

Report

Cloning, Sequencing, and Analysis of Inv8 Chromosome Breakpoints Associated with Recombinant 8 Syndrome

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Rec8 syndrome (also known as “recombinant 8 syndrome” and “San Luis Valley syndrome”) is a chromosomal disorder found in individuals of Hispanic descent with ancestry from the San Luis Valley of southern Colorado and northern New Mexico. Affected individuals typically have mental retardation, congenital heart defects, seizures, a characteristic facial appearance, and other manifestations. The recombinant chromosome is $\text{rec}(8)\text{dup}(8\text{q})\text{inv}(8)(\text{p}23.1\text{q}22.1)$, and is derived from a parental pericentric inversion, $\text{inv}(8)(\text{p}23.1\text{q}22.1)$. Here we report on the cloning, sequencing, and characterization of the 8p23.1 and 8q22 breakpoints from the inversion 8 chromosome associated with Rec8 syndrome. Analysis of the breakpoint regions indicates that they are highly repetitive. Of 6 kb surrounding the 8p23.1 breakpoint, 75% consists of repetitive gene family members—including *Alu*, LINE, and LTR elements—and the inversion took place in a small single-copy region flanked by repetitive elements. Analysis of 3.7 kb surrounding the 8q22 breakpoint region reveals that it is 99% repetitive and contains multiple LTR elements, and that the 8q inversion site is within one of the LTR elements.

Rec8 syndrome (also known as “recombinant 8 syndrome” and “San Luis Valley syndrome” [MIM 179613]) was first reported (Fujimoto et al. 1975) in a Hispanic female infant who died from heart failure at age 6 wk. Since that time, multiple other cases have been identified, almost all of known Hispanic ancestry with ancestral origin in the San Luis Valley of southern Colorado and northern New Mexico (Fujimoto et al. 1975; Sujansky et al. 1993). Rec8 syndrome is associated with significant morbidity and mortality, and affected individuals typically have congenital heart defects, mental retardation, seizures, a characteristic facial appearance, and other manifestations. Cardiac defects are found in 93% of individuals with Rec8 syndrome and are a major contributor to the mortality of children with Rec8 syndrome, who have an average age at death of ~6 years (Sujansky et al. 1993).

The recombinant chromosome is $\text{rec}(8)\text{dup}(8\text{q})\text{inv}(8)(\text{p}23.1\text{q}22.1)$ and is deleted for 8p23.1→pter and duplicated for 8q22→qter (fig. 1). In all cases, at least one

of the parents of a child with Rec8 syndrome carries a pericentric inversion, $\text{inv}(8)(\text{p}23.1\text{q}22.1)$. Carriers of the Inv8 chromosome are apparently phenotypically normal, and it has been estimated that an Inv8 carrier has a 6.2% chance in each pregnancy of having a child with Rec8 syndrome (Smith et al. 1987).

It is apparent that not all regions of the genome are equally likely to participate in chromosomal rearrangements, and there are several indications that 8p23.1 may be a relatively unstable region of the human genome. Multiple cases of chromosomal rearrangement involving 8p23.1 have been described in the literature, including duplications, deletions, and inverted duplications. Several cases of isolated $\text{del}(8\text{p}23.1)$ have been described (Fryns et al. 1989; Fagan et al. 1988; Blennow and Bronnum-Nielsen 1990; Pecile et al. 1990; Hutchinson et al. 1992; Pettenati et al. 1992; Wu et al. 1996). In general, clinical findings in these patients are variable and may include mental retardation, behavior problems, congenital heart defects, seizures, and genitourinary abnormalities. Multiple cases of “inverted duplication” of 8p have been described, producing a chromosome with a deletion of material distal to the 8p23.1 breakpoint and an inverted duplication of a variable amount of material proximal to the 8p23.1 breakpoint, with clinical manifestations ranging from insignificant to significant de-

