



Review

Immune evasion in cancer: Mechanistic basis and therapeutic strategies



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ARTICLE INFO

Article history:

Available online 25 March 2015

Keywords:

Cancer
Immune evasion
T cells
Therapy

ABSTRACT

Cancer immune evasion is a major stumbling block in designing effective anticancer therapeutic strategies. Although considerable progress has been made in understanding how cancers evade destructive immunity, measures to counteract tumor escape have not kept pace. There are a number of factors that contribute to tumor persistence despite having a normal host immune system. Immune editing is one of the key aspects why tumors evade surveillance causing the tumors to lie dormant in patients for years through "equilibrium" and "senescence" before re-emerging. In addition, tumors exploit several immunological processes such as targeting the regulatory T cell function or their secretions, antigen

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¹ These authors contributed to the cross-validation activity.

<http://dx.doi.org/10.1016/j.semcan.2015.03.004>

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presentation, modifying the production of immune suppressive mediators, tolerance and immune deviation. Besides these, tumor heterogeneity and metastasis also play a critical role in tumor growth. A number of potential targets like promoting Th1, NK cell, $\gamma\delta$ T cell responses, inhibiting Treg functionality, induction of IL-12, use of drugs including phytochemicals have been designed to counter tumor progression with much success. Some natural agents and phytochemicals merit further study. For example, use of certain key polysaccharide components from mushrooms and plants have shown to possess therapeutic impact on tumor-imposed genetic instability, anti-growth signaling, replicative immortality, dysregulated metabolism etc. In this review, we will discuss the advances made toward understanding the basis of cancer immune evasion and summarize the efficacy of various therapeutic measures and targets that have been developed or are being investigated to enhance tumor rejection.

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1. Introduction

Cancer remains one of the leading causes of death globally, with an estimated 12.7 million cases around the world affecting both sexes equally. This number is expected to increase to 21 million by 2030. The immune system interacts intimately with tumors over the entire process of disease development and progression to metastasis. This complex cross talk between immunity and cancer cells can both inhibit and enhance tumor growth and is now classified as a hallmark of cancer [1]. The balance of these actions between and across the hallmarks determines the eventual outcome, which in the case of clinically overt cancer results from evasion of the destructive elements of the immune response by the tumor. Mechanisms resulting in evasion of immune attack include the selection of tumor variants resistant to immune effectors (sometimes designated “immunoediting”) and progressive formation of an immune suppressive environment within the tumor. Although considerable knowledge has been accumulated on how tumors avoid immune destruction, discovering effective cancer therapies still remains a daunting task for the researcher and clinician. In this report, we will briefly present an overview of how tumors evade immune surveillance by focusing on how the immune system reacts to the development of tumors, how certain cancers evade immunity, and what measures can be taken to eradicate cancer. We will address important aspects of tumor and host immune interactions as set out below.

2. Tumors and immunity

The involvement of the host immune system in cancer progression is well established, although greater emphasis has been placed on tumor eradication by immunity than tumor immune potentiation, which may be equally important. These interactions between the immune system and the tumor occur through complex events that usually eventually climax either in successful tumor eradication or immune evasion by the tumor [2].

2.1. Relationship between tumor formation and immune responses

Tumor development and survival is a chaotically governed process involving the interplay between cancer cells, normal stromal cells and host defense mechanisms. Several other factors such as cellular changes due to infection or disease-induced stress may also contribute to tumor growth or tumor suppression. Generally, CD8⁺ cytotoxic T cells (CTL) and CD4⁺ helper T (Th)1 cells curb cancer development via mechanisms commonly involving their production of interferon (IFN)- γ and cytotoxins [3] but other factors such as chronic inflammation may override these effects to promote cancer development [4,5]. For example, the risk of overt hepatocellular carcinoma (HCC) appears to be closely linked to the duration of the Hepatitis B and C viral-induced inflammatory state

[6–9]. Compelling evidence has also documented, both in animal tumor models and in human cancers, that chronic inflammation plays a critical role in the development of colon and pancreatic cancers [6]. Therefore, when beneficial acute responses fail to resolve tumors/cancer, lingering chronic inflammation can lead to promotion of tumor cell growth and angiogenesis [6,10]. In addition, ongoing activity due to autoimmune disease has also been shown to support development of many cancers including lymphoma [6,10–12].

2.2. Tumor progression and immunity

Vital fundamental discoveries made over the last few decades have unequivocally shown that the immune system plays a critical role in maintaining an equilibrium between immune recognition and tumor development with a dual capacity to both promote and suppress tumor growth. These discoveries collectively support the concept of “immunoediting” and help to explain why tumors can sometimes lie dormant in patients for years before re-emerging, and why tumors grow despite the host having a fully functional immune system [13]. During cancer immune editing, the immune system is able to recognize and destroy the most immunologically vulnerable cancer cells because they present tumor antigens, resulting in their elimination [14]. Nonetheless, due to genetic instability, constant tumor cell division can generate with reduced immunogenicity that can evade immune elimination. This state of production of new tumor cell variants balanced by the elimination has been dubbed “equilibrium”, during which the cancer cells continue to divide, accumulating mutational changes by chance or in response to immune-induced inflammation. Thus, a balance between immune control and tumor growth is maintained, giving the appearance of tumor dormancy [15]. However, these processes eventually enable tumors to impair the capacity of the immune system to eradicate them by immune suppressive effects or by loss of target antigen expression. It is at this stage that tumor escape occurs, resulting in overt clinical cancer. Nonetheless, there may also be conditions under which tumor cells are truly dormant, for example by induction of “senescence”. In this case, they would be likely to remain dormant permanently, as replicative senescence is generally believed to be irreversible [16].

2.3. Factors that tumors exploit to avoid immune responses

2.3.1. Regulatory cells

Immune suppression in the tumor microenvironment, mediated by CD4⁺CD25⁺ FoxP3⁺ regulatory T cells (Tregs), or other types of suppressive cells, seems to be a major mechanism of tumor immune escape and can be a crucial hurdle for tumor immunotherapy [17]. A number of studies have shown that tumor-derived Tregs have comparatively higher suppressive activity than naturally occurring Tregs [18,19]. Tregs are drawn into the tumor microenvironment via tumor cell-mediated chemokine production [20,21]. Evidence

also suggests that transforming growth factor (TGF)- β , produced by tumor cells among other cells, aids conversion of CD4⁺ T cells into suppressive Tregs in situ [22]. Thus, elimination of Tregs by anti-CD25 monoclonal antibodies (mAbs) or by other means may promote tumor rejection. Myeloid cells, especially “myeloid-derived suppressor cells” (MDSCs), modulated dendritic cells (DCs) and alternatively-activated M1 and M2 macrophages create an inflammatory microenvironment and can also act as mediators of tumor initiation, angiogenesis, and metastasis [23,24]. Moreover, a vicious cycle may be instigated in that higher levels of inflammatory mediators confer resistance to apoptosis in MDSCs which would otherwise be subject to downregulation by T cells in complex interaction networks [25]. Thus, CD11b⁺Gr1⁺ MDSCs suppress CD8⁺ T cell-mediated antitumor immunity [26], one mechanism for which may be TCR ζ -chain downregulation. MDSCs with this phenotype accumulate in, for example, melanoma lesions in a manner intimately linked to the inflammatory milieu, implying that the tumor inflammatory microenvironment supports MDSC recruitment and immunosuppressive activity. Reduction of chronic inflammatory mediators by pharmacological means can reduce the amounts of MDSC and decrease immunosuppression [27]. CD11b⁺F4/80⁺ macrophages having an M2 phenotype produce high levels of TGF- β , IL-10, and vascular endothelial growth factor (VEGF) and promote tumor growth [28–30]. In addition, a number of tumor-derived factors and gangliosides have been reported to alter DC phenotype. These immature, functionally-impaired DCs have lower levels of CD80, CD86, CD40, and high indoleamine 2,3-dioxygenase (IDO) expression that also contributes to suppression of T cell immunity [31].

2.3.2. Defective antigen presentation

It is well established that another fundamental mechanism by which tumors evade immune surveillance is by down-modulating antigen processing machinery affecting the major histocompatibility complex (MHC) I pathway, proteasome subunits latent membrane protein (LMP)2 and LMP7, transporter associated with antigen processing (TAP) protein, and tapasin [32–37]. Thus, expression of tumor antigen is downregulated, which can lead to enhanced tumor incidence and metastasis because cytotoxic T lymphocyte (CTL) can no longer recognize target antigens on the tumor cells [38].

2.3.3. Immune suppressive mediators

As alluded to above, tumors can evade immune surveillance by crippling CTL functionality via production of several immune suppressive cytokines, either by the cancer cells or by the non-cancerous cells present in the tumor microenvironment, especially including immune cells and epithelial cells. TGF- β is a chief mediator of this activity [39]. In addition, tumor necrosis factor (TNF)- α , IL-1, IL-6, colony stimulating factor (CSF)-1, IL-8, IL-10, and type I IFNs can also significantly contribute to cancer growth [40–44]. In addition to immune suppressive cytokines, other factors such as VEGF produced by tumors, inhibit the differentiation of progenitors into DCs [45], thus affecting efficient uptake and antigen presentation. VEGF and IL-10 and TGF- β are also known to inhibit maturation of DCs. DCs retaining the immature phenotype are tolerogenic as they do not present antigen in the proper context with appropriate costimulation to T cells [46]. Other factors such as tumor gangliosides and receptor-binding cancer-associated surface antigen (RCAS1) also contribute to tumor progression [47,48]. Additional studies revealed that expression of RCAS1 is associated with apoptosis of tumor infiltrating lymphocytes (TILs) [49,50]. Similarly, ganglioside antigens, on cell surface or shed from cells surface, are known to suppress CTL and DC function [51]. Immunosuppressive enzymes such as IDO, arginase, and inhibitor of nuclear factor kappa-B kinase (IKK)2 may also contribute significantly to tumor

progression [52–55] via direct actions on tumor cell proliferation or through induction of T cell tolerance/suppression [56–58].

2.3.4. Tolerance and immune deviation

Most tumor cells fail to express costimulatory molecules and can thereby induce anergy or tolerance in T cells by engaging the T cell receptor in the absence of costimulation [59]. Tumors are also known to evade immune attack by shifting the balance from Th1 to Th2 (immune deviation) in a TGF- β - and IL-10-dependent manner [60]. In addition, tumor expression of inhibitory molecules like programmed cell death (PD)-L1/B7H1 has been shown to cause deletion or anergy on tumor reactive cells [61,62]. There is also evidence that down regulation of death receptors prevents death ligand-mediated killing of tumor cells by both CTLs and natural killer (NK) cells [63]. Slavin-Chiorini et al. [64] have demonstrated that CTL studies in conjunction with antibody blocking studies enhanced antitumor effector activity mainly through CD54. There are reports to show that p53 tumor suppressive gene is implicated in the regulation of tumor cell death by CTLs [65]. Thus, factors promoting tolerance and immune deviation are significant contributors to cancer immune evasion.

2.3.5. Apoptosis

A number of studies have shown that cancer cells delete tumor-specific CTLs through apoptosis [66,67]. The different influences governing tumor growth and immune evasion strategies are briefly outlined in Fig. 1.

3. Tumor heterogeneity and immune responses

Cells of the immune system can inhibit tumor growth and progression through the recognition and rejection of malignant cells containing initiation mutations. Though tumors originate from a single transformed cell, due to genetic instability, they commonly become genetically heterogeneous, exhibiting multiple phenotypes both in terms of morphology and physiology. They also display striking heterogeneity in cell surface molecule expression, proliferative and angiogenic potential [68], which is believed to stem from morphological and epigenetic plasticity. Thus, the tumor cells express a wide variety of antigens including some which may be tumor-specific or tumor-associated, differentiation antigens, and lectin-binding sites. These antigens are unevenly distributed on tumor subpopulations and induce different immune responses to the same determinant [69]. This tumor antigenic heterogeneity has a significant effect on genotype, gene expression, cellular morphology, metabolic activity, motility, and behavior such as proliferation rate, antigen expression, drug response and metabolic potential [70–74]. Such heterogeneity has important implications for diagnosis, treatment efficacy, and the identification of potential targets [70,75]. The key aspects of tumor heterogeneity and its subsequent effects on tumor growth are briefly outlined in Fig. 2.

