

OsteoArthritis and Cartilage (2004) 12, 277–283

© 2004 Published by Elsevier Ltd on behalf of OsteoArthritis Research Society International.

doi:10.1016/j.joca.2004.01.001

Osteoarthritis and Cartilage

**International
Cartilage
Repair
Society**

Hyaluronic acid levels may have predictive value for the progression of knee osteoarthritis

K. Pavelka*, Š. Forejtová, M. Olejárová, J. Gatterová, L. Šenolt, P. Špaček, M. Braun, M. Hulejová, J. Štovíčková and A. Pavelková

Institute of Rheumatology Prague and Clinic of Rheumatology, Charles University 1st Medical School, Prague, Czech Republic

Summary

Study objectives: To study prognostic value of different biochemical markers for morphological progression of early knee osteoarthritis.**Design:** A total of 89 patients with knee osteoarthritis (OA) were enrolled into the study. The follow-up period was 2 years. Radiological OA progression was evaluated by measuring joint space width. Pentosidine was detected using the HPLC method described earlier, cartilage oligomeric matrix protein (COMP) using the method published by our team. MMP-9, tissue inhibitors of metalloproteinases (TIMP), YKL-40 and hyaluronic acid were detected using commercially available kits.**Results:** In the group of patients suffering from knee OA, higher serum levels of pentosidine ($P=0.04$), MMP-9 ($P=0.02$), TIMP ($P=0.04$) and COMP ($P=0.05$) were detected compared with healthy control subjects. Using a correlation analysis method, it has been found that the patients with higher basic serum levels of hyaluronic acid had a faster radiological progression ($r=0.56$, $P<0.005$), as well as the patients with higher basic serum pentosidine levels ($r=0.30$, $P<0.005$). Other biochemical markers had no statistically significant prognostic value.**Conclusions:** In our study, serum levels of hyaluronic acid and pentosidine had a predictive value for further development of knee OA in that further joint space narrowing was detected in the patients with knee OA in the next 2 years.

© 2004 Published by Elsevier Ltd on behalf of OsteoArthritis Research Society International.

Key words: Osteoarthritis, progression, biochemical markers, hyaluronic acid, pentosidine.

Introduction

Osteoarthritis (OA) is the most common joint condition affecting up to 12% of the population¹. Its prevalence in the population of 65 years of age and over is almost 50%. OA often causes disability resulting in enormous financial costs of the therapy, including final surgical treatment.

The American College of Rheumatology (ACR) has published the criteria for diagnosis and classification of OA. However, these are not convenient for the detection of early OA stages^{2–4}.

Also, the evaluation of OA progression has not been successfully addressed despite different organizations such as OARS (Osteoarthritis Research Society International)⁵ or OMERACT (Outcome Measures In Rheumatoid Arthritis Clinical Trials)⁶ publishing their 'Recommendations'. These outcomes may be classified as indicators evaluating the symptoms (pain and functions), and indicators of the morphological seriousness and progression of the disease⁷. There has been a common view that in order to be able to evaluate the effectiveness of chondroprotective therapy, or 'structurally modifying drugs' as they are also called⁵, it is necessary to identify a

morphological criterion. Although such methods as MRI scan⁸ and ultrasound are being tested at present, the only validated indicator is still a flat X-ray image.

Recently a debate has been stirred concerning the most advantageous radiological equipment and techniques for OA progression evaluation⁹. The debate mainly concerns knee joints. There have been reports that most investigators now prefer imaging in semiflexion¹⁰, both assisted and unassisted, with the help of fluoroscopy, and a digital computer assessment of the joint space width¹¹. However, no clearly defined X-ray protocol has yet been published.

Biochemical OA markers represent a relatively new and rapidly developing research field. The possibility for using these markers for early OA diagnosis and prognostic evaluation as well as the methods of OA progression were proposed more than 15 years ago. The international GREESS group has proposed the following points of interest which should be addressed by future trials studying biological markers¹²:

- use them as surrogate indicators for diagnosis, in addition to X-ray,
- identify healthy persons with a possible risk of OA,
- predict the development of early stages (both treated and untreated),
- evaluate OA progression.

The most complicated task is to identify and select suitable biochemical markers. OA is a structurally complex

*Address correspondence to: Professor Karel Pavelka, M.D., Ph.D., Institute of Rheumatology, Na slupi 4, 128 50 Prague 2, Czech Republic. Tel: +420 234075244; Fax: +420 224914451; E-mail: pavelka@revma.cz

Received 14 March 2003; revision accepted 9 January 2004.

disease manifesting itself by the destruction of joint cartilage, subchondral bone, sclerosis and cysts, osteophytes and synovial inflammation. Cartilage destruction is considered to be the most important and primary event and therefore cartilage attracts most of the attention. However, it is also important to take other joint tissues into account.