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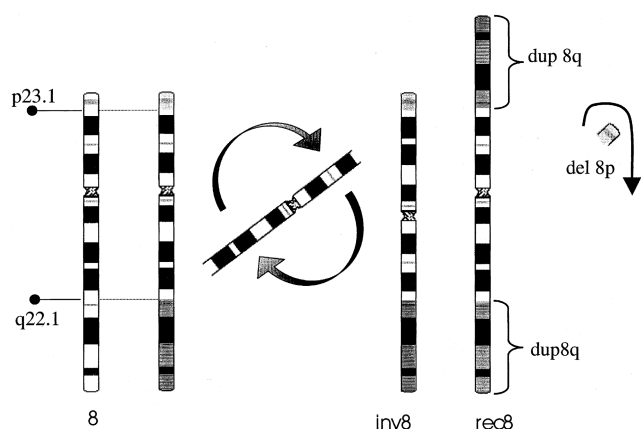


Figure 1 Chromosome 8 rearrangements. The normal chromosome 8 is shown, with 8p23.1 and 8q22 breakpoints indicated. The Inv8 chromosome results from a pericentric inversion. The Rec8 chromosome contains a duplication of 8q22→8qter, whereas the 8p23.1→8pter fragment is lost.

velopmental delay (Dill et al. 1987; Nevin et al. 1990; Henderson et al. 1992; Feldman et al. 1993; Minelli et al. 1993; Barber et al. 1994; Engelen et al. 1994; Mitchell et al. 1994; Guo et al. 1995; Florida et al. 1996; Barber et al. 1998).

In addition to its tendency to participate in chromosomal rearrangements, 8p23.1 has been noted by Ohashi et al. (1994) to contain a sequence that can function as a neocentromere. 8p23.1 also represents a region of the genome that is evolutionarily unstable, as evidenced by comparison of human and macaque banding patterns and FISH analysis (Small et al. 1985; Weinberg et al. 1992).

In spite of this evidence that 8p23.1 is an unstable segment of the genome, the exact molecular basis of these rearrangements has not been defined. Here we report on the cloning and characterization of the 8p23.1 and 8q22 breakpoints from the Inv8 chromosome associated with Rec8 syndrome. Analysis of the sequences surrounding these breakpoints indicates that they contain multiple repetitive elements that may have mediated the original inversion event.

Material from two Rec8 syndrome patients and three Inv8 carriers was used to construct lymphoblastoid cell lines (Neitzel et al. 1986) and hamster-human hybrid cell lines (Moore et al. 1977), segregating the abnormal chromosome 8 from other chromosome 8 material. Rec8-1 is a female Rec8 syndrome patient with a 46,XX,rec(8)dup(8q)inv(8)(p23.1q22.1)pat karyotype. During the neonatal period, the patient was noted to be cyanotic and to have a heart murmur, brachycephaly, and lowset ears. Cardiac evaluation at age 2 d revealed a ventricular septal defect, patent ductus arteriosus, and pulmonary atresia. On karyotypic analysis, the father

(Inv8-1) and brother of Rec8-1 were found to be Inv8 carriers, with 46,XY,inv(8)(p23q22). The mother's karyotype was normal. Rec8-2 is a Rec8 syndrome female with a karyotype of 46,XX,rec(8)dup(8q)inv(8)(p23.1q22.1). Rec8-2 had profound mental retardation, moderate conductive hearing loss, atrial septal defect, patent ductus arteriosus, ventricular septal defect, and pulmonary stenosis. Inv8-2 is a female Inv8 carrier with a karyotype of 46,XX,inv(8)(p23.1q22) and the mother of a female child with Rec8 syndrome diagnosed at age 2 years. Inv8-3 is a female inversion 8 carrier, with a karyotype of 46,XX,inv(8)(p23.1q22). An "Inv8 mapping panel" was created to facilitate mapping, and, in addition to the Inv8 and Rec 8 LCLs and hybrids mentioned earlier, includes HeLa; CHO Gly-B (Jones et al. 1981); Cl17, a hamster-human hybrid line containing normal chromosome 8 as its only human material (Jones et al. 1981); and 21q+ (Drabkin et al. 1985), a hamster-human hybrid containing 8q22-8qter translocated to human chromosome 21.

