 2.2 [53]. The back and lateral walls, as well as the floor cage, were coated with a black adhesive plastic paper. In order to facilitate behavioural recording, the front wall of the test cage had a mirror with the same dimensions arranged at a 45° angle in relation to the vertical plane. This mirror arrangement also prevented the animal from seeing its reflection in the mirror. At the end of the recording period, any food that occasionally spilled on the cage floor was recovered and weighed with the food that remained in the feeder. The difference between food or water weight at the beginning and at the end of the recording period was taken as the amount of food or water consumed. During the 30 min experimental session, the ingestive behaviours such as the latency to start feeding and drinking, the feeding and drinking duration, as well as the feeding and drinking frequency were evaluated. The duration and frequency of five non-ingestive behaviours (grooming, locomotion, rearing, resting and sniffing) were also assessed. The behaviour duration assessed in this study represents the sum of each episode occurred during the experimental session. All behaviour categories were defined in previous studies [54] and are described in Table 1.

An additional experimental group (experiment 3) was carried out in order to confirm whether the PHE effects on feeding behaviour were site-specific. In this group, the PHE dose that evoked feeding behaviour changes in experiments 1 and 2 was injected into areas in the vicinity of

Table 1
The behavioural categories used for behavioural analysis.
Based on Halford and Blundell [54].

Category	Description
Eating (food intake)	Biting, gnawing or swallowing food from wet mash dish directly or from front paws.
Drinking (water intake)	Licking the spout water bottles.
Grooming	Licking of the body, feet, and genitals. Scratching of coat or head with hind leg. Stroking whiskers with paws. Biting of the tail.
Locomotion	Walking around the cage or circling. Movements' involving all four paws.
Rearing	Front paws raised from the tank floor and either placed on the side of the tank or placed in front of the body.
Sniffing	Rapid wrinkling of nose (twitching of vibrissae) directed at some aspect of the environment. Head movement. Rear limbs immobile.
Resting	Animal inactive. Relaxed position with head curled to body or resting on the bottom of the cage, stretched out either on side or belly.

the MnR such as the pontine nuclei (Pn), pontine nuclei oral part (PnO), paramedia raphe nucleus (PMnR) reticulotegmental nucleus pons (RtTg), trigeminothalamic tract (tth), medial lemniscus (ml), and the feeding behaviour of both food-restricted and overnight fasted rats was evaluated.

2.4. Histological analysis

At the end of the experiments, the animals were deeply anaesthetised and transcardially perfused with 0.9% saline, followed by 10% formalin. The brains were removed and subsequently cut (on a vibratome) in the transverse plane (100 µm thick sections). Sections were stained with cresyl violet and the cannula loci were examined and documented through a camera lucida attached to a light microscope. Cannula placements were mapped onto the corresponding atlas drawings of Paxinos and Watson [52]. Data from the rats with misplaced cannulae in the MnR were not included in the analyses.

2.5. Statistical analysis

Separate one-way ANOVA tests were carried out to analyse the food-restricted data, for the different doses of PHE or CLON (0, 6 and 20 nmol). The same statistical procedures were employed for data analyses of the overnight fast group. The effects of treatment with saline (0) or PHE (20 nmol) in the MnR and non-median raphe nucleus (Pn, PnO, PMnR, RtTg, tth and ml) were analysed by two-way ANOVA (drug-dose × injected area). All these tests were followed, when appropriate, by Duncan's post-hoc test, and a $p < 0.05$ was accepted as being statistically significant in these procedures. All statistical procedures were performed using the Statistica 9 software for Windows (StatSoft, Tulsa, OK, USA).

3. Results

Fig. 1 illustrates the injection sites within the MnR in coronal sections of the brain. Histological analysis indicated that 77 points of injection were within the MnR distributed between AP – 7.44 mm and AP – 8.40 mm. The misplaced injection sites included points in the Pn ($n = 11$), PnO ($n = 10$), RtTg ($n = 4$), PMnR ($n = 2$), tth ($n = 2$) and ml ($n = 2$).

3.1. Experiment 1: food-restricted rats (15 g chow/day for 7 days) – drug dose effects on ingestive and non- ingestive behaviours

The one-way ANOVA test revealed that food intake was affected by the drug PHE [$F(2,18) = 10.36$; $p < 0.001$]. Duncan's test revealed

that both doses of PHE (6 and 20 nmol) injected into the MnR of food-restricted rats decreased food intake when compared with the control group. This hypophagic effect was accompanied by a decrease in feeding duration [$F(2,18) = 3.65$; $p < 0.046$] only after treatment of the MnR with the highest dose of PHE (Fig. 2). The latency to start feeding and the feeding frequency were not affected by injections of either dose of PHE into the MnR (Fig. 2). Drinking behaviour was also not modified by PHE treatment (Table 2). Except for resting and sniffing behaviours, the other non-ingestive behaviours did not change after PHE administration into the MnR. The highest dose of PHE (20 nmol) increased resting duration [$F(2,18) = 5.27$; $p < 0.015$] and resting frequency [$F(2,18) = 3.69$; $p < 0.045$] (Table 3). Both PHE doses increased sniffing duration [$F(2,18) = 4.76$; $p < 0.021$] (Table 3). Feeding (Fig. 3), drinking (Table 2) and non-ingestive behaviours remained unchanged after administration of both doses of CLON into the MnR (Table 3).

3.2. Experiment 2: overnight fast group – drug dose effects on ingestive and non- ingestive behaviours

The one-way ANOVA test demonstrated that both food intake [$F(2,16) = 17.56$; $p < 0.00009$] and the latency to start feeding [$F(2,16) = 3.81$; $p < 0.044$] were significantly affected by PHE treatment in the MnR. Feeding duration was marginally affected by PHE [$F(2,16) = 3.30$; $p < 0.062$]. Food intake decreased after injection of both doses of PHE into the MnR of overnight fasted rats (Fig. 4). This hypophagic response was evoked by both doses of PHE and was accompanied by an increase in the latency to start feeding (Fig. 4). A reduction in feeding duration was observed only after treatment of the MnR with the 20 nmol dose of PHE (Fig. 4). Drinking behaviour was not modified by PHE treatment (Table 2). Non-ingestive behaviours remained unchanged after the administration of both doses of PHE into the MnR (Table 4), except for locomotion duration, which was marginally affected by the drug [$F(2,16) = 3.19$; $p < 0.067$], and sniffing frequency which was significantly affected by PHE treatment [$F(2,16) = 6.83$; $p < 0.007$]. Both locomotion duration and sniffing frequency increased after injection of the highest dose PHE into the MnR (Table 4). Treatment of the MnR with both doses of CLON did not change feeding (Fig. 5), drinking (Table 2) and non-ingestive behaviours of overnight fasted rats (Table 4).