Biochemical markers may be classified according to several aspects. According to their origin, it is possible to classify the markers as originating from cartilage, bone, or as markers reflecting synovial inflammation. The advantage of the marker would be its specificity to a certain tissue. However, this is mostly not the case. The markers may further be specified as indicators of degradation, indicators of synthesis and pure indicators of inflammation. Another classification of the markers is according to the compartment in which they may be detected: serum, urine and synovial fluid. During the last year, pentosidine a new potential indicator has been identified¹³.

Increased biochemical marker levels have been identified e.g. in the serum or urine of OA patients compared with healthy control subjects in proteoglycan monomer fragments¹⁴, keratansulphate¹⁵, cartilage oligomeric matrix protein (COMP)¹⁶, C telopeptide collagen II¹⁷, hyaluronic acid¹⁸, CRP¹⁹, YKL 40²⁰, metalloproteinases²¹ and pyridinoline²². Predictive biochemical marker values for OA progression have been published for COMP²³, hyaluronic acid, CRP and TIMP^{24,25}.

The main objectives of our study were to: (a) perform a longitudinal clinical and morphological study of OA progression in early/moderate OA and look for a correlation between clinical and different morphological indicators (RTG, MRI, ultrasound and scintigraphy), (b) evaluate a battery of potential biochemical OA markers and to express a standpoint with regard to their value as predictive morphological progression indicators. In this study, the results will be presented which correlate biochemical markers with structural damage. The correlation with clinical parameters will be the subject of the next publication.

Methods

In the longitudinal study, patients were followed for 2 years. The patients were examined at the beginning and at the end of the evaluated time period.

PATIENTS

Patients with knee OA according to the ACR criteria were enrolled into the study^{2,3} all had baseline radiographs and have proven osteophytes. These were symptomatic patients with a reported pain during the last month. The disease duration (from its diagnosing) did not exceed 3 years. Each patient has provided his/her written consent with the participation in the study. In the control group we used 20 volunteers employed in the Institute whose mean age was the same as that of the studied group and who did not suffer from clinically manifest osteoarthritis of the knee or other joints. Blood sera from healthy volunteers were only taken at baseline for ethical reasons.

RADIOLOGICAL EVALUATION

For X-ray imaging, a technique with knee in extension was used in order to achieve maximum standardization, as we described earlier²⁷.

The joint space width was measured at the narrowest point in the medial compartment of the femorotibial joint. For measuring, a 0.1 mm caliper and magnifying glasses was used. Each image was read by two experienced radiologists who did not know which of the images was first or last. This method, described by Lequesne²⁸, has been modified at our Institute²⁹. When using this method, a so-called intraobserver error was 3.6% and interobserver error was 6.5%. In addition to the measurement of the joint space when evaluating OA progression, we also used the atlas published by R. Altman³⁰. For bilateral affection, the knee which was in a worse clinical state was selected for examination.

BIOCHEMICAL MARKERS

After the collection, the blood samples were centrifuged and the serums were frozen at -70°C until their final processing.

COMP was determined using a sandwich ELISA monoclonal antibodies 17-C10 and 16-F12³¹. Intra and inter assay variance of the method has been established as 8% and 10% respectively.

ELISA MMP-9, MMP-13 AND TIMP-1

The levels of matrix metalloproteinases MMP-9 and MMP-13 and of their tissue inhibitor TIMP-1 were detected by the ELISA method, using kits produced by the Biotrak/Amersham Biosciences Company. To detect MMP-9 and TIMP-1, ELISA plates were covered with the serums diluted at 1:20, to detect MMP-13 undiluted serums were used. Each sample was applied into two pits. The concentrations were evaluated using a computer program of the ELISA reader Sun Rise supplied by the Schoeller Instruments company.