Previous studies by Shechter et al. (1994) reported the isolation of YAC B20C12, encoding the squalene synthase (SS) gene on chromosome 8. Additional SS YACs 737E5 and 779E9 were identified through screening the CEPH megaYAC library (Green and Olson 1990; Chumakov 1992), and SS P1 clones P36G5 and P72A2 by screening the Dupont/Merck P1 library (Sternberg 1990; Pierce and Sternberg 1992). FISH analyses (Lichter et al. 1990; Patterson et al. 1993) of SS YACs B20C12, 737E5, and 779E9 and SS P1 clones P36G5 and P72A2, against metaphase chromosomes from an Inv8 LCL, revealed that these clones span the 8p23.1 breakpoint (fig. 2). To further limit the breakpoint candidate region, breakpoint-spanning SS P1 clone P36G5 was digested with *Bam*HI, and fragments were subcloned into λ ZAP or pUC19 and were analyzed. The ends of individual subclones were sequenced, and the sequences were used to design primer pairs for detection of each end (for primer pairs, see table 1 in online edition). PCR analysis of the Inv8 mapping panel reveals that the 6-kb subclone P36-21 spans the breakpoint (data not shown). P36-21 was completely sequenced (partial sequence found in fig. 3; complete sequence GenBank accession number AF181099), and additional PCR analyses of the Inv8 mapping panel limited the breakpoint to a 25-bp region (data not shown). Sequences flanking this region were used to design probes just proximal and distal to the breakpoint.

Approximately 5×10^5 plaques from an Inv8 hybrid λ DASH library were screened with breakpoint-flanking probes, and a single clone (Inv8/8p-prox) was isolated. Clone Inv8/8p-prox was partially sequenced and was compared to P36-21 to identify the breakpoint. Novel sequence of Inv8/8p-prox (representing presumptive 8q material) was used to probe a HeLa λ DASH library, and



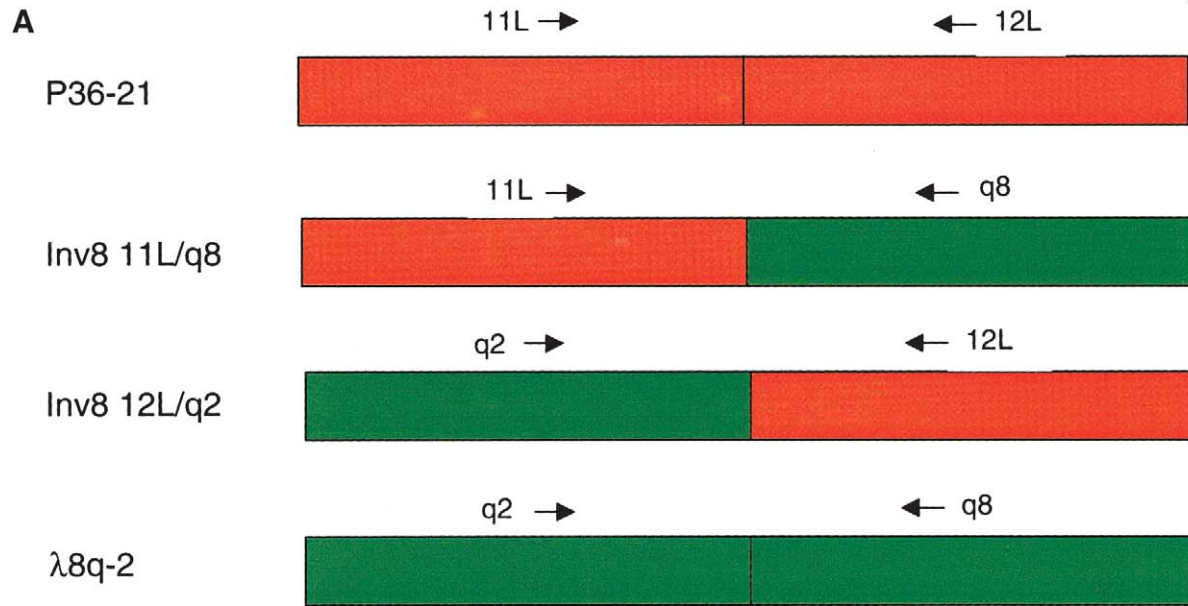
Figure 2 FISH with P1 clone P36G5 against Inv8 lymphoblastoid cell line metaphase chromosomes. *Thick arrow*, hybridization to the normal chromosome 8; *thin arrows*, double signal on the Inv8 chromosome.