3.3. Experiment 3: PHE injections into the vicinity of the MnR

The highest dose of PHE (20 nmol) injected into the MnR of both food-restricted rats and overnight fasted rats, which had evoked a decrease in food intake, was also administered to other regions in the vicinity of the MnR (Pn, PnO, PMnR, RtTg, tth, ml). These injection sites were grouped as the non-median raphe region and food intake data after PHE treatment were used to confirm the specificity of the MnR as the site of PHE injections that evoke a hypophagic response. The two-way ANOVA test revealed that the food intake exhibited by food-restricted rats was affected by the drug [$F(1,26) = 10.40$; $p = 0.003$] and there was also a significant interaction between the drug and the injection site [$F(1,26) = 11.89$; $p = 0.001$]; in addition, the food intake response exhibited by overnight fasted rats was significantly affected by the drug [$F(1,26) = 7.24$; $p = 0.012$] and there was also a significant interaction between the drug and the injection site [$F(1,26) = 14.20$; $p = 0.0008$]. The food intake decrease evoked by the highest dose of PHE was circumscribed to the MnR in both food-restricted and overnight fasted rats (Fig. 6).

4. Discussion

The results of the present study revealed that while the injection of PHE into the MnR decreased food intake in both food-deprived conditions, treatment with CLON in the MnR did not affect feeding

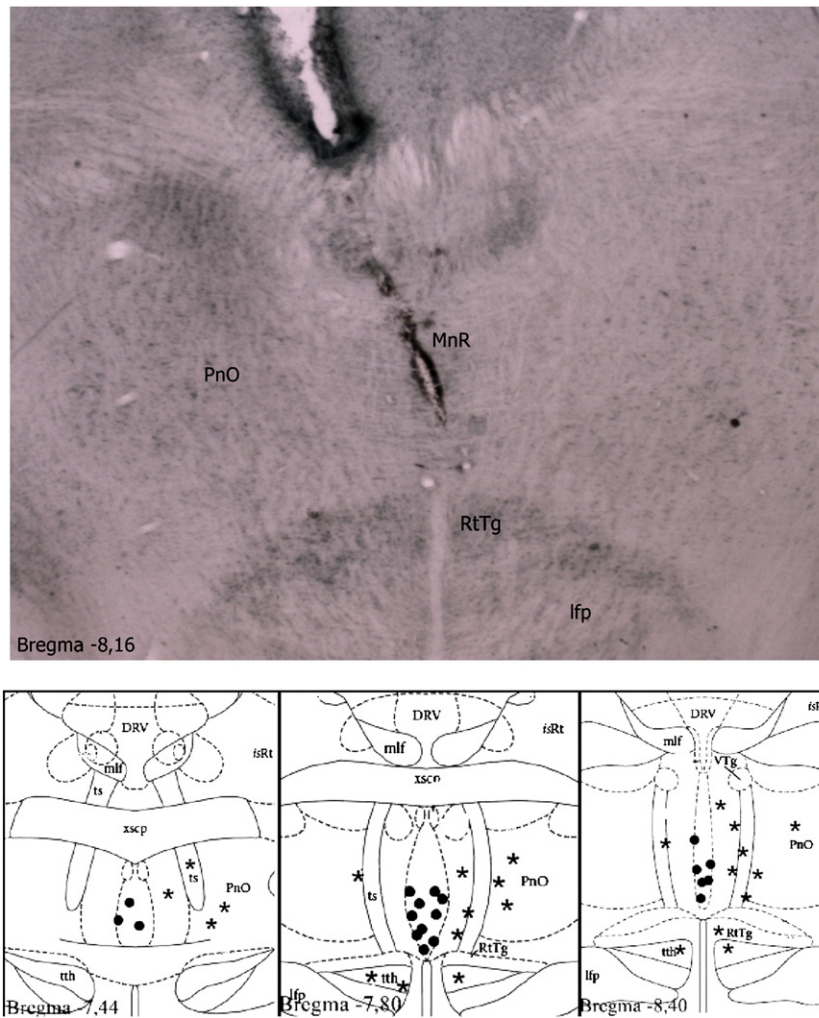


Fig. 1. Photomicrographs and schematic drawings of coronal sections through the median raphe nucleus (MnR) of the rats, illustrating the approximate sites (●) of phenylephrine (PHE)/clonidine (CLON) injections (at doses of 0, 6 and 20 nmol), and (★) phenylephrine (PHE) injections into the vicinity of the MnR. Arrows point to injection sites. Numbers at the top refer to anteroposterior stereotaxic coordinates from the rat brain atlas (Adapted from Paxinos and Watson, 2007). Scale bar = 100 μ m. DRV = dorsal raphe nu, ventral part; isRt = isthmic reticular formation; lfp = longitudinal fasciculus pons; mlf = medial longitudinal fasciculus; MnR = median raphe nucleus; PnO = Pontine reticular nu, oral part; RtTg = reticulotegmental nu pons; ts = tectospinal tract; tth = trigeminothalamic tract; VTg = ventral tegmental nu; xscp = decussation sup cerebellar peduncle.

behaviour. This hypophagic effect evoked by PHE is specifically mediated by α 1-adrenergic receptors within MnR, since treatment with the highest dose of PHE, which decreased food intake when administered into the MnR, did not change the feeding response when injected into the vicinity of the MnR. Opposite feeding responses were induced by the injection of α -adrenergic receptor agonists into the MnR of free-feeding rats. CLON injection into the MnR evoked hyperphagia while the PHE treatment did not change feeding behaviour [46,48].

Based on these results, we suggest that the hyperphagic effect evoked by CLON in free-feeding rats could be attributed to the inhibition of NA release from presynaptic stores, thus removing the α 1-adrenergic stimulatory tone on serotonergic neurons within the MnR, since α 2-adrenergic receptors seem to exert a tonic inhibitory influence on 5-HT release in the MnR [30]. The absence of feeding effects after treatment with PHE in the MnR of free-feeding rats could be due to elevated adrenergic inputs on serotonergic neurons [39,48]. It seems that the reduction of an adrenergic inhibitory influence in food-deprived rats allows for α 1-adrenergic receptor activation by PHE, resulting in 5-HT release, since facilitatory control of 5-HT release is attributed to α 1-adrenoceptors [21,30] situated on MnR serotonergic cell bodies [55]. In agreement with this suggestion, we have recently reported [47] that the blockade of α 1-

adrenergic receptors within the MnR of free-feeding rats resulted in hyperphagia accompanied by a reduction in the latency to start eating, an increase in feeding duration and an increase in feeding frequency. These data suggest the presence of tonic inhibitory inputs mediated by α 1-adrenergic receptors on MnR neurons that restrain food intake under free-feeding conditions. The 5-HT released by MnR α 1-adrenergic receptor activation could inhibit food intake through projections that terminate in the prosencephalic areas involved in the control of food intake such as the hypothalamus or amygdala [56–59].

