



Chlorophyll *a* concentration across a trophic gradient of lakes: An estimator of phytoplankton biomass?

Peter Kasprzak^{a,b,*}, Judit Padisák^c, Rainer Koschel^{a,b}, Lothar Krienitz^{a,b},
Frank Gervais^{a,d}

^aLeibniz-Institute of Freshwater Ecology & Inland Fisheries, Berlin, Germany

^bDepartment of Limnology of Stratified Lakes, Alte Fischerhütte 2, D-16775 Neuglobsow, Germany

^cDepartment of Limnology, University of Pannonia, P.O.B. 158, H-8200 Veszprém, Hungary

^dDepartment of Limnology of Shallow Lakes, Müggelseedamm 301, D-12587 Berlin, Germany

Received 23 June 2008; accepted 3 July 2008

Dedicated to Prof. Jürgen Benndorf on the occasion of his 65th birthday.

Abstract

Chlorophyll *a* (*chl a*) concentration was evaluated as a predictor of phytoplankton biomass across a broad trophic gradient of lakes (oligotrophic – highly eutrophic). First, a literature survey was conducted to collect information on the proportion of *chl a* in phytoplankton biomass. As a result of this study ($n = 21$) a mean value of $0.505\% \pm 0.197$ S.D. *chl a* per unit wet weight of phytoplankton was calculated. Second, analyses were performed on 756 paired measurements from an unpublished database where the specific *chl a* content of phytoplankton biomass was regressed against phytoplankton standing stocks and *chl a* concentration. Within an interval of $0.1\text{--}50\text{ g m}^{-3}$ of phytoplankton wet weight, a substantial decrease in *chl a* proportion from approximately 2.5% to 0.18% was found. Likewise, the proportion in phytoplankton wet weight decreased from 0.7% to 0.15% across a *chl a* concentration interval of $0.001\text{--}0.150\text{ g m}^{-3}$. These results had a significant impact both on *chl a*-based biomass calculations and the subsequent comparison with phytoplankton biomasses resulting from microscopic counts. Assuming the microscopic method was a measure of the “true” phytoplankton standing stocks, then the precision by which phytoplankton biomass might be predicted based on *chl a* measurements is clearly better when using variable proportions as compared to a constant conversion factor. The same holds for temporal coherence between annual records of phytoplankton biomass. The temporal fit was apparently better when relating the results of microscopic counts and biomass estimation based on variable proportions of *chl a* in phytoplankton biomass. Nevertheless, this effect diminished as the trophic status of the lakes increased. Because of their variable specific *chl a* content, separate taxonomic groups of phytoplankton differently affected the proportion of *chl a* in total

*Corresponding author at: Department of Limnology of Stratified Lakes, Alte Fischerhütte 2, D-16775 Neuglobsow, Germany.
Tel.: +49 33082 69914; fax: +49 33082 69917.

E-mail address: daphnia@igb-berlin.de (P. Kasprzak).

phytoplankton wet weight. Chlorophyceae, Cryptophyceae and cyanobacteria had a high impact, while Bacillariophyceae, Dinophyceae and Chrysophyceae were of lesser importance.

© 2008 Elsevier GmbH. All rights reserved.

Keywords: Phytoplankton; Biomass estimation; Comparison of methods; Microscopic counts; Specific chlorophyll *a* proportion; Conversion factors; Lakes; Trophic gradient

Introduction

Estimating phytoplankton biomass is one of the most useful measurements in limnology and oceanography. Although frequently performed, the approach is not trivial and the results are sometimes hard to interpret (Tolstoy, 1977; Wasmund, 1984; Stich and Brinker, 2005). This is especially true if information from various methods is being compared (Hallegraeff, 1977; Halfson, 1984; Schmid et al., 1998).

Methods to determine phytoplankton standing crops have been developed for quite some time and can be categorised into two general groups: (1) particle counting (Utermöhl, 1923, 1958), and (2) measurement of chemical constituents (Richards and Thompson, 1952; Strickland and Parsons, 1960) with flow-cytometry being a combination of both (Töpel et al., 2004). Over the past decades both approaches were heavily refined and modified. Nevertheless, some of the basic methodological problems have not been resolved (Padisák et al., 1999; Wright et al., 1997). The most important are: (i) methodological flaws, (ii) variable *chl a* proportions per unit phytoplankton biomass, (iii) the taxonomic composition of the phytoplankton community and finally (iv) seasonal aspects.

Microscopic examination and counting of phytoplankton species in collected samples is time-consuming and requires extensive taxonomic experience by the investigator (Banse, 1977; Krienitz et al., 1996). Chemical preservation of the samples can alter the size frequency distribution of the phytoplankton cells (Verity et al., 1992). Moreover, autotrophic picoplankton (APP) may sometimes contribute significantly to total phytoplankton biomass but are not often recorded (Padisák et al., 1997). To overcome these problems, particle counters and image analysis systems have been utilised, but their performance in estimating phytoplankton biomass as compared to microscopic methods is still questioned (Hillebrand et al., 1999).

Concerning *chl a* extraction and the subsequent photometric or HPLC measurements, several authors have shown that there is no ideal protocol (Párista et al., 2002; Stich and Brinker, 2005). Depending on the taxonomic structure of the phytoplankton sample being analysed, different extraction solvents may have different extraction efficiencies (Vollenweider, 1974; Wright et al., 1997).

Finally, various studies have found that *chl a* content per unit of phytoplankton biomass decreases as phytoplankton standing stocks increase (Desortova, 1981; Ahlgren, 1983; Wojciechowska, 1989; Watson et al., 1992; Talling, 1993; Chow-Fraser et al., 1994; Schmid et al., 1998; Felip and Catalan, 2000; Sandu et al., 2003; Kiss et al., 2006). This phenomenon may be influenced by lake trophic status (Harris, 1986), phytoplankton community structure (Bursche, 1961; Nusch and Palme, 1975), the size frequency distribution of the algal cells (Watson and McCauley, 1988), and by seasonal shifts within the plankton community (Loth, 1985; Vanni et al., 1993).

Notwithstanding these problems and limitations, we examined whether *chl a* concentration across a trophic gradient of lakes (oligotrophic – eutrophic) can be used as a predictor of phytoplankton biomass. *Chl a*-based calculations of phytoplankton biomass were performed by applying constant conversion factors as determined from the literature and by using variable ratios gained from a comprehensive database of the Leibniz-Institute of Freshwater Ecology & Inland Fisheries (IGB, Neuglobsow, Germany). Moreover, we tested the precision and temporal coherence by which time series of phytoplankton biomass of various lakes can be predicted using these conversion factors as compared to the results of microscopic counts.

Material and methods

Investigation sites

The five lakes included in this study are located within the eastern part of Germany's glacial Baltic lake region (53°15'N, 13°10'E) approximately 100 km north of Berlin. They are seepage lakes with ground water and rainfall being the major sources of water. The lakes thermally stratify from May until at least September. Mean temperature of the mixed layer varies between 4 °C (January) and 20 °C (August). Global radiation ranges between 200 J cm⁻² d⁻¹ (December) and 1700 J cm⁻² d⁻¹ (June; German Weather Service, unpublished results). The lakes have significantly different morphometric and chemical characteristics. Their trophic status spans from oligotrophic to highly eutrophic (cf. Table 1). For more information about the five study lakes see Casper (1985), Kasprzak et al.

