



ELSEVIER

Available online at www.sciencedirect.com

ScienceDirect

Current Opinion in
Neurobiology

GABA actions and ionic plasticity in epilepsy

Kai Kaila^{1,2}, Eva Ruusuvuori^{1,2}, Patricia Seja^{1,2}, Juha Voipio¹ and Martin Puskarjov^{1,2}

Concepts of epilepsy, based on a simple change in neuronal excitation/inhibition balance, have subsided in face of recent insights into the large diversity and context-dependence of signaling mechanisms at the molecular, cellular and neuronal network level. GABAergic transmission exerts both seizure-suppressing and seizure-promoting actions. These two roles are prone to short-term and long-term alterations, evident both during epileptogenesis and during individual epileptiform events. The driving force of GABAergic currents is controlled by ion-regulatory molecules such as the neuronal K-Cl cotransporter KCC2 and cytosolic carbonic anhydrases. Accumulating evidence suggests that neuronal ion regulation is highly plastic, thereby contributing to the multiple roles ascribed to GABAergic signaling during epileptogenesis and epilepsy.

Addresses

¹Department of Biosciences, University of Helsinki, FI-00014 Helsinki, Finland

²Neuroscience Center, University of Helsinki, FI-00014 Helsinki, Finland

Corresponding author: Kaila, Kai (Kai.Kaila@helsinki.fi)

Current Opinion in Neurobiology 2014, 26:34–41

This review comes from a themed issue on **Inhibition: synapses, neurons and circuits**

Edited by **Gordon Fishell** and **Gábor Tamás**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 30th November 2013

0959-4388 © 2013 The Authors. Published by Elsevier Ltd. Open access under [CC BY license](#).

<http://dx.doi.org/10.1016/j.conb.2013.11.004>

Much of the neurobiological research on epilepsies has focused on the role of GABAergic transmission in various phases of disease progression. Alterations in GABAergic signaling, which include changes in the properties of interneurons and in their quantitative as well as qualitative postsynaptic effects, are intimately involved in the development and chronic manifestations of epileptiform activity. In this review, we focus on GABA_A receptor (GABA_AR) functions and the ion transporters which affect the reversal potential of GABA_AR-mediated currents (E_{GABA}). ‘Ionic plasticity’ [1] (Figure 1) refers to changes in neuronal signaling related to the operation and functional modulation of plasmalemmal ion transporters

(Figure 2a) which set E_{GABA} either directly (Cl^- and/or HCO_3^- transporters) or indirectly (the Na-K ATPase).

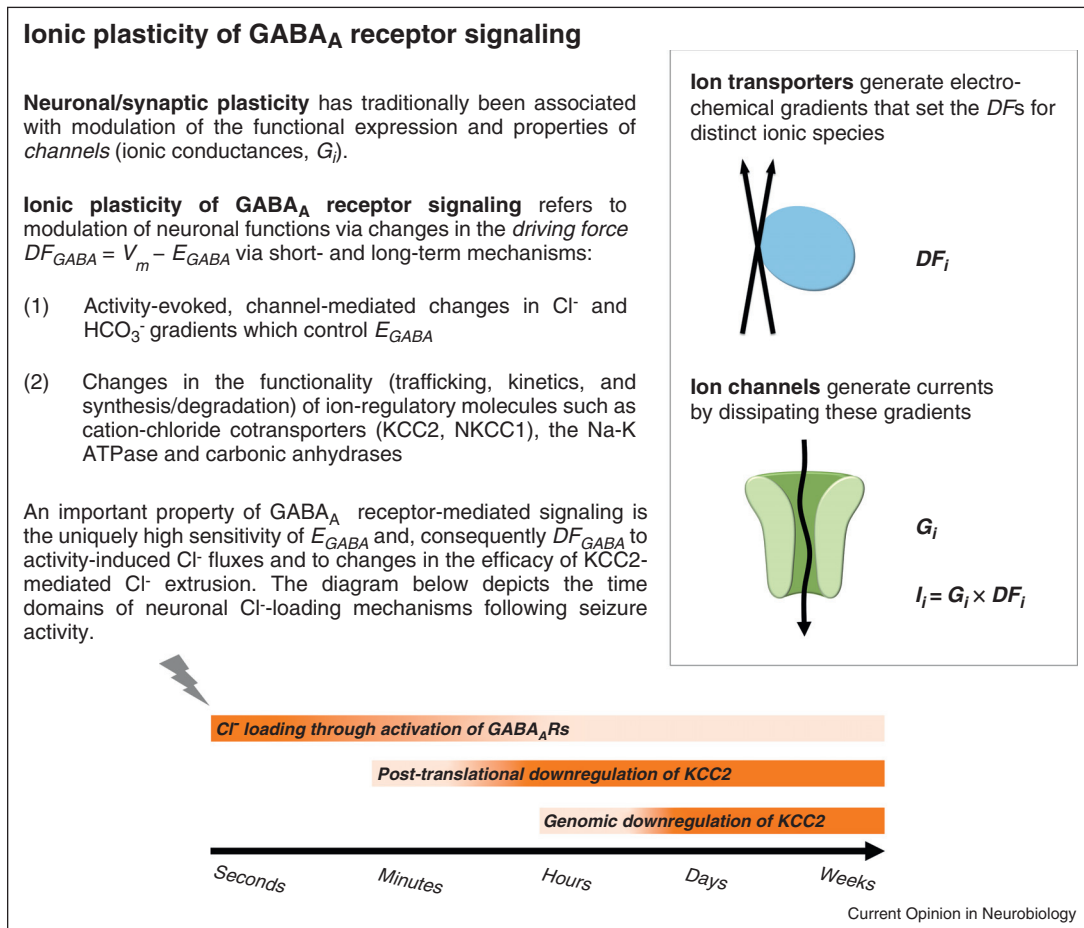
Epilepsies have turned out to be a spectrum disorder with a range of etiologies and comorbidities. Concepts of epilepsy and epileptogenesis seem likely to undergo revisions [2,3*]. Here we will mainly focus on mesial temporal lobe epilepsy (TLE) which is the most common type of refractory epilepsy. The primary cause leading to TLE is typically an insult to the brain (traumatic brain injury, inflammation, status epilepticus), but in patients the nature of the initial insult remains often unknown because of long delays between the insult and appearance of the recurrent seizures characteristic of TLE [4].