PENTOSIDINE

Serum pentosidine was detected by a method published by us earlier^{13,40} which is based on high-pressure liquid chromatography (HPLC) in reversion phase. Pentosidine (PEN) monitoring was performed using a fluorescent detector at excitation and emission wavelength $\lambda_{\text{EX}}/\lambda_{\text{EM}}=335/385$ nm, column temperature was 40°C , mobile phase flow rate was 0.5 ml/min. A brief method description: 1 ml of the sample (serum, urine) was mixed with 1 ml of concentrated HCl (35%) and hydrolyzed at 105°C over the period of 16 h. Preliminary hydrolyzate refining (0.5 ml) was performed with spheric cellulose CC31 by selective sorption in 15 ml of n-butylalcohol – acetic acid – water mixture (8-1-1) and by desorption using 3 ml 0.05 M HCl. The hydrolyzate was evaporated and after being dissolved in 250 μl of the mobile phase, was applied into the HPLC column in the amount of 10 μl . The reproducibility of the HPLC evaluation was 1%, reproducibility of the entire method (i.e. including hydrolyzation and refining) 4.44%, recovery was $77\pm 3.5\%$ and sensitiveness limit was 17.6 femtomols.

STATISTICAL ANALYSIS

The comparison of OA patients with healthy control subjects was performed using the *t*-test.

The correlation of biochemical markers with joint space narrowing was performed using a correlation coefficient in linear regression.

Table I

	Demographic characteristic of the OA patients with knee joints (N=89)	
	Mean±SD	Controls
Males/females	30/59	8/12
Age (year)	56.7±7.2	51.7±6.4
Disease duration (years)	2.9±1.8	–
BMI	28.6±4.6	27.8±5.2
Joint space at the beginning of the study	4.95±1.46	–
Joint space narrowing over 2 years (mm)	-0.40±0.79	–

Table II
Biochemical markers in patients with knee OA*

Indicator	Study beginning		Study end		Controls
	Value	P 1	Value	P 2	
Hyaluronic acid (ng/ml)	30.2±19.6	NS	27.1±16.6	0.1	32.5±20.6
Pentosidine (nmol/l)	143.4±99.0	0.04	125.6±76.9	NS	115.3±34.0
YKL-40 (ng/ml)	59.0±34.9	NS	64.0±41.7	NS	65.8±35.3
MMP-9 (ng/ml)	147.1±101.4	0.02	150.7±131.9	0.05	97.9±67.0
TIMP (ng/ml)	872.7±155.1	0.04	833.8±161.8	NS	781.8±145.7
COMP (ng/ml)	1063.2±626.4	0.05	1235.5±647.5	0.04	1022.9±646.0

* N=89. MMP-9: Metalloproteinase 9; MMP-13: metalloproteinase 13; TIMP: tissue inhibitors of metalloproteinases; COMP: cartilage oligomeric matrix protein; P 1: significance, t-test of the original value compared with control values; P 2: final values compared with control values.

Results

A total of 100 patients were enrolled into the study of which 89 patients presented themselves for the final examination after 2 years. There were 59 females and 30 males (Table I). The mean age was approximately 57 years and the mean disease duration was 2.9 years.

RADIOLOGICAL PROGRESSION EVALUATION

The joint space width was 4.95±1.46 mm at the beginning of the study and 4.55±1.58 mm at the end of the study. The mean joint space narrowing was 0.4±0.79 mm over the period of 2 years, which corresponds with a mean yearly progression of 0.2 mm.

BIOCHEMICAL MARKERS

Table II shows the comparison of biochemical marker values at the beginning and end of the study between the entire group of patients with osteoarthritis and the healthy control subjects. At the beginning of the study, higher values of pentosidine (143.4±99.0 nmol/l vs 115.3±34.3 nmol/l, $P=0.04$), MMP-9 (147.1±101.4 ng/ml vs 97.9±67.0, $P=0.02$), TIMP (872.7±155.1 ng/ml vs 781.8±145.7 ng/ml, $P=0.04$) and of COMP (1063.2±626.4 ng/ml vs 1022.9±646.0 ng/mol, $P=0.05$) were detected. At the end of the study, only the values of the following markers were significant compared with control subjects: MMP-9 (150.7±131.9 vs 97.9±67 ng/ml, $P=0.05$) and COMP (1235.5±647.5 vs 1022.9±646.0 ng/ml, $P=0.04$). No significant changes were registered during the 2-year period.

We have also performed a correlation of individual biochemical marker values in relation to the narrowing of the joint space. A significant relation has been detected for the correlation between the initial values of hyaluronic acid and

joint space narrowing during the next 2 years ($r=0.56$) ($P<0.005$) but not for hyaluronic acid values at the end of the study ($r=0.06$) (Fig. 1). Basic pentosidine levels ($r=0.30$) ($P<0.005$) had a significant relation to further joint space narrowing and a positive relation was also detected at the end of the study ($r=0.34$, $P=0.005$) (Fig. 2). No correlation has been found between initial and final COMP levels and joint space narrowing (Fig. 3).