Table 1. Morphometric and chemical characteristics of the investigation sites according to Koschel et al. (1985), Casper (1985), Koschel and Kasprzak (1994) and unpublished data of the authors

Feature	Stechlin	Kleiner Väter	Großer Väter	Tiefwaren	Feldberger Haus
Area (km ²)	4.23	0.10	0.12	1.41	1.36
Volume (10 ⁶ m ³)	98.70	0.51	0.63	13.80	8.15
Mean depth (m)	23.3	5.0	5.2	9.6	6.0
Maximum depth (m)	69.5	13.3	11.5	23.0	12.0
Sampled layer (m)	0–20	0–6	0–6	0–5	0–5
Total P (µg L ⁻¹)	8.6–16.7	16.2–22.7	18.6–29.2	24.8–124.6	93.1–1292.5
Total N (µg L ⁻¹)	300–585	750–820	860–950	980–1200	1300–1900
Chlorophyll <i>a</i> (mean) (µg L ⁻¹)	0.7–9.5 (2.3)	1.1–8.1 (4.0)	0.9–15.6 (4.6)	1.4–29.5 (6.2)	1.3–175.9 (24.2)
Secchi transparency (m)	8.4	4.3	3.9	4.7	1.8
Trophic status	Oligotrophic	Mesotrophic	Mesotrophic	Meso-eutrophic	Eutrophic

Total phosphorus and nitrogen concentrations represent the minimum and maximum of annual means, respectively, within the mixed layer of the lakes. Concerning chlorophyll *a* concentration, the grand mean of all observations is additionally denoted in parenthesis. The classification of the trophic status was conducted following OECD recommendations (Premazzi and Chiaudani, 1992). Water clarity was calculated based on all available Secchi readings.

(2000), Kasprzak et al. (2003), Koschel and Adams (2003) and Koschel et al. (2006).

Field sampling

Data used in this study were collected during 1985–2006 from the deepest location of each lake. However, the number of investigation years and consequently the number of samples were different (min. Tiefwareensee, $n = 74$, max. Feldberger Haussee, $n = 265$, cf. Table 3). During most years, samples were taken at least once a month although samples were also collected biweekly from May until September in a number of years. Samples were either taken at discrete depths or as composite samples, and as such roughly represented the euphotic zone of the study lakes (cf. Table 1). All samples were split for *chl a* and phytoplankton biomass measurements.

Field sampling and sample treatment followed widely accepted protocols (see below). Still, over the years technical equipment (e.g. microscopes, sedimentation chambers, photometers and chemicals) varied and several people performed the analyses (especially phytoplankton counts). For this study, we did not compare results produced during various periods or by different investigators. Rather, we assumed that the methods had no major bias and correctly reflected the magnitude of each value in the time series.

Chlorophyll *a* determination

Chl a samples were processed following recommendations of the German Standard Methods of Water and Wastewater Analyses (Deutsche Einheitsverfahren, 1983–1985). Appropriate aliquots (200–2000 ml) were either filtered through membrane filters (pore size

1.2 µm; Lake Stechlin, Tiefwareensee, Feldberger Haussee) or through Whatman GF/C glass fibre filters (Kleiner Vätersee, Großer Vätersee). If not analysed immediately, the two types of filters were frozen at -20 and -80 °C, respectively, until further treatment. Prior to extraction, filters were homogenised to enhance extraction efficiency. Extraction was performed by either adding 99.8% acetone (Lake Stechlin, Tiefwareensee, Feldberger Haussee) or 90% hot ethanol (78 °C; Kleiner Vätersee, Großer Vätersee) followed by either filtration or centrifugation to produce a supernatant with minimal turbidity. For Lake Stechlin, Tiefwareensee and Feldberger Haussee, the absorbance of the processed samples was recorded at three different wavelengths (630, 665 and 750 nm) following the protocol of Strickland and Parsons (1960) for calculating *chl a* concentration. For Kleiner Vätersee and Großer Vätersee, *chl a* concentration was determined using the absorption maximum of 665 nm exclusively.

Phytoplankton biomass estimation

Phytoplankton biomass was estimated in four separate ways: (1) by microscopic counts, (2) based on a constant *chl a* proportion in phytoplankton wet weight, (3) by applying variable *chl a* ratios in phytoplankton wet weight as related to phytoplankton biomass and (4) by using variable *chl a* ratios in phytoplankton wet weight as related to *chl a* concentration.

Microscopic counts

Phytoplankton samples were preserved in Lugol's solution and counted under an inverted microscope (Utermöhl, 1958). Biovolume was calculated based on cell- or colony volumes suggested by several authors (Willén, 1992; Hamilton, 1990; Gosselain and Hamilton, 2000; Hoehn et al., 1998) and subsequently converted

into wet weight assuming a specific gravity of 1 g cm^{-3} (Rott, 1978). APP was preferably counted immediately after sampling in unpreserved samples. If counting was not possible, either unpreserved samples were deep-frozen and enumerated within one month or sub-samples preserved in 0.2% formaldehyde were filtered on to $0.4 \mu\text{m}$ pore size polycarbonate membranes and processed later. APP was only estimated for Lake Stechlin, Kleiner Vätersee and Großer Vätersee. Although its biomass may increase as phytoplankton standing stocks rise, the proportion of APP to total phytoplankton tended to diminish as the lake trophic status increased from oligotrophic to eutrophic (see Fig. 2c, p. 72, in Callieri and Stockner, 2002). With respect to the total phosphorus concentration of Tiefwareensee and highly eutrophic Feldberger Haussee, the proportion of APP within total phytoplankton may range between 3% and 5%. We therefore decided to not correct any of the phytoplankton measurements collected from these lakes. For further details concerning phytoplankton analysis see Krienitz et al. (1996), Gervais et al. (1997), Padisák et al. (1997) and Padisák et al. (2003).

Constant *chl**a*/wet weight ratio as related to phytoplankton biomass

Based on the results of 21 studies reported in the literature a mean proportion of *chl**a* in phytoplankton wet weight was estimated. Using this factor the *chl**a* data considered in our study were converted into phytoplankton wet weight. Numbers on specific *chl**a* content originate from North American and European lakes and reservoirs, respectively. Their trophic status represents a broad spectrum ranging from oligotrophic to eutrophic conditions. In two instances information collected from algal cultures was integrated.

Variable *chl**a*/wet weight ratio as related to phytoplankton biomass

A total of 756 paired observations (IGB database) on the ratio of *chl**a*/phytoplankton wet weight was regressed over phytoplankton wet weight. The regression function was subsequently used to calculate phytoplankton biomass.

Variable *chl**a*/wet weight ratio as related to *chl**a* concentration

Based on the same collection of 756 paired observations (IGB database) the ratio of *chl**a*/phytoplankton wet weight was regressed against *chl**a* concentration. By applying the regression function, phytoplankton biomass was estimated.

Microscopic estimates of phytoplankton biomass are designated BM_{count} . Phytoplankton biomass calculated by constant or variable per wet weight *chl**a* proportions

are denoted $\text{BM}_{\text{chl}a\text{-con}}$, $\text{BM}_{\text{chl}a\text{-var}(1)}$, and $\text{BM}_{\text{chl}a\text{-var}(2)}$, respectively.

Statistical analyses

To qualify and compare different ways of calculating phytoplankton biomass, Pearson's product-moment regression coefficient (r), mean systematic deviation (MSD, bias) and standard deviation (S.D.) were calculated. Regression analysis was performed in order to relate the results of phytoplankton calculations based on *chl**a* measurements ($\text{BM}_{\text{chl}a\text{-con}}$, $\text{BM}_{\text{chl}a\text{-var}(1)}$, $\text{BM}_{\text{chl}a\text{-var}(2)}$) and microscopic counts (BM_{count}). A graphical check for homoscedasticity indicated variances were not homogeneous. Therefore, prior to regression analysis both *chl**a* concentration and phytoplankton biomass values were \log_{10} -transformed. Kolmogorov–Smirnov one-sample-test indicated the residuals of the \log_{10} -transformed data to be approximately normal distributed. Spearman rank correlation coefficient was applied to comprehend the impact of separate taxonomic groups of phytoplankton on the relation of *chl**a*/ BM_{count} as related to BM_{count} . All statistical calculations were performed using SPSS 9.0 (SPSS Inc., Chicago, Bühl and Zöfel, 2000).

Results

The three ways of calculating the proportion of *chl**a* in phytoplankton wet weight (*chl**a*/ BM_{count}) led to remarkably different findings. The literature survey results indicated the *chl**a*/ BM_{count} content of several phytoplankton populations (Table 2). Depending on the parameter used for the calculation, the proportion amounted to 0.505% (mean) and 0.447% (median). Overall the results varied between 0.158% (minimum) and 0.900% (maximum).