two clones were identified ($\lambda 8q-2$ and $\lambda 8q-8$). In addition, this probe was used to screen the CEPH megaYAC library (Chumakov 1992) by PCR and two YACs were identified: 811C1 and 900B3. Both of these YACs have been positioned in the Whitehead Institute database and are part of chromosome 8q contig WC-100, localized to 8q22.1–22.3. Comparison of $\lambda 8q-2$ sequence (partial sequence in fig. 3; complete sequence GenBank accession number AF181100) with Inv8 junction fragment and P36-21 sequences verified the molecular location of the rearrangement (fig. 3).

To characterize the rearrangement and determine whether Inv8 and Rec8 chromosomes all contained the same breakpoints, we used sequences from P36-21 (normal 8p23.1) and $\lambda 8q-2$ (normal 8q22) to design PCR assays specifically for the rearranged chromosomes. Primer combination 12L and q2 is specific for the novel junction fragment proximal-8p/distal-8q and detects both the Inv8 and Rec8 chromosomes, whereas combination 11L and q8 is specific for the distal-8p/proximal-8q novel junction fragment and detects only the Inv8 chromosome (fig. 3). PCRs were done with 12L/q2 and 11L/q8 on Inv8 LCLs Inv8-3 and Inv8-2, Inv8 hybrid Inv8-1, Rec8 hybrid Rec8-2, and Rec8 LCL Rec8-1, and products were purified and sequenced. All Inv8 and Rec8 samples produced a product of identical

size and sequence with primers 12L/q2 (fig. 3b), verifying that these breakpoints are identical and supporting the hypothesis of a founder effect leading to dispersion of a single Inv8 chromosome throughout this population. As expected, primers 11L/q8 produce a product only with the Inv8 LCLs and hybrids and fail to produce a product with Rec8 LCL Rec8-1 and Rec8 hybrid Rec8-2. The Inv8 products were all of identical size and sequence (fig. 3c). Comparison of the rearranged sequences found in 12L/q2 with that of normal 8p and 8q reveals that there is only a 2-bp region of identity at the breakpoint, and that formation of this product did not result in the loss of any material. Comparison of the rearranged sequences found in p11L/q8 with that of normal 8p and 8q reveals that the best alignment is obtained by crossing over in a 7-bp region of identity, and that formation of this product did not result in the loss of any material. The 2-bp region of identity found in 12L/q2 is not contained within the 7-bp region of identity found in 11L/q8, but is 4 bp away on P36-21 and 9 bp away on $\lambda 8q-2$, indicating that the rearrangements took place at slightly offset locations.

Approximately 6 kb of P36-21 (8p23.1) sequence and 4 kb of $\lambda 8q-2$ (8q22) sequence were analyzed by the BLAST algorithm (Altschul et al. 1990), RepeatMasker (A. F. A. Smit and P. Green), and MAR-Finder. BLAST