Seizures in the non-epileptic brain

Seizures can take place in disease states other than epilepsy. Refractory status epilepticus (SE), a life-threatening epileptic crisis characterized by prolonged recurrent seizures which do not respond to diazepam [5], is caused by factors such as inflammation and stroke, and it is seen in a minority of patients with established epilepsy. Much of our knowledge on the mechanisms and consequences of seizures come from *in vivo* and *in vitro* work on animals with no previous history of epilepsy or epileptogenesis. Induction of experimental SE has been shown to produce fast and robust changes in neuronal plasticity and a fast development of pharmacoresistance to conventional antiepileptic drugs which enhance GABAergic transmission. A straightforward explanation (see [6,7]) is that recurrent seizures lead to a progressive internalization of postsynaptic GABA_ARs and to a consequent erosion of inhibition. Work on brain slices has shown that the efficacy of feedforward inhibition declines rapidly after recurrent seizure-like activity, leading to a loss of the powerful inhibitory surround that is initially associated with these paroxysmal events [8,9*,10]. Seizures evoked in healthy adult brain tissue induce a fast decrease in the expression of KCC2 [11–13], the main neuronal Cl^- extruder. This molecule underlies classical, ‘Eccles-type’ hyperpolarizing inhibitory postsynaptic potentials (IPSPs) in central neurons [14]. GABAergic inhibition also has a shunting effect on electrical signals in the postsynaptic membrane (Box 1). The downregulation of GABA_ARs and KCC2 in response to trauma and/or intense seizure activity (Figure 2b) leads to a long-lasting decrease in the efficacy of both shunting and voltage inhibition, respectively.

The extrusion capacity of KCC2 can saturate even in the absence of functional downregulation [15*] of the transporter. Therefore, *the loss of voltage inhibition in response to*

Figure 1

Ionic plasticity of GABA_A receptor signaling.