In Fig. 1 (upper panel), the ratio of *chl**a*/ BM_{count} is plotted against BM_{count} for the 756 paired observations obtained from the IGB data collection. Over a range of BM_{count} roughly spanning between 0.1 and 50 g ww m^{-3} , a substantial decrease in specific *chl**a* content from approximately 2.5% to 0.18% was found (cf. Eq. (1)). Likewise, within a concentration interval of 0.001– 0.150 g m^{-3} *chl**a* (Fig. 1, lower panel) the proportion of *chl**a*/ BM_{count} decreased from approximately 0.7% to 0.15% (cf. Eq. (2)):

$$\log\left(\frac{\text{chl}a}{\text{BM}_{\text{count}}}\right) = -0.403 - 0.482 \log \text{BM}_{\text{count}} + 0.229(\log \text{BM}_{\text{count}})^2 - 0.040(\log \text{BM}_{\text{count}})^3$$

$$(r^2 = 0.550, \text{ S.D.} = 0.243, P < 0.001, \text{ chl}a - \text{g m}^{-3}, \text{ BM}_{\text{count}} - \text{g ww m}^{-3}), \quad (1)$$

Table 2. Proportion of chlorophyll *a* (chl_a) in phytoplankton wet weight (ww) calculated based on published literature information

chl _a /ww [%]	<i>n</i>	Parameter	Reference
0.900	25	Median	Nicholls and Dillon (1978, p. 146), Table 3, lakes, natural phytoplankton
0.560	4	Mean	Tolstoy (1977, p. 16), lakes, natural phytoplankton (10–90 μg L ⁻¹)
0.420	7	Median	Vörös and Padišák (1991, p. 113), Table 1, lakes, natural phytoplankton (0.1–238 μg L ⁻¹)
0.691	19	Median	Reynolds (1984, p. 30), Table 4, lakes, single species of natural populations
0.630	4	Mean	Rott (1978, p. 16), Table 1, lakes, natural phytoplankton (3.2–5.5 μg L ⁻¹)
0.230	16	Median	Wojciechowska (1989, p. 66), Table 1, lakes, natural phytoplankton (3–12 μg L ⁻¹)
0.690	4	Mean	Loth (1985, p. 323), Table 1, reservoirs, natural phytoplankton (1–8 μg L ⁻¹)
0.720	11	Median	Desortova (1981, p. 160), Table 1, reservoirs, natural phytoplankton (2.4–23.1 μg L ⁻¹)
0.364	1	–	Schellenberger et al. (1985, p. 219), lakes, natural phytoplankton
0.447	14	Median	Chow-Fraser et al. (1994, pp. 2060–2061), Figs. 6b and 9, lakes, natural phytoplankton (0.6–100 μg L ⁻¹)
0.355	6	Median	Watson et al. (1992, p. 2607), Fig. 1b, lakes, natural phytoplankton (1–100 μg L ⁻¹)
0.390	96	Median	Bursche (1961, pp. 617–645), Tables. 2–11, laboratory cultures, various species (10–70 μg L ⁻¹)
0.280	33	Median	Montagnes et al. (1994, p. 1047), Table 1, laboratory cultures, various species (0.1–100 μg L ⁻¹)
0.780	23	Median	Ahlgren (1983, p. 497), Table 3, lakes, natural phytoplankton (6–118 μg L ⁻¹)
0.600	17	Median	Schmidt et al. (1998, p. 1658), Fig. 1a, lakes, natural phytoplankton (5–25 μg L ⁻¹)
0.158	38	Median	Sandu et al. (2003, S. 393), Table 2, lakes, natural phytoplankton (13.3–125.3 μg L ⁻¹)
0.643	1	–	Felip and Catalan (2000, p. 91–105), lake, natural phytoplankton (1.1–5.7 μg L ⁻¹)
0.619	20	Median	Kiss et al. (2006, p. 2052), Table 1, lakes, natural phytoplankton (8.7–64.8 μg L ⁻¹)
0.400	5	Mean	Lampert and Schober (1978, p. 370), Abb. 2b, lake, natural phytoplankton (10–35 μg L ⁻¹)
0.420	24	Median	Talling (1993, p. 90), Table 3, lakes, natural phytoplankton (1–120 μg L ⁻¹)
0.300	1	–	Padišák et al. (1999, p. 369)
0.505	Mean		
0.197	S.D.		
0.447	Median		
0.158	Minimum		
0.900	Maximum		

n refers to the number of observations derived from the references cited. Regardless of *n*, the results concerning chl_a/ww [%] were not frequency-weighted but used as separate numbers to calculate grand mean, median and standard deviation. In many cases the values have been estimated from chlorophyll *a* and cell volume data given in figures and tables. When provided, the interval of chlorophyll *a* concentration is given in parenthesis.

$$\log\left(\frac{\text{chl}_a}{\text{BM}_{\text{count}}}\right) = -0.979 - 0.282 \log \text{BM}_{\text{count}}$$

$$(r^2 = 0.164, \text{ S.D.} = 0.027, P < 0.001,$$

$$\text{chl}_a - \text{g m}^{-3}, \text{ BM}_{\text{count}} - \text{g ww m}^{-3}). \quad (2)$$

Spearman's rank correlation coefficient pointed to different effects of separate taxonomic groups of phytoplankton to the changes in the ratio of chl_a/BM_{count}. Except for chrysophyceans all other taxa significantly affected the specific chl_a content of phytoplankton wet weight ($p < 0.001$; Chlorophyceae: -0.436 , Cryptophyceae: -0.268 , cyanobacteria: -0.214 , Bacillariophyceae: -0.208 , Dinophyceae: -0.170 and Chrysophyceae -0.062).

The results portrayed above are clearly reflected in Fig. 2 where phytoplankton wet weight was calculated based on chl_a content (BM_{chl_a-con}, BM_{chl_a-var(1)}, BM_{chl_a-var(2)}) and regressed against the results of microscopic counts (BM_{count}). If related to BM_{count}, the before mentioned constant proportion of 0.505% chl_a per unit phytoplankton wet weight resulted in a significant under-

estimation of phytoplankton biomass and a greater variability along the regression line (Fig. 2, upper panel, slope 0.617, $p < 0.0001$, S.E. 0.013, adjusted r^2 0.729). When applying variable proportions of chl_a per unit phytoplankton wet weight (Eq. (1)), the ratio between BM_{chl_a-var(1)} and BM_{count}, respectively, was close to 1, and the variability along the regression line was smaller (Fig. 2, central panel, slope 1.036, $p < 0.0001$, S.E. 0.012, adjusted r^2 0.906). Concerning the results calculated using Eq. (2), the difference between BM_{chl_a-var(2)} and BM_{count} was somewhat smaller (Fig. 2, lower panel, slope 0.781, $p < 0.0001$, S.E. 0.027, adjusted r^2 0.722) as was found for BM_{chl_a-con}. Still, the estimated phytoplankton biomass was clearly lower than BM_{count}.

The predictive power of BM_{chl_a-con}, BM_{chl_a-var(1)} and BM_{chl_a-var(2)} as related to BM_{count} are summarised in Table 3. Looking at the results obtained from all lakes, the closest relationship was found between BM_{count} and BM_{chl_a-var(1)}. This was true for correlation (r 0.952), MSD (-0.003) and variability (S.D. 0.217). The quality of the prediction of phytoplankton biomass based on BM_{chl_a-con} and BM_{chl_a-var(2)} was less precise.