It is estimated that 60% of MnR neurons are non-serotonergic [60]. It is possible that GABAergic or glutamatergic neurotransmission in the MnR [10,13,61] could be an alternative mechanism involved in the hypophagic response evoked by PHE in food-deprived rats, since intra-MnR injection of muscimol [1,62,63] and baclofen [1] or the glutamatergic agonist (NMOLDA) [64] and glutamatergic antagonists (kynurenic acid and 2-amino-5-phosphonovaleric acid) increased food intake in free-feeding rats [65]. Raphe/neurokinin pathways could also mediate the feeding effects induced by PHE treatment in the MnR since the activation of neurokinin-3 (NK-3) receptors located on MnR serotonin cell bodies has been shown to decrease food intake [63].

Except for the latency to start feeding, the hypophagic response evoked by PHE injections into the MnR of overnight fasted rats

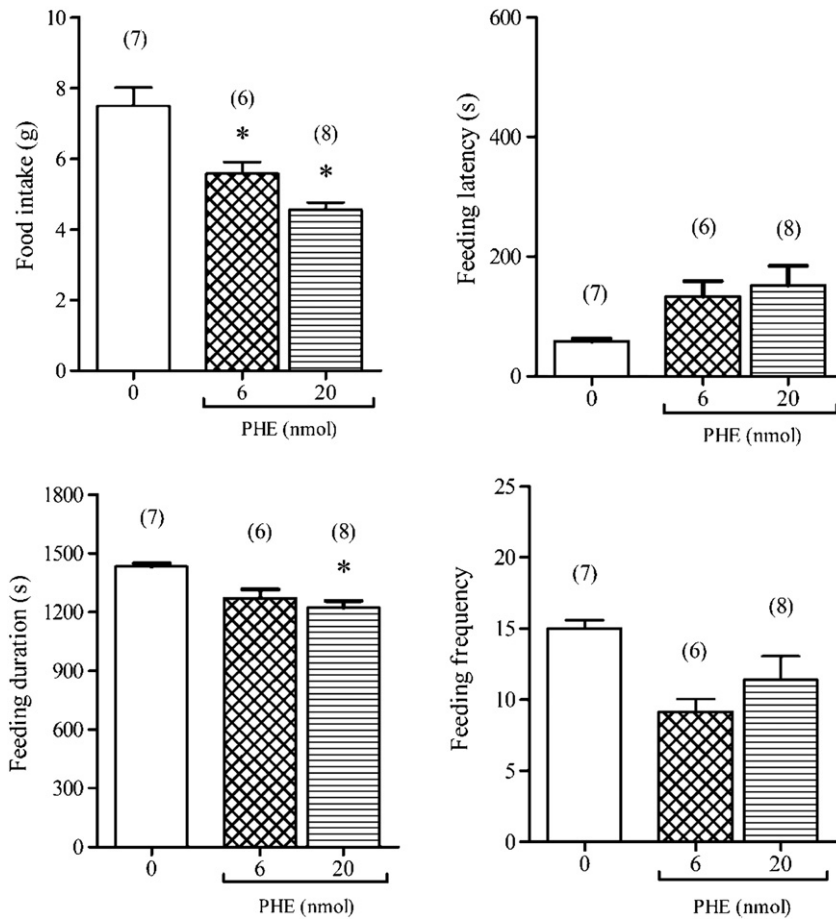


Fig. 2. Changes on food intake, feeding latency, feeding duration and feeding frequency after injection of vehicle (0) or phenylephrine (PHE, 6 nmol and 20 nmol) into the median raphe nucleus (MnR) of food restricted rats (15 g chow/day during 7 days). Values are mean \pm S.E.M. * $p < 0.05$ as compared to vehicle treatment (one-way ANOVA followed by Duncan's test for multiple comparisons). Numbers in parentheses indicate the number of animals per group.

was similar to that evoked by the same treatment in the MnR of food-restricted rats. The feeding duration decreased after the injection of the highest PHE dose into the MnR and feeding frequency was not affected by PHE treatment. In contrast, the latency to start feeding remained unchanged after treatment with PHE in the MnR of food-restricted rats, while both doses of PHE delayed the initiation of feeding in the overnight fasted rats. Feeding duration and the latency to start eating have been associated with different aspects of the control of feeding [66]. Changes in the latency to begin feeding may be associated with changes in satiety mechanisms or

in the processing of those signals that inhibit eating between feeding. In this sense, the influence of MnR $\alpha 1$ -adrenergic receptors on satiety mechanisms was presented only in the overnight fasted but not in the food-restricted rats. The reason for these discrepancies could be attributed to metabolic/hormone changes or patterns of orexigenic/anorexigenic neuropeptide gene expression associated with adaptations to a food restriction regimen [67–70]. On the other hand, changes in the feeding duration seemed to relate to satiation processes, or the mechanisms set in motion by nutrient-related signals that cause the end of a feeding bout. In this sense, we could

Table 2

Drinking behaviour after injection of phenylephrine (PHE) and clonidine (CLON) into the median raphe nucleus (MnR) of food restricted (15 g chow/day during 7 days) and overnight fast rats during 30 min. The number of animals used in each drug dose is illustrated in Figs. 2–4.

Behaviour	Food restricted rats			Overnight fast		
		PHE	CLON		PHE	CLON
Water intake	0	0.0 \pm 0.0	0.25 \pm 0.1	0	0.1 \pm 0.1	0.08 \pm 0.08
	6	0.08 \pm 0.08	0.50 \pm 0.2	6	0.0 \pm 0.0	0.3 \pm 0.3
	20	0.12 \pm 0.81	0.3 \pm 0.2	20	0.1 \pm 0.1	0.2 \pm 0.2
Latency (s)	0	1800.0 \pm 116.1	1577.7 \pm 138.4	0	1681.3 \pm 106.4	1627.4 \pm 263.7
	6	1745.6 \pm 125.5	1435.9 \pm 138.4	6	1800.0 \pm 114.9	1405.0 \pm 244.2
	20	1547.0 \pm 108.7	1639.4 \pm 138.4	20	1632.0 \pm 114.9	1296.8 \pm 263.7
Duration (s)	0	0.0 \pm 0.0	4.0 \pm 2.7	0	2.2 \pm 1.5	0.3 \pm 0.3
	6	0.79 \pm 0.79	5.2 \pm 3.9	6	0.0 \pm 0.0	1.4 \pm 0.8
	20	10.6 \pm 10.1	15.8 \pm 14.7	20	3.0 \pm 1.9	0.9 \pm 0.6
Frequency (turn)	0	0.0 \pm 0.0	0.8 \pm 0.5	0	0.4 \pm 0.3	0.2 \pm 0.2
	6	0.3 \pm 0.3	1.2 \pm 0.8	6	0.0 \pm 0.0	0.4 \pm 0.2
	20	1.2 \pm 1.1	1.2 \pm 0.8	20	0.6 \pm 0.6	0.3 \pm 0.2

Values are mean \pm S.E.M.

Table 3

Duration of non-ingestive behaviours 30 min after injection of vehicle (0) phenylephrine (PHE) or clonidine (CLON) into the median raphe nucleus of food restricted rats (15 g chow/day during 7 days). The number of animals used in each drug dose is illustrated in Figs. 2 and 3.