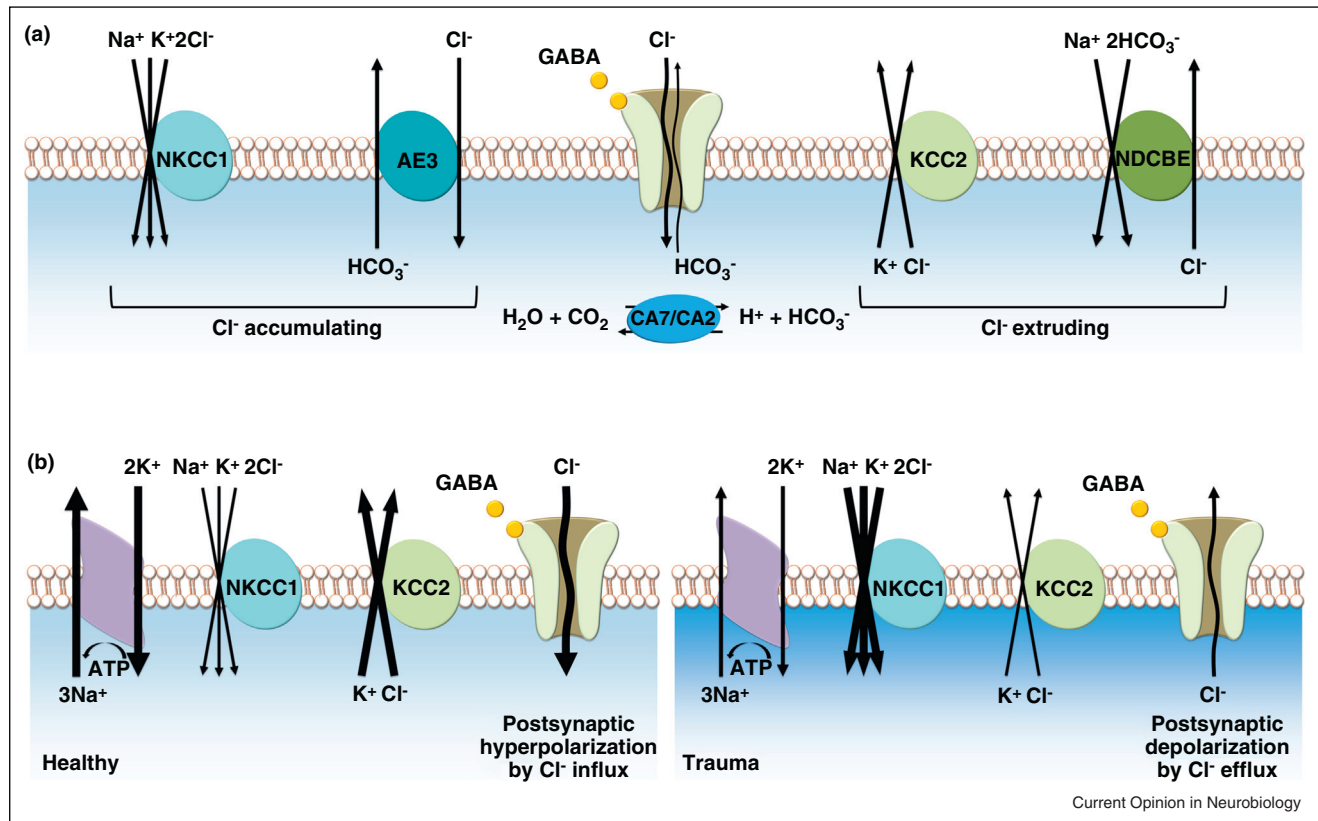
seizure activity is likely to be much faster than the loss of shunting. This is because the intense activation of interneurons will lead to a massive Cl^- influx which is aided by the depolarizing currents mediated by HCO_3^- across GABA_ARs and by coactivation of glutamatergic ionotropic receptors [16], as shown by continuous monitoring of E_{GABA} [17]. In fact, the depolarization mediated by the HCO_3^- current can drive a Cl^- influx that is large enough to induce a qualitative change in GABA_AR-mediated signaling from inhibitory to excitatory [17,18*]. During intense GABA_AR activation, the driving force of the inward HCO_3^- current is much more stable than that of the outward Cl^- current because the intracellular HCO_3^- is effectively replenished by neuronal cytosolic carbonic anhydrase [19,20*]. Indeed, under experimental conditions with enhanced synaptic release of GABA, pharmacologically isolated GABA_AR-mediated transmission alone is able to produce spontaneous paroxysmal activity in the brain slice [21]. The effect of the HCO_3^- -dependent anion shift is augmented by KCC2-mediated net accumulation of K^+ in the interstitial space [18*], leading to further, non-synaptic depolarization and excitation of neurons. This

probably includes (the at its time enigmatic observation of) GABA-driven antidromic spiking [22]. The membrane potential of glial cells is highly sensitive to extracellular K^+ , and the above positive feedback loop may act in synergy with glutamate release [23] from depolarized glia [17] to sustain a seizure. GABA_ARs and the Na-K-2Cl cotransporter NKCC1 (but not KCC2) are expressed in presynaptic terminals [14], and thus the antidromic spiking might also involve a direct presynaptic GABA_AR depolarization following an activity-dependent anion shift within the terminal. The role of extracellular K^+ is a classical focus in epilepsy research [24], and the time is ripe to readdress this topic with the novel insight pointing to GABAergic K^+ transients as a major pro-convulsant mechanism [25,26].

Seizures in the epileptic brain

It is likely that seizure mechanisms in chronically epileptic tissue differ dramatically from those evoked in brain tissue in healthy experimental animals or observed in patients with no previous history of epilepsy. Even if no cognitive defects are detectable during seizure-free periods of time, the cortex of chronically epileptic patients generates

Figure 2



Neuronal ion regulation sets the driving force for GABA_A receptor-mediated currents. **(a)** The cation chloride cotransporters govern neuronal Cl⁻ regulation. In mature neurons the K-Cl cotransporter isoform 2, KCC2, mediates Cl⁻ extrusion driven by the K⁺ gradient while the Na⁺-K-2Cl cotransporter isoform 1, NKCC1, mediates Na⁺ driven Cl⁻ uptake. In addition to this, the Na⁺-independent and Na⁺-dependent Cl⁻-HCO₃⁻ exchangers AE3 and NDCBE, respectively, may modulate intracellular Cl⁻ levels. The main function of these HCO₃⁻ transporters, together with the ubiquitous Na-H exchanger (not depicted), is to keep the intracellular pH level significantly more alkaline than what is predicted on the basis of passive distribution of H⁺ and HCO₃⁻ ions. Hence, the equilibrium potential for HCO₃⁻ is much more positive than the resting membrane potential and HCO₃⁻ invariably mediates a depolarizing current across GABA_ARs. Intracellular HCO₃⁻ concentration is rapidly replenished even during prolonged GABA_AR activation via the activity of carbonic anhydrase isoforms 2 and 7 (CA2 and CA7) which catalyze the formation of HCO₃⁻ from CO₂ [20]. **(b)** The cation-chloride cotransporters shown in panel (a) are fueled by the Na⁺ and K⁺ gradients generated by the Na-K ATPase. The conventional, GABA_AR-mediated hyperpolarizing IPSPs seen in mature neurons depend on the functional expression of KCC2 (left) that maintains a low intracellular Cl⁻ level that favors conductive Cl⁻ influx. Seizure-induced post-translational changes in CCC functional expression, e.g. via altered membrane expression, commence in tens of minutes. Prolonged changes at the levels of post-translational modification and transcription convert GABA_AR signaling back to its immature, depolarizing/excitatory mode of action during the course of epileptogenesis (right). The positive shift in E_{GABA} assists in reducing energy consumption during 'energy crisis' by reducing the driving forces of temporally overlapping and mutually counteracting excitatory and inhibitory postsynaptic ion fluxes (see [16]). For further details, see text.