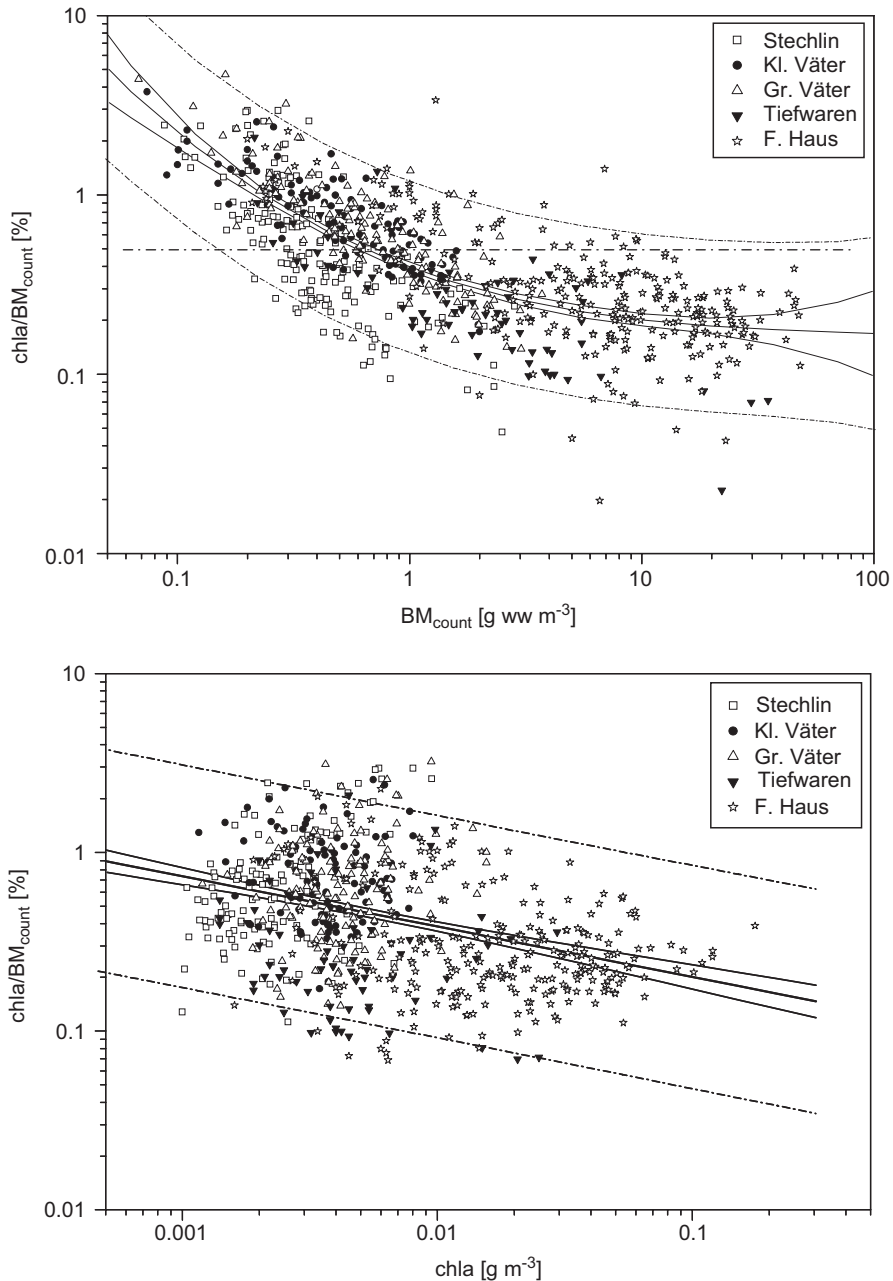


Fig. 1. Proportion of chlorophyll *a* per phytoplankton wet weight ($\text{chla}/\text{BM}_{\text{count}}$) regressed against phytoplankton wet weight (BM_{count} , upper panel) as obtained from microscopic counts and against chla concentration (lower panel, including 95% confidence bands and prediction bands of the regression line, cf. Eqs. (1) and (2)). The dotted horizontal line in the upper panel indicates the 0.505% constant ratio of per wet weight chlorophyll *a* content in phytoplankton biomass (cf. Table 2).

Furthermore, the difference in the predictive power of $\text{BM}_{\text{chla-con}}$ and $\text{BM}_{\text{chla-var}(2)}$ was small. Clear differences were only found concerning MSD where $\text{BM}_{\text{chla-var}(2)}$ was superior to $\text{BM}_{\text{chla-con}}$. Looking at separate lakes, the results were varying. Nevertheless, even then a clear tendency of better correlation, smaller deviation and lesser variation was detected when phytoplankton biomass was calculated based on $\text{BM}_{\text{chla-var}(1)}$ as compared to the use of $\text{BM}_{\text{chla-con}}$ and $\text{BM}_{\text{chla-var}(2)}$.

Qualified by Pearson's product-moment correlation coefficient (r), Fig. 3 rates the temporal coherence between BM_{count} and $\text{BM}_{\text{chla-con}}$, $\text{BM}_{\text{chla-var}(1)}$ and $\text{BM}_{\text{chla-var}(2)}$, respectively, as was found for the time series of separate lakes. It is obvious that by using $\text{BM}_{\text{chla-var}(1)}$ a higher temporal coherence could have been achieved as compared to the application of $\text{BM}_{\text{chla-con}}$ or $\text{BM}_{\text{chla-var}(2)}$. Moreover, this tendency was less pronounced in eutrophic lakes.

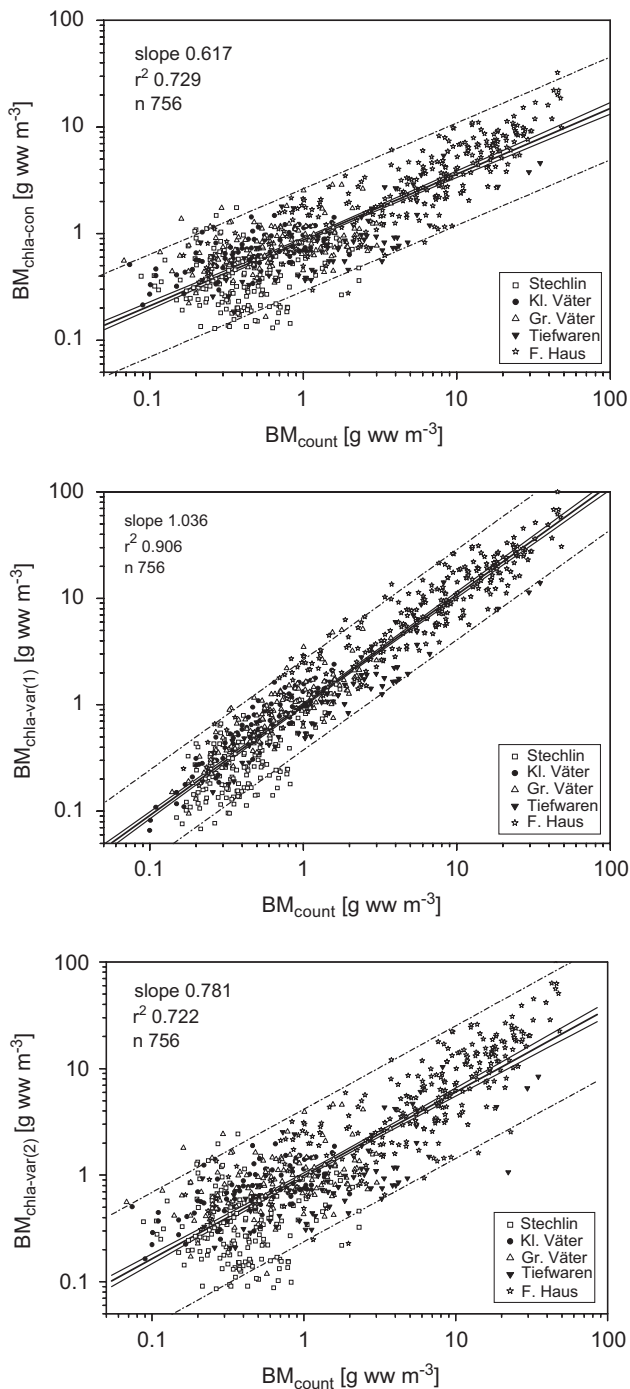


Fig. 2. Relation between phytoplankton wet weight based on microscopic counts (BM_{count}) and phytoplankton wet weight calculated using constant ($BM_{\text{chla-con}}$, upper panel) and variable ($BM_{\text{chla-var}(1)}$, central panel, $BM_{\text{chla-var}(2)}$, lower panel) proportions within phytoplankton biomass, respectively.

Finally, the example of three separate lakes (Fig. 4) provides specific information on the annual record of temporal coherence between BM_{count} , $BM_{\text{chla-con}}$, $BM_{\text{chla-var}(1)}$ and $BM_{\text{chla-var}(2)}$. Again, as indicated by Pearson's product-moment correlation coefficient (r), there was a general tendency of higher temporal

coherence when using, $BM_{\text{chla-var}(1)}$ as compared to $BM_{\text{chla-con}}$ and $BM_{\text{chla-var}(2)}$. The best fit of annual records in individual lakes was found for Feldberger Haussee in the year 2000 (r 0.989).