B

Sample	Sequence
p36-21	TTTAAATCTCTCTTCCATTAAATCTGCTTCTCCTGTG GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
Rec8-1	tggatttcccttctgggcaccctcactctcataaga GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
Rec8-2	tggatttcccttctgggcaccctcactctcataaga GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
Inv8-1	tggatttcccttctgggcaccctcactctcataaga GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
Inv8-2	tggatttcccttctgggcaccctcactctcataaga GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
Inv8-3	tggatttcccttctgggcaccctcactctcataaga GA TAGCATTAAACAGCCTCTCAGTTTGTAAAT
λ8q-2	tggatttcccttctgggcaccctcactctcataagagagaaagctg <u>ttctcct</u> ttctctttctttt

C

Sample	Sequence
p36-21	TTTAAATCTCTCTTCCATTAAATCTGCT TTCTCCT GTGGATAGCATTAAACAGCCTCTCAGTTTGTAAAT
Inv8-1	TTTAAATCTCTCTTCCATTAAATCTGCT TTCTCCT ttctctttcttttgcctattcaacctctgctcct
Inv8-2	TTTAAATCTCTCTTCCATTAAATCTGCT TTCTCCT ttctctttcttttgcctattcaacctctgctcct
Inv8-3	TTTAAATCTCTCTTCCATTAAATCTGCT TTCTCCT ttctctttcttttgcctattcaacctctgctcct
λ8q-2	ccctcactctcataagagagaaagctg ttctcct ttctctttcttttgcctattcaacctctgctcct

Figure 3 A, Schematic illustrating structure of P36-21 (red) and λ8q-2 (green) and novel Inv8 junction fragments detected with primer pairs 11L/q8 and 12L/q2. B, Sequence analysis of 12L/q2 PCR products. Sequence of P36-21 (normal 8p23.1) is indicated in upper case, and of λ8q-2 (normal 8q22) in lower case. Sequence from Rec8 patients and Inv8 carriers is indicated with material corresponding to P36-21 in upper case and to λ8q-2 in lower case. The boxed and bolded “GA” indicates the site of recombination. Underlined “TTCTCCT” denotes site of recombination found in 11L/q8 PCR products. C, Sequence analysis of 11L/q8 PCR products. Sequence of P36-21 (normal 8p23.1) is indicated in uppercase, and of λ8q-2 (normal 8q22) in lowercase. Sequence from Inv8 carriers is indicated with material corresponding to P36-21 in upper case and to λ8q-2 in lower case. The boxed and bolded “TTCTCCT” indicates the site or recombination. Underlined “GA” denotes site of recombination found in 12L/q2 products.

analysis did not produce hits of any significant homology outside of repetitive elements (data not shown). RepeatMasker analysis revealed that both P36-21 and $\lambda 8q-2$ contain multiple repetitive elements (fig. 4). P36-21 is 6,062 bp in length and is 75% repetitive, containing four *Alu* elements, two LINE2 elements, and five LTR elements. Formation of the Inv8 chromosome takes place at two sites that are slightly offset, one at bp 2,618–2,624 (11L/q8) and one at bp 2,628–2,629 (12L/q2), within a small 370-bp unique sequence region flanked by a LINE/L2 element and an *Alu* element. A single potential matrix attachment region (MAR) was identified in P36-21, between 800 bp and 1400 bp, by use of the MAR-Finder program. This same region was also shown by RepeatMasker to contain an LTR10C element and an *Alu*Sq element. The sequence of $\lambda 8q-2$ is 3724 bp in length and contains multiple LTR elements, totaling 3,688 bp, or 99.03% of the sequence. Recombination at the 8q22 break occurs at two places that are slightly offset, one at bp 2,560–2,566 (11L/q8) and one at bp 2,550–2,551 (12L/q2) within an LTR element.