abnormal interictal activities which are seen as brief (tens of milliseconds) spikes in the EEG. In hippocampal tissue from TLE patients, the *in vitro* counterpart of interictal activity is highly sensitive to bumetanide [27], a drug that selectively blocks Cl⁻ uptake mediated by NKCC1 in neurons *in vitro* [26]. In a manner similar to the depolarizing GABA_AR actions and associated NKCC1-dependent network events in the immature rodent hippocampus [14], interictal activity *in vitro* shows an obligatory dependence on excitatory GABAergic and glutamatergic excitatory synaptic drive [28,29**]. In hippocampal tissue resected from human TLE patients, intracellular recordings have revealed a subpopulation of pyramidal neurons where KCC2 levels are low and GABA has an excitatory action

[27,28] (see also [13]). Thus, GABA_AR signaling appears to resume its immature, depolarizing/excitatory mode of action at least in some pyramidal neurons during the course of epileptogenesis. However, while the changes in cation-chloride cotransporter (CCC) expression levels (low KCC2, high NKCC1) in these cells provide an explanation for the generation of interictal activity, there are no data to suggest that ictogenesis is based on these mechanisms (for review see [26]).

Intriguingly, it seems to be more difficult to evoke seizures in human TLE tissue than in brain tissue from healthy animals [30*]. This is a difficult paradox, since surgically obtained human TLE tissue typically has a

Box 1 Synaptic and extrasynaptic GABA_AR signaling

Synaptic GABA_AR-mediated inhibition is based on shunting and hyperpolarization of the postsynaptic membrane. *Shunting inhibition* has a duration set by the GABA_AR channels' open time, and the associated increase in conductance acts to suppress the temporal and spatial summation of incoming excitatory synaptic signals, as well as intrinsic pro-excitatory currents generated in the dendritic tree. *Voltage inhibition*, which hyperpolarizes the postsynaptic membrane, is dependent on the inwardly-directed electrochemical gradient of Cl⁻, maintained by KCC2, and it counteracts excitatory mechanisms for a longer time period, which is set by the time constant of the cell membrane. Unlike shunting, voltage inhibition does not take place in all types of mature CNS neurons because of cell-type specific lack of KCC2. The postsynaptic GABA_ARs in the neocortex and hippocampus consist of α(1-3), β(x) and γ2 subunits [80] whereof the of α1β2γ2 is the most common one.

Tonic GABA_AR-mediated signaling is based on the activation of high-affinity *extrasynaptic GABA_AR*s (with a subunit combination consisting of α5βγ2, α4βδ or α1βδ in neocortical and hippocampal neurons) by ambient GABA. The subunit composition of GABA_AR undergoes marked changes during epileptogenesis with consequent changes in the abundance of postsynaptic and extrasynaptic receptors [81], often followed by an increase in tonic inhibition [82]. Tonic GABAergic signaling is highly sensitive to changes in the efficacy of GABA uptake transporters (GAT1-4) [83], and it produces a spatially extended shunting effect in the target neurons, with voltage changes set by DF_{GABA}. Excessive tonic inhibition is known to promote absence seizures by inducing slow-wave discharges in thalamo-cortical networks, while enhancing tonic inhibition has an anticonvulsant action in partial seizures and catamenial epilepsy [84]. Synaptic GABAergic signaling is often called 'phasic' (e.g. 'phasic inhibition') to underscore its distinct properties versus tonic signaling. Both phasic and tonic GABA_AR-mediated signaling can be functionally inhibitory or excitatory, depending on the ion-regulatory mechanisms which set E_{GABA} and DF_{GABA}, on the GABA_AR-mediated conductance; and on the intrinsic electrophysiological properties of the target neuron.

sclerotic CA1 region, and such macroscopic differences in the properties of the healthy versus chronically diseased circuitry will compromise evaluation of the effects of distinct CCCs or changes in their expression. In slices from human TLE tissue, seizure-like events are not preceded by interictal but rather by 'pre-ictal' events which are largely based on recurrent glutamatergic signaling [29^{••},31]. These bursts have a wide spatial extent and a high propagation speed which probably makes them particularly effective in activating interneurons [30[•]]. While dendritic GABAergic synapses are lost in animal models of chronic epilepsy [32], a wealth of data suggest that parvalbumin-positive, perisomatically targeting interneurons shape the rhythmicity and synchrony which makes it possible for the seizures to effectively spread across wide cortical territories [33].