Overall the conclusion seems reasonable that $BM_{\text{chla-var}(1)}$ was clearly a better estimator of phytoplankton biomass than $BM_{\text{chla-con}}$ or $BM_{\text{chla-var}(2)}$. Calculations based on $BM_{\text{chla-var}(1)}$ and microscopic counts (BM_{count}) lead to similar results both with respect to precision and temporal coherence. In contrast, estimation of phytoplankton biomass using $BM_{\text{chla-con}}$ resulted in significant over- (oligotrophic lakes) and underestimation (mesotrophic, eutrophic lakes) of phytoplankton biomass, respectively. As compared to $BM_{\text{chla-con}}$, phytoplankton biomass calculated from $BM_{\text{chla-var}(2)}$ was more similar to the results of microscopic counts. Still, the deviations in terms of precision and temporal coherence were clearly greater as was found for $BM_{\text{chla-var}(1)}$. Of course, these conclusions can only be considered valid if BM_{count} represented the "true" phytoplankton biomass.

Discussion

The microscopic elaboration of phytoplankton samples including subsequent calculation of algal biomass are labour-intensive and require sound taxonomic skills of the investigator (Utermöhl, 1958; Hillebrand et al., 1999). Consequently, *chl a* concentration began being used as a quick and easy-to-measure surrogate of phytoplankton biomass (e.g. Richards and Thompson, 1952; Strickland and Parsons, 1960; Kamoto, 1966; Dillon and Rigler, 1974). Nevertheless, besides the fact that *chl a* measurements lack any information on phytoplankton community structure, our investigations indicate that constant conversion factors are inappropriate for calculating phytoplankton biomass. In oligotrophic lakes, *chl a*-based phytoplankton biomass calculations tend to be higher than biomass derived from microscopic counts, while an increasing trend of underestimation was found for mesotrophic and eutrophic lakes, respectively. Using constant conversion factors, in a study on the relation of *chl a* content and phytoplankton biomass in eutrophic Danube delta lakes, Sandu et al. (2003) found significantly higher results by microscopic counts as compared to estimates based on constant *chl a* proportions.

Taking into account the 756 IGB paired observations of *chl a*/ BM_{count} , the conclusion instead seems reasonable for a functional relationship between *chl a* and BM_{count} . The higher BM_{count} the lower is the proportion of *chl a* per unit of BM_{count} . Beyond the above-mentioned multitude of variables (cf. Introduction section), there are two major factors responsible for dropping *chl a*/ BM_{count} as BM_{count} increases.

Table 3. Statistical indices qualifying the results of various approaches to calculate phytoplankton biomass based on *chl a* measurements as compared to the results of microscopic counts

Lake	<i>chl a</i> /BM _{count} (%) min., max. (mean)	BM _{count} vs. BM _{chl a-con}			BM _{count} vs. BM _{chl a-var(1)}			BM _{count} vs. BM _{chl a-var(2)}			<i>n</i>
		<i>r</i>	MSD	S.D.	<i>r</i>	MSD	S.D.	<i>r</i>	MSD	S.D.	
Stechlin (o)	0.047–2.954 (0.699)	0.045	0.038	0.361	0.624	0.172	0.229	0.126	0.087	0.408	170
K. Väter (m)	0.173–3.757 (0.860)	0.639	−0.126	0.248	0.918	−0.069	0.128	0.499	−0.154	0.246	103
G. Väter (m)	0.138–4.676 (0.796)	0.443	−0.053	0.305	0.847	−0.071	0.166	0.387	−0.095	0.319	144
Tiefwaren (eu)	0.023–2.093 (0.344)	0.562	0.188	0.647	0.930	0.147	0.173	0.582	0.133	0.637	74
F. Haus (eu)	0.020–3.372 (0.380)	0.801	0.286	0.319	0.898	−0.085	0.189	0.798	0.076	0.331	265
All lakes	0.020–4.676 (0.594)	0.853	0.100	0.396	0.952	−0.003	0.217	0.850	0.020	0.390	756

BM_{count} – phytoplankton wet weight obtained from microscopic counts, BM_{chl a-con} – phytoplankton wet weight assuming a constant ratio between chlorophyll *a* and phytoplankton wet weight (*chl a*/ww = 0.505%, cf. Table 2), BM_{chl a-var(1)} – phytoplankton wet weight assuming a variable ratio between chlorophyll *a* and phytoplankton wet weight (cf. Fig. 1, upper panel, Eq. (1)), BM_{chl a-var(2)} – phytoplankton wet weight assuming a variable ratio between chlorophyll *a* and phytoplankton wet weight (cf. Fig. 1, lower panel, Eq. (2)), *r* – Pearson’s product moment correlation coefficient, MSD – mean systematic deviation (bias), S.D. – standard deviation, *n* – number of observations. The acronyms in the first column of the table refer to oligotrophic (o), mesotrophic (m) and eutrophic (eu).

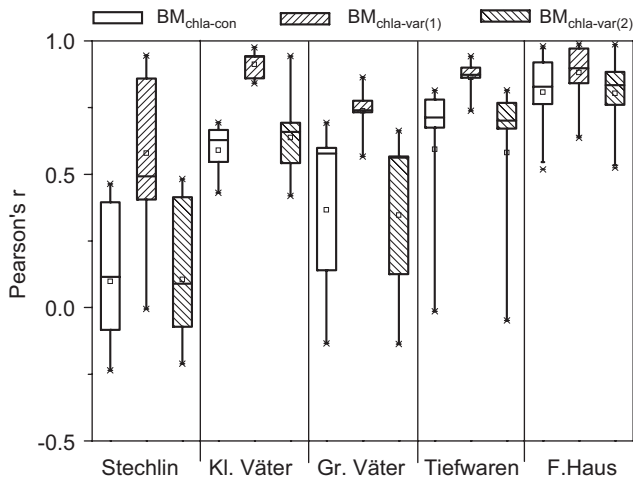


Fig. 3. Temporal coherence as qualified by Pearson’s product-moment correlation coefficient (*r*) of annual phytoplankton biomass records, estimation based on microscopic counts (BM_{count}) and on the assumption of constant (BM_{chl a-con}) and variable (BM_{chl a-var(1)}, BM_{chl a-var(2)}) ratios of per wet weight chlorophyll *a* content in phytoplankton biomass calculated for separate lakes. Number of recorded lake years: Lake Stechlin – 10, Kleiner Vätersee – 6, Großer Vätersee – 8, Tiefwareensee – 5 and Feldberger Haussee – 18. Box-whisker-plots indicate minimum, maximum, percentiles (10, 25, 75, 90%), mean and median of (*r*).

First, some studies provide convincing evidence that the proportion of *chl a* per unit of BM_{count} is inversely related to algal cell- or colony volume (Harris, 1986; Wojciechowska, 1989; Vörös and Padisák, 1991). Therefore, a given biomass unit of “small” algal cells is likely to contain more *chl a* than does the same amount of “big” phytoplankton cells. Because increasing mean cell- or colony volume is characteristic of eutrophic lakes (Watson and McCauley, 1988), decreasing *chl a*/BM_{count} is an apparent consequence of nutrient enrich-

ment (Watson et al., 1992; Chow-Fraser et al., 1994). Also, because mean phytoplankton cell size typically increases during seasonal succession (Sommer et al., 1986), seasonal aspects might be relevant as well.

Furthermore, besides our own investigation a number of other studies indicate decreasing *chl a*/BM_{count} in natural phytoplankton populations as BM_{count} increases (Desortova, 1981; Wojciechowska, 1989; Talling, 1993; Schmid et al., 1998; Felip and Catalan, 2000; Kiss et al., 2006). This trend does seem to be valid both for natural phytoplankton communities but also for laboratory cultures of certain species as well (*Planktothrix agardhii*; Ahlgren, 1983). Reynolds (2006) argues that a high surface-to-volume ratio is beneficial for nutrient acquisition. Because nutrient limitation tends to diminish as nutrient concentrations rise, eutrophication might be the ultimate factor allowing the mean particle size of natural phytoplankton communities to increase. Thus, growing phytoplankton biomass in concert with increasing mean cell- or colony volume might necessarily result in decreasing *chl a*/BM_{count} ratio as the lakes develop from oligotrophic into eutrophic conditions.