Discussion

The objective of our study was mainly to identify the importance of so-called biochemical markers for OA progression. A group of patients suffering from primary knee OA of a relatively early stage (<3 years) were selected for the study.

Joint space narrowing over a 2-year period was 0.4 mm, which corresponds with 0.2 mm a year. Literature data on a mean yearly joint space narrowing differ substantially (10 times) ranging from 0.06 to 0.6 mm/year³². In our previous studies, we detected a yearly joint space narrowing rate of only 0.1 mm/year^{27,29}. Apart from the used technique, the yearly joint space narrowing rate also depends on patients selected for the study. In our study, the patients suffered from more severe symptoms than in previous studies. As to radiological technique, the patients' knees were in extension when making X-ray images, the reason for it being an uncertainty at the beginning of the study (and virtually to the present time) as to which semiflexion techniques is the most advantageous for longitudinal patient studies³³.

One of the possibly important features of biochemical OA markers is their potential as a diagnostic tool. In accordance with a published literature, we found higher COMP¹⁶, TIMP²⁶ and MMP 9³⁴ values in patients with OA. The finding of higher pentosidine levels in OA patients compared to healthy control subjects is relatively new.

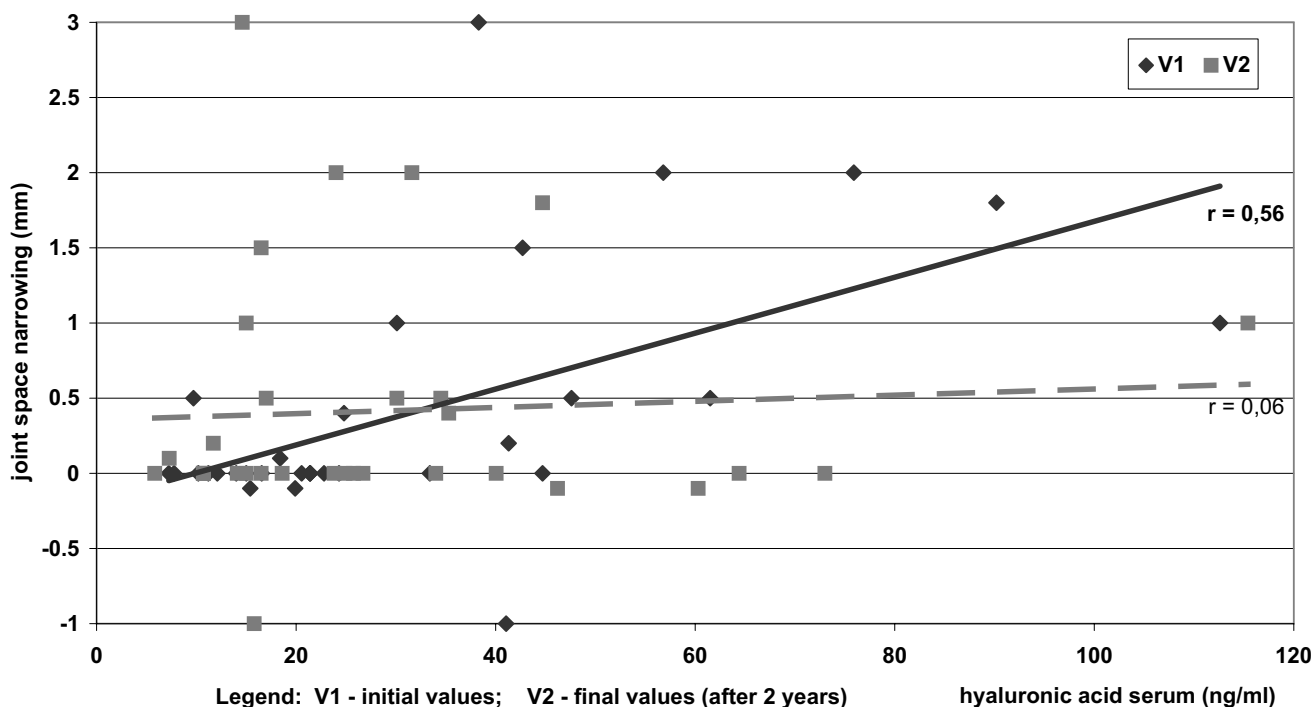


Fig. 1. Correlation between serum hyaluronic acid levels and knee joint space narrowing in patients with OA during 2 years.

