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Original article

Hepatoprotective standardized EtOH—water extract from the seeds of *Fraxinus rhynchophylla* Hance

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ABSTRACT

Fraxinus rhynchophylla Hance (Oleaceae), its stem barks are known as Cortex fraxini (秦皮 gín pí) listed in Chinese Pharmacopoeia. Phytochemical study has indicated that methanol extracts from Qinpi has protective effect on acute liver injury. The present study investigates the hepatoprotective activity of EtOH-water extract from the seeds of F. rhynchophylla Hance against carbon tetrachloride-induced liver injury in mice. The EtOH-water extract significantly alleviated liver damage as indicated by the decreased levels of serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST), the malondialdehyde (MDA) content, and increased the levels of superoxide dismutase (SOD), glutathione (GSH) and glutathione peroxidase (GSH-Px), and reduced the pathological tissue injury induced by CCl₄. Quantitative analysis of seven major constituents (1-7) in EtOH-water extract (EWE) was developed by high performance liquid chromatography-diode-array detector (HPLC-DAD). The current research indicates that the EWE from the seeds of F. rhynchophylla Hance decreased liver index, inhibited the increase of serum aminotransferase induced by CCl₄, and decreased hepatic MDA content, SOD and GSH-Px activities. These results suggested that the pretreatment with EWE protected mice against CCl₄-induced liver injuries. Based on the results, the EtOH-water extract from the seeds of F. rhynchophylla Hance is efficacious for prevention and treatment of CCl₄-induced hepatic injury in mice. Secoiridoid and tyrosol glucosides might be the active ingredients responsible for the biological and pharmacological activities of hepatoprotection.

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1. Introduction

The liver plays an important role in human metabolism and detoxification of endogenous and exogenous chemicals.¹ Liver injuries or dysfunctions have been recognized as serious health problem. Especially acute and chronic liver injuries resulted from the exposure to toxic chemicals, drugs, and virus infiltration from ingestion or infection, have gained more attention in recent years.^{2–4} Corticosteroids and interferon has been used for the treatment of hepatic diseases, however, these synthetic chemical

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drugs are not well accepted by patients due to limited therapeutic efficacy and serious complications.⁵ Therefore, more effective complementary and therapeutic drugs with low or no side-effects are needed for the treatment of liver diseases.^{6–9} In recent years, some effective and safe dietary ingredients for liver-protection have been isolated from traditional medicinal plants, such as glycyrrhizin,¹⁰ curcumin,¹¹ resveratrol,¹² as well as silvbin and silvmarin.^{13,14} Fraxinus rhynchophylla Hance (Oleaceae) is a commonly used Chinese traditional medicinal plant, mainly distributed in China and Korea.¹⁵ Its stem bark also known as *Cortex fraxini* (Qinpi) is Chinese herbal drug for treating diseases such as acute conjunctivitis and psoriasis; arresting discharges; curing chronic bronchitis; and bacillary dysentery, diuretic, antirheumatic, analgesic, antiperspiratory effects, and enhancing eyesight.¹⁶ Phytochemical study has indicated that methanol extracts from Qinpi has protective effect on acute liver injury.¹⁷ Many natural products such

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as secoiridoid glucosides, coumarins, lignans, sesquilignans, and coumarinolignans have been identified from this plant.^{18–23} Recently, several pharmacological activities of phytochemical constituents isolated from the barks and leaves of F. rhynchophylla have been carried out, including anti-diabetes effects, 24-28 anti-Toxoplasma gondii effects, 29 including antioxidant enzymes, 30 inhibiting amyloid-β-induced neuronal cell damage,³¹ and inhibiting nitric oxide synthesis activities.³² Hydrangeside B, along with other secoiridoid glucosides showed hepatoprotective activities against DL-galactosamine-induced toxicity in human hepatocyte HL-7702 (HL-7702) cells.³³ Secoiridoid glycoside, oleuropein, showed antihepatitis B virus (HBV) activity and effectively blocked hepatitis B surface antigen (HBsAg) secretion with an IC₅₀ of 23.2 μ g/mL in HepG2.2.15 cells with no significant cytotoxicity.³⁴ The hepatoprotective activity of oleuropein against carbon tetrachloride (CCl₄)-induced liver damage in mice was achieved through the NF-E2-related factor 2-mediated induction of heme oxygenase-1.35 Recently, Peng et al.³⁶ investigated the effect of *Fraxinus rhyncho*phylla ethanol extract (FR_{EtOH}) on liver fibrosis induced by CCl₄ in rats. However, the hepatoprotective activity of the seeds of F. rhynchophylla Hance has not been evaluated so far.