The impact of species-specific differences on *chl a* per unit of phytoplankton biomass is controversial. Reynolds (1984, p. 38) reported on significant differences in volume-related *chl a* proportion between different taxonomic groups of phytoplankton. Working with laboratory cultures, Bursche (1961) and Nusch and Palme (1975) found significant differences between systematic groups as well with highest proportions in green algae and lowest in cyanobacteria. Desortova (1981) and Talling (1993) confirmed these result for natural phytoplankton communities during those periods when one or another taxonomic group dominated. Kohl and Nicklisch (1988) concluded that because of the structure of the light-harvesting pigment–protein complex, the specific *chl a* content is high in green algae, low

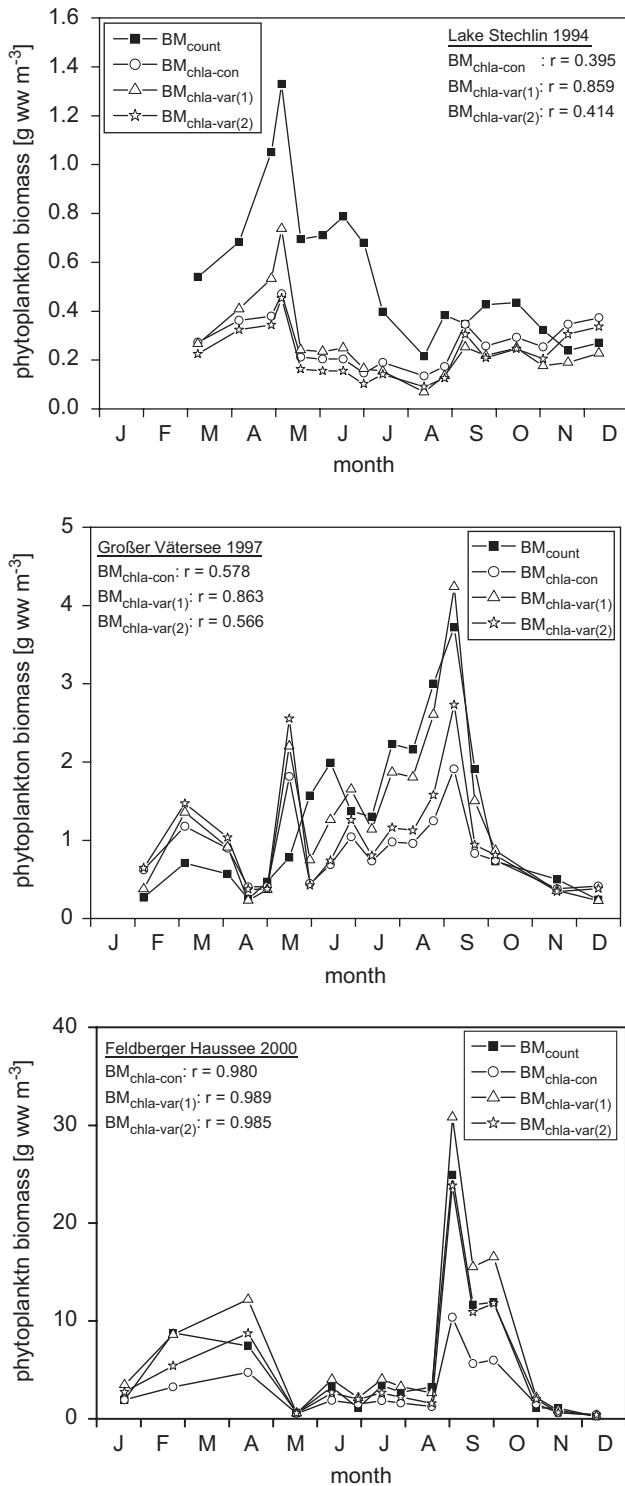


Fig. 4. Selected examples of annual records of phytoplankton standing stocks in various lakes estimated based on microscopic counts (BM_{count}) and on the assumption of constant ($BM_{chla-con}$) and variable ($BM_{chla-var(1)}$, $BM_{chla-var(2)}$) ratios of per wet weight chlorophyll *a* content in phytoplankton biomass, respectively. Pearson's product-moment correlation coefficient (r) has been used to qualify temporal coherence.

in cyanobacteria and intermediate in chromophyta. However, other studies doubt the impact of taxonomic composition of natural phytoplankton communities on $chla/BM_{count}$ ratio (Schellenberger et al., 1985; Vörös and Padisák, 1991).

Nutrient enrichment of lakes is usually accompanied by characteristic shifts within the phytoplankton community. During eutrophication small flagellated taxa are replaced by increasing proportions of green algae, with cyanobacteria finally predominating (Reynolds, 1984; Richman et al., 1984; McQueen et al., 1986). From algological studies concerning the lakes under consideration it is known that the phytoplankton population of eutrophic Feldberger Haussee during a number of years was occupied by filamentous cyanobacteria (Kasprzak et al., 1993). Especially during the early 1990s, sometimes 90% of total phytoplankton biomass was represented by this group (Krienitz et al., 1996). Therefore, a high percentage of cyanobacteria within total phytoplankton biomass may additionally have contributed to low $chla/BM_{count}$ when BM_{count} was high. Nevertheless, our study at least from a statistical point of view indicates that the proportion of other major taxonomic groups (Chlorophyceae, Cryptophyceae) may also significantly affect the ratio of $chla/BM_{count}$. The missing impact of chrysophyceans, however, might be a result of mixotrophic nutrition (Jones and Rees, 1994) and large relative proportions during the clear-water phase (Fott et al., 1980). Within such periods, photosynthesis is not the principal source of carbon acquisition of the phytoplankton community. Therefore, a minor influence of this group on $chla/BM_{count}$ is likely.

Our results also point out a closer temporal coherence between BM_{count} on one hand, and $BM_{chla-con}$, $BM_{chla-var(1)}$, and $BM_{chla-var(2)}$ on the other hand, as the lakes develop from oligotrophic into eutrophic conditions. Moreover, temporal coherence clearly increased when phytoplankton biomass was calculated by variable $chla$ proportions as opposed to constant ratios. At least to some extent these differences might be a result of methodological shortcomings. Some authors criticised the extraction method of $chla$ estimation in combination with photometric extinction measurements for being imprecise when $chla$ concentration was low (Vollenweider, 1974; Hallegraeff, 1977; Schmid et al., 1998). Additionally, our study has shown that the proportion of $chla$ within phytoplankton biomass may significantly decrease as phytoplankton biomass increases. Since $chla$ represents only a minor proportion of phytoplankton biomass, biased measurements may result in significant miscalculations of phytoplankton standing stocks (Tolstoy, 1977; Ahlgren, 1983). Furthermore, methodological problems related to microscopic examinations of phytoplankton samples in concert with incorrect calculation of phytoplankton biovolume may additionally contribute to poor temporal coherence and

precision of the measurements (Utermöhl, 1958; Verity et al., 1992; Hillebrand et al., 1999).

In summary we conclude that chlorophyll *a* concentration might be used with caution as a predictor of phytoplankton biomass. Regardless of whether constant or variable proportions of *chl a* have been applied to calculate phytoplankton standing stocks ($BM_{chl a-con}$, $BM_{chl a-var(1)}$, $BM_{chl a-var(2)}$), for extensive numbers of observations ($n = 756$) collected across a broad trophic gradient of lakes (oligotrophic – eutrophic) we found a close statistical correlation with the results of microscopic biomass estimation ($r^2 = 0.729, 0.906$ and 0.722). However, if the results of microscopic estimates (BM_{count}) are considered the “true” phytoplankton biomass, constant proportions of chlorophyll *a* ($BM_{chl a-con}$) tend to either over- or underestimate BM_{count} depending on the trophic status of the lakes. In contrast, estimations based on variable proportions of chlorophyll *a* ($BM_{chl a-var(1)}$) and BM_{count} were found to be similar. Moreover, using BM_{count} as a reference, our results applied to annual records of phytoplankton in lakes of various trophic status clearly indicate, that $BM_{chl a-var(1)}$ is a much better predictor than $BM_{chl a-con}$. The results using $BM_{chl a-var(2)}$ are ambiguous. Although the correlation with BM_{count} is somewhat better as compared to $BM_{chl a-con}$, the calculated biomass is clearly lower as was found for microscopic counts. Nevertheless, both precision and temporal coherence of the prediction are the better the higher the trophic status of a given lake no matter if it is based on $BM_{chl a-con}$, $BM_{chl a-var(1)}$ or $BM_{chl a-var(2)}$.