Behaviour	Phenylephrine (PHE)			Clonidine (CLON)		
	Dose (nmol/0.2 µl)	Duration (s)	Frequency (turn number/30 min)	Dose (nmol/0.2 µl)	Duration (s)	Frequency (turn number/30 min)
Locomotion	0	124.8 ± 19.4	33.4 ± 4.1	0	231.2 ± 48.7	60.5 ± 8.7
	6	158.2 ± 30.2	42.3 ± 9.3	6	180.7 ± 42.5	48.5 ± 12.5
	20	125.4 ± 29.9	30.5 ± 8.3	20	170.6 ± 19.1	42.8 ± 5.4
Resting	0	8.1 ± 4.3	2.7 ± 1.6	0	10.4 ± 5.5	2.8 ± 1.3
	6	10.0 ± 6.4	1.8 ± 0.7	6	22.8 ± 15.1	3.5 ± 1.6
	20	115.6 ± 40.9*	8.5 ± 2.4*	20	61.9 ± 36.7	3.3 ± 2.6
Rearing	0	57.8 ± 11.7	18.8 ± 2.7	0	86.8 ± 15.8	36.6 ± 7.9
	6	101.8 ± 29.7	26.6 ± 7.8	6	105.6 ± 24.7	29.7 ± 5.9
	20	39.5 ± 11.8	15.5 ± 4.8	20	84.5 ± 15.9	25.2 ± 4.0
Sniffing	0	70.6 ± 13.6	20.2 ± 3.8	0	99.9 ± 13.5	32.0 ± 3.8
	6	151.2 ± 30.0*	29.6 ± 4.1	6	155.9 ± 43.6	35.5 ± 9.0
	20	159.8 ± 23.1*	25.5 ± 4.7	20	132.5 ± 25.5	29.5 ± 3.6
Grooming	0	16.4 ± 4.2	2.2 ± 0.4	0	35.7 ± 10.3	4.5 ± 1.4
	6	31.9 ± 4.5	4.0 ± 0.4	6	61.0 ± 13.9	4.5 ± 1.1
	20	50.0 ± 15.9	4.7 ± 1.6	20	74.0 ± 24.0	5.2 ± 1.6

Values are mean ± S.E.M. *p < 0.05 as compared to the vehicle treatment (one-way ANOVA followed by Duncan's post-hoc test).

suggest that α1-adrenergic receptors in the MnR can be associated with neural circuits involved in satiation induced by signals arising from the digestive tract [71–74], characterised in this study by a reduction in the meal size and shortening feeding duration in both food-restricted and overnight fasted rats.

The hypophagia evoked by PHE injection into the MnR of 7-day food-restricted rats could be attributed to an increase in the duration

of sniffing behaviour, an indication of ambient exploration. This increase in exploratory behaviour induced by both doses of PHE could have interfered with the feeding response such that the animal stopped eating not because he was satiated, but because the drug could have stimulated a different behaviour such as exploration. However, the noticeable increase in resting (or sleep-like) behaviour after PHE injection into the MnR may be interpreted as an integral part

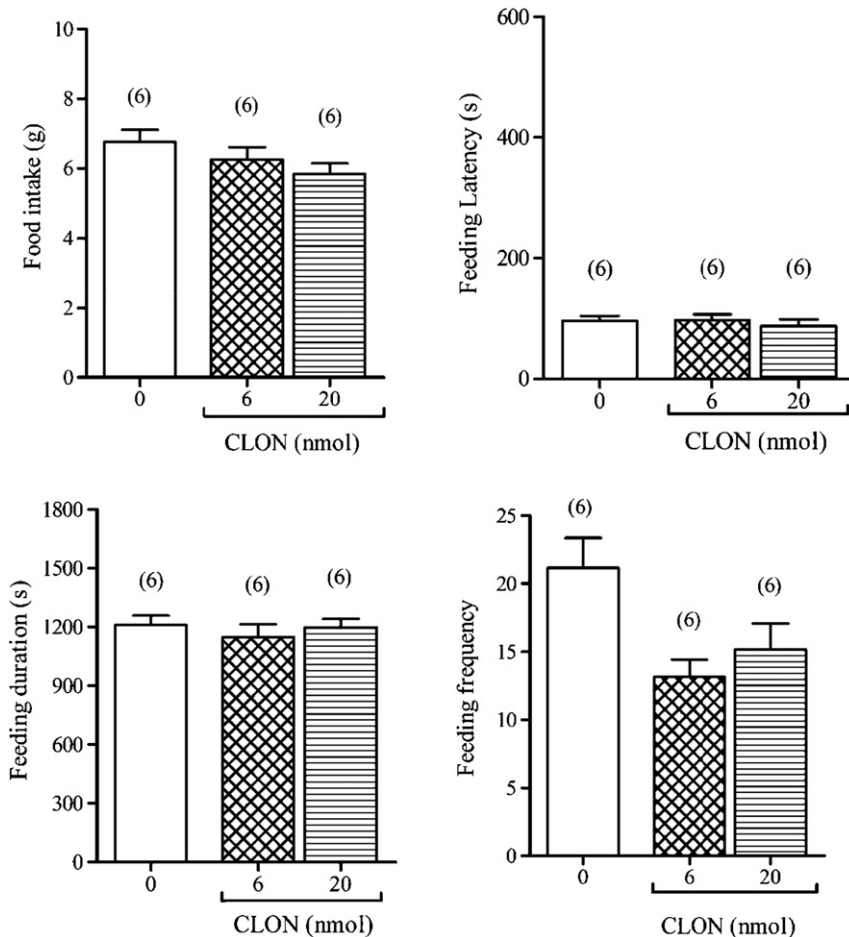


Fig. 3. Changes on food intake, feeding latency, feeding duration and feeding frequency after injection of vehicle (0) or clonidine (CLON, 6 nmol and 20 nmol) into the median raphe nucleus (MnR) of food restricted rats (15 g chow/day during 7 days). Values are mean ± S.E.M. Numbers in parentheses indicate the number of animals per group.