What triggers seizures in TLE? The diversity of recent explanations shows that this fundamental problem has still not been satisfactorily solved. We note that the term 'trigger' is ambiguous in that it has been used to describe (i) extrinsic factors that increase the propensity of seizures (e.g. hyperventilation or fever in children) and (ii) the

intrinsic neuronal and network mechanisms that act as immediate causes for seizure generation. Here, we will consider the latter. There is evidence that local desynchronization of neuronal activity is needed for the initiation of seizure activity [34]. Thus, a simple working hypothesis for the generation of TLE-related seizures and the role of KCC2 therein might be constructed as follows: In the seizure-triggering 'kernel' of diseased tissue, pre-ictal activity leads to a loss of phasic hyperpolarizing IPSPs (Box 1), which results (over short times) from the high Cl⁻ load [35,36] and is enhanced and consolidated largely by post-translational downregulation of membrane-associated KCC2 [37[•],38[•]] and later (cf. [37[•]]) by block of KCC2 transcription (Figure 1). The lack of hyperpolarizing IPSPs will lead, in turn, to the local desynchronization and promotion of seizures [34]. This idea is consistent with and supported by the actions of hyperpolarizing inhibition on spike probability and timing in healthy tissue [39].

TrkB and calpain as coordinating factors in epileptogenesis and epilepsy

BDNF-TrkB signaling has been put forward as a coordinating factor in epileptogenesis [40–42]. Indeed, the parallel loss of postsynaptic GABA_ARs and KCC2 after recurrent seizures may imply a shared mechanism, which most likely consists of signaling cascades down-stream of the tropomyosin-related kinase B (TrkB) receptor [40], the main target of brain-derived neurotrophic factor (BDNF). Seizures enhance BDNF secretion and the activation of TrkB [41] but, notably, BDNF itself is not always responsible for seizure-induced TrkB activation (cf. [43]). Conditional knockout of TrkB [44], transient inhibition of TrkB [45] or uncoupling of TrkB from the PLCγ1 cascade [46[•]] are all reported to suppress epileptogenesis. Enhanced TrkB activation in mature neurons rapidly decreases surface expression of GABA_ARs [47,48] and downregulates KCC2 [12,49]. There exist close parallels between the role of BDNF-TrkB signaling in epilepsy and in chronic pain [50]. In both cases, inflammation may induce BDNF secretion from activated microglia, and cause a downregulation of KCC2 in adjacent neurons [50]. Notably, inflammation is also a major cause of SE and epileptogenesis [51].

Fast, seizure-induced downregulation of KCC2 activity (over tens of minutes to hours), depends on post-transcriptional mechanisms [15[•],37[•]], including protein phosphatase 1-mediated dephosphorylation of KCC2 at serine 940 [38[•],52] and cleavage by the protease calpain [37[•],52], which is activated by Ca²⁺ and/or BDNF (for review, see [53]). A decrease in KCC2 mRNA occurs within hours of a seizure [12] and may contribute to consolidate KCC2 downregulation (Figure 1), but is not needed for chronic suppression of KCC2 protein expression or Cl⁻ extrusion [54]. Indeed, the increased level of calpain expression observed in TLE tissue [55,56] could account

for the chronic suppression of hippocampal KCC2 expression observed in patients with chronic epilepsy (see above). Interestingly, up-regulation of the gene encoding for the endogenous calpain inhibitor calpastatin is observed during the acute and latent phase of limbic epileptogenesis, whereas this enhancement appears to be lost in the chronic phase, characterized with spontaneous recurrent seizures [57]. Notably, calpain cleaves not only KCC2 [37,52,54] but also other proteins involved in GABAergic transmission, including GAD65 [58], VGAT/VIIAT [59], GAT1 [60] and gephyrin [61]. Thus, mounting evidence suggests that activation of calpain is another coordinating factor in epileptogenesis with important effects on GABAergic signaling (see also [62]).

Developmental stage and seizure mechanisms

Neuronal signaling mechanisms are radically different in developing and mature brain. One of the best examples is GABA_AR-mediated signaling, which undergoes a well-known ‘developmental shift’ from depolarizing/excitatory to hyperpolarizing (for review, see [14,63]). Initially neuronal Cl⁻ accumulation by NKCC1 is dominant, and KCC2 is expressed later during neuronal maturation [14]. The maturation of KCC2-dependent hyperpolarizing inhibition is accompanied by the expression of neuronal carbonic anhydrase isoform 7 (CA7), at around postnatal day (P) 12 in rodent hippocampus [64], followed by neuronal expression of the ubiquitous CA isoform 2 (CA2) at ~P20 [20^{*}]. The *simultaneous presence* of KCC2 and CA activity is crucial both for the generation of GABA-dependent neuronal Cl⁻ loads in response to interneuronal activity, and for paroxysmal extracellular K⁺ transients [18^{*}]. The key role of NKCC1 in immature, depolarizing GABAergic transmission and GABA-driven network events has led to a number of studies in neonatal rodents on the possible therapeutic use of the NKCC1 blocker, bumetanide. This work has been largely disappointing, as described elsewhere [26]. Cortical development is much more advanced in the human newborn than in the rodent [65]. Unlike in neonate rodents, CA7 and KCC2 are both expressed at high levels in hippocampus and neocortex of full term human babies [15^{*},20^{*},66,67]. This major species difference has numerous implications for translational work on the mechanisms of GABAergic signaling and seizures.