Acknowledgements

Thanks to M. Papke, G. Heisig, K. Kalis, R. Kruspe, M. Sachtleben and R. Degebrot for sample collection and sample treatment. We are grateful to J. Gladitz and T. Petzoldt for support in statistical analyses. The results used in this study have in part been produced within several projects supported by the Bundesministerium für Bildung und Forschung (BMBF), by the German Research Foundation (DFG) and by grants provided by the Bundesland Mecklenburg-Vorpommern. Thanks also to the German Weather Service (Offenbach, Neuglobsow) for providing unpublished data. Richard C. Lathrop significantly enhanced language and style of the manuscript. Finally, two anonymous reviewers clearly helped to improve the paper.

References

Ahlgren, G., 1983. Comparison of methods for estimation of phytoplankton carbon. *Arch. Hydrobiol.* 98, 489–508.
 Banse, K., 1977. Determining carbon to chlorophyll ratio of natural phytoplankton. *Mar. Biol.* 41, 199–212.

Bühl, A., Zöfel, O., 2000. SPSS Version 9, Einführung in die moderne Datenanalyse unter Windows. Addison-Wesley Verlag, München, pp. 275–301, pp. 370–413.
 Bursche, E.-M., 1961. Änderungen im Chlorophyllgehalt und im Zellvolumen bei Planktonalgen, hervorgerufen durch unterschiedliche Lebensbedingungen. *Int. Rev. Ges. Hydrobiol.* 46, 610–652.
 Callieri, C., Stockner, G., 2002. Freshwater autotrophic picoplankton – a review. *J. Limnol.* 61, 1–14.
 Casper, S.J., 1985. Lake Stechlin – A Temperate Oligotrophic Lake. *Monographiae Biologicae*, vol. 58. Dr. W. Junk Publishers, Dordrecht, Boston, Lancaster.
 Chow-Fraser, P., Trew, D.O., Findley, D., Stainton, M., 1994. A test of the hypothesis to explain the sigmoidal relationship between total phosphorus and chlorophyll *a* concentrations in Canadian lakes. *Can. J. Fish. Aquat. Sci.* 51, 2052–2065.
 Desortova, B., 1981. Relationship between chlorophyll-*a* concentration and phytoplankton biomass in several reservoirs in Czechoslovakia. *Int. Rev. Ges. Hydrobiol.* 66, 153–169.
 Deutsche Einheitsverfahren, 1983–1985. Deutsche Einheitsverfahren zur Wasser-, Abwasser- und Schlammuntersuchung. Verlag Chemie, Weinheim.
 Dillon, P.J., Rigler, F.H., 1974. The phosphorus–chlorophyll relationship in lakes. *Limnol. Oceanogr.* 19, 767–773.
 Felip, M., Catalan, J., 2000. The relationship between phytoplankton biovolume and chlorophyll in a deep oligotrophic lake: decoupling in their spatial and temporal maxima. *J. Plankton Res.* 22, 91–105.
 Fott, J., Pechar, L., Pražáková, M., 1980. Fish as a factor controlling water quality in ponds. *Dev. Hydrobiol.* 2, 255–261.
 Gervais, F., Padišák, J., Koschel, R., 1997. Do light quality and low nutrient concentration favour picocyanobacteria below the thermocline of the oligotrophic Lake Stechlin. *J. Plankton Res.* 19, 771–781.
 Gosselain, V., Hamilton, P.B., 2000. Algamica – revision to a key-based computerized counting program for free-living, attached and benthic algae. *Hydrobiologia* 438, 139–142.
 Halfson, E., 1984. The composition of particulate organic matter in the euphotic zone of Lake Superior. *J. Great Lakes Res.* 10, 299–306.
 Hallegraef, G.M., 1977. A comparison of different methods for the quantitative evaluation of biomass of freshwater phytoplankton. *Hydrobiologia* 55, 145–165.
 Hamilton, P.B., 1990. The revised edition of a computerized plankton counter for plankton, periphyton and sediment analyses. *Hydrobiologia* 194, 23–30.
 Harris, G.P., 1986. *Phytoplankton Ecology. Structure, Function and Fluctuations. The Concept of Limiting Nutrients.* Capman and Hall, London, pp. 137–165.
 Hillebrand, H., Durselen, C.D., Kirschtel, D., Pollinger, U., Zohary, T., 1999. Biovolume calculation for pelagic and benthic microalgae. *J. Phycol.* 35, 403–424.
 Hoehn, E., Clasen, J., Scharf, W., Ketelaars, H.A.M., Nienhäuser, A.E., Horn, H., Kerksen, H., Ewig, B., 1998. Erfassung und Bewertung von Planktonorganismen. Arbeitsgemeinschaft der Trinkwassertalsperren, Arbeitskreis Biologie, Technische Information 7, 2. Auflage. Siegburg.