The present study aimed to evaluate the hepatoprotective activity of EtOH—water extract from the seeds of *F. rhynchophylla* Hance employing a widely used CCl_4 -induced liver damage model in mice and quantitative analysis of six secoiridoid glucosides (**1**–**6**) and one tyrosol glucoside (**7**) by high performance liquid chromatography-diode-array detector (HPLC-DAD) method.

2. Materials and methods

2.1. Chemicals and reagents

CH₃OH (HPLC grade), CH₃CH₂OH (HPLC grade), and CH₃CN (HPLC grade) were purchased from Sigma-Aldrich (St. Louis, MO, USA). All other organic solvents used in the current study, such as CH₃OH, ethyl acetate (EtOAc), acetone, and chloroform (CHCl₃) were of analytical grade. They are commercially available from Hengxing Chemical Reagent Co., Ltd. (Tianjin, China). Chemical standards of oleoside dimethyl ester (**1**), ligstroside (**2**), nuzhenide (**3**), 10-hydroxoleoside dimethyl ester (**4**), GI3 (**5**), GI5 (**6**), and salidroside (**7**) were prepared in our laboratory. The purity of each compound was >98%, determined by HPLC analysis. The chemical structures of these reference compounds are shown in Fig. 1.

2.2. Materials

The seeds of *F. rhynchophylla* Hance were provided in August 2013 from Baoji City, Shaanxi province, China. The herbariums of *F. rhynchophylla* Hance (FRH001) were deposited in Room 612, Department of Pharmaceutical Engineering, College of Chemical Engineering, Northwest University.

2.3. HPLC analysis conditions

HPLC analysis was performed on an Agilent 1260 separation module connected to a G1315D DAD detector using a Synergi 4u Hydro-RP 80R column ($250 \times 4.6 \text{ mm}$, 4 µm, 100 Å) with a flow rate of 1.0 mL/min. Solvent system: 0 min: 95% A (1% phosphoric acid) and 5% B (acetonitrile), 2 min: 95% A (1% phosphoric acid) and 5% B (acetonitrile), 5 min: 80% A (1% phosphoric acid) and 20% B (acetonitrile), 25 min: 75% A (1% phosphoric acid) and 25% B (acetonitrile), 27 min: 95% A (1% phosphoric acid) and 25% B (acetonitrile), 30 min: 95% A (1% phosphoric acid) and 5% B (acetonitrile). At the end of the run, 100% of acetonitrile was allowed to flush the column for 10 min, and an additional 10 min of post run time was set to allow for equilibration of the column with the starting eluant. The UV detector was operating at 230 nm, and the column temperature was maintained at 30 °C.

2.4. Calibration curves

Methanol stock solutions containing the seven standard compounds **1–7** were prepared and diluted to five different final concentrations. A calibration curve was constructed for each of the compounds by plotting peak areas versus compound concentrations.

2.5. Preparation of seeds of F. rhynchophylla Hance extract

The 5 kg air-dried seeds of *F. rhynchophylla* Hance were percolated twice with absolute ethyl alcohol at room temperature. The ethyl alcohol was evaporated under vacuum. The herb residue was then percolated twice with water at room temperature and made the SFR-water extracts. Finally, mixed the above two extracts, and the *in vivo* bioactivity study of seeds of *F. rhynchophylla* Hance was carried on by this sample. The HPLC spectrum of the extract is shown in Fig. 2.