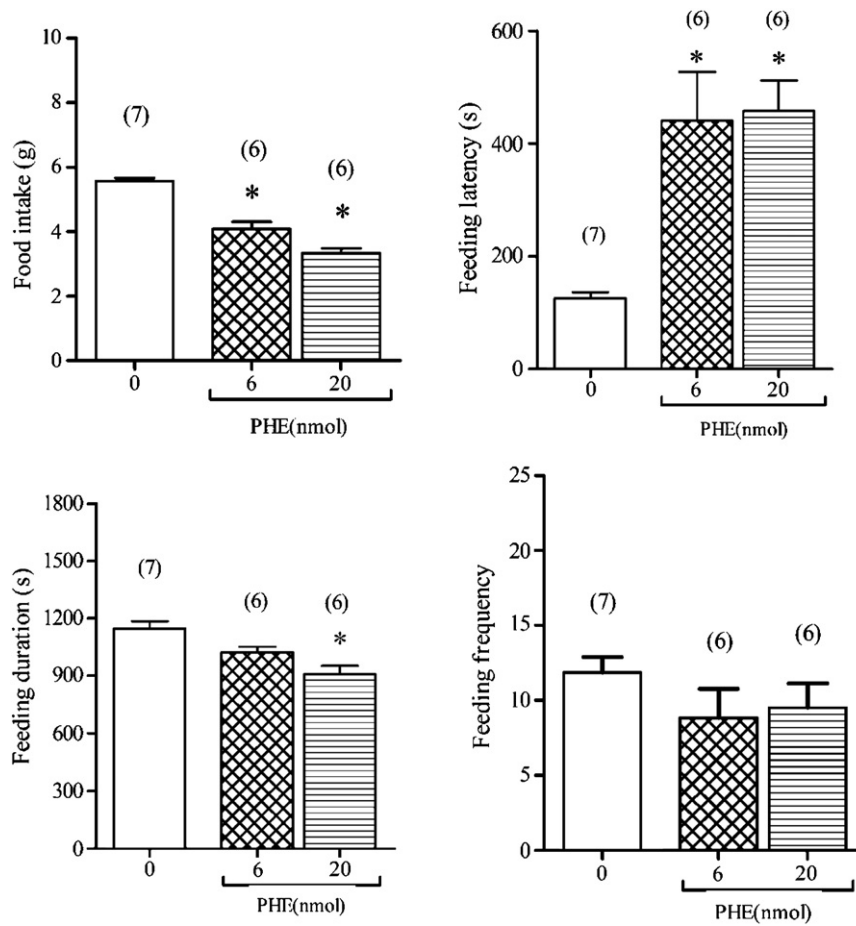


Fig. 4. Changes on food intake, feeding latency, feeding duration and feeding frequency after injection of vehicle (0) or phenylephrine (PHE, 6 nmol and 20 nmol) into the median raphe nucleus (MnR) of overnight fasted rats. Values are mean \pm S.E.M. * $p < 0.05$ as compared to vehicle treatment (one-way ANOVA followed by Duncan's test for multiple comparisons). Numbers in parentheses indicate the number of animals per group.

of the normal behavioural satiety sequence (feeding–maintenance–resting) [54,75,76]. Furthermore, the highest dose of PHE injected into the MnR of overnight fasted rats evoked an increase in both the sniffing frequency and duration of locomotion, but the lowest PHE dose evoked a hypophagic response with no changes in non-ingestive behaviours.

Taken together, these data suggest that $\alpha 1$ -adrenergic receptors within the MnR seem to specifically participate in the control of food intake.

We have previously shown that injection of AD into the MnR of free-feeding rats decreased the latency to start drinking [39]. These data suggest the involvement of MnR adrenergic receptors in circuits

Table 4

Duration of non-ingestive behaviours 30 min after injection of vehicle (0) phenylephrine (PHE) or clonidine (CLON) into the median raphe nucleus of overnight fasted rats. The number of animals used in each drug dose is illustrated in Figs. 4 and 5.

Behaviour	Phenylephrine (PHE)			Clonidine (CLON)		
	Dose (nmol/0.2 μ l)	Duration (s)	Frequency (turn number/30 min)	Dose (nmol/0.2 μ l)	Duration (s)	Frequency (turn number/30 min)
Locomotion	0	181.8 \pm 14.6	50.8 \pm 5.5	0	145.1 \pm 15.6	41.6 \pm 4.2
	6	230.1 \pm 12.3	52.6 \pm 4.4	6	181.5 \pm 14.7	51.4 \pm 3.2
	20	252.7 \pm 31.4*	65.0 \pm 9.2	20	183.3 \pm 26.7	46.2 \pm 6.5
Resting	0	6.3 \pm 3.1	2.0 \pm 1.1	0	26.1 \pm 11.5	7.3 \pm 2.7
	6	69.0 \pm 45.0	6.5 \pm 3.0	6	12.9 \pm 3.7	4.7 \pm 1.4
	20	55.9 \pm 28.8	8.3 \pm 2.0	20	12.7 \pm 4.3	4.3 \pm 1.2
Rearing	0	124.9 \pm 19.5	35.8 \pm 3.7	0	77.2 \pm 8.6	28.2 \pm 2.3
	6	126.3 \pm 40.0	32.3 \pm 5.6	6	95.6 \pm 14.1	33.0 \pm 3.6
	20	93.4 \pm 11.8	32.3 \pm 3.6	20	91.6 \pm 13.9	28.2 \pm 4.3
Sniffing	0	197.1 \pm 48.4	36.0 \pm 4.0	0	104.3 \pm 11.5	31.0 \pm 3.4
	6	169.5 \pm 18.1	37.1 \pm 5.0	6	131.8 \pm 15.2	34.9 \pm 4.1
	20	305.1 \pm 34.0	60.3 \pm 6.5*	20	142.3 \pm 38.2	33.2 \pm 7.5
Grooming	0	62.1 \pm 13.9	6.6 \pm 1.4	0	43.1 \pm 15.1	4.0 \pm 1.5
	6	75.6 \pm 11.3	4.5 \pm 0.5	6	87.2 \pm 18.2	4.7 \pm 0.5
	20	112.8 \pm 23.0	9.3 \pm 1.7	20	54.3 \pm 12.0	3.3 \pm 0.5

Values are mean \pm S.E.M.

* $p < 0.05$ as compared to the vehicle treatment (one-way ANOVA followed by Duncan's post-hoc test).