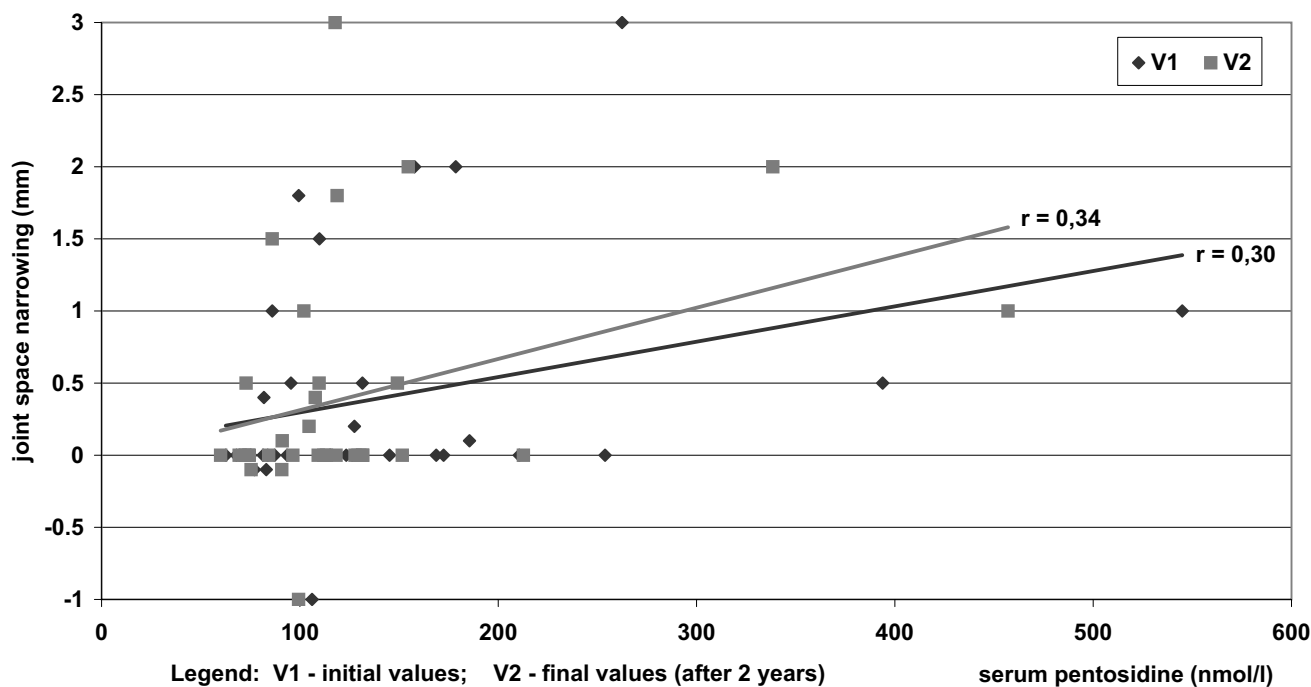


Fig. 2. Correlation between serum pentosidine levels and knee joint space narrowing in patients with OA during 2 years.

However, despite the fact that these values were significantly higher in the group with OA compared with the control group, the clinical importance of the observation is disputable. The differences are higher only to a certain extent and in so-called healthy control subjects there is such a variance in the values that their importance for an individual patient is rather low.

Apart from the diagnostic importance of biochemical markers also their prognostic value is being considered.

In this study, a higher progression risk of OA progression was posed by increased serum hyaluronic acid values, which is, however, predominantly a marker of synovial inflammation. A prognostic value of serum hyaluronic acid values has already been published¹⁸.