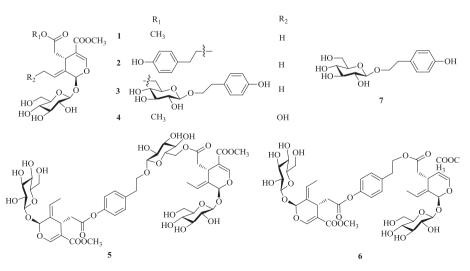


Fig. 1. Structures of compounds 1-7.

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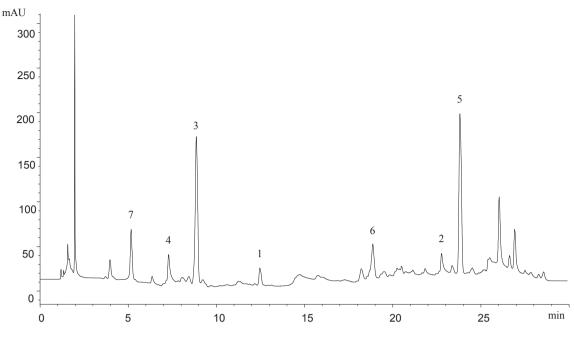


Fig. 2. The HPLC chromatogram of EtOH-Water extract.

2.6. The hepatoprotective activities assays

2.6.1. Animals

Male Kunming mice (weighing 18–22 g), obtained from the Experimental Animal Center of Xi'an Jiaotong University, were used. The animal ethical approval communication number is SCXK 2012-003 (Northwest University, Xi'an, Shaanxi, China). All animals were grouped and housed in polyacrylic cages ($29 \times 18 \times 16$ cm) with no more than six animals per cage and maintained under standard laboratory conditions (temperature 25 ± 2 °C, relative humidity $50 \pm 10\%$) with dark and light cycle (14/10 h). The mice were acclimatized to laboratory condition for 5 days before commencement of experiment. All the experiments were performed in accordance with the Regulations of Experimental Animal Administration issued by the State Committee of Science and Technology of People's Republic of China.

2.6.2. CCl₄-induced hepatotoxicity model

Mice were randomly divided into five groups of 12 animals each. In the control group (group I) and CCl₄-intoxicated group (group II), mice were given a single daily dose of distilled water (0.2 mL/10 g, intragastrically (ig)). In the test groups, mice were given 2.0 g (group IV) and 4.0 g (group V) of EtOH—water extract (EWE) per kg body weight (BW) once daily. In group III, silymarin was served as a positive control, and mice were given 0.1 g/kg BW. All administrations were conducted for 7 weeks. On the 49th day, all mice except those in the control group were given simultaneously a CCl₄/peanut oil mixture (0.1:100, intraperitoneally, 0.1 mL/10 g, ig) 2 h after the last administration, while the control group received peanut oil alone. Then all the animals were fasted for 16 h and were subsequently tested for the following analysis.

2.6.3. Liver index

Liver index was determined as percent of wet liver weight to body weight.

2.6.4. Assessment of liver function

After blood collection, serum was separated by centrifugation at 604 *g* at room temperature for 20 min. The serum alanine amino-transferase (ALT) and aspartate aminotransferase (AST) values were measured with commercially available diagnostic kits produced by Nanjing Jiancheng Bioengineering Institute (Nanjing, China).

2.6.5. Determination of MDA, SOD and GSH-Px activity

After the animals were sacrificed, livers were immediately excised. With the exception of a portion of the left lobe to be used for histopathological examination, the livers were homogenized in phosphate buffer (50 mM, pH 7.4) and centrifuged at 420 g for 20 min at 4 °C. The malondialdehyde (MDA) content, superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px) activities along with protein levels in the supernatant were measured according to commercially available diagnostic kits.

2.6.6. Histopathological examinations

A left lobe portion of the liver was incubated for 24 h in 10% neutral formalin solution. Based on standard procedures, we obtained 5 μ m sections for histopathological studies using hematoxylin and eosin (H&E) staining.

2.6.7. Statistical analysis

The data were analyzed in triplicate, using SAS software, version 8.1 (SAS Institute, Cary, NC, USA). Tukey's test was used to determine statistical significance. The values were considered significantly different when the *P*-value was lower than 0.05.