Adaptive mechanisms

From an evolutionary point of view, it is easy to understand why neurons and neuronal networks are endowed with adaptive response patterns which promote their survival under various insults [68]. Adaptive mechanisms can be detected, for instance, in the slowing of disease progression induced by application of proconvulsant drugs such as atipamezole and rimonabant soon after an insult [69,70]. Moreover, whether disease stage-related expression patterns of the endogenous calpain

inhibitor calpastatin [57] (see above) are causally involved here, is an interesting question for future work. Clearly, empirical information is required to judge whether a disease-related change at the molecular, cellular or network level is a ‘dysfunction’ (maladaptive, pro-epileptogenic) or an adaptive (anti-epileptogenic) process.

After trauma, neurons undergo processes of de-differentiation, seen as a shift in gene expression patterns to those of earlier developmental stages [1,71]. The adaptive value of such processes may be best explained from the factors involved in neuronal survival after trauma: (i) downregulation of energy metabolism under conditions of an ‘energy crisis’ (see [72]); as well as the presence of (ii) sufficient connectivity and (iii) trophic factor signaling, which promote neuronal survival. With regard to (i), the mammalian brain works close to theoretical limits on energy consumption, with most of it used to maintain the ionic driving forces which are needed for electrical signaling [16,73]. Changes in the functions and expression patterns of ion transporters and channels may thus have evolved as adaptive mechanisms to protect neurons during states of energy crisis. This idea fits well with the fast downregulation of both ion transporters and channels (e.g. GABA_ARs and KCC2) in response to seizures (see above and Figure 2b). Furthermore, the Na-K ATPase, the major ion-regulatory and energy-consuming molecule of the brain, is functionally downregulated after trauma or seizure [26,74]. The Na-K ATPase and KCC2 are functionally tightly linked, and there is evidence that the two molecules form a structural ion-transport metabolon [75,76]. We note also that shutting down Cl⁻ permeable GABA_ARs will reduce the energy-metabolic load imposed by cation-based glutamatergic signaling [16].

Conclusions

Changes in excitation–inhibition (E/I) balance are often used to explain epileptogenesis and seizure generation but, as should be obvious from the work reviewed above, the explanatory value of the E/I balance in the context of epilepsy is limited. Moreover, the postulated cause (E/I imbalance) is deduced from the outcome (seizures), which is an obvious circular argument. A *gross* change in the E/I imbalance is not, either, consistent with the fact that seizures in chronic epilepsy can occur infrequently and unpredictably, with intervening periods of intact cognitive and mnemonic cortico-hippocampal functions. The studies reviewed presently demonstrate that GABA_AR signaling has multiple, context-specific and age-specific actions which can prevent or promote epileptogenesis and seizure generation. A context-specificity and age-specificity is true also for intracellular signaling cascades such as those down-stream of TrkB receptors [76–78], which exert a strong influence on neuronal plasticity. This context-dependent diversity in cellular

and molecular signaling is not only a major challenge for basic research on the etiology and mechanisms of epileptiform syndromes, but also for the design of novel, genuinely antiepileptic drugs [79] which, instead of having solely symptomatic anticonvulsant actions, would halt and even reverse the progression of epilepsy.