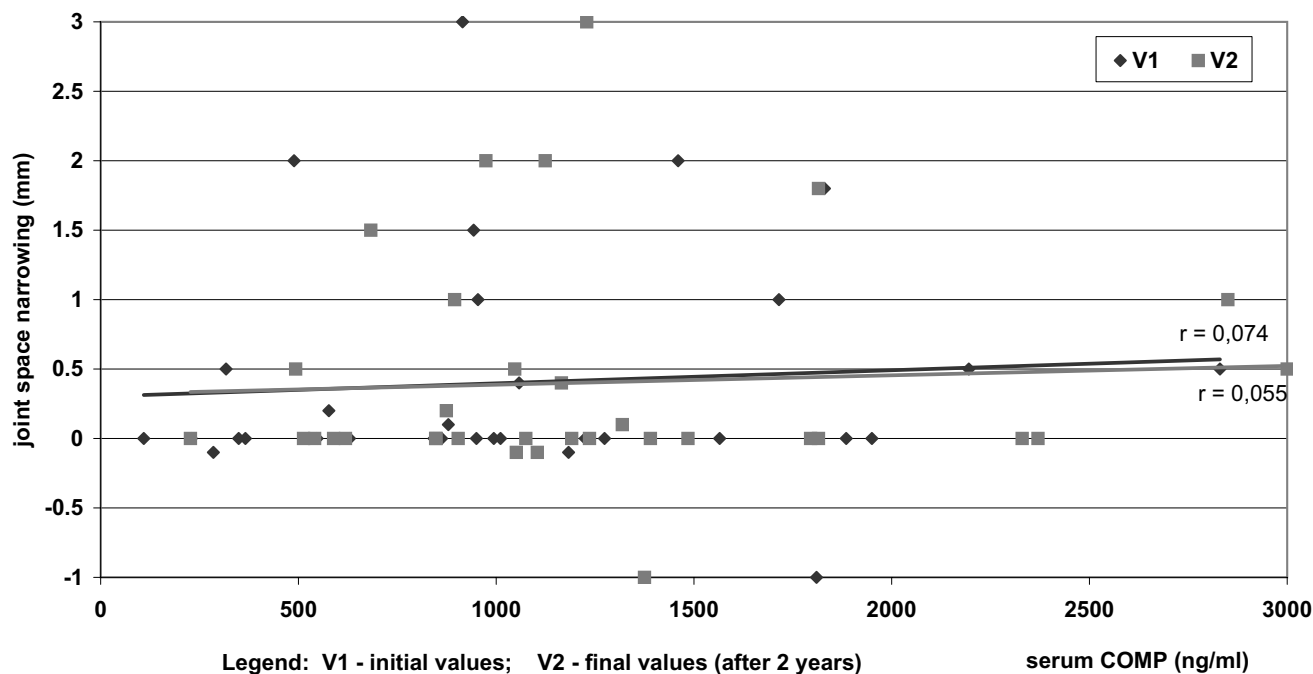


Fig. 3. Correlation between serum COMP levels and knee joint space narrowing in patients with OA during 2 years.

Final glycation products (advanced glycation end-products – AGE) have enjoyed the attention of researchers in the last 10 years³⁵. Of the AGE group, one of the best-defined products is pentosidine detectable by the HPLC method. AGE have been studied in connection with aging³⁶ but have also been detected in a number of other diseases such as renal insufficiency³⁷ or rheumatoid arthritis³⁸. Recently, De Groot *et al.*³⁹ has found AGE accumulation in joint cartilage which had a negative impact on the balance of synthesis and degradation of extracellular matrix proteoglycans. Recently, Špacek *et al.* has detected only insignificantly higher urine pentosidine levels in knee OA patients compared with healthy control subjects. However, after the surgery of the joint affected by the arthritic process, a significant decrease in urine pentosidine levels was detected¹³. Pentosidine serum levels are not tissue specific, but were found in another study correlation of pentosidine and COMP in synovial fluid⁴¹. We have found in the same study, correlation of pentosidine in synovial fluid and serum ($r^2=0.7$, $P<0.001$) but no correlation in COMP levels in synovial fluid and serum ($r^2=0.00092$). We suggest that pentosidine may be a more suitable putative local marker of cartilage pathology than COMP.

To establish a potential prognostic value of pentosidine, which has been reported in this study, further trials will be required.

Acknowledgements

The study was performed based on the Grant of the Ministry of Health of the Czech Republic (MZCR) No. NK/5366-4

References

1. Lawrence R, Helmick G, Arnett DF, Dey RA, Felson DT, Giannini EH, *et al.* Estimates of the prevalence of arthritis and selected musculoskeletal disorders in the United States. *Arthritis Rheum* 1998;41:778–88.
2. Altman R, Alarcon G, Appelreuth D, Block D, Borenstein D, Brandt K, *et al.* The American College of Rheumatology criteria for the classification and reporting of osteoarthritis of the hip. *Arthritis Rheum* 1991;34:505–14.
3. Altman R, Asch E, Block CD, Bole D, Borenstein D, Brandt K, *et al.* Development of criteria for the classification of osteoarthritis of the knee. *Arthritis Rheum* 1986;29:1039–49.
4. Altman R, Alarcon G, Appelreuth D, Block D, Borenstein D, Brandt K. The American College of Rheumatology criteria for the classification and reporting of osteoarthritis of the hand. *Arthritis Rheum* 1990;33:505–14.
5. Altman R, Brandt K, Hochberg M, Moskowitz R, Bellamy N, Tugwell P. Design and conduct of clinical trials in patients with osteoarthritis: recommendations from a task force of the Osteoarthritis Research Society. *Osteoarthritis Cart* 1996;4:217–43.
6. Bellamy N, Kirwan J, Boers M. Recommendations for a core set of outcome measures for future phase III. Clinical trials in the knee, hip and hand osteoarthritis. Consensus development at OMERACT III. *J Rheumatol* 1997;24:799–802.
7. Bellamy N. Osteoarthritis clinical trials: candidates' variables and demimetric properties. *J Rheumatol* 1997;24:768–78.
8. Eckstein F, Westhoff J, Sittek H, Maag KP, Haubner M, Faber S. In vivo reproducibility of three-dimensional