3. Results

3.1. HPLC method validation

All calibration curves made by the HPLC method exhibited excellent linear regressions with the determination coefficients (r^2) ranging from 0.99 to 0.9999, and the calibration ranges adequately covered variations (Table 1).

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Table 1

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Calibration curves and LOD and LOQ data of 7 compounds investigated by HPLC.										
No.	Calibration curves ^a	r ²	Linear range (mg/mL)	LOD ^b (mg/mL)	LOQ ^b (mg/mL)					
1	Y = -26.3302 + 15081.4372X	0.9998	0.02-0.48	0.0005	0.0016					
2	Y = 40.3439 + 13692.4304X	0.9986	0.01-0.24	0.0048	0.0158					
3	Y = 667.3964 + 4408.5855X	0.9999	0.07-3.50	0.0012	0.0035					
4	Y = -1578.7230 + 15651.8470X	0.9993	0.10-1.20	0.0330	0.0992					
5	Y = 481.0792 + 6299.1942X	0.9984	0.20-3.20	0.0010	0.0034					
6	Y = -2.0423 + 1721.8740X	0.9999	0.05-1.20	0.0038	0.0119					
7	Y = 1228.5054 + 2292.3870X	0.9999	0.20-2.40	0.0250	0.0840					

^a Y is the value of peak area, and X is the value of the reference compound's concentration (μ g/mL).

^b LOD and LOQ were determined at S/N of about 3 and 10, respectively.

3.2. Quantification of the seven compounds

According to these results, the content of compound **3** (377.280 mg/g) was higher than other compounds, and the content of compound **5** was 330.960 mg/g in EtOH–water extract from the seed of *F. rhynchophylla* Hance (Table 2). The content of total analytes is 778.086 mg/g or 77.81% in EtOH–water extract from the seed of *F. rhynchophylla* Hance.

3.3. The hepatoprotective activities assays

3.3.1. EtOH-water extract (EWE) decreased liver index

Liver swelling induced by CCl₄ was manifested on the increase of liver index. As shown in Fig. 3, in the model group (CCl₄-intoxicated group), liver index was 4.96, which was significantly higher than control group (P < 0.05). Pretreatment with **EWE** significantly decreased liver index compared with model group (P < 0.05).

3.3.2. EWE inhibited the increase of serum aminotransferase induced by CCl_4

The results of hepatoprotective effect of **EWE** on the serum ALT and AST activities are shown in Fig. 4. In CCl₄-intoxicated group, serum ALT and AST activities were 239.46 and 11.97 U/L, respectively, whereas the values of control group were only 46.79 and 2.34 U/L, respectively. Therefore, a significant increase in the activities of serum ALT and AST was observed when liver was exposured to CCl₄ (P < 0.05). Administration with different doses of **EWE** for 49 days significantly reduced the activities of serum ALT and AST as compared to the control group (P < 0.05) in a dosedependent manner. Silymarin, which has been used as medicine for liver disease, had superior effects on decreasing serum ALT and AST activities.

3.3.3. EWE decreased hepatic MDA content, SOD and GSH-Px activities

The levels of MDA, SOD and GSH-Px are indicators of oxidative stress. A 30.77% increase of MDA content was observed in model group compared with control group (Fig. 5). Pretreatment with EWE and silymarin could completely inhibit CCl₄-induced increase of liver MDA content. As shown in Figs. 6 and 7, hepatic SOD and GSH-Px activities were decreased by 44.21% and 59.96%,

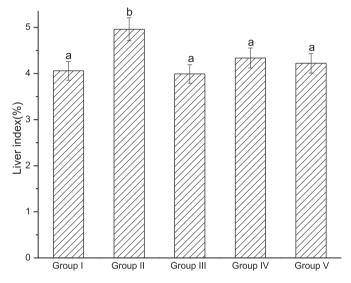


Fig. 3. Effects of **EWE** on liver index. Different lower case letters correspond to significant differences at P < 0.05. Group I was the control group. Group II was CCl₄-treated group. Group III was given CCl₄ + silymarin. Group IV and V were given CCl₄ + 2.0 and 4.0 g/kg BW of **EWE** respectively.

respectively, after CCl₄ treatment. However, pretreatment with **EWE** significantly increased SOD and GSH-Px activities (P < 0.05).