The junction sequences of several constitutional chromosomal rearrangements have been characterized, leading to speculation on the molecular mechanisms behind such rearrangements. In some cases, such as deletion of an α -globin gene leading to α -thalassemia (Ottolenghi et al. 1974; Taylor et al. 1974), recombination is between homologous segments of two members of a gene family. In other cases, such as in the LDL-receptor (Hobbs et al. 1986), α -globin (Nicholls et al. 1987), β -globin (Vanin et al. 1983; Henthorn et al. 1986), and apolipoprotein B genes (Huang et al. 1989), rearrangement has been shown to be mediated by *Alu* repetitive elements. It has been proposed that *Alu* elements function as recombinatorial hotspots (Calabretta et al. 1982; Barsh et al. 1983), and Rudiger et al. (1995) have speculated that a highly conserved 26-bp core sequence en-

hances recombination. Other repetitive elements, such as *LINE-1* (*L1*), have also been shown to participate in chromosomal rearrangements (Toriello et al. 1996). In still other cases (e.g., Budarf et al. 1995), there is minimal or no obvious sequence homology to mediate the rearrangement, and the cause is speculative. In at least one case of a constitutional translocation t(8;17), resulting in isolated lissencephaly (Kurahashi et al. 1998), there was no sequence homology between the parental chromosomes, but the region immediately surrounding the 17p13.3 breakpoint had five *Alu* repetitive elements and the region surrounding the 8p11.2 breakpoint contained three *L1* sequences, leading to speculation that the repetitive elements themselves contribute to instability and recombination.

In the case of the Inv8 chromosome, we have shown that the inversion event takes place between two highly repetitive regions, and that the exact break occurred in a small single-copy region of 8p23.1 and in the midst of an expanse of LTR elements in 8q22, supporting the hypothesis that repetitive elements are themselves recombinogenic.

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Electronic-Database Information

Accession numbers and URLs for data in this article are as follows:

GenBank, (for complete P36-21 sequence [accession number

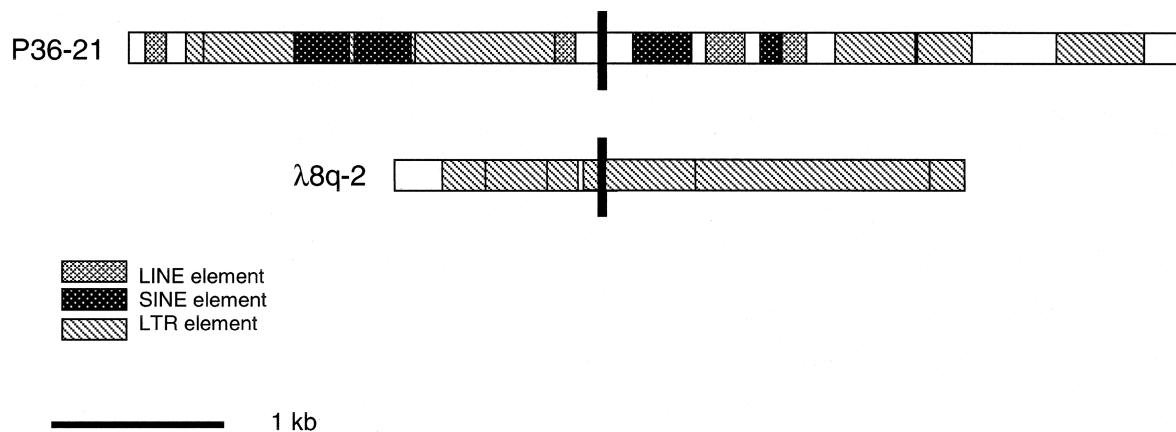


Figure 4 RepeatMasker analysis of P36-21 and $\lambda 8q-2$. Repetitive elements are as indicated. Breakpoint location is indicated by a solid, vertical bar.

AF181099] and complete λ 8q-2 sequence [accession number AF181100]),
 MAR-Finder, <http://www.ncgr.org/MarFinder/intro.html>
 Online Mendelian Inheritance in Man (OMIM), <http://www.ncbi.nlm.nih.gov/Omim> (for Rec8 syndrome [MIM 179613])
 RepeatMasker, <http://ftp.genome.washington.edu/RM/RepeatMasker.html>
 Whitehead Institute for Biomedical Research/MIT Center for Genome Research, <http://www-genome.wi.mit.edu/>

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