- cartilage volume and thickness measurements with MR imaging. *AJR* 1998;170:593–7.
9. Buckland-Wright C, Bird C, Toukin C, Hangartner T, Ritter-Hrncirik C, Cline G, *et al.* X-ray technologist reproducibility in radiography of osteoarthritis knees for a multicenter clinical trial. *Arthritis Rheum* 2001;44(Suppl. 5):385.
 10. Buckland-Wright C, Wolfe F, Ward RJ, Flowers N, Hayne L, Bénite P, *et al.* Substantial superiority of semiflexed (MTP) views in knee osteoarthritis: a comparative radiographic study, without fluoroscopy, of standing extended, semiflexed AP and schuss views. *J Rheumatol* 1999;26:2664–74.
 11. Brandt K, Mazzuca S, Courozier T, Bénite P, Dacre JE, Peterfy CH, *et al.* Which is the best radiographic protocol for a clinical trial of a structure modifying drug in patients with knee osteoarthritis? *J Rheumatol* 2002;29:1308–20.
 12. Vignon E, Garnerio P, Delmas P, Avonac B, Bettica P, Boers M, *et al.* Recommendations for the registration of drugs used in the treatment of osteoarthritis: an update on biochemical markers. *Osteoarthritis Cart* 2001;9:289–93.
 13. Špaček P, Adam M. HPLC method for pentosidine determination in urine, serum and tissues as a marker of glycation and oxidation loading of the organism. *J Liquid Chrom (Rel Technol)* 2002;25(12):1807–20.
 14. Lohmander S, Hoerner LA, Lark W. Metalloproteinases tissue inhibitor and proteoglycan fragments in knee synovial fluid in human osteoarthritis. *Arthritis Rheum* 1993;36:181–9.
 15. Pavelka K, Seibel M. Quantifikation Nachweis Keratansulphat-spezifischer Epitope im Synovialpunktat bei entzündlichen und degenerativen Gelenkerkrankungen. *Z Rheumatol* 1989;48(6):294–300.
 16. Clark AG, Jordan JM, Vilím V, Renner JG, Dragomir A, Luta G, *et al.* Serum cartilage oligomeric matrix protein reflects osteoarthritis presence and severity: The Johnston County osteoarthritis project. *Arthritis Rheum* 1999;42:2356–64.
 17. Woodworth TG, Otterness IG, Johnson K, Pickering E, Saltarelli M, Gaoton M, *et al.* Urinary type II collagen neoepitope in osteoarthritis is associated with disease activity. *Arthritis Rheum* 1999;42(Suppl.):258.
 18. Laurent TC, Larent UBG, Fraser RE. Serum hyaluronan as a disease marker. *Ann Med* 1995;28:241–53.
 19. Spector TD, Hart DJ, Nandra D. Low levels increases in serum C reactive protein are present in early osteoarthritis of the knee and predict progressive disease. *Arthritis Rheum* 1997;40:723–7.
 20. Johansen JS, Hvolris J, Hansen M. Serum YKL-40 levels in healthy children and adults. Comparison with serum and synovial fluid levels of YKL-40 in patients with osteoarthritis or trauma of the knee point. *Br J Rheumatol* 1996;35:553–9.
 21. Naito K, Takahashi K, Suzuji M, Kushida K, Ohismi T, Miura M, *et al.* Measurement of matrix metalloproteinases (MMPs) and tissue inhibitor of metalloproteinase-I (TIMP-I) in patients with generalised osteoarthritis. *Rheumatology* 1999;38:510–5.
 22. Astbury C, Bird HA, Mc Laren AM, Robins SP, *et al.* Urinary excretion of pyridinoline crosslinks of collagen correlated with point damage in arthritis. *Br J Rheumatol* 1994;33:11–5.
 23. Conrozier T, Saxne T, Shan Sei Fan C, Mathieu P, Tron AM, Heinegard D, *et al.* Serum concentrations of cartilage oligomeric matrix protein and bone sialoprotein in hip OA. A one-year study. *Ann Rheum Dis* 1998;9:527–32.