3.4. Histopatholgy

The histological observations of the hepatoprotective effects of **EWE** on CCl₄-induced liver damage are illustrated in Fig. 8. Liver sections from control mice displayed regular cellular morphology (Fig. 8A). Compared with the normal liver tissues of control mice, liver tissue in CCl₄-intoxicated mice revealed liver injury characterized by dilated sinusoidal spaces, and inflammatory cell infiltration (Fig. 8B). However, the hepatic lesions induced by treatment with CCl₄ were remarkably ameliorated in hypertrophy of hepatocytes, dilated sinusoidal spaces and inflammatory cell infiltration by treatment with silymarin and different dose of **EWE** (Fig. 8C–E).

Table 2

Contents of seven compounds in the EtOH-water extract of seeds of F. rhynchophylla Hance.

No	Contents of analytes ^a (mg/g, $n = 3$)							Extraction rate (w %) ^b
	1	2	3	4	5	6	7	
EtOH-water	2.247	2.930	377.280	11.244	330.960	27.368	26.057	9.08

^a Content = $\frac{X \frac{V_1 \text{ (injection volume of standard solution)}}{V_2 \text{ (injection volume of sample solution)}} \times V \text{ (sample volume)}}{M \text{ (sample amount)}}$

^b Extraction rate = solids content of the extract/sample volume.

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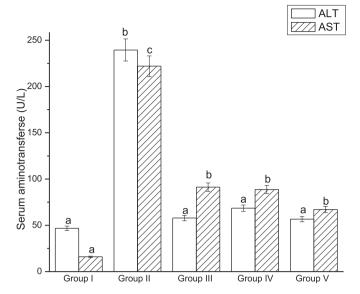


Fig. 4. Effects of **EWE** on serum ALT and AST. Different lower case letters correspond to significant differences at P < 0.05. Group I was the control group. Group II was CCl₄-treated group. Group III was given CCl₄ + silymarin. Group IV and V were given CCl₄ + 2.0 and 4.0 g/kg BW of **EWE** respectively.

4. Discussion

The therapeutic benefits of Chinese herbal medicine have been recognized for centuries on the basis of clinical experience and practice. Qinpi, the stem barks of *F. rhynchophylla* Hance, is a traditional Chinese herbal drug for treating various types of chronic diseases.

ALT and AST are aminotransferases and associated with liver parenchymal cell. The enzymes normally present in the cytosol and are leaked out into the blood stream, when the hepatocellular plasma membrane is damaged. Thus, ALT and AST serum levels are very sensitive markers employed in the diagnosis of liver diseases. Rats administered once with CCl₄ exhibited liver injury, as indicated by significant elevation in serum markers for hepatic cell

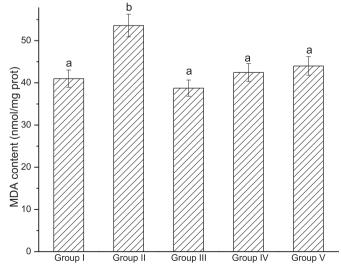


Fig. 5. Effects of **EWE** on hepatic MDA. Different lower case letters correspond to significant differences at P < 0.05. Group I was the control group. Group II was CCl₄-treated group. Group III was given CCl₄ + silymarin. Group IV and V were given CCl₄ + 2.0 and 4.0 g/kg BW of **EWE** respectively.