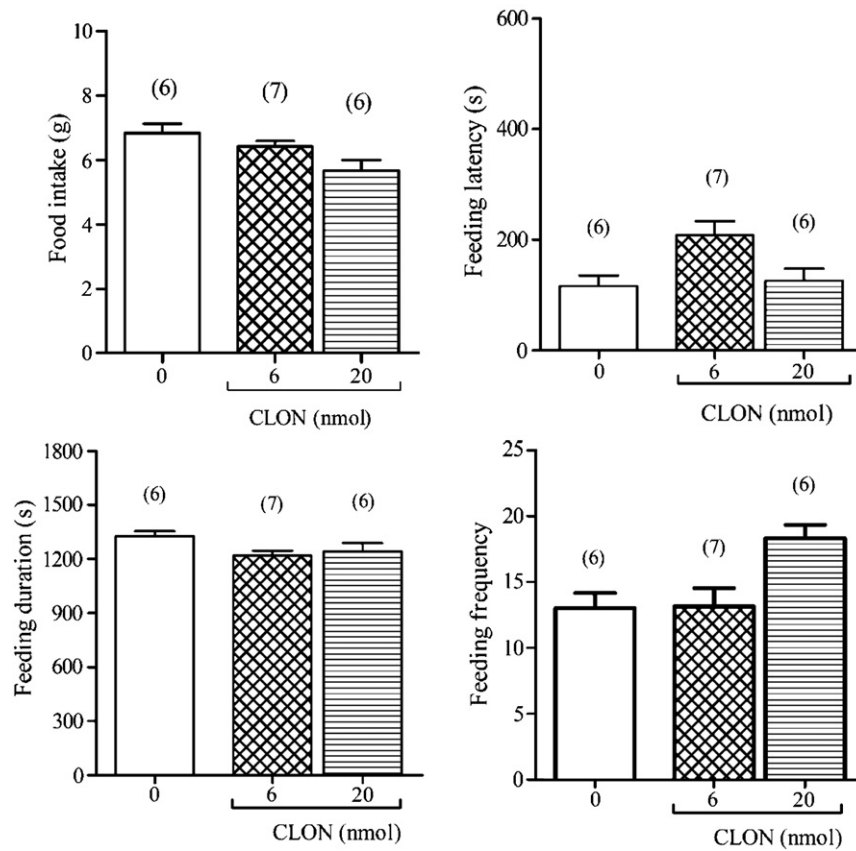


Fig. 5. Changes on food intake, feeding latency, feeding duration and feeding frequency after injection of vehicle (0) or clonidine (CLON, 6 nmol and 20 nmol) into the median raphe nucleus (MnR) of overnight fast. Values are mean \pm S.E.M. Numbers in parentheses indicate the number of animals per group.

controlling water intake. The present results exclude the participation of α -adrenergic receptors within the MnR in the control of water intake since treatment with either α 1- or α 2-adrenergic receptor agonists in the MnR failed to affect drinking behaviour. Furthermore, the adrenergic control of drinking and feeding behaviours seems to include separate circuits in the MnR since α 1-adrenergic receptor activation in this nucleus increased food intake but was not associated with changes in drinking behaviour.

In conclusion, the present results reinforce the inhibitory functional role of α 1-adrenergic receptors within the MnR on feeding

behaviour. This inhibitory influence tonically restrains food intake under free-feeding conditions since MnR α 1-adrenergic receptor blockade increased food intake in rats with free access to lab chow [47]; the intensity of this inhibitory mechanism seemed to decline as food availability decreased.

Financial disclosure

Anderson Savaris Ribas, Rafael Appel Flores, Aparecida Marcelino de Nazareth, Moacir Serralvo Faria, Mariana Graciela Terenzi, José Marino-

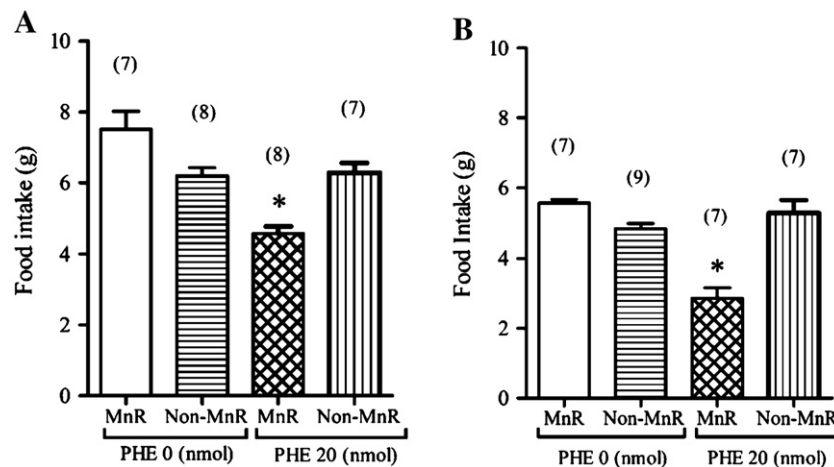


Fig. 6. Changes on food intake after injection of vehicle (0) or phenylephrine (PHE, 20 nmol) into the median raphe nucleus (MnR) and surrounding area (non-MnR) of food restricted rats (15 g chow/day during 7 days) (A) and overnight fast rats (B). Values are mean \pm S.E.M. * p < 0.05 as compared to vehicle treatment in both areas and the PHE treatment in the non-MnR (two-way ANOVA followed by Duncan's test for multiple comparisons). Numbers in parentheses indicate the number of animals per group.

Neto, Marta Aparecida Paschoalini report no biomedical financial interests or other potential conflicts of interest.

Acknowledgements

This study was financed by CNPq proc (134770/2009-9).