4. Immune system and cancer metastasis

It is fascinating how cancer cells migrate throughout the body from their original location to establish themselves at a new location [76]. How this exodus of tumor cells occurs is only now beginning to be understood. In general, cancer cells detach from the primary tumor and travel through the surrounding tissues and basement membranes, avoid immune destruction and metastasize to distant organs [77,78]. This metastatic process is what is responsible for most cancer deaths [79–82]. Although there are several underlying mechanisms of tumor dissemination and colonization [83], the “progression model” which suggests that a series of mutational events occur either in a subpopulation of primary

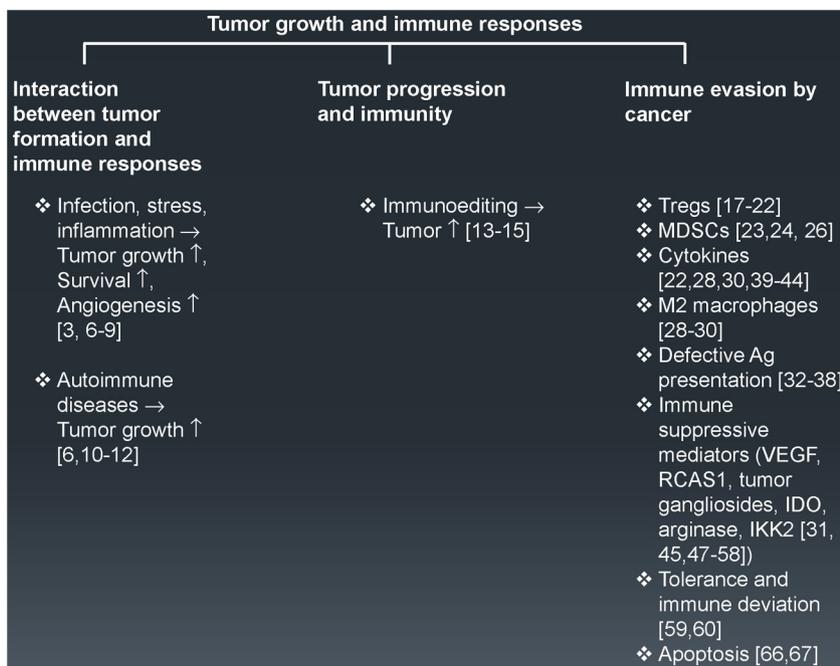


Fig. 1. Tumor growth and immune response. An overview of the different key factors governing tumor formation, progression, and immune evasion. The numbers in parentheses represent the relevant references in support of the statements made.

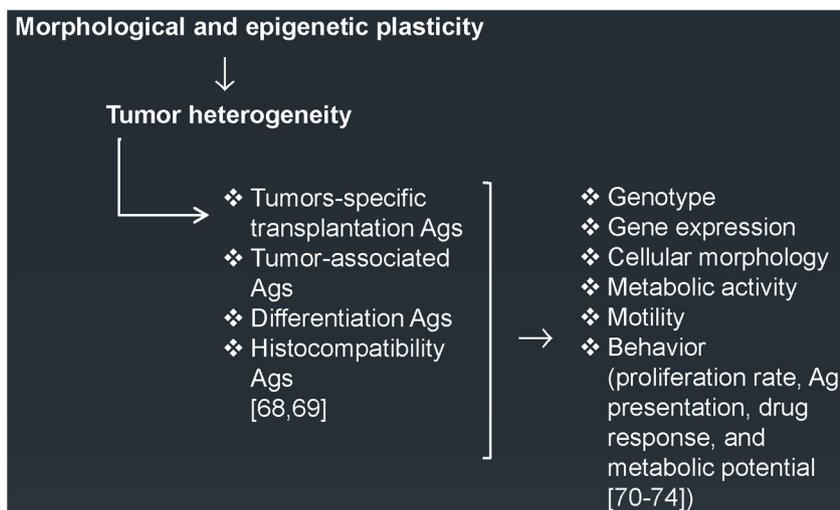


Fig. 2. Tumor heterogeneity and immune response. Shown here are important sequential events leading to tumor heterogeneity and its consequences for the various aspects of the immune response. The numbers in parentheses are the relevant literature cited.

tumor or in disseminated cells, resulting in a small fraction of the cells that acquire full metastatic potential is a well-accepted theory [84]. This view has been corroborated by a number of investigations [85–88]. Among other factors, once again, it is the TGF- β secreted by the cancer cells that makes a major contribution to tumor metastasis [89]. In addition, the vasculature also plays an important role in metastatic seeding at different sites. It has been shown that tumor vasculature hyperstimulated by VEGF often has reduced pericyte coverage and that looser association of such pericytes with the endothelium facilitates metastatic dissemination [90]. In addition, hypoxia in and around tumor vessels also contributes to metastatic dissemination of cancer cells in a hypoxia inducible factor (HIF)-, VEGF-, and inducible nitric oxide synthase (iNOS)-dependent manner [91, 92]. Notably, hypoxia promotes the formation of pre-metastatic niches through the production of lysyl oxidase [93]. Hypoxia further conditions pre-metastatic niches by

recruiting MDSCs and suppressing NK cell functions [94]. In support of a role for immunosurveillance in controlling tumor metastasis, a recent study revealed that high expression of Irf7-regulated genes in primary human breast tumors is associated with prolonged bone metastasis-free survival [95]. A brief overview of the events promoting tumor metastasis and the involvement of immune responses is provided in Fig. 3.

5. Conventional cancer therapy and the immune system

Although a variety of agents have been screened for their anti-tumor effects and some have been approved for the treatment of cancer patients, chemotherapy, radiation therapy, and surgery remain the mainstays of standard cancer therapeutic strategies. A downside to these therapies is their ability to cause a transient immune suppression which in turn increases the risk of infection

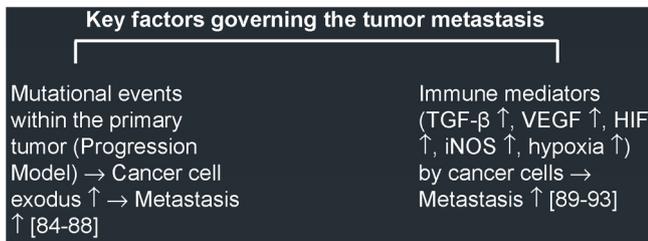


Fig. 3. Immune system and tumor metastasis. Depicted here are the key sequential events based on the “Progression Model” leading to cancer cells exodus from the primary location and subsequent establishment at a distant location and the possible role of various immune modulators that aid this process. The numbers in parentheses are the relevant literature cited.

and is also likely to decrease the immune system’s ability to inhibit further development of cancer. For example, standard chemotherapy decreases the host’s native immune competent cells including T cell populations. However, this transient loss of immune activity has been shown to return 2–3 weeks after chemotherapy [96]. In addition, patients are at risk for viral, fungal, and parasitic infections, and when chemotherapy continues long-term, these patients may permanently lose their cell-mediated immune function [97]. Nevertheless, recent evidence suggests that some chemotherapeutic drugs rely on the induction of anticancer immune responses for therapeutic activity by inducing a type of tumor cell death that is “immunogenic” [98]. The immune-stimulating property of some chemotherapeutic drugs, such as anthracyclines and oxaliplatin, requires preapoptotic translocation of calreticulin (CRT) on the tumor cell surface, post-apoptotic release of the chromatin-binding protein high mobility group B1 (HMGB1), and extracellular release of ATP. Interestingly, phosphohydrolysis of extracellular ATP by ecto-nucleotidases (i.e. CD39 and CD73) acts as a counterbalancing process to chemotherapy-induced immunogenic cell death [99]. Other chemotherapies appear to alter the phenotype of surviving tumor cells making them better targets for immune cells [100,101]. Radiation therapy has also been shown to impact cell-mediated immunity. On the one hand, radiotherapy can suppress antitumor immunity, presumably due to the high radiosensitivity of lymphocytes [102]. There are also reports to suggest that high doses of total lymphocyte irradiation increase T suppressor cell activity and loss of the ability to recognize autoantigens [103]. On the other hand, low doses of radiation result in the generation of reactive oxygen species (ROS) leading to the activation of intracellular signaling pathways that induce T cell proliferation and differentiation [104,105]. Radiation has been shown to alter the phenotype of cells resulting in increased expression of death receptors [106], chemokines [107], adhesion molecules such as intercellular adhesion molecule (ICAM)-1 and MHC-I [108], and costimulatory molecules [109,110] on tumor cells. Moreover, tumor cells surviving radiation have also been shown to be more sensitive to cytolysis by T cells [108,111]. Radiation has also been shown to result in the increased expression of proinflammatory cytokines such as TNF- α and IL-1 β that activate antigen presenting cells (APCs) [112,113]. Radiotherapy can thus trigger significant antitumor immune responses, related to the well-known abscopal effect, that is, the regression of metastases upon irradiation of the primary tumor, despite the metastasis being outside of the radiation field [114,115]. It is indeed generally accepted that radiotherapy depends to some degree on the activation of antitumor immune responses for its efficacy [116].

Finally, trauma due to surgical resection of tumors has profound effects on the immune system because of increased production of proinflammatory cytokines and other immune modulators like IL-6, C-reactive protein (CRP), TNF- α , IL-1 β etc. [117,118]. Also, decreased delayed-type hypersensitivity (DTH) reactions,

due to surgery, pose a risk for infection [119]. To overcome surgery-mediated transient immune suppression, the introduction of laparoscopic methods may reduce such suppression and thus decrease tumor growth [120]. Conversely, surgery has also been shown to induce danger/damage that enhances antitumor efficacy and reduces metastasis [121]. There is evidence that tumor growth control can actually potentiate rather than curb metastasis, again illustrating the general finding that very similar pathways can have either inhibitory or facilitatory activity on tumor growth. A case in point is that chemotherapy, radiotherapy, and biological/targeted therapies can promote tumor metastasis via the so-called tumor bed effect [122,123]. Currently, both primary and metastatic cancers are treated by similar approaches where radiation is often the mainstay choice of therapy [124]. Surgery is rarely performed on metastatic lesions. Thus, these standard anticancer therapies, although they can be effective alone, will have enhanced therapeutic efficacy when combined with agents that boost the weakened immune system, if we are able to learn how to avoid potential tumor growth stimulatory effects.

6. Strategies for cancer immunotherapy

Tumor cells have developed multiple mechanisms for evading immune surveillance. Current treatments for cancer include chemotherapy, radiation therapy, immunotherapy, targeted therapy, and surgery which all have limitations and detrimental side effects [125]. Recent investigations have identified several classes of anticancer agents that are targeted, efficient, and have less adverse side effects. An increasing number of clinical trials are currently underway to stimulate the immune system to combat cancer. Important among these include vaccination with peptides [126], vaccination with DCs [127], vaccination with viral-based vectors [128,129] and immunotherapy with autoreactive effector cells [130]. Interestingly, there are also studies to show that administration of bacteria can increase tumor immunogenicity [131]. For example, treatment with *Clostridium novyi-NT* is shown to attract many inflammatory cells such as neutrophils, monocytes, and lymphocytes that can kill tumor cells [132]. Especially important will be the extended use of immunomodulatory antibodies which have recently yielded such dramatic effects in highly refractory tumors (see below). Many clinical trials of all these approaches, and especially combinations thereof, are currently ongoing and hold great promise.

6.1. Cellular targets

In addition to the obvious targets, the tumor cells themselves, some of the several regulatory cells including regulatory B cells or their products implicated in tumor escape are currently being targeted to promote tumor rejection. For example, IDO is an immunoregulatory enzyme which suppresses T-cell immunity but can be targeted in the tumor microenvironment by IDO-reactive CD8⁺ T cells. It was shown that IDO-specific T cells could enhance tumor immunity by eliminating IDO⁺ suppressive cells and changing the regulatory microenvironment [133].

As mentioned above, important among suppressive cells are Tregs, which are powerful inhibitors of antitumor immunity and an impediment to successful immunotherapy [22]. In support of this, inhibition of Tregs by monoclonal antibodies has been shown to decrease tumor development [134,135]. In addition, other regulatory cell populations such as MDSCs which accumulate in spleen, blood, tumors, and bone marrow of tumor-bearing mice and cancer patients [136,137] have been considered as important targets for therapeutic intervention [138]. MDSCs secrete IL-10 and TGF- β and enhance angiogenesis and metastasis by inducing Treg production

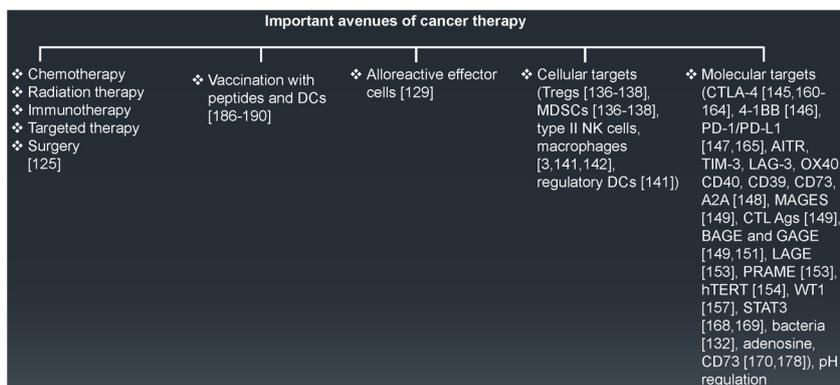


Fig. 4. Cancer therapy. A brief overview of the various available therapeutic options for cancer. A few of these have entered clinical trials some of which have been approved for treatment of specific types of cancers. The numbers in parentheses are the literature cited.

[23,139]. Increasing evidence suggests that the M2 macrophages promote tumor growth and metastasis, and strategies to target these cells are also being developed [140]. Type II NK cells are also known to contribute to tumor development via their secretion of characteristic cytokines. About 60% of murine NK cells express Ly49 and CD94/NKGA inhibitory receptors, the blockade of which augments antitumor activity [3,141,142]. In addition, regulatory DCs (expressing CD25, PD-1, PD-L1, IL-10, TGF- β , kynurenine, IDO, cyclooxygenase (Cox)-2, and arginase (Arg)-1) play a significant role in tumor development [143] and therapies directed against these cells have also been investigated [144].