Acknowledgements

We thank Profs. Wolfgang Löscher and Richard Miles for constructive comments on an early version of this paper. The authors' original work was supported by the Academy of Finland, the Sigrid Jusélius Foundation, the Jane and Aatos Erkko Foundation and the Letten Foundation.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Rivera C, Voipio J, Kaila K: **Two developmental switches in GABAergic signalling: the K⁺-Cl⁻ cotransporter KCC2 and carbonic anhydrase CAVII.** *J Physiol* 2005, **564**:953.
2. Jensen FE: **Epilepsy as a spectrum disorder: implications from novel clinical and basic neuroscience.** *Epilepsia* 2011, **52**(Suppl. 1):1-6.
3. Maguire J, Salpekar JA: **Stress, seizures, and hypothalamic-pituitary-adrenal axis targets for the treatment of epilepsy.** *Epilepsy Behav* 2013, **26**:352-362.
- This review provides an excellent overview of the important roles which stress and the HPA axis play in epilepsy, and highlights stress-related pathological responses as potential therapeutic targets.
4. Fox CK, Glass HC, Sidney S, Lowenstein DH, Fullerton HJ: **Acute seizures predict epilepsy after childhood stroke.** *Ann Neurol* 2013, **74**:249-256.
5. Rossetti AO, Lowenstein DH: **Management of refractory status epilepticus in adults: still more questions than answers.** *Lancet Neurol* 2011, **10**:922-930.
6. Naylor DE, Liu H, Wasterlain CG: **Trafficking of GABA(A) receptors, loss of inhibition, and a mechanism for pharmacoresistance in status epilepticus.** *J Neurosci* 2005, **25**:7724-7733.
7. Wasterlain CG, Liu H, Naylor DE, Thompson KW, Suchomelova L, Niquet J, Mazarati AM, Baldwin RA: **Molecular basis of self-sustaining seizures and pharmacoresistance during status epilepticus: the receptor trafficking hypothesis revisited.** *Epilepsia* 2009, **50**(Suppl. 12):16-18.
8. Trevelyan AJ, Sussillo D, Yuste R: **Feedforward inhibition contributes to the control of epileptiform propagation speed.** *J Neurosci* 2007, **27**:3383-3387.
9. Trevelyan AJ, Schevon CA: **How inhibition influences seizure propagation.** *Neuropharmacology* 2013, **69**:45-54.
- A lucid description of the multiple roles of GABAergic interneurons in the generation, suppression and propagation of seizures.
10. Cammarota M, Losi G, Chiavegato A, Zonta M, Carmignoto G: **Fast spiking interneuron control of seizure propagation in a cortical slice model of focal epilepsy.** *J Physiol* 2013, **591**:807-822.
11. Lee HH, Jurd R, Moss SJ: **Tyrosine phosphorylation regulates the membrane trafficking of the potassium chloride cotransporter KCC2.** *Mol Cell Neurosci* 2010, **45**:173-179.
12. Rivera C, Li H, Thomas-Crusells J, Lahtinen H, Viitanen T, Nanobashvili A, Kokaia Z, Airaksinen MS, Voipio J, Kaila K, Saarna M: **BDNF-induced TrkB activation down-regulates the K⁺-Cl⁻ cotransporter KCC2 and impairs neuronal Cl⁻ extrusion.** *J Cell Biol* 2002, **159**:747-752.
13. Pathak HR, Weissinger F, Terunuma M, Carlson GC, Hsu FC, Moss SJ, Coulter DA: **Disrupted dentate granule cell chloride regulation enhances synaptic excitability during development of temporal lobe epilepsy.** *J Neurosci* 2007, **27**:14012-14022.
14. Blaesse P, Airaksinen MS, Rivera C, Kaila K: **Cation-chloride cotransporters and neuronal function.** *Neuron* 2009, **61**:820-838.
15. Kahle KT, Deeb TZ, Puskarjov M, Silayeva L, Liang B, Kaila K, Moss SJ: **Modulation of neuronal activity by phosphorylation of the K-Cl cotransporter KCC2.** *Trends Neurosci* 2013 <http://dx.doi.org/10.1016/j.tins.2013.08.006>. in press.
- A compact update on post-translational mechanisms which regulate KCC2 function under physiological and pathophysiological conditions.
16. Buzsaki G, Kaila K, Raichle M: **Inhibition and brain work.** *Neuron* 2007, **56**:771-783.
17. Kaila K, Lamsa K, Smirnov S, Taira T, Voipio J: **Long-lasting GABA-mediated depolarization evoked by high-frequency stimulation in pyramidal neurons of rat hippocampal slice is attributable to a network-driven, bicarbonate-dependent K⁺ transient.** *J Neurosci* 1997, **17**:7662-7672.
18. Viitanen T, Ruusuvaari E, Kaila K, Voipio J: **The K⁺-Cl cotransporter KCC2 promotes GABAergic excitation in the mature rat hippocampus.** *J Physiol* 2010, **588**:1527-1540.
- The authors demonstrate that during intense interneuronal activity, carbonic anhydrase-dependent chloride loading of neurons leads to a KCC2-mediated increase in extracellular K⁺ which promotes nonsynaptic excitation and is likely to contribute to paroxysmal activity.
19. Kaila K: **Ionic basis of GABA(A) receptor channel function in the nervous system.** *Prog Neurobiol* 1994, **42**:489-537.
20. Ruusuvaari E, Huebner AK, Kirilkin I, Yukin AY, Blaesse P, Helmy M, Kang HJ, El MM, Hennings JC, Voipio J, Sestan N, Hubner CA, Kaila K: **Neuronal carbonic anhydrase VII provides GABAergic excitatory drive to exacerbate febrile seizures.** *EMBO J* 2013, **32**:2275-2286.
- This study describes the developmental expression patterns of carbonic anhydrase isoforms CA2 and CA7 in the mouse hippocampus, and shows that GABAergic excitation mediated by the neuron-specific isoform CA7 contributes to experimental febrile seizures in rodents. CA7 expression patterns in the human brain were found to be consistent with a similar role in febrile seizures in children.
21. Uusisaari M, Smirnov S, Voipio J, Kaila K: **Spontaneous epileptiform activity mediated by GABA(A) receptors and gap junctions in the rat hippocampal slice following long-term exposure to GABA(B) antagonists.** *Neuropharmacology* 2002, **43**:563-572.
22. Stasheff SF, Mott DD, Wilson WA: **Axon terminal hyperexcitability associated with epileptogenesis in vitro. II. Pharmacological regulation by NMDA and GABA(A) receptors.** *J Neurophysiol* 1993, **70**:976-984.
23. Angulo MC, Kozlov AS, Charpak S, Audinat E: **Glutamate released from glial cells synchronizes neuronal activity in the hippocampus.** *J Neurosci* 2004, **24**:6920-6927.
24. Bazhenov M, Timofeev I, Frohlich F, Sejnowski TJ: **Cellular and network mechanisms of electrographic seizures.** *Drug Discov Today Dis Models* 2008, **5**:45-57.
25. Miles R, Blaesse P, Huberfeld G, Wittner L, Kaila K: **Chloride homeostasis and GABA signaling in temporal lobe epilepsy.** In *Jasper's Basic Mechanisms of the Epilepsies*, edn 4. Edited by Noebels JL, Avoli M, Rogawski MA, Olsen RW, Delgado-Escueta AV. Oxford University Press; 2012.
26. Löscher W, Puskarjov M, Kaila K: **Cation-chloride cotransporters NKCC1 and KCC2 as potential targets for novel antiepileptic and antiepileptogenic treatments.** *Neuropharmacology* 2013, **69**:62-74.
27. Huberfeld G, Wittner L, Clemenceau S, Baulac M, Kaila K, Miles R, Rivera C: **Perturbed chloride homeostasis and GABAergic signaling in human temporal lobe epilepsy.** *J Neurosci* 2007, **27**:9866-9873.
28. Cohen I, Navarro V, Clemenceau S, Baulac M, Miles R: **On the origin of interictal activity in human temporal lobe epilepsy in vitro.** *Science* 2002, **298**:1418-1421.
29. Huberfeld G, Menendez de la Prida L, Pallud J, Cohen I, Le Van QM, Adam C, Clemenceau S, Baulac M, Miles R:
 - of outstanding interest