References

- Wirtshafter D. The control of ingestive behavior by the median raphe nucleus. *Appetite* 2001;36:99–105.
- Azmitia EC, Segal M. An autoradiographic analysis of the differential ascending projections of the dorsal and median raphe nuclei in the rat. *J Comp Neurol* 1978;179:641–68.
- Imai H, Steindler DA, Kitai ST. The organization of divergent axonal projections from the midbrain raphe nuclei in the rat. *J Comp Neurol* 1986;243:363–80.
- Jacobs BL, Azmitia EC. Structure and function of the brain serotonin system. *Physiol Rev* 1992;72:165–229.
- Descarries L, Watkins KC, Garcia S, Beaudet A. The serotonin neurons in nucleus raphe dorsalis of adult rat: a light and electron microscope radioautographic study. *J Comp Neurol* 1982;207:239–54.
- Kiss J, Csaki A, Bokor H, Kocsis K, Kocsis B. Possible glutamatergic/aspartatergic projections to the supramammillary nucleus and their origins in the rat studied by selective [³H] D-aspartate labelling and immunocytochemistry. *Neuroscience* 2002;111:671–91.
- Köhler C, Steinbusch H. Identification of serotonin and non-serotonin containing neurons of the mid-brain raphe projecting to the entorhinal area and the hippocampal formation. A combined immunohistochemical and fluorescent retrograde tracing study in the rat brain. *Neuroscience* 1982;7:951–75.
- Li YQ, Li H, Kaneko T, Mizuno N. Morphological features and electrophysiological properties of serotonergic and non-serotonergic projection neurons in the dorsal raphe nucleus. An intracellular recording and labeling study in rat brain slices. *Brain Res* 2001;900:110–8.
- Van Bockstaele EJ, Biswas A, Pickel VM. Topography of serotonin neurons in the dorsal raphe nucleus that send axon collaterals to the rat prefrontal cortex and nucleus accumbens. *Brain Res* 1993;624:188–98.
- Calizo LH, Akanwa A, Ma X, Pan Y, Lemos JC, Craig C, et al. Raphe serotonin neurons are not homogenous: electrophysiological, morphological and neurochemical evidence. *Neuropharmacology* 2001;61:524–43.
- Amilhon B, Lepicard E, Renoir T, Mongeau R, Popa D, Poirel O, et al. VGLUT3 (vesicular glutamate transporter type 3) contribution to the regulation of serotonergic transmission and anxiety. *J Neurosci* 2010;30:2198–210.
- Fremeau Jr RT, Kam K, Qureshi T, Johnson J, Copenhagen DR, Storm-Mathisen J, et al. Vesicular glutamate transporters 1 and 2 target to functionally distinct synaptic release sites. *Science* 2004;304:1815–9.
- Fu W, Le Maître E, Fabre V, Bernard JF, David Xu ZQ, Hokfelt T. Chemical neuroanatomy of the dorsal raphe nucleus and adjacent structures of the mouse brain. *J Comp Neurol* 2010;518:3464–94.
- Ma QP, Bleasdale C. Modulation of brain stem monoamines and gammaaminobutyric acid by NK1 receptors in rats. *Neuroreport* 2002;13:1809–12.
- Allers KA, Sharp T. Neurochemical and anatomical identification of fast and slow firing neurons in the rat dorsal raphe nucleus using juxtacellular labelling methods in vivo. *Neuroscience* 2003;122:193–204.
- Commons KG, Connolly KR, Valentino RJ. A neurochemically distinct dorsal raphe-limbic circuit with a potential role in affective disorders. *Neuropharmacology* 2003;28:206–15.
- Day HE, Greenwood BN, Hammack SE, Watkins LR, Fleshner M, Maier SF, et al. Differential expression of 5HT-1A, alpha 1b adrenergic, CRFR1, and CRF-R2 receptor mRNA in serotonergic, gamma-aminobutyric acidergic, and catecholaminergic cells of the rat dorsal raphe nucleus. *J Comp Neurol* 2004;474:364–78.
- Cardin JA, Carlen M, Meletis K, Knoblich U, Zhang F, Deisseroth K, et al. Targeted optogenetic stimulation and recording of neurons in vivo using cell-type-specific expression of Channelrhodopsin-2. *Nat Protoc* 2010;5:247–54.
- Varga V, Losonczy A, Zemelmann BV, Borhegyi Z, Nyiri G, Domonkos A, et al. Fast synaptic subcortical control of hippocampal circuits. *Science* 2009;326:449–53.
- Hopwood SE, Stamford JA. Noradrenergic modulation of serotonin release in rat dorsal and median raphe nuclei via α -1 and α -2 adrenoceptors. *Neuropharmacology* 2001;423(41):433–42.
- Adell A, Celada P, Abellán MT, Artigas F. Origin and functional role of the extracellular serotonin in the midbrain raphe nuclei. *Brain Res Rev* 2002;39:154–80.
- Cryan JF, Page ME, Lucki I. Noradrenergic lesions differentially alter the antidepressant-like effects of reboxetine in a modified forced swim test. *Eur J Pharmacol* 2002;436:197–205.
- Kia HK, Miquel MC, Brisorgueil MJ, Daval G, Riad M, El Mestikawy S, et al. Immunocytochemical localization of serotonin_{1A} receptors in the rat central nervous system. *J Comp Neurol* 1996;365:289–305.
- Trojniar W, Staszewska M. Bilateral lesions of the pedunculopontine tegmental nucleus affect feeding induced by electrical stimulation of the ventral tegmental area. *Acta Neurobiol Exp* 1995;55:201–6.
- Currie PJ, Coscina DV. Diurnal variations in the feeding response to 8-OHDPAT injected into the dorsal or median raphe. *Neuro Report* 1993;4:1105–7.
- Bendotti C, Samanin R. 8-Hydroxy-2-(di-n-propylamino) tetralin (8-OHDPAT) elicits eating in free-feeding rats by acting on central serotonin neurons. *Eur J Pharmacol* 1986;121:147–50.
- Currie PJ, Fletcher PJ, Coscina DV. Administration of 8-OHDPAT into the midbrain raphe nuclei: effects on medial hypothalamic NE-induced feeding. *Am J Physiol* 1994;266:1645–51.
- Fletcher PJ. Dopamine receptor blockade in nucleus accumbens or caudate nucleus differentially affects feeding induced by 8-OH-DPAT injected into dorsal or median raphe. *Brain Res* 1991;552:181–9.
- Funk D, Li Z, Fletcher J, Le AD. Effects of injections of 8-hydroxy-2-(di-n-propylamino) tetralin or muscimol in the median raphe nucleus on c-fos mRNA in the rat brain. *Neuroscience* 2005;131:475–9.
- Adell A, Artigas F. Regulation of the release of 5-hydroxytryptamine in the median raphe nucleus of the rat by catecholaminergic afferents. *Eur J Neurosci* 1999;11:2305–11.
- Talley EM, Rosin DL, Lee A, Guyenet PG, Lynch KR. Distribution of alpha 2 adrenergic receptor-like immunoreactivity in the rat central nervous system. *J Comp Neurol* 1996;372:111–34.
- Nnerstall JR, Fernandez I, Orensanz LM. The alpha-adrenergic receptor: radiohistochemical analysis of functional characteristics and biochemical differences. *Pharmacol Biochem Behav* 1985;22:859–74.
- Costall B, Naylor RJ, Marsden CD, Pycocck CJ. Serotonergic modulation of the dopamine response from the nucleus accumbens. *J Pharm Pharmacol* 1976;28:523–6.
- Ennis M, Aston-Jones G. Two physiologically distinct populations of neurons in the ventrolateral medulla innervate the locus coeruleus. *Brain Res* 1987;425:275–82.
- Freitas RL, Ferreira CM, Ribeiro SJ, Carvalho AD, Elias-Filho DH, Garcia-Cairasco N, et al. Intrinsic neural circuits between dorsal midbrain neurons that control fear-induced responses and seizure activity and nuclei of the pain inhibitory system elaborating postictal antinociceptive process: a functional neuroanatomical and neuropharmacological study. *Exp Neurol* 2005;191:225–42.
- Morgane PJ, Galler JR, Mokler DJ. A review of systems and networks of the limbic forebrain/limbic midbrain. *Prog Neurobiol* 1995;75:143–60.
- Vertes RP, Fortin WJ, Crane AM. Projections of the median raphe nucleus in the rat. *J Comp Neurol* 1999;407:555–82.
- Maidel S, Lucinda AM, Aquino VW, Faria MS, Paschoalini MA. The adrenaline microinjection into the median raphe nucleus induced hypophagic effect in rats submitted to food restriction regimen. *Neurosci Letters* 2007;422:123–7.
- Dos Santos RLD, Mansur SS, Steffens SM, Faria MS, Marino Neto J, Paschoalini MA. Food intake increased after injection of adrenaline into the median raphe nucleus of free-feeding rats. *Behav Brain Res* 2009;197:411–6.
- De Vry J, Schreiber R. Effects of selected serotonin 5-HT(1) and 5-HT(2) receptor agonists on feeding behavior: possible mechanisms of action. *Neurosci Biobehav Rev* 2000;24:341–53.
- Dourish CT, Hutson PH, Curzon G. Characteristics of feeding induced by the serotonin agonist 8-hydroxy-2-(di-n-propylamino) tetralin (8-OH-DPAT). *Brain Res Bull* 1985;15:377–84.
- Schwartz MW, Woods SC, Porte Jr D, Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature* 2000;404:661–771.
- Woods SC, Schwartz MW, Baskin DG, Seeley RJ. Food intake and the regulation of body weight. *Annu Rev Psychol* 2000;51:255–77.
- Ebenezer IS, Arkle MJ, Tite RM. 8-Hydroxy-2-(di-n-propylamino)-tetralin inhibits food intake in fasted rats by an action at 5-HT_{1A} receptors. *Methods Find Exp Clin Pharmacol* 2007;29:269–72.
- López-Alonso VE, Mancilla-Díaz JM, Rito-Domingo M, González-Hernández B, Escartin-Pérez RE. The effects of 5-HT_{1A} and 5-HT_{2C} receptor agonists on behavioral satiety sequence in rats. *Neurosci Lett* 2007;416:285–8.
- Mansur SS, Terenzi MG, Marino-Neto J, Faria MS, Paschoalini MA. Changes in food intake and anxiety-like behaviors after clonidine injected into the median raphe nucleus. *Behav Brain Res* 2010;212:71–7.
- Mansur SS, Terenzi MG, Marino-Neto J, Faria MS, Paschoalini MA. Alpha 1 receptor antagonist in the median raphe nucleus evoked hyperphagia in free-feeding rats. *Appetite* 2011;57(2):498–503.
- Mansur SS, Terenzi MG, Marino-Neto J, Faria MS, Paschoalini MA. Phenylephrine into the median raphe nucleus evokes an anxiolytic-like effect in free-feeding rats but does not alter food intake in free feeding rats. *Behav Brain Res* 2011;217:209–14.
- Happe HK, Coulter CL, Gerety ME, Sanders JD, O'Rourke M, Bylund DB, et al. Alpha-2 adrenergic receptor development in rat CNS: Na autoradiographic study. *Neuroscience* 2004;123:167–78.
- Kaye W. Neurobiology of anorexia and bulimia nervosa. *Physiol Behav* 2008;94:121–35.
- Kelley AE, Baldo BA, Pratt WE, Will MJ. Corticostriatal-hypothalamic circuitry 433 and food motivation: integration of energy, action and reward. *Physiol Behav* 2005;434(86):773–95.
- Paxinos G, Watson C. The rat brain in stereotaxic coordinates. 6th ed. New York: Academic Press & Elsevier Inc.; 2007.
- Ottoni EB. Etholog 2.2: a tool for the transcription and timing of behavior observation sessions. *Behav Res Methods Instrum Comput* 2000;32(468):446–9.
- Halford JCG, Wanninayake SCD, Blundell JE. Behavioral satiety sequence (BSS) for the diagnosis of drug action on food intake. *Pharmacol Biochem Behav* 1998;61:159–68.
- Gallagher DW, Aghajanian GK. Effect of antipsychotic drugs on the firing of dorsal raphe cells. I. Role of adrenergic system. *Eur J Pharmacol* 1976;39:341–55.
- Canteras NS, Simerly RB, Swanson LW. Organization of projections from the medial nucleus of the amygdala: a PHAL study in the rat. *J Comp Neurol* 1995;360:213–45.