6.2. Molecular targets

In addition to cellular targets, several molecular targets including cytotoxic T-lymphocyte-associated protein 4 (CTLA)-4 [145], 4-1BB [146], PD1/PD-L1 [147], and activation-inducible TNFR (AITR), T cell immunoglobulin mucin (TIM)-3, Lymphocyte-activation gene (LAG)-3, OX40, CD40, CD39, CD73, A2A [148] and cancer antigens of different types, such as melanoma-associated antigen (MAGE) family members and NY-ESO-1, human telomerase reverse transcriptase (hTERT) and Wilm's tumor (WT)1 have been considered as important antitumor targets [149]. In melanoma, MAGE, B melanoma antigen (BAGE), and G antigen (GAGE) family antigens have been targeted for therapeutic vaccination [150,151]. The L antigen family member (LAGE)-1 gene closely related to NY-ESO-1 may also be an appropriate target [152]. The preferentially expressed antigen in melanoma (PRAME) is also a melanoma-associated antigen recognized by CTL [153]. Human telomerase activity and hTERT expression are detected in a majority (>90%) of human cancer cells [154]. To increase potential efficacy, hTERT promoters have been utilized for cancer gene therapy [155,156]. Wilms' tumor gene WT1 is expressed in several different cancers and illustrates the general principle that tumor escape from immunity as a result of downregulation of target antigen expression is unlikely to occur when the gene product has an essential role in tumorigenesis [157]. A number of studies suggest that the WT1 protein is a promising target for cancer immunotherapy [158,159].

Targeting cell surface molecules other than tumor antigen targets for antibody-based therapeutic intervention of cancer is becoming an important available option for the clinician. Of these, so far only anti-CTLA-4 (ipilimumab) has been approved for clinical use in the USA, Canada, United Kingdom, and European Union [160,161], but PD1 and PD-L1-specific antibodies will surely be licensed very soon. Ipilimumab is currently in phase III clinical trials for the treatment of prostate cancer [162] and for cancers of the lung [163] and kidney [164] as well as melanoma. In one recent trial, administration of the anti-PD-1 antibody

nivolumab showed unprecedented therapeutic objective responses in 18–28% of patients with advanced non-small-cell lung carcinoma, melanoma, and renal cell carcinoma [165]. While CTLA-4 and PD-1/PD-L1 blocking Abs have shown efficacy by blocking inhibitory signals to responding T cells, agonist Ab to OX40 and 4-1BB propel T-cell immunity by sending stimulatory signals. Several clinical trials are underway investigating their therapeutic properties [166]. Targeting Tregs by anti-CD25 antibodies showed inhibition of neuroblastoma tumors in mice [167]. There are also data demonstrating that activation of the signal transducer and activator of transcription (STAT)3 signaling pathway supports tumor development by inducing accumulations of MDSCs and inhibition of DC differentiation [168]; hence its inactivation leads to inhibition of cancer development by a DC- and Treg-dependent mechanism [169].

Targeting immunosuppression by soluble mediators is another attractive approach for cancer immunotherapy. A plethora of immunosuppressive factors has been associated with tumorigenesis, including TGF- β , IDO, arginase, prostaglandin-E2 (PGE2) and extracellular adenosine. Recent studies have shown that extracellular adenosine, essentially produced by the ecto-nucleotidase CD73, plays an important role in tumor development and metastasis [170,171,99,172–174]. These findings are corroborated by studies using mice deficient in CD73 or the high affinity A2A adenosine receptor [173–176]. These animals exhibit increased CTL-mediated antitumor immunity [177]. Inhibition of pH regulatory molecules and certain heat shock proteins limit cancer cell-mediated immune suppression. Targeting these molecules could simultaneously counteract the metastatic potential of cancer cells and restore antitumor immune surveillance. The above-mentioned cancer therapeutic targets and their beneficial effects are briefly outlined in Fig. 4.

6.3. Vaccination therapy (peptide, DNA, and DC)

Several studies demonstrated the efficacy of therapeutic viral vaccines [178]. Peptide vaccines derived from tumor-associated antigens (TAA) may significantly contribute to immune enhancement or tumor regression. Many TAAs have been identified and molecularly characterized. However, so far only a limited number of TAA peptides, mostly recognized by CD8⁺ T cells in melanoma patients, have been clinically tested. In some clinical trials, partial or complete tumor regression was observed in 10–30% of patients [179]. Peptides such as melan-A/MART-1_{27–35} and gp100, which readily activate specific T cells in vitro [180] and in vivo [181,182], show limited immunogenicity when used as vaccines for cancer patients [183,184]. Alternatively, DNA cancer vaccines may also represent an effective approach [185]. Such vaccines, although

Table 1

Cross-validation of potential targets that may enhance anticancer immune responses to other cancer hallmarks.

Potential targets for immune-modulation	(Promote/enhance) Th1 responses via increase number of NK cells	(Promote/enhance) $\gamma\delta$ T cell activities	(Promote/activate) macrophages	(Inhibit) Treg lymphocytes	(Promote/enhance) NK cell activity	(Promote/induce) IL-12
Other cancer hallmarks						
Genomic Instability Sustained	0	0	0	0	0	0
Proliferative Signaling	0	0	–	0	0	0
Tumor-Promoting Inflammation	–	–	+	+/-	+	+
Evasion of Anti-growth Signaling	[207,208]	[209]	[210]	[211–213]	[214,215]	[216,217]
Resistance to Apoptosis	0	+	0	+	+	0
Replicative Immortality	[224]	[218]	[221]	[219]	[220]	[223]
Dysregulated Metabolism	0	0	0	0	0	0
Angiogenesis	+	–	+/-	+	+	+
Tissue Invasion and Metastasis	[225–228]	[229–232]	[233]	[234]	[225]	[235]
Tumor Microenvironment	[236]	[237]	[238]	[239]	[240]	[241]
	[242]	[243]	[244–246]	[247]	[248]	[249]

The symbols presented above represent as follows: +, complementary; –, contrary; +/-, controversial; 0, no known relationship.

having many variants, utilize the same basic principle involving the isolation of DNA from cancer cells and subsequent transfer, most commonly via the intramuscular route, into tumor-bearing individuals. It has been shown that the administration of DNA vaccines via the intramuscular route effectively primes both the adaptive as well as innate arms of the immune system [186]. While naked DNA is quite sturdy and stable at different temperatures, and retains immune activating abilities, plasmid DNA vaccines are less immunogenic [187]. Refinements to the existing DNA vaccination strategies are showing promising results. Among these, the use of an electrical pulse, commonly called electroporation, electroporation or electrotransfer [188] is currently used in preclinical protocols and has been shown to have strong immune activating abilities [189]. Recent therapeutic studies involving DNA vaccines have shown promise, for example, for the treatment of glioma. Incorporation of cancer cell DNA into healthy immune competent cells and subsequent transfer into tumor-bearing mice showed decreased tumor burden and increased survival of both spontaneous as well as established tumors. Further analysis revealed that DNA vaccine-mediated antitumor activity in the above case involved over-production of IFN- γ and participation of T and NK/lymphokine activated killer (LAK) cells [190,191]. Adoptive transfer of peptide-pulsed DC [192] is also an option. In all cases, it takes a long time to develop such therapies and the newest results now being published suggest that peptide vaccinations with selected multi-peptide vaccines, combined with immunomodulatory agents, may indeed achieve impressive results. Thus, a phase II multi-center granulocyte macrophage colony stimulating factor (GM-CSF)-adjuvanted multi-peptide vaccine for refractory late-stage renal cancer patients has yielded unprecedented 3-year survival benefits especially in those patients able to respond to more than one peptide, provided they had received a pulse of low-dose cyclophosphamide prior to vaccination. It was proposed that the cyclophosphamide reduced the Tregs in the patients, for which some evidence was presented [193,194]. The United States FDA has approved the use of sipuleucel-T, a cellular product made of blood APCs cultured with a fusion protein of prostatic acid phosphatase (PAP) and GM-CSF [195]. Efficacy studies revealed a

4-month extended median survival in patients with prostate cancer [196].

6.4. Cross validation

A cross-validation team conducted a peer-reviewed literature review of the targets and approaches listed in Tables 1 and 2, and these evidences of cross-hallmark activity are referenced accordingly. This process led to the creation of two unique matrices, whereby a series of candidate compounds and molecular/cellular targets were identified for having immune system evasion mechanistic relevance. The complete mapping of these candidate targets and actions was screened for known complementary, contrary or combinations of actions across all cancer hallmarks described in Hanahan and Weinberg [1]. For example, inhibiting or stimulating an immune evasion target may or may not have been examined in other hallmark mechanism. Each potential target-hallmark or approach-hallmark interaction was considered to have either a pro- or anti-chemotherapeutic effect. There were also mixed indications or many instances where no known relationship existed. In summary, the findings gathered in this effort varied considerably by each hallmark. These tables provide information that can serve as a starting point for future basic and translational research on phytochemical combinations for immune evasion targets and for chemotherapeutic applications.

6.5. Phytochemicals

Besides these conventional immunotherapeutic approaches, several phytochemicals have been shown to facilitate tumor regression. Prominent among these are isothiocyanate, curcumin, genistein, epigallocatechin gallate, lycopene, resveratrol, and glucosinolates. Some have entered clinical trials and are beginning to yield encouraging results [197]. There are other natural, plant-derived or nutrient substances, including flavonoids, omega-3 fatty acids, zinc, and vitamin C, that are purported to strengthen the immune system [198–201], yet their roles as nutrients to resolve inflammation or assist in suppressing tumorigenesis are not clear

Table 2
Cross-validation of phytochemicals that may enhance anticancer immune responses to other cancer hallmarks.

Approach	<i>Ganoderma lucidum</i> (polysaccharide fraction)	<i>Trametes versicolor</i> (protein-bound polysaccharide-k)	<i>Astragalus membranaceus</i> (polysaccharide fraction)	<i>Lentinus edodes</i> (polysaccharide L-II, lentinan)	Astaxan-thin	Polyphenol-resveratrol analog HS-1793
Other cancer hallmarks						
Genomic Instability	0	0	0	0	0	0
Sustained Proliferative Signaling	+	0	0	+	+	0
Tumor-Promoting Inflammation	+	0	+	+	+	0
Evasion of Anti-growth Signaling	+	+	+	+	+	0
Resistance to Apoptosis	+	0	+	+	+	+
Replicative Immortality	+	0	0	0	0	0
Dysregulated Metabolism	0	0	0	0	0	0
Angiogenesis	+	0	–	+	0	+
Tissue Invasion and Metastasis	+	0	+	+	+	+
Tumor Microenvironment	+	+	+	+	+	+

The symbols presented above represent as follows: +, complementary; –, contrary; +/-, controversial; 0, no known relationship.

from human studies. Too often, these alternative or complementary agents are not evaluated with standard sets of clinical outcomes that are needed to advance our understanding of how nutritional components and phytochemicals may enhance tumoricidal immunity or inhibit tumor immune evasion mechanisms described above. While some dietary supplements have been shown to enhance the ability of NK cells to identify and destroy dysfunctional cells, such as infected or cancerous cells [202,203], these studies have not comprehensively assessed increased T cell production of cytokines such as IFN and TNF, or reduced secretion of immune suppressive factors from tumors. The emerging evidence for dietary supplement doses that far exceed physiological nutrient exposures suggests that some bioactive food components can even be hazardous [204], and are now largely discouraged for consumption during cancer treatment [205]. Table 2 summarizes potential targets and approaches that may enhance anticancer immune responses.

6.6. Adoptive T cell therapy

Autoreactive T cells are potentially tolerant to self-tissues, due to diverse mechanisms in the periphery [291]. Adoptive T cell therapy involves the isolation and expansion of autologous T cells specific for tumor antigen and their subsequent re-infusion into the patient. Tumor-reactive T cells such as tumor-infiltrating lymphocytes (TIL) combined with IL-2 showed potentially interesting results already in the 1980s, but objective response rate was low in metastatic melanoma patients [292,293]. In 2002, Rosenberg and colleagues [294,295] introduced a lymphodepletion regimen before administering adoptive T cell therapy, resulting in elimination of the immune-suppressive cells, increase of key cytokines for T cells such as IL-7 and IL-15, and creation of a space for T-cell proliferation. When lymphopenia is induced, remaining peripheral T cells initiate homeostatic proliferation to reconstitute the lost T cells, and the tolerant autoreactive CD8⁺ T cells acquire an opportunity to proliferate and become functional [296,297]. This may be one mechanism by which self-tumor Ag-specific T cells are increased in cancer patients after chemo- or radio-therapy [298,299]. This lymphodepletion treatment markedly improved the clinical efficacy of adoptive cell therapy using TILs, with an objective response in ~70% of melanoma patients and complete durable regressions were

found in ~50% [300]. Rosenberg et al. [301] have demonstrated objective cancer regression in patients with metastatic melanoma. Though good clinical outcome has been observed by Rosenberg et al. [301], generating T cells for adoptive T cell therapy is a cumbersome process. There have been many efforts to develop a practical protocol to produce autologous self-tumor Ag-specific T cells, but most of them are still complicated and time-consuming because self-tumor Ag-reactive T cells exist as a minor population. Recently, however, an efficient method has been developed to produce tumor-specific CD8⁺ T cells from ~50 ml of peripheral blood mononuclear cells based upon the unique property of 4-1BB (CD137) to be selectively expressed on antigen-engaged T cells [302]. Clinical trials with various solid tumors are underway to test the safety and efficacy of the CTLs thus generated. To overcome major hurdles in the preparation of autologous self-tumor Ag-specific T cells, gene-modified T cells like TCR or chimeric Ag receptor (CAR)-modified T cells were developed [303]. Currently, these gene-modified T cells are being tested for safety and efficacy in the clinic and clinical results will tell us whether adoptive T cell therapy could provide a new opportunity for cancer patients who failed to respond to standard therapies. However, the many mechanisms of tumor escape discussed above (tumor suppression, downregulation of target antigens etc.) need to be considered and counteracted in combination with these modalities.