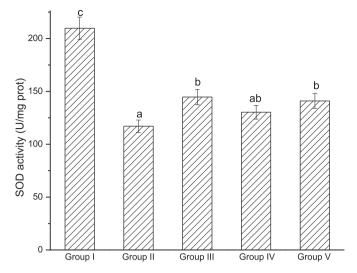


Fig. 6. Effects of **EWE** on hepatic SOD activity. Different lower case letters correspond to significant differences at P < 0.05. Group I was the control group. Group II was CCl_4 -treated group. Group III was given CCl_4 + silymarin. Group IV and V were given $CCl_4 + 2.0$ and 4.0 g/kg BW of **EWE** respectively.

damage (ALT and AST).³⁷ At present study, in EWE-treated groups (2.0 and 4.0 g/kg) of rats, the studies showed that treatment with EWE significantly and dose-dependently decreased the elevations of serum levels of ALT and AST. Furthermore, the improvement of the hepatic impairment was achieved and could be observed by EWE intervention in CCl₄-induced liver injury. Thus, the results showed the protective effects of EWE on hepatic pathology and orally administered EWE exerted therapeutic effects on CCl₄-induced liver injury in rats.

Oxidative stress associated with lipid peroxidation is involved in liver development. The extracts and the identified compounds from *F. rhynchophylla* Hance had been reported anti-oxidative activity.^{30–32} MDA is an aldehyde, which is a product of poly-unsaturated fatty acid peroxidation, and is a highly toxic molecule considered as more than just a marker of lipid peroxidation. In this

Fig. 7. Effects of **EWE** on hepatic GSH-Px activity. Different lower case letters correspond to significant differences at P < 0.05. Group I was the control group. Group II was CCl₄-treated group. Group III was given CCl₄ + silymarin. Group IV and V were given CCl₄ + 2.0 and 4.0 g/kg BW of **EWE** respectively.

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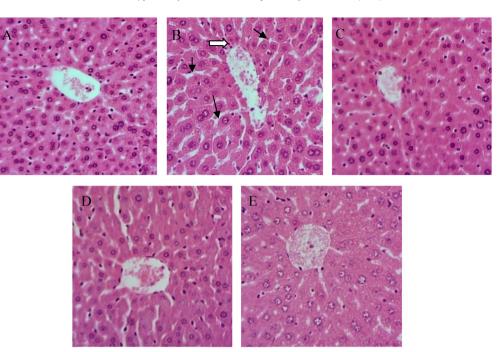


Fig. 8. Effects of EWE on hepatic morphological analysis (×200 H&E): control mice (A), CCl₄-treated mice (B), mice pretreated with silymarin prior to CCl₄ (C), mice pretreated with 2.0 (D) and 4.0 g/kg (E) BW of EWE respectively prior to CCl₄. (→ dilated sinusoidal spaces, ⇔ inflammatory cell infiltration).

study, the elevated hepatic MDA level was decreased by pretreatment of EWE, so it indicated that EWE could protect the liver against CCl₄-induced lipid peroxidation. GSH and SOD are a host of antioxidant systems to protect the hepatocyte from oxidative stress. GSH, or GSH-Px, and SOD work in concert to control the cascades of uncontrolled lipid peroxidation and protect cell from oxidative damage by scavenging of reactive oxygen species (ROS) or the toxic free radicals.^{38–40} In this study, the CCl₄-model rats present lower GSH-Px and SOD level than the control group. Thus, it was supposed that EWE possessed the properties to enhance GSH-Px and SOD, and could prevent hepatice impairment via inhibiting oxygen-free radicals and increasing anti-oxidation in CCl₄-induced liver injury in rats.

5. Conclusion

In conclusions, the current research indicates that the EtOH—water extract (EWE) from the seeds of *F. rhynchophylla* Hance decreased liver index, inhibited the increase of serum amino-transferase induced by CCl₄, and decreased hepatic MDA content, SOD and GSH-Px activities. These results suggested that the pre-treatment with EWE protected mice against CCl₄-induced liver injuries. Further hepatoprotective effect and the possible mechanism of active constituents of the seeds of *F. rhynchophylla* Hance will be studied.

Conflict of interests

The authors declare that they have no competing interest.

