

cutaneous abnormalities caused by activated RAS and to understand the basic functions of RAS regulation in development, we generated a mouse model in which *Kras* is constitutively active in the skin. Ectodermal activation of *Kras* caused multiple skin abnormalities which phenocopy the spectrum and pattern of cutaneous defects in human Costello syndrome. In the epidermis, activated RAS increased the production of epidermal progenitors leading to an overall expansion of the skin and the appearance of the characteristic Costello redundant skin phenotype. In contrast, we found that *Kras* inhibited hair growth through defects in proliferation. Analysis of genes involved in regulating hair growth revealed a striking downregulation of Sonic hedgehog (*Shh*) gene expression. Likewise, we found that initiation of *Shh* expression during the new hair cycle was also inhibited by *Kras*. These findings suggest that at least two of the defining phenotypic abnormalities in Costello and other RAS-related syndromes involve insufficient expression of *Shh* and more generally, that RAS signals may play a role in a negative feedback inhibition of the *Shh*-signaling center in the hair follicle.

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#### Program/Abstract # 454

##### The LIM-domain binding protein *Ldb1* is required for proper endocardial cushion formation during heart development in *Mus musculus*

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Congenital heart defects are the most common type of major human birth defect, affecting more than 30,000 births in the United States each year. There is clear evidence that LIM-domain binding protein *Ldb1* is crucial for heart formation: *Ldb1* knockout mice never form a heart, and die at E9.5–10.0. Additionally, our evidence suggests that *Ldb1* is required for events throughout heart development. In order to elucidate these roles, we employed a conditional (floxed) knockout of *Ldb1* driven by *Tie2-Cre* (a.k.a. *Tek-Cre*). *Tie2-Cre* is expressed from E9.5 in endothelial tissues. *Tie2-Cre; Ldb1* (floxed) embryos arrest development at E12.5–13.5, and die by E15.5. Through histological and immunohistochemical analyses of the conditionally mutant hearts we have found defects in the atrioventricular (AV) endocardial cushion, the endocardium and the myocardium. The AV endocardial cushion appears hypocellular, while the endocardium is hypercellular. However, there is no increase in apoptosis apparent in the AV cushion. Together with the hypercellular endocardium, the lack of increased cell death suggests rather a failure of the epithelial-to-mesenchyme transition that leads to the observed AV cushion defects. Our results demonstrate that *Ldb1*-mediated transcriptional events are crucial not only during early cardiogenesis, but also for AV endocardial cushion formation and endocardial regulation.

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#### Program/Abstract # 455

##### Monocilia in the embryonic mouse heart imply a direct role for cilia in cardiac morphogenesis

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Blood flow and cardiac function are essential for cardiac morphogenesis: however, how these mechanical signals are sensed by cardiac

cells during development remains unclear. Cilia function as mechanosensors in other fluid-filled organs, thus cilia could also be fluid flow sensors in heart development. They have an indirect role in heart development via the requirement for cilia at the embryonic organizer (node) in the development of global left–right asymmetry. We present evidence that cilia also have a direct role in cardiac morphogenesis after the establishment of LR asymmetry. Cilia are found in the mouse embryo heart at e8.5–e12.5. We demonstrate abnormal development of the endocardial cushions (ECCs) and compact myocardium (CM) in e9.5 mouse embryos with absent cilia due to mutation of the heterotrimeric kinesin component *Kif3a* or abnormal ciliary mechanosensing due to mutation in *polycystin2*. In contrast, hearts from embryos with abnormal LR development due mutation in left–right dynein resulting in paralyzed, but structurally normal cilia, show less penetrant ECC defects and normal CM. These observations support a role for cilia in cardiac development distinct from their early function in LR development. Cilia may function as mechanosensors in heart development, integrating flow, cardiac function and morphogenesis.

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#### Program/Abstract # 456

##### Investigating Bmp-signaling functions in second heart field

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Secondary heart field (SHF) contributes to outflow tract (OFT) and right ventricular myocardium, OFT endocardium, and vascular smooth muscle. Previous data from chick and mouse models implicated that Bmp signaling may play an important role in SHF development. To investigate functions of Bmp signaling in SHF diversification, we inactivated *Bmp2* and *Bmp4* specifically in SHF using conditional null alleles and the *Mef2c* *AHF cre* allele. We also used a doxycycline regulated *Bmp4* allele to induce expanded *Bmp4* specifically in SHF. We found that there are quantitative requirements for *Bmp2* and *Bmp4*-mediated signaling. The most sensitive *Bmp2,4*-responsive event is epithelial mesenchymal transition (EMT) in proximal OFT while expansion of CNC in OFT requires intermediate doses of *Bmp2,4*. Expansion and differentiation of the SHF itself is relatively resistant to loss of *Bmp2,4* signaling while pharyngeal endoderm and branchial arch artery (BAA) remodeling retain normal. *eHand* was abolished in OFT of *Bmp2,4* double loss-of-function mutants, indicating that Bmp signaling are required for CNC patterning and *eHand* could be a direct downstream target of Bmp signaling since it contains several *Smad* binding sites. *Nkx2.5* was dramatically elevated in *Bmp2, 4* double loss-of-function mutants and down-regulated in expanded *Bmp4* mutants, suggesting Bmp signaling negatively regulates *Nkx2.5*. Our findings uncover that *Bmp2* and *Bmp4*-mediated signaling play a crucial positive role in SHF diversification and potentially function by regulating *eHand* and *Nkx2.5*.

**Keywords:** secondary heart field (SHF), outflow tract (OFT), Bmp signaling

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#### Program/Abstract # 457

##### Nodal dependent and independent axis conversions during asymmetric morphogenesis of the zebrafish heart

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