7. Conclusions

Here we wish to emphasize that immunotherapeutic approaches may advance via the inclusion of holistic or integrative therapy of cancer. Especially, we want to emphasize that dual approaches which seek to (1) eliminate immune suppressing factors, and (2) enhance tumor-killing activities will be necessary to achieve successful cancer therapy. In view of the immune suppressive factors present in the tumor microenvironment from the very earliest stages of tumor formation, nontoxic agents that control or eliminate the immunosuppressive factors can be used for therapy of cancer or also utilized as cancer control and chemopreventive agents. A tumor-killing agent requires us to aim at cross-clonal common targets, which overcome the intra- and inter-tumoral heterogeneity.

An in-depth understanding of how tumors evade immune surveillance will help develop effective therapeutic strategies that can be used for the benefit of cancer patients.

Disclosure

BSK: patents for methods regarding anti-CD137 and adoptive CTL therapeutics. RLW: patent for use of IGFBP-3 as anticancer therapy; FDA murine work for Arrium Corporation for Omega 3 fatty acid anti-adhesion product; Consultant for Ethicon Endosurgery and Olympus Corporation regarding surgical staplers and advanced endoscopic polypectomy methods. TL: worked with Medtronic in developing a passive immunotherapeutic strategy for treatment of Alzheimer's disease. WKD: patent for methods and compositions regarding Th-1 dendritic cells; Consultant of Gerson Lehrman Group; Medical advisor of Texans for stem cell research.

Acknowledgments

This work was supported by grants from the National Cancer Center, Korea (NCC-1310430-2) and the National Research Foundation (NRF-2005-0093837). W. Nicol Keith & Alan Bilslund were supported by the University of Glasgow, Beatson Oncology Centre Fund, and Cancer Research UK (<http://www.cancerresearchuk.org>) grant C301/A14762.

References

- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144:646–74.
- Becker JC, Andersen MH, Schrama D, Thor Straten P. Immune-suppressive properties of the tumor microenvironment. *Cancer Immunol Immunother* 2013;62:1137–48.
- Zamarron BF, Chen W. Dual roles of immune cells and their factors in cancer development and progression. *Int J Biol Sci* 2011;7:651–8.
- Blackwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2002;357:539–45.
- Rakoff-Nahoum S. Why cancer and inflammation. *Yale J Biol Med* 2007;79:123–30.
- Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010;140:883–99.
- Karin M. Nuclear factor-kappaB in cancer development and progression. *Nature* 2006;441:431–6.
- Karin M, Lawrence T, Nizet V. Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. *Cell* 2006;124:823–35.
- Park EJ, Lee JH, Yu GY, He G, Ali SR, Holzer RG, et al. Dietary and genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell* 2010;140:197–208.
- de Martel C, Franceschi S. Infections and cancer: established associations and new hypotheses. *Crit Rev Oncol Hematol* 2009;70:183–94.
- Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–7.
- Coussens LM, Werb Z. Inflammatory cells and cancer: think different! *J Exp Med* 2001;193:F23–6.
- Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoeediting: from immunosurveillance to tumor escape. *Nat Immunol* 2002;3:991–8.
- Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoeediting. *Annu Rev Immunol* 2004;22:329–60.
- Sawann JB, Smyth MJ. Immune surveillance of tumors. *J Clin Investig* 2007;117:1137–46.
- Shay JW, Roninson IB. Hallmarks of senescence in carcinogenesis and cancer therapy. *Oncogene* 2004;23:2919–33.
- Jacobs JF, Nierkens S, Figdor CG, de Vries IJ, Adema GJ. Regulatory T cells in melanoma: the final hurdle towards effective immunotherapy? *Lancet Oncol* 2012;13:e32–42.
- Yokokawa J, Cereda V, Remondo C, Gulley JL, Arlen PM, Schlom J, et al. Enhanced functionality of CD4+CD25(high)FoxP3+ regulatory T cells in the peripheral blood of patients with prostate cancer. *Clin Cancer Res* 2008;14:1032–40.
- Gasparoto TH, de Souza Malaspina TS, Benevides L, de Melo Jr EJ, Costa MR, Damante JH, et al. Patients with oral squamous cell carcinoma are characterized by increased frequency of suppressive regulatory T cells in the blood and tumor microenvironment. *Cancer Immunol Immunother* 2010;59:819–28.
- Curjel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004;10:942–9.
- Lee I, Wang L, Wells AD, Dorf ME, Ozkaynak E, Hancock WW. Recruitment of Foxp3+ T regulatory cells mediating allograft tolerance depends on the CCR4 chemokine receptor. *J Exp Med* 2005;201:1037–44.
- Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol* 2006;6:295–307.
- Murdoch C, Muthana M, Coffelt SB, Lewis CE. The role of myeloid cells in the promotion of tumour angiogenesis. *Nat Rev Cancer* 2008;8:618–31.
- Shozaei F, Zhong C, Wu X, Yu L, Ferrara N. Role of myeloid cells in tumor angiogenesis and growth. *Trends Cell Biol* 2008;18:372–8.
- Ostrand-Rosenberg S, Sinha P, Chornoguz O, Ecker C. Regulating the suppressors: apoptosis and inflammation govern the survival of tumor-induced myeloid-derived suppressor cells (MDSC). *Cancer Immunol Immunother* 2012;61:1319–25.
- Seung LP, Rowley DA, Dubey P, Schreiber H. Synergy between T-cell immunity and inhibition of paracrine stimulation causes tumor rejection. *Proc Natl Acad Sci USA* 1995;92:6254–8.
- Umansky V, Sevko A. Overcoming immunosuppression in the melanoma microenvironment induced by chronic inflammation. *Cancer Immunol Immunother* 2012;61:275–82.
- Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol* 2010;22:231–7.
- van Kempen LC, Ruiters DJ, van Muijen GN, Coussens LM. The tumor microenvironment: a critical determinant of neoplastic evolution. *Eur J Cell Biol* 2003;82:539–48.
- Sica A, Larghi P, Mancino A, Rubino L, Porta C, Totaro MG, et al. Macrophage polarization in tumour progression. *Semin Cancer Biol* 2008;18:349–55.
- Munn DH, Sharma MD, Lee JR, Jhaveri KG, Johnson TS, Keskin DB, et al. Potential regulatory function of human dendritic cells expressing indoleamine 2,3-dioxygenase. *Science* 2002;297:1867–70.
- Garrido F, Ruiz-Cabello F, Cabrera T, Pérez-Villar JJ, López-Botet M, Duggan-Keen M, et al. Implications for immunosurveillance of altered HLA class I phenotypes in human tumours. *Immunol Today* 1997;18:89–95.
- Hicklin DJ, Marincola FM, Ferrone S. HLA class I antigen downregulation in human cancers: T-cell immunotherapy revives an old story. *Mol Med Today* 1999;5:178–86.
- Johnsen AK, Templeton DJ, Sy M, Harding CV. Deficiency of transporter for antigen presentation (TAP) in tumor cells allows evasion of immune surveillance and increases tumorigenesis. *J Immunol* 1999;163:4224–31.
- Restifo NP, Esquivel F, Kawakami Y, Yewdell JW, Mulé JJ, Rosenberg SA, et al. Identification of human cancers deficient in antigen processing. *J Exp Med* 1993;177:265–72.
- Rotem-Yehudai R, Groettrup M, Soza A, Kloetzel PM, Ehrlich R. LMP-associated proteolytic activities and TAP-dependent peptide transport for class I MHC molecules are suppressed in cell lines transformed by the highly oncogenic adenovirus 12. *J Exp Med* 1996;183:499–514.
- Seliger B, Mauerer MJ, Ferrone S. TAP off-tumors on. *Immunol Today* 1997;18:292–9.
- Mauerer MJ, Gollin SM, Martin D, Swaney W, Bryant J, Castelli C, et al. Tumor escape from immune recognition: lethal recurrent melanoma in a patient associated with downregulation of the peptide transporter protein TAP-1 and loss of expression of the immunodominant MART-1/Melan-A antigen. *J Clin Investig* 1996;98:1633–41.
- Pasche B. Role of transforming growth factor beta in cancer. *J Cell Physiol* 2001;186:153–68.
- Lind MH, Rozell B, Wallin RP, van Hogerlinden M, Ljunggren HG, Toftgård R, et al. Tumor necrosis factor receptor 1-mediated signaling is required for skin cancer development induced by NF-kappaB inhibition. *Proc Natl Acad Sci USA* 2004;101:4972–7.
- Lin EY, Gouon-Evans V, Nguyen AV, Pollard JW. The macrophage growth factor CSF-1 in mammary gland development and tumor progression. *J Mammary Gland Biol Neoplasia* 2002;7:147–62.
- Klein SC, Jücker M, Abts H, Tesch H. IL6 and IL6 receptor expression in Burkitt's lymphoma and lymphoblastoid cell lines: promotion of IL6 receptor expression by EBV. *Hematol Oncol* 1995;13:121–30.
- Matsuda M, Salazar F, Petersson M, Masucci G, Hansson J, Pisa P, et al. Interleukin 10 pretreatment protects target cells from tumor- and allo-specific cytotoxic T cells and downregulates HLA class I expression. *J Exp Med* 1994;180:2371–6.
- Sotomayer EM, Fu YX, Lopez-Cepero M, Herbert L, Jimenez JJ, Albarracín C, et al. Role of tumor-derived cytokines on the immune system of mice bearing a mammary adenocarcinoma. II. Downregulation of macrophage-mediated cytotoxicity by tumor-derived granulocyte-macrophage colony-stimulating factor. *J Immunol* 1991;147:2816–23.
- Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, et al. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. *Nat Med* 1996;2:1096–103.
- Gabrilovich D. Mechanisms and functional significance of tumor induced dendritic-cell defects. *Nat Rev Immunol* 2004;4:941–52.
- McKallip R, Li R, Ladisch S. Tumor gangliosides inhibit the tumor-specific immune response. *J Immunol* 1999;163:3718–26.
- Nakashima M, Sonoda K, Watanabe T. Inhibition of cell growth and induction of apoptotic cell death by the human tumor-associated antigen RCAS1. *Nat Med* 1999;5:938–42.
- Nakabayashi H, Nakashima M, Hara M, Toyonaga S, Yamada SM, Park KC, et al. Clinic-pathological significance of RCAS1 expression in gliomas: a potential mechanism of tumor immune escape. *Cancer Lett* 2007;246:182–9.