- [57] Petrovich GD, Risold PY, Swanson LW. Organization of projections from the basomedial nucleus of the amygdala: a PHAL study in the rat. *J Comp Neurol* 1996;374:387–420.
- [58] Simansky KJ. Serotonergic control of the organization of feeding and satiety. *Behav Brain Res* 1996;73:37–42.
- [59] Leibowitz SF, Alexander JT. Hypothalamic serotonin in control of eating behavior, meal size, and body weight. *Biol Psychiatry* 1998;44:851–64.
- [60] Leger L, Wiklund L. Distribution and numbers of indoleamine cell bodies in the rat brainstem determined with Falck-Hillarp fluorescence histochemistry. *Brain Res Bull* 1982;9:245–51.
- [61] Sotelo C, Cholley B, Mestikawy SE, Gozlan H, Hamon M. Direct immunohistochemical evidence of the existence of 5-HT_{1A} autoreceptors on serotonergic neurons in the midbrain raphe nuclei. *Eur J Neurosci* 1990;2:1144–54.
- [62] Klitenick MA, Wirtshafter D. Elicitation of feeding, drinking, and gnawing following microinjections of muscimol into the median raphe nucleus of rats. *Behav Neural Biol* 1989;51:436–41.
- [63] Paris JM, Mitsushio H, Lorens SA. Intra-midbrain raphe injections of the neurokinin-3 agonist senktide inhibit food and water intake in the rat. *Pharmacol Biochem Behav* 1991;38:223–6.
- [64] Wirtshafter D, Krebs JC. Control of food intake by kainate/quisqualate receptors in the median raphe nucleus. *Psychopharmacology* 1990;101:137–41.
- [65] Wirtshafter D, Trifunovic R. Stimulation of ingestive behaviors following injections of excitatory amino acid antagonists into the median raphe nucleus. *Pharmacol Biochem Behav* 1988;30:529–33.
- [66] Blundell JE. Serotonin manipulations and the structure of feeding behaviour. *Appetite* 1986(Suppl. 7):39–56.
- [67] Masoro EJ. Caloric restriction and aging: an update. *Exp Gerontol* 2000;35:299–305.
- [68] Kinzig KP, Hargrave SL, Tao EE. Central and peripheral effects of chronic food and weight restoration in the rat restriction. *Am J Physiol Endocrinol Metab* 2009;296:E282–90.
- [69] Pesic V, Marinkovic P, Janac B, Ignjatovic S, Popic J, Kanazir S, et al. Changes of behavioral parameters during long-term food restriction in middle-aged Wistar rats. *Physiol Behav* 2010;101:672–8.
- [70] Sucajtyś-Szulc E, Goyke E, Korczynska J, Stelmanska E, Rutkowski B, Swierczynski J. Chronic food restriction differentially affects NPY mRNA level in neurons of the hypothalamus and in neurons that innervate liver. *Neurosci Lett* 2008;433:174–7.
- [71] Moran TH. Gastrointestinal signals: satiety. *Encyclopedia of neuroscience* 2009:571–6.
- [72] Chaudhri OB, Field BCT, Bloom SR. Gastrointestinal satiety signals. *International Journal of obesity* 2008;32:S28–31.
- [73] Sharkey KA. Peripheral satiety signals: view from the chair. *International Journal of Obesity* 2009;33:S3–6.
- [74] Schwartz GJ. Integrative capacity of the caudal brainstem in the control of food intake. *Philos Trans R Soc Lond B Biol Sci* 2006;361:1275–80.
- [75] Blundell JE, Latham CJ. Serotonergic influences on food intake: effect of 5-HT on parameters of feeding behaviour in deprived and free-feeding rats. *Pharmacol Biochem Behav* 1979;11:431–7.
- [76] Blundell JE, Latham CJ. Behavioural pharmacology of feeding. In: Silverstone T, editor. *Drugs and appetite*. London: Academic Press; 1980. p. 41–80.