Acknowledgment

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References

- 1. Alter MJ. Epidemiology of viral hepatitis and HIV coinfection. J Hepatol. 2006;44:S6.
- Lee CP, Shih PH, Hsu CL, et al. Hepatoprotection of tea seed oil (*Camellia oleifera* Abel.) against CCl₄-induced oxidative damage in rats. *Food Chem Toxicol*. 2007;45:888–895.
- Cemek M, Aymelek F, Buyukokuroglu ME, et al. Protective potential of Royal Jelly against carbon tetrachloride induced-toxicity and changes in the serum sialic acid levels. *Food Chem Toxicol.* 2010;48:2827–2832.
- Jia XY, Zhang QA, Zhang ZQ, et al. Hepatoprotective effects of almond oil against carbon tetrachloride induced liver injury in rats. *Food Chem.* 2011;125: 673–678.
- Stickel F, Schuppan D. Herbal medicine in the treatment of liver diseases. *Dig Liver Dis*. 2007;39:293–304.
- Ghosh N, Ghosh R, Mandal V, et al. Recent advances in herbal medicine for treatment of liver diseases. *Pharm Biol.* 2011;49:970–988.
- Srivastava A, Shivanandappa T. Hepatoprotective effect of the root extract of Decalepis hamiltonii against carbon tetrachloride-induced oxidative stress in rats. Food Chem. 2010;118:411–417.
- 8. Bishayee A, Darvesh AS, Politis T, et al. Resveratrol and liver disease: from bench to bedside and community. *Liver Int*. 2010;30:1103–1114.
- Lal AA, Murthy PB, Pillai KS. Screening of hepatoprotective effect of a herb mixture against CCl₄ induced hepatotoxicity in Swiss albino mice. J Environ Biol. 2007;28:201–207.
- Yoshikawa M, Matsui Y, Kawamoto H, et al. Effects of glycyrrhizin on immunemediated cytotoxicity. J Gastroenterol Hepatol. 1997;12:243–248.
- 11. Ravindran PN, Nirmal Babu K, Sivaraman K. *Turmeric: The Genus Curcuma*. Boca Raton, BR: CRC Press; 2007.
- 12. Sener G, Toklu HZ, Sehirli AQ, et al. Protective effects of resveratrol against acetaminophen-induced toxicity in mice. *Hepatol Res.* 2006;35:62–68.
- **13.** Sturm S, Stuppner H. Analysis of iridoid glycosides from *Picrorhiza kurroa*, by capillary electrophoresis and high performance liquid chromatography-mass spectrometry. *Chromatographia*. 2001;53:612–618.
- Li CC, Hsiang CY, Wu SL, et al. Identification of novel mechanisms of silymarin on the carbon tetrachloride-induced liver fibrosis in mice by nuclear factor-kB bioluminescent imaging-guided transcriptomic analysis. *Food Chem Toxocol*. 2012;50:1568–1575.
- **15.** Wang H, Dou Y, Tian J, et al. Research on medical specialty of traditional Chinese medicines using dot-immunoblotting method based on polyclonal antibody prepared from traditional Chinese medicines with hot/cold nature. *Zhongguo Zhongyao Zazhi*. 2009;34:438–442.
- 16. National Pharmacopoeia Committee. *Pharmacopoeia of People's Republic of China. I.* 2010. Appendix 271.
- Yin MH, Lu HZ, Jiang LJ, et al. Protective effect of ash bark extract against acute liver injury in mice. *Lishizhen Med Materia*. 2007;18:590–591.