- [50] Sonoda K, Miyamoto S, Nakashima M, Wake N. The biological role of unique molecule RCAS1: a bioactive marker that induces connective tissue remodeling and lymphocyte apoptosis. *Front Biosci* 2008;13:1106–16.
- [51] Birkle S, Zeng G, Gao L, Yu RK, Aubry J. Role of tumor-associated gangliosides in cancer progression. *Biochimie* 2003;85:455–63.
- [52] Muller AJ, Prendergast GC. Marrying immunotherapy with chemotherapy: why say IDO? *Cancer Res* 2005;65:8065–8.
- [53] Mills CD, Shearer J, Evans R, Caldwell MD. Macrophage arginine metabolism and the inhibition or stimulation of cancer. *J Immunol* 1992;149:2709–14.
- [54] Boutard V, Havouis R, Fouqueray B, Philippe C, Moulinoux JP, Baud L. Transforming growth factor-beta stimulates arginase activity in macrophages. Implications for the regulation of macrophage cytotoxicity. *J Immunol* 1995;155:2077–84.
- [55] Xia Y, Yeddula N, Leblanc M, Ke E, Zhang Y, Oldfield E, et al. Reduced cell proliferation by IKK2 depletion in a mouse lung-cancer model. *Nat Cell Biol* 2012, <http://dx.doi.org/10.1038/ncb2428>.
- [56] Prendergast GC. Immune escape as a fundamental trait of cancer: focus on IDO. *Oncogene* 2008;27:3889–900.
- [57] Munder M. Arginase: an emerging key player in the mammalian immune system. *Br J Pharmacol* 2009;158:638–351.
- [58] Kim HJ, Hawke N, Baldwin AS. NF- κ B and IKK as therapeutic targets in cancer. *Cell Death Differ* 2006;13:738–47.
- [59] Staveley-O'Carroll K, Sotomayor E, Montgomery J, Borrello I, Hwang L, Fein S, et al. Induction of antigen-specific T cell anergy: an early event in the course of tumor progression. *Proc Natl Acad Sci USA* 1998;95:1179–83.
- [60] Maeda H, Shiraishi A. TGF-beta contributes to the shift toward Th2-type responses through direct and IL-10-mediated pathways in tumor-bearing mice. *J Immunol* 1996;156:73–8.
- [61] Driessens G, Kline J, Gajewski TF. Costimulatory and inhibitory receptors in anti-tumor immunity. *Immunol Rev* 2009;229:126–44.
- [62] Topalian SL, Drake CG, Pardoll DM. Targeting PD-1/B7-H1 (PD-L1) pathway to activate antitumor immunity. *Curr Opin Immunol* 2012;24:207–12.
- [63] French LE, Tschopp J. Defective death receptor signaling as a cause of tumor immune escape. *Semin Cancer Biol* 2002;12:51–5.
- [64] Slavin-Chiorini DC, Catalfamo M, Kudo-Saito C, Hodge JW, Scholm J, Sabzevari H. Amplification of the lytic potential of effector/memory CD8+ cells by vector-based enhancement of ICAM-1(CD54) in target cells: implications for intratumoral vaccine therapy. *Cancer Gene Ther* 2004;11:665–80.
- [65] Chouaib S, meslin F, Thiery J, Mami-Chouaib F. Tumor resistance to specific lysis: a major hurdle for successful immunotherapy of cancer. *Clin Immunol* 2009;130:34–40.
- [66] Bogen B. Peripheral T cell tolerance as a tumor escape mechanism: deletion of CD4+ T cells specific for a monoclonal immunoglobulin idiotype secreted by a plasmacytoma. *Eur J Immunol* 1996;26:2671–9.
- [67] Lauritzsen GF, Hofgaard PO, Schenck K, Bogen B. Clonal deletion of thymocytes as a tumor escape mechanism. *Int J Cancer* 1998;78:216–22.
- [68] Marusky A, Polyak K. Tumor heterogeneity: causes and consequences. *Biochim Biophys Acta* 2010;1805:105–17.
- [69] Miller FR. Intratumor immunologic heterogeneity. *Cancer Metastasis Rev* 1982;1:319–34.
- [70] Campbell LL, Polyak K. Breast tumor heterogeneity: cancer stem cells or clonal evolution? *Cell Cycle* 2007;6:2322–8.
- [71] Dick JE. Stem cell concepts renew cancer research. *Blood* 2008;112:4793–807.
- [72] Fidler IJ, Hart IR. Biological diversity in metastatic neoplasms: origins and implications. *Science* 1982;217:998–1003.
- [73] Heppner GH. Tumor heterogeneity. *Cancer Res* 1984;44:2259–65.
- [74] Nicolson GL. Generation of phenotypic diversity and progression in metastatic tumor cells. *Cancer Metastasis Rev* 1984;3:25–42.
- [75] Merlo LM, Pepper JW, Reid BJ, Maley CC. Cancer as an evolutionary and ecological process. *Nat Rev Cancer* 2006;6:924–35.
- [76] Ardiani A, Gameiro SR, Palena C, Hamilton DH, Kwilas A, King TH, et al. Vaccine-mediated immunotherapy directed against a transcription factor driving the metastatic process. *Cancer Res* 2014;74:1945–57.
- [77] Butler TP, Gullino PM. Quantitation of cell shedding into efferent blood of mammary adenocarcinoma. *Cancer Res* 1975;35:512–6.
- [78] Tarin D, Price JE, Kettlewell MG, Souter RG, Vass AC, Crossley B, et al. Mechanisms of human tumor metastasis studied in patients with peritoneovenous shunts. *Cancer Res* 1984;44:3584–92.
- [79] Chambers AF, Naumov GN, Varghese HJ, Nadkarni KV, MacDonald IC, Groom AC. Critical steps in hematogenous metastasis: an overview. *Surg Oncol Clin N Am* 2001;10:243–55.
- [80] Fidler IJ. Critical determinants of cancer metastasis: rationale for therapy. *Cancer Chemother Pharmacol* 1999;43:53–10.
- [81] Folkman J. The role of angiogenesis in tumor growth. *Semin Cancer Biol* 1992;3:65–71.
- [82] Woodhouse EC, Chuaqui RF, Liotta LA. General mechanisms of metastasis. *Cancer* 1997;80:1529–37.
- [83] Hunter KW, Crawford NPS, Alsarraj J. Mechanisms of metastasis. *Breast Cancer Res* 2008;10, <http://dx.doi.org/10.1186/bcr1988>.
- [84] Nowell PC. The clonal evolution of tumor cell populations. *Science* 1976;194:23–8.
- [85] Fidler IJ, Kripke ML. Metastasis results from preexisting variant cells within a malignant tumor. *Science* 1977;197:893–5.
- [86] Kang Y, Siegel PM, Shu W, Drobnjak M, Kakonen SM, Cordon-Cardo C, et al. A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell* 2003;3:537–49.
- [87] Minn AJ, Gupta GP, Siegel PM, Bos PD, Shu W, Giri DD, et al. Genes that mediate breast cancer metastasis to lung. *Nature* 2005;436:518–24.
- [88] Minn AJ, Kang Y, Serganova I, Gupta GP, Giri DD, Doubrovin M, et al. Distinct organ-specific metastatic potential of individual breast cancer cells and primary tumors. *J Clin Invest* 2005;115:44–55.
- [89] Mundy GR. Mechanisms of bone metastasis. *Cancer* 1997;80:1546–56.
- [90] Cooke VG, LeBleu VS, Keskin D, Khan Z, O'Connell JT, Teng Y, et al. Pericyte depletion results in hypoxia-associated epithelial-to-mesenchymal transition and metastasis mediated by met signaling pathway. *Cancer Cell* 2012;21:66–81.
- [91] Branco-Price C, Zhang N, Schnelle M, Evans C, Katschinski DM, Liao D, et al. Endothelial cell HIF-1 α and HIF-2 α differentially regulate metastatic success. *Cancer Cell* 2012;21:52–65.
- [92] Kashiwagi S, Izumi Y, Gohongi T, Demou ZN, Xu L, Huang PL, et al. NO mediates mural cell recruitment and vessel morphogenesis in murine melanomas and tissue-engineered blood vessels. *J Clin Invest* 2005;115:1816–27.
- [93] Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, et al. Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. *Cancer Cell* 2009;15:35–44.
- [94] Sceneay J, Chow MT, Chen A, Halse HM, Wong CS, Andrews DM, et al. Primary tumor hypoxia recruits CD11b+Ly6Cmed/Ly6G+ immune suppressor cells and compromises NK cell cytotoxicity in the premetastatic niche. *Cancer Res* 2012;72:3906–11.
- [95] Bidwell BN, Slaney CY, Withana NP, Forster S, Cao Y, Loi S, et al. Silencing of Irf7 pathways in breast cancer cells promotes bone metastasis through immune escape. *Nat Med* 2012;18(August(8)).
- [96] Mackall CL. T-cell immunodeficiency following cytotoxic antineoplastic therapy: a review. *Stem Cells* 2000;18:10–8.
- [97] Noonan FP, Halliday FJ, Wall DR, Clunie GJA. Cell-mediated immunity and serum blocking factors in cancer patients during chemotherapy and immunotherapy. *Cancer Res* 1977;37:2473–80.
- [98] Galluzzi L, Senovilla L, Zitvogel L, Kroemer G. The secret ally: immunostimulation by anticancer drugs. *Nat Rev Drug Discov* 2012;11:215–33.
- [99] Loi S, Pommey S, Haibe-Kains B, Beavis PA, Darcy PK, Smyth MJ, et al. CD73 promotes anthracycline resistance and poor prognosis in triple negative breast cancer. *Proc Natl Acad Sci USA* 2013;110:11091–6.
- [100] Garnett CT, Scholm J, Hodge JW. Combination of docetaxel and recombinant vaccine enhances T-cell responses and anti-tumor activity; effects of docetaxel on immune enhancement. *Clin Cancer Res* 2008;14:3536–44.
- [101] Hodge JW, Garnett CT, Farsaci B, Palena C, Tsang KY, Ferrone S, et al. Chemotherapy-induced immunogenic modulation of tumor cells enhances killing by cytotoxic T lymphocytes and is distinct from immunogenic cell death. *Int J Cancer* 2013;133:624–36.
- [102] Uzawa A, Suzuki G, Kakata Y, Akashi M, Ohyama Hakanuma A. Radiosensitivity of CD45RO+ memory and CD45RO+ naive T cells in culture. *Radiat Res* 1994;137:25–33.
- [103] Ferguson RM, Sutherland DE, Kim T, Simmons RL, Najarian JS. The *in vitro* assessment of the immunosuppressive effect of fractionated total lymphoid irradiation in renal allotransplantation. *Transplant Proc* 1981;13:1673–5.
- [104] Lander HM, Tauras JM, Ogiste JS, Hori O, Moss RA, Schmidt AM. Activation of the receptor for advanced glycation end products triggers a p21(ras)-dependent mitogen-activated protein kinase pathway regulated by oxidant stress. *J Biol Chem* 1997;272:17810–4.
- [105] Kasid U, Suy S, Dent P, Whiteside TL, Sturgill TW. Activation of Raf by ionizing radiation. *Nature* 1996;382:813–6.
- [106] Ifeadi V, Garnett-Benson C. Sub-lethal irradiation of colorectal tumor cells imparts enhanced and sustained susceptibility to multiple death receptor signaling pathways. *PLoS ONE* 2012;7:e31762.
- [107] Matsumura S, Demaria S. Up-regulation of the proinflammatory chemokine CXCL16 is a common response of tumors cells to ionization radiation. *Radiat Res* 2010;173:418–25.
- [108] Garnett CT, Palena C, Chakraborty M, Tsang K, Scholm J, Hodge JW. Sub-lethal irradiation of human tumor cells modulates phenotype resulting in enhanced killing by cytotoxic T lymphocytes. *Cancer Res* 2004;64:7985–94.
- [109] Kumari A, Cacan E, Greer SF, Garnett-Benson C. Turning T cells on: epigenetically enhanced effector T-cell activity following tumor cells irradiation. *J Immunother Cancer* 2013;1:17, <http://dx.doi.org/10.1186/2052-1426-1-17>.
- [110] Bernstein M, Garnett CT, Zhang H, Velcich A, Wattenberg M, Gameiro S, et al. radiation-induced modulation of costimulatory and coinhibitory T-cell signaling molecules on human prostate carcinoma cells promotes productive anti-tumor immune interactions. *Cancer Biother Radiopharm* 2014;29:153–61.
- [111] Chakraborty M, Abrams SI, Coleman CN, Camphausen K, Scholm J, Hodge JW. External beam radiation of tumors alters phenotype of tumor cells to render them susceptible to vaccine-mediated T-cell killing. *Cancer Res* 2004;64:4328–37.
- [112] McBride WH, Chiang CS, Olson JL, Wang CC, Hong JH, Pajonik F, et al. A sense of danger from radiation. *Radiat Res* 2004;162:1–19.
- [113] Ishihara H, Tanaka I, Nemoto K, Tsunooka K, Cheeramakara C, Yoshida K. Immediate-early, transient induction of the interleukin-1 beta gene in mouse spleen macrophages by ionizing radiation. *J Radiat Res* 1995;36:112–24.
- [114] Ehlers G, Fridman M. Abscopal effect of radiation in papillary adenocarcinoma. *Br J Radiol* 1973;46:220–2.
- [115] Demaria S, Ng B, Devitt ML, Babb JS, Kawashima L, Liebes L, et al. Inozation radiation of distinct unrelated tumors (abscopal effect) is immune mediated. *Int J Radiat Oncol Biol Phys* 2004;58:862–70.