 24. Sharif M, George L, Shepstone J, Thonar E, Kundson V, Cushnaghan J, *et al.* Serum hyaluronic acid level as a predictor of disease progression in osteoarthritis of the knee. *Arthritis Rheum* 1995;38:760–7.
 25. Conrozier T, Chappuis-Cellier C, Richard M. Increase of C reactive protein levels by immunonephelometry in patients with rapidly destructive hip OA. *Rev Rhum (Engl edn)* 1998;12:759–65.
 26. Chavalier X, Conrozier T, Gehrman M, Claudepierre P, Mathieut P, Unger S, *et al.* Tissue inhibitor of metalloproteinase-1 (TIMP-I) serum level may predict progression of the hip osteoarthritis. *Osteoarthritis Cart* 2001;9:300–7.
 27. Pavelka K, Gatterová J, Gollerová V, *et al.* A 5-year randomized controlled, double-blind study of glycosaminoglycan polysulphuric acid complex (Rumalon) as a structure modifying therapy in osteoarthritis of the hip and knee. *Osteoarthritis Cart* 2000;8:335–42.
 28. Lequesne M. Chondrometry quantitative evaluation of point space width and rate of joint space loss in osteoarthritis of the hip. *Rev Rhum (Engl edn)* 1995;62:155–8.
 29. Pavelka K, Gatterová J, Altman RD. Radiographic progression of knee osteoarthritis in a Czech cohort. *Clin Exp Rheum* 2000;18:473–7.
 30. Altman R, Hochberg M, Murphy WA. Atlas of individual radiographic features in osteoarthritis. *Osteoarthritis Cart* 1996;3(Suppl. A):3–70.
 31. Vilím V, Vobúrka Z, Vytášek R, Šenolt L, Tchetverikov I, Kraus VB, *et al.* Monoclonal antibodies to human cartilage oligomeric matrix protein: epitope mapping and characterization of sandwich ELISA *Clin Chim Acta* 2002;3019:1–11.
 32. Mazzuca SA, Brandt KD, Katz BP. Is conventional radiography suitable for evaluation of a disease – modifying drugs in patients with knee osteoarthritis? *Osteoarthritis Cart* 1997;5:217–26.
 33. Working Group Brandt KD. Which is the best radiographic protocol for a clinical trial of a structure-modifying drug in patients with knee osteoarthritis? *J Rheumatol* 2002;29:1308–20.
 34. Matrisian LM. Metalloproteinases and their inhibitors in matrix remodelling. *TIG* 1990;4:121–5.
 35. Reiser KM. Nonenzymatic glycation of collagen in aging and diabetes. *Proc Soc Exp Biol Med* 1998;218:23–7.
 36. Bailey AJ, Paul RG, Knott L. Mechanism of maturation and aging of collagen. *Mech Ageing Dev* 1998;106:1–56.
 37. Miyata T, Veda Y, Horie K, Nangaku M, Tanaka S, Ypersele C, *et al.* Renal catabolism of advanced glycation end products the fate of pentosidine. *Kidney Int* 1998;53:416–22.
 38. Miyata T, Ishiguro N, Yasuda Y, Yasada Y, Ito T, Nangaku M, *et al.* Increased pentosidine, an advanced glycation end products, in plasma and synovial fluid from patients with rheumatoid arthritis and its relation with inflammatory markers. *Biochem Biophys Res Commun* 1998;266:45–9.
 39. De Groot J, Verzijel N, Jacobs MG, Budde M, Bank RA, Bjijsma JW, *et al.* Accumulation of advanced

- glycation end products reduces chondrocyte – mediated extracellular matrix turnover in human articular cartilage. *Osteoarthritis Cart* 2001;9:720–6.
40. Špacek P, Adam M. Pentosidine in osteoarthritis: HPLC determination in body fluids and in tissues. *Arthritis Rheum* 2004.
41. Šenolt L, Vilím V, Braun M, Špaček P, Pavelka K. Pentosidine, well-characterized advanced glycation and product, in serum and synovial fluid in patients with primary knee osteoarthritis (Abstract Lisbon). *Ann Rheum Dis* 2003;129.
-