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- Kostova I, Iossifova T. Chemical components of *Fraxinus* species. *Fitoterapia*. 2007;78:85–106.
- 19. Kwon YS, Kim CM. A study on the chemical constituents from leaves of. *Fraxinus rhynchophylla, Canis Et Felis.* 1996;8:52–63.
- Si CL, Zhang Y, Zhu ZY, et al. Isolation and structure elucidation of secoiridoid glucosides from *Fraxinus rhynchophylla* leaves. *Chem Nat Compd.* 2009;45: 814–816.
- Si CL, Liu Z, Su YF, et al. Coumarins and secoiridoid glucosides from bark of Fraxinus rhynchophylla Hance. Holzforschung. 2008;62:553–555.
- Ahn JH, Hwang BY, Lee MK. Simultaneous quantitation of nine constituents of Fraxinus rhynchophylla using high performance liquid chromatography-diode array detector. Nat Prod Sci. 2013;19:236–241.
- 23. Wu ZB, Liu Y, Tian SS, et al. Chemical constituents of the stem bark of *Fraxinus* rhynchophylla. Chem Nat Compd. 2014;49:1162–1163.
- 24. Shin E, Choi KM, Yoo HS, et al. Inhibitory effects of coumarins from the stem barks of *Fraxinus rhynchophylla* on adipocyte differentiation in 3T3-L1 cells. *Biol Pharm Bull.* 2010;33:1610–1614.
- **25.** Choi KM, Shin E, Liu Q, et al. Hydroxyframoside B, a secoiridoid of *Fraxinus rhynchophylla*, inhibits adipocyte differentiation in 3T3-L1 cells. *Planta Med*. 2011;77:1020–1023.
- **26.** Ahn JH, Shin E, Liu Q, et al. Lignan derivatives from *Fraxinus rhynchophylla* and inhibitory activity on pancreatic lipase. *Nat Prod Sci.* 2012;18:116–120.
- Ahn JH, Shin E, Liu Q, et al. Secoiridoids from the stem barks of *Fraxinus rhynchophylla* with pancreatic lipase inhibitory activity. *Nat Prod Res.* 2013;27: 1132–1135.
- Xiao K, Song QH, Zhang SW, et al. Water-soluble constituents of the root barks of *Fraxinus rhynchophylla* (Chinese drug Qinpi). J Asian Nat Prod Res. 2008;10: 205–210.
- Jiang JH, Jin CM, Kim YC, et al. Anti-toxoplasmosis effects of oleuropein isolated from Fraxinus rhynchophylla. Biol Pharm Bull. 2008;31:2273–2276.

- Thuong PT, Pokharel YR, Lee MY, et al. Dual anti-oxidative effects of Fraxetin isolated from Fraxinus rhynchophylla. Biol Pharm Bull. 2009;32: 1527–1532.
- Yang EJ, Kim SI, Ku HY, et al. Syringin from stem bark of *Fraxinus rhynchophylla* protects Aβ_(25–35)-induced toxicity in neuronal cells. *Arch Pharm Res.* 2010;33: 531–538.
- Kim NY, Pae HO, Ko YS, et al. In vitro inducible nitric oxide synthesis inhibitory active constituents from Fraxinus rhynchophylla. Planta Med. 1999;65:656–658.
- **33.** Shi J, Li CJ, Yang JZ, et al. Hepatoprotective coumarins and secoiridoids from *Hydrangea paniculata*. *Fitoterapia*. 2014;96:138–145.
- Zhao G, Yin Z, Dong J. Antiviral efficacy against hepatitis B virus replication of oleuropein isolated from *Jasminum officinale* L. var.grandiflorum. *J Ethnopharmacol.* 2009;125:265–268.
- **35.** Domitrović R, Jakovac H, Marchesi VV, et al. Preventive and therapeutic effects of oleuropein against carbon tetrachloride-induced liver damage in mice. *Pharmacol Res.* 2012;65:451–464.
- 36. Peng WH, Tien YC, Huang CY, et al. Fraxinus rhynchophylla ethanol extract attenuates carbon tetrachloride-induced liver fibrosis in rats via downregulating the expressions of uPA, MMP-2, MMP-9 and TIMP-1. J Ethnopharmacol. 2010;127:606–613.
- 37. Carobene A, Braga F, Roraas T, et al. A systematic review of data on biological variation for alanine aminotransferase, aspartate aminotransferase and y-glutamyl transferase. *Clin Chem Lab Med.* 2013;51:1997–2007.
- Valko M, Rhodes CJ, Moncol J, et al. Free radicals, metals and antioxidants in oxidative stress-include cancer. *Chem Biol Interact*. 2006;160:1–40.
- Owen JB, Butterfield DA. Measurement of oxidized/reduced glutathione ratio. Methods Mol Biol. 2010;648:269–277.
- Hayes JD, Flanagan JU, Jowsey IR. Glutathione transferases. Annu Rev Pharmacol Toxicol. 2005;45:51–88.