- [116] Durante M, Reppingen N, Held KD. Immunologically augmented cancer treatment using modern radiotherapy. *Trends Mol Med* 2013;(13):00096–8, pii: S1471–4914.
- [117] Sylla P, Kirman I, Whelan RL. Immunological advantages of advanced laparoscopy. *Surg Clin N Am* 2005;85:1–18.
- [118] Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 1999;340:448–54.
- [119] Lennard TW, Shenton BK, Borzotta A, Donnelly PK, White M, Gerrie LM, et al. The influence of surgical operations on components of the human immune system. *Br J Surg* 1985;72:771–6.
- [120] Whelan RL, Franklin M, Holubar SD, Donahue J, Fowler R, Munger C. Postoperative cell mediated immune response is better preserved after laparoscopic vs open colorectal resection in humans. *Surg Endosc* 2003;17:972–8.
- [121] Zhang P, Cote AL, de vriess VC, Usherwood EJ, Turk MJ. Induction of post-surgical immunity and T-cell memory by a poorly immunogenic tumor. *Cancer Res* 2007;67:6468–76.
- [122] Belanich M, Randall T, Pastor MA, Kibitel JT, Alas LG, Dolan ME, et al. Intracellular localization and intercellular heterogeneity of the human DNA repair protein O(6)-methylguanine-DNA methyltransferase. *Cancer Chemother Pharmacol* 1996;37:547–55.
- [123] Hotta T, Saito Y, Fujita H, Mikami T, Kurisu K, Kiya K, et al. O6-alkylguanine-DNA alkyltransferase activity of human malignant glioma and its clinical implications. *J Neurooncol* 1994;21:135–40.
- [124] Sleeman J, Steeg PS. Cancer metastasis as a therapeutic target. *Eur J Cancer* 2010;46:1177–80.
- [125] Melief CJ, Toes RE, Medema JP, van der Burg SH, Ossendorp F, Offringa R. Strategies for immunotherapy of cancer. *Adv Immunol* 2002;75:235–82.
- [126] Itok K, Yamada A. Personalized peptide vaccines: a new therapeutic modality for cancer. *Cancer Sci* 2006;97:970–6.
- [127] Gilboa E. DC-based cancer vaccines. *J Clin Invest* 2007;117:1195–203.
- [128] Hodge JW, Higgins J, Schlom J. Harnessing the unique local immunostimulatory properties of modified vaccinia Ankara (MVA) virus to generate superior tumor-specific immune responses and anti-tumor activity in a diversified prime and boost vaccine regimen. *Vaccine* 2009;27:4475–82.
- [129] Scholm J, Hodge JW, Palena C, Tsang KY, Jochems C, greiner JW. Therapeutic cancer vaccines. *Adv Cancer Res* 2014;121:67–124.
- [130] June CH. Adoptive T cell therapy for cancer in the clinic. *J Clin Invest* 2007;117:1466–76.
- [131] Patyar S, Joshi R, Byrard DS, Prakash A, Medhi B, Das BK. Bacteria in cancer therapy: a novel experimental strategy. *J Biomed Sci* 2010;17, doi: 1186/1423-0127-17-21.
- [132] Xu J, Liu XS, Zhou SF, Wei MQ. Combination of immunotherapy with anaerobic bacteria for immunogene therapy of solid tumors. *Gene Ther Mol Biol* 2009;13:36–52.
- [133] Andersen MH. The specific targeting of immune regulation: T-cell responses against indoleamine 2,3-dioxygenase. *Cancer Immunol Immunother* 2012;61:1289–97.
- [134] Byrne WL, Mills KH, Lederer JA, O'Sullivan GC. Targeting regulatory T cells in cancer. *Cancer Res* 2011;71:6915–20.
- [135] Rasku MA, Clem AL, Telang S, Taft B, Gettings K, Gragg H, et al. Transient T cell depletion causes regression of melanoma metastases. *J Transl Med* 2008;6:12, <http://dx.doi.org/10.1186/1479-5876-6-12>.
- [136] Marigo I, Dolcetti L, Serafini P, Zanovello P, Bronte V. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunol Rev* 2008;222:162–79.
- [137] Ostrand-Rosenberg S. Myeloid-derived suppressor cells: more mechanisms for inhibiting antitumor immunity. *Cancer Immunol Immunother* 2010;59:1593–600.
- [138] Kim YS, Kim YJ, Lee JM, Kim EK, Park YJ, Choe SK, et al. Functional changes in myeloid-derived suppressor cells (MDSCs) during tumor growth: FKBP51 contributes to the regulation of the immunosuppressive function of MDSCs. *J Immunol* 2012;188:4226–34.
- [139] Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009;9:162–74.
- [140] Melancon MP, Lu W, Huang Q, Thapa P, Zhou D, Ng C, et al. Targeted imaging of tumor-associated M2 macrophages using a macromolecular contrast agent PG-Gd-NIR813. *Biomaterials* 2010;31:6567–73.
- [141] Barten R, Torkar M, Haude A, Trowsdale J, Wilson MJ. Divergent and convergent evolution of NK-cell receptors. *Trends Immunol* 2001;22:52–7.
- [142] George TC, Ortaldo JR, Lemieux S, Kumar V, Bennett M. Tolerance and alloreactivity of the Ly49D subset of murine NK cells. *J Immunol* 1999;163:1859–67.
- [143] Schmidt SV, Nino-Castro AC, Schultze JL. Regulatory dendritic cells: there is more than just immune activation. *Front Immunol* 2012;3:274, <http://dx.doi.org/10.3389/fimmu.2012.00274>.
- [144] Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. *Nat Rev Cancer* 2012;12:265–77.
- [145] Weber J. Review: anti-CTLA-4 antibody ipilimumab: case studies of clinical response and immune-related adverse events. *Oncologist* 2007;12:864–72.
- [146] Vinay DS, Kwon BS. Immunotherapy of cancer with 4-1BB. *Mol Cancer Ther* 2012;11:1062–70.
- [147] Topalian SL, Drake CG, Pardoll DM. Targeting the PD-1/B7-H1(PD-L1) pathway to activate antitumor immunity. *Curr Opin Immunol* 2012;24:207–12.
- [148] Kim YS, Jung HW, Choi J, Kwon BS, Ham SY, Jung AK, et al. Expression of AITR and AITR ligand in breast cancer patients. *Oncol Rep* 2007;18:1189–94.
- [149] Shashidharamurthy R, Bozeman EN, Patel J, Kaur R, Meganathan J, Selvaraj P. Immunotherapeutic strategies for cancer treatment: a novel protein transfer approach for cancer vaccine development. *Med Res Rev* 2012;32:1197–219.
- [150] Rosenberg SA. A new era for cancer immunotherapy based on the genes that encode cancer antigens. *Immunity* 1999;10:281–7.
- [151] Van den Eynde BJ, van der Bruggen P. T cell defined tumor antigens. *Curr Opin Immunol* 1997;9:684–93.
- [152] Chen YT, Scanlan MJ, Sahin U, Türeci O, Gure AO, Tsang S, et al. A testicular antigen aberrantly expressed in human cancers detected by autologous antibody screening. *Proc Natl Acad Sci USA* 1997;94:1914–8.
- [153] Ikeda H, Lethé B, Lehmann F, van Baren N, Baurain JF, de Smet C, et al. Characterization of an antigen that is recognized on a melanoma showing partial HLA loss by CTL expressing an NK inhibitory receptor. *Immunity* 1997;6:199–208.
- [154] Liu L, Lai S, Andrews LG, Tollefsbol TO. Genetic and epigenetic modulation of telomerase activity in development and disease. *Gene* 2004;340:1–10.
- [155] Gu J, Kagawa S, Takakura M, Kyo S, Inoue M, Roth JA, et al. Tumor-specific transgene expression from the human telomerase reverse transcriptase promoter enables targeting of the therapeutic effects of the Bax gene to cancers. *Cancer Res* 2000;60:5359–64.
- [156] Koga S, Hirohata S, Kondo Y, Komata T, Takakura M, Inoue M, et al. A novel telomerase-specific gene therapy: gene transfer of caspase-8 utilizing the human telomerase catalytic subunit gene promoter. *Hum Gene Ther* 2000;11:1397–406.
- [157] Oji Y, Ogawa H, Tamaki H, Oka Y, Tsuboi A, Kim EH, et al. Expression of the Wilms' tumor gene WT1 in solid tumors and its involvement in tumor cell growth. *Jpn J Cancer Res* 1999;90:194–204.
- [158] Oka Y, Elisseeva OA, Tsuboi A, Ogawa H, Tamaki H, Li H, et al. Human cytotoxic T-lymphocyte responses specific for peptides of the wild-type Wilms' tumor gene (WT1) product. *Immunogenetics* 2000;51:99–107.
- [159] Oka Y, Tsuboi A, Kawakami M, Elisseeva OA, Nakajima H, Udaka K, et al. Development of WT1 peptide cancer vaccine against hematopoietic malignancies and solid cancers. *Curr Med Chem* 2006;13:2345–52.
- [160] Wolchok J. How recent advances in immune therapy are changing the standard care for patients with metastatic melanoma. *Ann Oncol* 2012;23, viii5–viii21.
- [161] Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010;363(8):711–23.
- [162] Drake CG, Antonarakis ES. Current status of immunological approaches for the treatment of prostate cancer. *Curr Opin Urol* 2012;22:197–202.
- [163] Lynch TJ, Bondarenko I, Luft A, Serwatowski P, Barlesi F, Chacko R, et al. Ipilimumab in combination with Paclitaxel and Carboplatin as first-line treatment in stage IIIB/IV non-small-cell lung cancer: results from a randomized, double-blinded, multicenter phase II study. *J Clin Oncol* 2012;30:2046–54.
- [164] Yang JC, Hughes M, Kammula U. Ipilimumab (anti-CTLA-4 antibody) causes regression of metastatic renal cell cancer as dissociated with enteritis and hypophysitis. *J Immunother* 2007;30:825–30.
- [165] Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* 2012;366:2443–54.
- [166] Melero I, Grimaldi AM, Perez-Gracia JL, Ascierio PA. Clinical development of immunostimulatory monoclonal antibodies and opportunities for combination. *Clin Cancer Res* 2013;19:997–1008.
- [167] Jing W, Yan X, Hallett WH, Gershon JA, Johnson BD. Depletion of CD25⁺ T cells from hematopoietic stem cell grafts increases posttransplantation vaccine-induced immunity to neuroblastoma. *Blood* 2011;117:6952–62.
- [168] Nefedova Y, Huang M, Kusmartsev S, Bhattacharya R, Cheng P, Salup R, et al. Hyperactivation of STAT3 is involved in abnormal differentiation of dendritic cells in cancer. *J Immunol* 2004;172:464–74.
- [169] Wang X, Crowe PJ, Goldstein D, Yang JL. STAT3 inhibition, a novel approach to enhancing targeted therapy in human cancers. *Int J Oncol* 2012;41:1181–91.
- [170] Stagg J, Smyth MJ. Extracellular adenosine triphosphatase and adenosine in cancer. *Oncogene* 2010;29:5346–58.
- [171] Beavis PA, Divisekera U, Paget C, Chow MT, John LB, Devaud C, et al. Blockade of A2A receptors potently suppresses the metastasis of CD73⁺ tumors. *Proc Natl Acad Sci USA* 2013;110:14711–6.
- [172] Allard B, Pommey S, Smyth MJ, Stagg J. Targeting CD73 enhances the anti-tumor activity of anti-PD-1 and anti-CTLA-4 mAbs. *Clin Can Res* 2013;19(20):5626–35.
- [173] Beavis PA, Stagg J, Darcy PK, Smyth MJ. CD73: a potent suppressor of antitumor immune responses. *Trends Immunol* 2012;33:231–7.
- [174] Stagg J, Divisekera U, McLaughlin N, Sharkey J, Pommey S, Denoyer D, et al. Anti-CD73 antibody therapy inhibits breast tumor growth and metastasis. *Proc Natl Acad Sci USA* 2010;107:1547–52.
- [175] Stagg J, Divisekera U, Duret H, Sparwasser T, Teng MW, Darcy PK, et al. CD73-deficient mice have increased antitumor immunity and are resistant to experimental metastasis. *Cancer Res* 2011;71:2892–900.
- [176] Stagg J, Divisekera U, Duret H, Sparwasser T, Teng MW, Darcy PK, et al. CD73-deficient mice are resistant to carcinogenesis. *Cancer Res* 2011;71:2892–900.
- [177] Ohta A, Gorelik E, Prasad SJ, Ronchese F, Lukashev D, Wong MKK, et al. A2A adenosine receptor protects tumors from anti-tumor T cells. *Proc Natl Acad Sci USA* 2006;103:13132–7.
- [178] Gulley JL, Madan RA, Tsang KY, Jochems C, Marte JL, Farsaci B, et al. Immune impact induced by PROSTVAC (PSA-TRICOM) a therapeutic vaccine for prostate cancer. *Cancer Immunol Res* 2013;2:133–41.