- Jones, R.I., Rees, S., 1994. Influence of temperature and light on particle ingestion by the fresh-water phytoflagellate. *Dinobryon*. *Arch. Hydrobiol.* 132, 203–211.
- Kamoto, S.M., 1966. Primary production by phytoplankton community in some Japanese lakes and its dependence on lake depth. *Arch. Hydrobiol.* 62, 1–28.
- Kasprzak, P., Krienitz, L., Koschel, R., 1993. Changes in the plankton of Lake Feldberger Haussee (Germany, Mecklenburg-Vorpommern) in response to biomanipulation. *Arch. Hydrobiol.* 128, 149–168.
- Kasprzak, P., Gervais, F., Adrian, R., Weiler, W., Radke, R., Jäger, I., Riest, S., Siedel, U., Schneider, B., Böhme, M., Eckmann, R., Walz, N., 2000. Trophic characterisation, pelagic food web structure and comparison of two mesotrophic lakes in Brandenburg (Germany). *Int. Rev. Hydrobiol.* 85, 167–189.
- Kasprzak, P., Koschel, R., Krienitz, L., Gonsiorczyk, T., Anwand, K., Laude, U., Wysujack, K., Brach, H., Mehner, T., 2003. Reduction of nutrient loading, planktivore removal and piscivore stocking as tools in water quality management: the Feldberger Haussee biomanipulation project. *Limnologica* 33, 190–204.
- Kiss, G., Dévai, G., Tóthmérész, B., Szabó, A., 2006. Multivariate analysis of long-term water quality changes of shallow Lake Balaton. *Verhandlungen der Internationalen Vereinigung für Theoretische und Angewandte Limnologie* 29, 2051–2055.
- Kohl, J.G., Nicklisch, A., 1988. *Ökophysiologie der Algen-Wachstum und Ressourcennutzung*. Akademie-Verlag, Berlin.
- Koschel, R., Adams, D.D., 2003. An approach to understanding a temperate oligotrophic lowland lake (Lake Stechlin, Germany). *Arch. Hydrobiol. Spec. Issues, Adv. Limnol.* 58, 1–9.
- Koschel, R., Casper, P., Gonsiorczyk, T., Rossberg, R., Wauer, G., 2006. Hypolimnetic Al and CaCO₃ treatments and aeration for restoration of a stratified eutrophic hardwater lake in Germany. *Verhandlungen der Internationalen Vereinigung für Theoretische und Angewandte Limnologie* 29, 2165–2171.
- Krienitz, L., Kasprzak, P., Koschel, R., 1996. Long term changes in phytoplankton communities in a Baltic hardwater lake (Feldberger Haussee) related to nutrient loading and biomanipulation. *Hydrobiologia* 330, 89–110.
- Loth, P., 1985. Zusammenhänge zwischen Chlorophyll a, Phytoplanktonvolumen, Seston, Sichttiefe und Trübung als Methoden zur Einschätzung der Wasserbeschaffenheit in Talsperren. *Acta Hydrochim. Hydrobiol.* 13, 319–329.
- McQueen, D., Post, J.R., Mills, E.L., 1986. Trophic relationships in freshwater pelagic ecosystems. *Can. J. Fish. Aquat. Sci.* 43, 1571–1581.
- Nusch, E.A., Palme, G., 1975. *Biologische Methoden für die Praxis der Gewässeruntersuchung*. Bestimmung des Chlorophyll-a und Phaeopigmentgehaltes in Oberflächenwasser. *Gas- u. Wasserfach, Wasser, Abwasser* 116, 562–565.
- Padisák, J., Krienitz, L., Koschel, R., Nedoma, J., 1997. Deep-lake autotrophic picoplankton maximum in the oligotrophic Lake Stechlin, Germany – origin, activity, development and erosion. *Eur. J. Phycol.* 32, 403–416.
- Padisák, J., Adrian, R., Nusch, E., Gerhardt, V., Bodemer, U., Zwirnmann, E., Dietrich, P., Soeder, C.J., Tümpling, W.v., Pftzner, S., Müller, D., Peter, S., Babenzien, H.-D., Sass, H., Bittl, T., Köhler, J., Koschel, R., Rossberg, R., G-Tóth, L., 1999. Biomasse und Bioaktivität. In: Tümpling, W.v., Friedrich, G. (Eds.), *Biologische Gewässeruntersuchung. Methoden der biologischen Wasseruntersuchung*. Gustav Fischer Verlag, Jena, Stuttgart, pp. 334–473.
- Padisák, J., Scheffler, W., Kasprzak, P., Koschel, R., Krienitz, L., 2003. Interannual variability in the phytoplankton composition of Lake Stechlin (1994–2000). *Arch. Hydrobiol. Spec. Issues, Adv. Limnol.* 58, 101–133.
- Párista, É., Ács, É., Böddi, B., 2002. Chlorophyll-a determination with ethanol – a critical test. *Hydrobiologia* 485, 191–198.
- Premazzi, G., Chiaudani, G., 1992. *Ecological Quality of Surface Waters. Quality Assessment Schemes for European Community Lakes*. Commission of the European Communities, Joint Research Centre, Ispra (Varese), Italy.
- Reynolds, C.S., 1984. *The Ecology of Freshwater Phytoplankton*. Cambridge Studies in Ecology. Cambridge University Press, Cambridge.
- Reynolds, C.S., 2006. *The Ecology of Phytoplankton*. Cambridge University Press, Cambridge.
- Richards, F.A., Thompson, T.G., 1952. The estimation and characterization of plankton populations by pigment analyses. 2. A spectrophotometric method for the estimation of plankton pigments. *J. Mar. Res.* 11, 156–172.
- Richman, S., Sager, P.E., Banta, G., Harvey, T.R., DeStasio, B., 1984. Phytoplankton standing stock, size distribution, species composition and productivity along a trophic gradient in Green Bay, Lake Michigan. *Verhandlungen der Internationalen Vereinigung für Theoretische und Angewandte Limnologie* 22, 460–469.
- Rott, E., 1978. Chlorophyll-a-Konzentration und Zellvolumen als Parameter der Phytoplanktonbiomasse. *Berichte des naturwissenschaftlich-medizinischen Vereins in Innsbruck* 65, 11–21.
- Sandu, C., Iacob, R., Nicolescu, N., 2003. Chlorophyll-a determination – a reliable method for phytoplankton biomass assessment. *Acta Bot. Hung.* 45, 389–397.
- Schellenberger, G., Stellmacher, R., Hoeg, S., Rohde, E., 1985. Zusammenhang zwischen Biomassen von einzelnen Algengruppen, Chlorophyllkonzentrationen und Fluoreszenzwerten im Müggelsee. *Acta Hydrophys.* 24, 211–220.
- Schmid, H., Bauer, F., Stich, H.B., 1998. Determination of algal biomass with HPLC pigment analysis from lakes of different trophic state in comparison to microscopically measured biomass. *J. Plankton Res.* 20, 1651–1661.
- Sommer, U., Gliwicz, Z.M., Lampert, W., Duncan, A., 1986. The PEG-model of seasonal succession of planktonic events in fresh waters. *Arch. Hydrobiol.* 106, 433–471.
- Stich, H.B., Brinker, A., 2005. Less is better – uncorrected versus phaeopigment-corrected photometric chlorophyll-a estimation. *Arch. Hydrobiol.* 162, 111–120.
- Strickland, J.D.H., Parsons, T.R., 1960. *A Manual of Sea Water Analysis*. Bulletin No. 25. Fisheries Research Board, Canada.
- Talling, J.F., 1993. Comparative seasonal changes, and their inter-annual variability and stability, in a 26-year record of

- total phytoplankton biomass in four English lake basins. *Hydrobiologia* 268, 65–98.
- Tolstoy, A., 1977. Chlorophyll a measurements of phytoplankton biomass. *Acta Univ. Upsal.* 416, 1–30.
- Töpel, J., Wilhelm, C., Meister, A., Beckert, A., Martinez-Ballesta, M.D., 2004. Cytometry of freshwater phytoplankton. In: *Methods in Cell Biology*. Elsevier Academic Press Inc., San Diego, pp. 375–407.
- Utermöhl, H., 1923. Das Nannoplankton der holsteinischen Seen. *Verhandlungen der Internationalen Vereinigung für Theoretische und Angewandte Limnologie* 1, 86–93.
- Utermöhl, H., 1958. Zur Vervollkommnung der quantitativen Planktonmethodik. *Mitteilungen der Internationalen Vereinigung für Theoretische und Angewandte Limnologie* 9, 1–38.
- Vanni, M.J., Temte, J., Allan, Y., Dodds, R., Howard, P.J., Leavit, P.R., Luecke, C., 1993. Herbivory, nutrients, and phytoplankton dynamics in Lake Mendota, 1987–89. In: Kitchell, J.F. (Ed.), *Food Web Management. A Case Study of Lake Mendota*. Springer, pp. 243–273.
- Verity, P.G., Robertson, C.Y., Tronzo, C.R., Andrews, M.G., Nelson, J.R., Sieracki, M.E., 1992. Relationships between cell-volume and the carbon and nitrogen content of marine photosynthetic nanoplankton. *Limnol. Oceanogr.* 37, 1434–1446.
- Vollenweider, R.A., 1974. Photosynthetic pigments. In: Vollenweider, R.A. (Ed.), *A Manual on Methods for Measuring Primary Production in Aquatic Environments*. Blackwell Scientific Publications, Oxford, pp. 21–26 (IBP Handbook no. 12).
- Vörös, L., Padisák, J., 1991. Phytoplankton biomass and chlorophyll-a in some shallow lakes in central Europe. *Hydrobiologia* 215, 111–119.
- Wasmund, N., 1984. Probleme der spektrophotometrischen Chlorophyllbestimmung. *Acta Hydrochim. Hydrobiol.* 12, 255–275.
- Watson, S., McCauley, E., 1988. Contrasting patterns of net- and nanoplankton production and biomass among lakes. *Can. J. Fish. Aquat. Sci.* 45, 915–920.
- Watson, S., McCauley, E., Downing, J.A., 1992. Sigmoid relationships between phosphorus, algal biomass, and algal community structure. *Can. J. Fish. Aquat. Sci.* 49, 2605–2610.
- Willén, E., 1992. Long-term changes in the phytoplankton of large lakes in response to changes during nutrient loading. *Nord. J. Bot.* 12, 575–587.
- Wojciechowska, W., 1989. Correlation between biomass, chlorophyll-a, photosynthesis and phytoplankton structure in a lake. *Ecol. Polska* 37, 59–82.
- Wright, S.W., Jeffrey, S.W., Mantoura, R.F., 1997. Evaluation of methods and solvents for pigment extraction. In: Wright, S.W., Jeffrey, S.W., Mantoura, R.F. (Eds.), *Phytoplankton Pigments in Oceanography – Guidelines to Modern Methods*. UNESCO Publishing, pp. 261–282.