- [179] Parmiani G, Castelli C, Dalerba P, Mortarini R, Rivoltini L, Marincola FM, et al. Cancer immunotherapy with peptide-based vaccines: what have we achieved? Where are we going? *J Natl Can Inst* 2002;94:805–18.
- [180] Rivoltini L, Kawakami Y, Sakaguchi K, Southwood S, Sette A, Robbins PF, et al. Induction of tumor-reactive CTL from peripheral blood and tumor-infiltrating lymphocytes of melanoma patients by *in vitro* stimulation with an immunodominant peptide of the human melanoma antigen MART-1. *J Immunol* 1995;154:2257–65.
- [181] Cormier JN, Salgaller ML, Prevette T, Barracchini KC, Rivoltini L, Restifo NP, et al. Enhancement of cellular immunity in melanoma patients immunized with a peptide from MART-1/Melan A. *Cancer J Sci Am* 1997;3:37–44.
- [182] Salgaller ML, Afshar A, Marincola FM, Rivoltini L, Kawakami Y, Rosenberg SA. Recognition of multiple epitopes in the human melanoma antigen gp100 by peripheral blood lymphocytes stimulated *in vitro* with synthetic peptides. *Cancer Res* 1995;55:4972–9.
- [183] Marchand M, van Baren N, Weynants P, Brichard V, Dréno B, Tessier MH, et al. Tumor regressions observed in patients with metastatic melanoma treated with an antigenic peptide encoded by gene MAGE-3 and presented by HLA-A1. *Int J Cancer* 1999;80:219–30.
- [184] Weber JS, Hua FL, Spears L, Marty V, Kuniyoshi C, Celis E. A phase I trial of an HLA-A1 restricted MAGE-3 epitope peptide with incomplete Freund's adjuvant in patients with resected high-risk melanoma. *J Immunother* 1999;22:431–40.
- [185] Rice J, Ottensmeier CH, Stevenson FK. DNA vaccines: precision tools for activating effective immunity against cancer. *Nat Rev Cancer* 2008;8:108–20.
- [186] Lin MA. DNA vaccines: an historical perspective and view to the future. *Immunol Rev* 2011;239:62–84.
- [187] Chiarella P, Fazio VM, Signori E. Electroporation in DNA vaccination protocols against cancer. *Curr Drug Metab* 2013;14(3):291–9.
- [188] Mir LM, Bureau MF, Gehl J, Rangara R, Rouy D, Caillaud JM, et al. High-efficiency gene transfer into skeletal muscle mediated by electric pulses. *Proc Natl Acad Sci USA* 1999;96:4262–7.
- [189] Chiarella P, Massi E, De Robertis M, Sibilio A, Parrella P, Fazio VM, et al. Electroporation of skeletal muscle induces danger signal release and antigen-presenting cell recruitment independently of DNA vaccine administration. *Expert Opin Biol Ther* 2008;8:1645–57.
- [190] Lichtor T, Glick RP, Lin H, O-Sullivan I, Cohen EP. Intratumoral injection of IL-secreting syngeneic/allogeneic fibroblasts transfected with DNA from breast cancer cells prolongs the survival of mice with intracerebral breast cancer. *Cancer Gene Ther* 2005;12:708–14.
- [191] Lichtor T, Glick RP, Feldman LA, Osawa G, Hardman J, O-Sullivan I, et al. Enhanced immunity to intracerebral breast cancer in mice immunized with a cDNA-based vaccine enriched for immunotherapeutic cells. *J Immunother* 2008;31:18–27.
- [192] Nestle FO. Dendritic cell vaccination for cancer therapy. *Oncogene* 2000;19:6673–9.
- [193] Walter S, Weinschenk T, Stenzl A, Zdrojowy R, Pluzanska A, Szczylik C, et al. Multipetide immune response to cancer vaccine IMA901 after single-dose cyclophosphamide associates with longer patient survival. *Nat Med* 2012;12:54–61.
- [194] Lutsiak ME, Semnani RT, De Pascalis R, Kashmiri SV, Scholm J, Szabzevari H. Inhibition of CD4+CD25+ T regulatory cell function implicated in enhanced immune response by low-dose cyclophosphamide. *Blood* 2005;105:2862–8.
- [195] Palucka K, Banchereau J. Dendritic-cell-based therapeutic cancer vaccines. *Immunity* 2013;39:38–48.
- [196] Kantoff PW, Higano CS, Shore ND, Berger ED, Small EJ, Penson DF, et al. Sipuleucicel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med* 2010;363:411–22.
- [197] Russo M, Spagnuolo C, Tedesco I, Russo GL. Phytochemicals in cancer prevention and therapy: truth or dare? *Toxins* 2010;2:517–51.
- [198] Middleton Jr E, Kandaswami C. Effects of flavonoids on immune and inflammatory cell functions. *Biochem Pharmacol* 1992;43:1167–79.
- [199] Calder PC. Immunomodulation by omega-3 fatty acids. *Prostaglandins Leukot Essent Fatty Acids* 2007;77:327–35.
- [200] Dardenne M. Zinc and immune function. *Eur J Clin Nutr* 2002;56:520–3.
- [201] Calder PC, Kew S. The immune system: a target for functional foods? *Br J Nutr* 2002;88:S165–76.
- [202] Hughes DA. Dietary carotenoids and human immune function. *Nutrition* 2001;10:823–7.
- [203] Kodama N, Komuta K, Nanba H. Effect of Maitake (*Grifola frondosa*) D-fraction on the activation of NK cells in cancer patients. *J Med Food* 2003;6:371–7.
- [204] Cole BF, Baron JA, Sandler RS, Haile RW, Ahnen DJ, Bresalier RS, et al. Folic acid for the prevention of colorectal adenomas: a randomized clinical trial. *JAMA* 2007;297:2351–9.
- [205] Rock CL. Multivitamin-multimineral supplements: who uses them? *Am J Clin Nutr* 2007;85:2775–95.
- [206] Kim J, Denu RA, Dollar BA, Escalante LE, Kuether JP, Callander NS, et al. Macrophages and mesenchymal stromal cells support survival and proliferation of multiple myeloma cells. *Br J Haematol* 2012;158:336–46.
- [207] Singh AK, Gaur P, Das SN. Natural killer T cell anergy, co-stimulatory molecules and immunotherapeutic interventions. *Hum Immunol* 2014;75:250–60.
- [208] Ito Y, Vela JL, Matsumura F, Hoshino H, Tyznik A, Lee H, et al. Helicobacter pylori cholesteryl α -glucosides contribute to its pathogenicity and immune response by natural killer T cells. *PLOS ONE* 2013;8(12):e78191.
- [209] Jameson J, Witherden D, Havran WL. T-cell effector mechanisms: gammadelta and CD1d-restricted subsets. *Curr Opin Immunol* 2003;15:349–53.
- [210] Cheng C, Huang C, Ma TT, Bian EB, He Y, Zhang L, et al. SOCS1 hypermethylation mediated by DNMT1 is associated with lipopolysaccharide-induced inflammatory cytokines in macrophages. *Toxicol Lett* 2014;225:488–97.
- [211] Strauss L, Bergmann C, Gooding W, Johnson JT, Whiteside TL. The frequency and suppressor function of CD4+CD25highFoxp3+ T cells in the circulation of patients with squamous cell carcinoma of the head and neck. *Clin Cancer Res* 2007;13:6301–11.
- [212] Stewart CA, Metheny H, Iida N, Smith L, Hanson M, Steinhagen F, et al. Interferon-dependent IL-10 production by Tregs limits tumor Th17 inflammation. *J Clin Invest* 2013;123:4859–74.
- [213] Whiteside TL, Schuler P, Schilling B. Induced and natural regulatory T cells in human cancer. *Expert Opin Biol Ther* 2012;12:1383–97.
- [214] Jia Y, Guan Q, Guo Y, Du C. Reduction of inflammatory hyperplasia in the intestine in colon cancer-prone mice by water-extract of *Cistanche deserticola*. *Phytother Res* 2012;26:812–9.
- [215] Elkabets M, Ribeiro VS, Dinarello CA, Ostrand-Rosenberg S, Di Santo JP, Apte RN, et al. IL-1 β regulates a novel myeloid-derived suppressor cell subset that impairs NK cell development and function. *Eur J Immunol* 2010;40:3347–57.
- [216] Albualescu R, Codrici E, Popescu ID, Mihai S, Necula LG, Petrescu D, et al. Cytokine patterns in brain tumour progression. *Mediat Inflamm* 2013;2013:979748.
- [217] Day SD, Enos RT, McClellan JL, Steiner JL, Velázquez KT, Murphy EA. Linking inflammation to tumorigenesis in a mouse model of high-fat-diet-enhanced colon cancer. *Cytokine* 2013;64:454–62.
- [218] Fowler DW, Copier J, Wilson N, Dalgleish AG, Bodman-Smith MD. Mycobacteria activate $\gamma\delta$ T-cell anti-tumour responses via cytokines from type 1 myeloid dendritic cells: a mechanism of action for cancer immunotherapy. *Cancer Immunol Immunother* 2012;61:535–47.
- [219] Fu Z, Zuo Y, Li D, Xu W, Li D, Chen H, et al. The crosstalk: tumor-infiltrating lymphocytes rich in regulatory T cells suppressed cancer-associated fibroblasts. *Acta Oncol* 2013;52:1760–70.
- [220] Pahl JH, Ruslan SE, Buddingh EP, Santos SJ, Szuhai K, Serra M, et al. Anti-EGFR antibody cetuximab enhances the cytolytic activity of natural killer cells toward osteosarcoma. *Clin Cancer Res* 2012;18:432–41.
- [221] Martinet W, De Meyer I, Verheye S, Schrijvers DM, Timmermans JP, De Meyer GR. Drug-induced macrophage autophagy in atherosclerosis: for better or worse? *Basic Res Cardiol* 2013;108:321.
- [222] Li N, Ma T, Han J, Zhou J, Wang J, Zhang J, et al. Increased apoptosis induction in CD4+CD25+ Foxp3+ T cells contributes to enhanced disease activity in patients with rheumatoid arthritis through IL-10 regulation. *Eur Rev Med Pharmacol Sci* 2014;18:78–85.
- [223] Kawabe K, Lindsay D, Braitch M, Fahey AJ, Showe L, Constantinescu CS. IL-12 inhibits glucocorticoid-induced T cell apoptosis by inducing GMEB1 and activating PI3K/Akt pathway. *Immunobiology* 2012;217:118–23.
- [224] Iannello A, Thompson TW, Ardolino M, Lowe SW, Raulet DH. p53-dependent chemokine production by senescent tumor cells supports NKG2D-dependent tumor elimination by natural killer cells. *J Exp Med* 2013;210:2057–69.
- [225] Yao L, Sgadari C, Furuke K, Bloom ET, Teruya-Feldstein J, Tosato G. Contribution of natural killer cells to inhibition of angiogenesis by interleukin-12. *Blood* 1999;93:1612–21.
- [226] Naldini A, Pucci A, Bernini C, Carraro F. Regulation of angiogenesis by Th1- and Th2-type cytokines. *Curr Pharm Des* 2003;9:511–9.
- [227] Larsen H, Muz B, Khong TL, Feldmann M, Paleolog EM. Differential effects of Th1 versus Th2 cytokines in combination with hypoxia on HIFs and angiogenesis in RA. *Arthritis Res Ther* 2012;14:R180 [Epub ahead of print].
- [228] Rankin EB, Yu D, Jiang J, Shen H, Pearce EJ, Goldschmidt MH, et al. An essential role of Th1 responses and interferon gamma in infection-mediated suppression of neoplastic growth. *Cancer Biol Ther* 2003;2:687–93.
- [229] Caccamo N, Dieli F, Meraviglia S, Guggino G, Salerno A. Gammadelta T cell modulation in anticancer treatment. *Curr Cancer Drug Targets* 2010;10:27–36.
- [230] Meraviglia S, Eberl M, Vermijlen D, Todaro M, Buccheri S, Cicero G, et al. *In vivo* manipulation of Vgamma9Vdelta2 T cells with zoledronate and low-dose interleukin-2 for immunotherapy of advanced breast cancer patients. *Clin Exp Immunol* 2010;161:290–7.
- [231] Gomes AQ, Martins DS, Silva-Santos B. Targeting $\gamma\delta$ T lymphocytes for cancer immunotherapy: from novel mechanistic insight to clinical application. *Cancer Res* 2010;70:10024–7.
- [232] Wakita D, Sumida K, Iwakura Y, Nishikawa H, Ohkuri T, Chamoto K, et al. Tumor-infiltrating IL-17-producing gammadelta T cells support the progression of tumor by promoting angiogenesis. *Eur J Immunol* 2010;40:1927–37.
- [233] Pakala R, Watanabe T, Benedict CR. Induction of endothelial cell proliferation by angiogenic factors released by activated monocytes. *Cardiovasc Radiat Med* 2002;3:95–101.
- [234] Facciabene A, Peng X, Hagemann IS, Balint K, Barchetti A, Wang LP, et al. Tumour hypoxia promotes tolerance and angiogenesis via CCL28 and T(reg) cells. *Nature* 2011;475:226–30.
- [235] Voest EE, Kenyon BM, O'Reilly MS, Truitt G, D'Amato RJ, Folkman J. Inhibition of angiogenesis *in vivo* by interleukin 12. *J Natl Cancer Inst* 1995;87:581–6.
- [236] Budhu A, Forgues M, Ye QH, Jia HL, He P, Zanetti KA, et al. Prediction of venous metastases, recurrence, and prognosis in hepatocellular carcinoma based on a unique immune response signature of the liver microenvironment. *Cancer Cell* 2006;10:99–111.
- [237] Ferrarini M, Ferrero E, Dagna L, Poggi A, Zocchi MR. Human gammadelta T cells: a nonredundant system in the immune-surveillance against cancer. *Trends Immunol* 2002;23:14–8.

- [238] Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* 2004;4:71–8.
- [239] DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, et al. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. *Cancer Cell* 2009;16:91–102.
- [240] Smyth MJ, Cretney E, Takeda K, Wiltrot RH, Sedger LM, Kayagaki N, et al. Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) contributes to interferon gamma-dependent natural killer cell protection from tumor metastasis. *J Exp Med* 2001;193:661–70.
- [241] Shen F, Li JL, Cai WS, Zhu GH, Gu WL, Jia L, et al. Interleukin-12 prevents colorectal cancer liver metastases in mice. *OncoTargets Ther* 2013;6:523–6.
- [242] Thakur A, Schalk D, Sarkar SH, Al-Khadimi Z, Sarkar FH, Lum LG. A Th1 cytokine-enriched microenvironment enhances tumor killing by activated T cells armed with bispecific antibodies and inhibits the development of myeloid-derived suppressor cells. *Cancer Immunol Immunother* 2012;61:497–509.
- [243] Ye J, Ma C, Wang F, Hsueh EC, Toth K, Huang Y, et al. Specific recruitment of $\gamma\delta$ regulatory T cells in human breast cancer. *Cancer Res* 2013;73:6137–48.
- [244] Lamagna C, Aurrand-Lions M, Imhof BA. Dual role of macrophages in tumor growth and angiogenesis. *J Leukoc Biol* 2006;80:705–13.
- [245] Schoppmann SF, Birner P, Stöckl J, Kalt R, Ullrich R, Caucig C, et al. Tumor-associated macrophages express lymphatic endothelial growth factors and are related to peritumoral lymphangiogenesis. *Am J Pathol* 2002;161:947–56.
- [246] Tsung K, Dolan JP, Tsung YL, Norton JA. Macrophages as effector cells in interleukin 12-induced T cell-dependent tumor rejection. *Cancer Res* 2002;62:5069–75.
- [247] Wu X, Peng M, Huang B, Zhang H, Wang H, Huang B, et al. Immune microenvironment profiles of tumor immune equilibrium and immune escape states of mouse sarcoma. *Cancer Lett* 2013;340:124–33.
- [248] Hayakawa Y, Sato-Matsushita M, Takeda K, Iwakura Y, Tahara H, Irimura T. Early activation and interferon- γ production of tumor-infiltrating mature CD27 high natural killer cells. *Cancer Sci* 2011;102:1967–71.
- [249] Kerker SP, Goldszmid RS, Muranski P, Chinnasamy D, Yu Z, Reger RN, et al. IL-12 triggers a programmatic change in dysfunctional myeloid-derived cells within mouse tumors. *J Clin Invest* 2011;121:4746–57.
- [250] Schepetkin IA, Quinn MT. Botanical polysaccharides: macrophage immunomodulation and therapeutic potential. *Int Immunopharmacol* 2006;6:317–33.
- [251] Jeff IB, Yuan X, Sun L, Kassim RM, Foday AD, Zhou Y. Purification and *in vitro* anti-proliferative effect of novel neutral polysaccharides from *Lentinus edodes*. *Int J Biol Macromol* 2013;52:99–106.
- [252] Zhao L, Xiao Y, Xiao N. Effect of lentinan combined with docetaxel and cisplatin on the proliferation and apoptosis of BGC823 cells. *Tumour Biol* 2013;34:1531–6.
- [253] Wakshlag JJ, Balkman CA, Morgan SK, McEntee MC. Evaluation of the protective effects of all-trans-astaxanthin on canine osteosarcoma cell lines. *Am J Vet Res* 2010;71:89–96.
- [254] Yasui Y, Hosokawa M, Mikami N, Miyashita K, Tanaka T. Dietary astaxanthin inhibits colitis and colitis-associated colon carcinogenesis in mice via modulation of the inflammatory cytokines. *Chem Biol Interact* 2011;193:79–87.
- [255] Tanaka T, Kawamori T, Ohnishi M, Makita H, Mori H, Satoh K, et al. Suppression of azoxymethane-induced rat colon carcinogenesis by dietary administration of naturally occurring xanthophylls astaxanthin and canthaxanthin during the postinitiation phase. *Carcinogenesis* 1995;16:2957–63.
- [256] Sliva D, Loganathan J, Jiang J, Jedinak A, Lamb JG, Terry C, et al. Mushroom *Ganoderma lucidum* prevents colitis-associated carcinogenesis in mice. *PLoS ONE* 2012;7:e47873.
- [257] Joseph S, Sabulal B, George V, Antony KR, Janardhanan KK. Antitumor and anti-inflammatory activities of polysaccharides isolated from *Ganoderma lucidum*. *Acta Pharm* 2011;61:335–42.
- [258] Yang B, Xiao B, Sun T. Antitumor and immunomodulatory activity of *Astragalus membranaceus* polysaccharides in H22 tumor-bearing mice. *Int J Biol Macromol* 2013;62:287–90.
- [259] Nishitani Y, Zhang L, Yoshida M, Azuma T, Kanazawa K, Hashimoto T, et al. Intestinal anti-inflammatory activity of lentinan: influence on IL-8 and TNFR1 expression in intestinal epithelial cells. *PLoS ONE* 2013;8:e62441.
- [260] Bisen PS, Baghel RK, Sanodiya BS, Thakur GS, Prasad GB. *Lentinus edodes*: a macrofungus with pharmacological activities. *Curr Med Chem* 2010;17:2419–30.
- [261] Speranza L, Pesce M, Patrino A, Franceschelli S, de Lutiis MA, Grilli A, et al. Astaxanthin treatment reduced oxidative induced pro-inflammatory cytokines secretion in U937: SHP-1 as a novel biological target. *Mar Drugs* 2012;10:890–9.
- [262] Hsieh TC, Wu JM. Suppression of proliferation and oxidative stress by extracts of *Ganoderma lucidum* in the ovarian cancer cell line OVCAR-3. *Int J Mol Med* 2011;28:1065–9.
- [263] Jiang J, Slivova V, Harvey K, Valachovicova T, Sliva D. *Ganoderma lucidum* suppresses growth of breast cancer cells through the inhibition of Akt/NF-kappaB signaling. *Nutr Cancer* 2004;49:209–16.
- [264] Hsieh TC, Wu P, Park S, Wu JM. Induction of cell cycle changes and modulation of apoptogenic/anti-apoptotic and extracellular signaling regulatory protein expression by water extracts of l-m-Yunity (PSP). *BMC Complement Altern Med* 2006;6:30.
- [265] Auyeung KK, Law PC, Ko JK. *Astragalus* saponins induce apoptosis via an ERK-independent NF-kappaB signaling pathway in the human hepatocellular HepG2 cell line. *Int J Mol Med* 2009;23:189–96.
- [266] Fang N, Li Q, Yu S, Zhang J, He L, Ronis MJ, et al. Inhibition of growth and induction of apoptosis in human cancer cell lines by an ethyl acetate fraction from shiitake mushrooms. *J Altern Complement Med* 2006;12:125–32.
- [267] Paloza F, Torelli C, Boninsegna A, Simone R, Catalano A, Mele MC, et al. Growth-inhibitory effects of the astaxanthin-rich alga *Haematococcus pluvialis* in human colon cancer cells. *Cancer Lett* 2009;283:108–17.
- [268] Gao Y, Gao H, Chan E, Tang W, Xu A, Yang H, et al. Antitumor activity and underlying mechanisms of ganopoly, the refined polysaccharides extracted from *Ganoderma lucidum*, in mice. *Immunol Investig* 2005;34:171–98.
- [269] Wang T, Xuan X, Li M, Gao P, Zheng Y, Zang W, et al. *Astragalus* saponins affect proliferation, invasion and apoptosis of gastric cancer BGC-823 cells. *Diagn Pathol* 2013;8:179.
- [270] Wang KP, Zhang QL, Liu Y, Wang J, Cheng Y, Zhang Y. Structure and inducing tumor cell apoptosis activity of polysaccharides isolated from *Lentinus edodes*. *J Agric Food Chem* 2013;61:9849–58.
- [271] Kavitha K, Kowshik J, Kishore TK, Baba AB, Nagini S. Astaxanthin inhibits NF-kB and Wnt/ β -catenin signaling pathways via inactivation of Erk/MAPK and PI3K/Akt to induce intrinsic apoptosis in a hamster model of oral cancer. *Biochim Biophys Acta* 2013;1830:4433–44.
- [272] Wesolowska O, Wisniewski J, Bielawska-Pohl A, Paprocka M, Duarte N, Ferreira MJ, et al. Stilbenes as multidrug resistance modulators and apoptosis inducers in human adenocarcinoma cells. *Anticancer Res* 2010;30:4587–93.
- [273] Yuen JW, Gohel MD, Au DW. Telomerase-associated apoptotic events by mushroom *ganoderma lucidum* on premalignant human urothelial cells. *Nutr Cancer* 2008;60:109–19.
- [274] Gao Y, Tang W, Gao H, Chan E, Lan J, Zhou S. *Ganoderma lucidum* polysaccharide fractions accelerate healing of acetic acid-induced ulcers in rats. *J Med Food* 2004;7:417–21.
- [275] Cao QZ, Lin ZB. Antitumor and anti-angiogenic activity of *Ganoderma lucidum* polysaccharides peptide. *Acta Pharmacol Sin* 2004;25:833–8.
- [276] Zhang L, Yang Y, Wang Y, Gao X. *Astragalus membranaceus* extract promotes neovascularisation by VEGF pathway in rat model of ischemic injury. *Pharmazie* 2011;66:144–50.
- [277] Sano B, Sugiyama Y, Kunieda K, Sano J, Saji S. Antitumor effects induced by the combination of TNP-470 as an angiogenesis inhibitor and lentinan as a biological response modifier in a rabbit spontaneous liver metastasis model. *Surg Today* 2002;32:503–9.
- [278] Kim DH, Hossain MA, Kim MY, Kim JA, Yoon JH, Suh HS, et al. A novel resveratrol analogue, HS-1793, inhibits hypoxia-induced HIF-1 α and VEGF expression, and migration in human prostate cancer cells. *Int J Oncol* 2013;43:1915–24.
- [279] Sliva D, Labarrere C, Slivova V, Sedlak M, Lloyd Jr FP, Ho NW. *Ganoderma lucidum* suppresses motility of highly invasive breast and prostate cancer cells. *Biochem Biophys Res Commun* 2002;298:603–12.
- [280] Liu X, Yang Y, Zhang X, Xu S, He S, Huang W, et al. Compound *Astragalus* and *Salvia miltiorrhiza* extract inhibits cell invasion by modulating transforming growth factor-beta/Smad in HepG2 cell. *J Gastroenterol Hepatol* 2010;25:420–6.
- [281] Ren L, Perera C, Hemar Y. Antitumor activity of mushroom polysaccharides: a review. *Food Funct* 2012;3:1118–30.
- [282] Ogasawara M, Matsunaga T, Suzuki H. Differential effects of antioxidants on the *in vitro* invasion, growth and lung metastasis of murine colon cancer cells. *Biol Pharm Bull* 2007;30:200–4.
- [283] Polonini HC, Lima LL, Gonçalves KM, do Carmo AM, da Silva AD, Raposo NR. Photoprotective activity of resveratrol analogues. *Bioorg Med Chem* 2013;21:964–8.
- [284] Mikstacka R, Ignatowicz E. Chemopreventive and chemotherapeutic effect of trans-resveratrol and its analogues in cancer. *Pol Merkur Lekarski* 2010;28:496–500.
- [285] Lu J, Sun LX, Lin ZB, Duan XS, Ge ZH, Xing EH, et al. Antagonism by *Ganoderma lucidum* polysaccharides against the suppression by culture supernatants of B16F10 melanoma cells on macrophage. *Phytother Res* 2014;28:200–6.
- [286] Ono Y, Hayashida T, Konagai A, Okazaki H, Miyao K, Kawachi S, et al. Direct inhibition of the transforming growth factor- β pathway by protein-bound polysaccharide through inactivation of Smad2 signaling. *Cancer Sci* 2012;103:317–24.
- [287] Li Q, Bao JM, Li XL, Zhang T, Shen XH. Inhibiting effect of *Astragalus* polysaccharides on the functions of CD4+CD25 high Treg cells in the tumor microenvironment of human hepatocellular carcinoma. *Chin Med J (Engl)* 2012;125:786–93.
- [288] Zong A, Cao H, Wang F. Anticancer polysaccharides from natural resources: a review of recent research. *Carbohydr Polym* 2012;904:1395–410.
- [289] Nagendraprabhu P, Sudhandiran G. Astaxanthin inhibits tumor invasion by decreasing extracellular matrix production and induces apoptosis in experimental rat colon carcinogenesis by modulating the expressions of ERK-2, NFkB and COX-2. *Investig New Drugs* 2011;29:207–24.
- [290] Choi YJ, Yang KM, Kim SD, Yoo YH, Lee SW, Seo SY, et al. Resveratrol analogue HS-1793 induces the modulation of tumor-derived T cells. *Exp Ther Med* 2012;3:592–8.
- [291] Goodnow CC, Sprent J, Fazekas de St Groth B, Vinuesa CG. Cellular and genetic mechanisms of self tolerance and autoimmunity. *Nature* 2005;435:590–7.
- [292] Rosenberg SA, Packard BS, Aebbersold PM, Solomon D, Topalian SL, Toy ST, et al. Use of tumor-infiltrating lymphocytes and interleukin-2 in the

- immunotherapy of patients with metastatic melanoma. A preliminary report. *N Engl J Med* 1988;319:1676–80.
- [293] Rosenberg SA, Yannelli JR, Yang JC, Topalian SL, Schwartzentruber DJ, Weber JS, et al. Treatment of patients with metastatic melanoma with autologous tumor-infiltrating lymphocytes and interleukin 2. *J Natl Cancer Inst* 1994;86:1159–66.
- [294] Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, et al. Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Science* 2002;298:850–4.
- [295] Rosenberg SA, Restifo NP, Yang JC, Morgan RA, Dudley ME. Adoptive cell transfer: a clinical path to effective cancer immunotherapy. *Nat Rev Cancer* 2008;8:299–308.
- [296] Schietinger A, Delrow JJ, Basom RS, Blattman JN, Greenberg PD. Rescued tolerant CD8 T cells are preprogrammed to reestablish the tolerant state. *Science* 2012;335:723–7.
- [297] King C, Ilic A, Koelsch K, Sarvetnick N. Homeostatic expansion of T cells during immune insufficiency generates autoimmunity. *Cell* 2004;117:265–77.
- [298] Lee PP, Yee C, Savage PA, Fong L, Brockstedt D, Weber JS, et al. Characterization of circulating T cells specific for tumor-associated antigens in melanoma patients. *Nat Med* 1999;5:677–85.
- [299] Nagorsen D, Scheibenbogen C, Marincola FM, Letsch A, Keilholz U. Natural T cell immunity against cancer. *Clin Cancer Res* 2003;9:4296–303.
- [300] Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res* 2011;17:4550–7.
- [301] Rosenberg SA, Yang C, White DE. Recombinant fowlpox viruses encoding the anchor-modified gp100 melanoma antigen can generate antitumor immune response in patients with metastatic melanoma. *Clin Cancer Res* 2003;9:2973–80.
- [302] Choi BK, Lee SC, Lee MJ, Kim YH, Kim YW, Ryu KW, et al. 4-1BB-based isolation and expansion of CD8+ T cells specific for self and non-self tumor antigens for adoptive T cell therapy. *J Immunother* 2014;37:225–36.
- [303] Kalos M. Muscle CARs and TcRs: turbo-charged technologies for the (T cell) masses. *Cancer Immunol Immunother* 2012;61:127–35.