Glutamatergic pre-ictal discharges emerge at the transition to seizure in human epilepsy. *Nat Neurosci* 2011, **14**:627-634.

This study, based on EEG and slice electrophysiology, is an excellent dissociation of the mechanisms underlying interictal and pre-ictal discharges in human TLE, and demonstrates that once established, recurrence of the pre-ictal discharges triggers seizures.

30. Huberfeld G, Le Duigou C, Le Van QM, Navarro V, Baulac M, Miles R: **The paradox of the paroxysm: can seizure precipitants help explain human ictogenesis?** *Neuroscientist* 2013, **19**:523-540.
- A thought-provoking article which aims at identifying common neuronal pathways for a wide diversity of seizure-precipitating factors, ranging from sleep deprivation and oscillations in hormonal levels to drug withdrawal.
31. Dzhalal VI, Staley KJ: **Excitatory actions of endogenously released GABA contribute to initiation of ictal epileptiform activity in the developing hippocampus.** *J Neurosci* 2003, **23**:1840-1846.
32. Cossart R, Dinocourt C, Hirsch JC, Merchán-Pérez A, De Felipe J, Ben Ari Y, Esclapez M, Bernard C: **Dendritic but not somatic GABAergic inhibition is decreased in experimental epilepsy.** *Nat Neurosci* 2001, **4**:52-62.
33. Freund TF, Katona I: **Perisomatic inhibition.** *Neuron* 2007, **56**:33-42.
34. Jiruska P, de Curtis M, Jefferys JG, Schevon CA, Schiff SJ, Schindler K: **Synchronization and desynchronization in epilepsy: controversies and hypotheses.** *J Physiol* 2013, **591**:787-797.
35. Deeb TZ, Nakamura Y, Frost GD, Davies PA, Moss SJ: **Disrupted Cl⁻ homeostasis contributes to reductions in the inhibitory efficacy of diazepam during hyperexcited states.** *Eur J Neurosci* 2013, **38**:2453-2467.
36. Lillis KP, Kramer MA, Mertz J, Staley KJ, White JA: **Pyramidal cells accumulate chloride at seizure onset.** *Neurobiol Dis* 2012, **47**:358-366.
37. Puskarjov M, Ahmad F, Kaila K, Blaesse P: **Activity-dependent cleavage of the K-Cl cotransporter KCC2 mediated by calcium-activated protease calpain.** *J Neurosci* 2012, **32**:11356-11364.
- The molecular mechanisms underlying seizure-induced erosion of inhibition at the level of neuronal Cl⁻ regulation are largely unknown. This study demonstrates a key role for calpain, a molecule with well-established roles in the control of neuronal plasticity, as a proximal mechanism responsible for downregulation of KCC2.
38. Lee HH, Deeb TZ, Walker JA, Davies PA, Moss SJ: **NMDA receptor activity downregulates KCC2 resulting in depolarizing GABA_A receptor-mediated currents.** *Nat Neurosci* 2011, **14**:736-743.
- As in the study above, NMDA receptors are found to have a key position in the activity-dependent run-down of KCC2. This work highlights the activity-dependent phosphorylation state of KCC2 as an important determinant of its functional stability and targeting for proteolytic pathways.
39. Haider B, McCormick DA: **Rapid neocortical dynamics: cellular and network mechanisms.** *Neuron* 2009, **62**:171-189.
40. McNamara JO, Huang YZ, Leonard AS: **Molecular signaling mechanisms underlying epileptogenesis.** *Sci STKE* 2006, **2006**:re12.
41. McNamara JO, Scharfman HE: **Temporal lobe epilepsy and the BDNF receptor, TrkB.** In *Jasper's Basic Mechanisms of the Epilepsies*, edn 4. Edited by Noebels JL, Avoli M, Rogawski MA, Olsen RW, Delgado-Escueta AV. Oxford University Press; 2012.
42. Goldberg EM, Coulter DA: **Mechanisms of epileptogenesis: a convergence on neural circuit dysfunction.** *Nat Rev Neurosci* 2013, **14**:337-349.
43. Rajagopal R, Chen ZY, Lee FS, Chao MV: **Transactivation of Trk neurotrophin receptors by G-protein-coupled receptor ligands occurs on intracellular membranes.** *J Neurosci* 2004, **24**:6650-6658.
44. He XP, Kotloski R, Nef S, Luikart BW, Parada LF, McNamara JO: **Conditional deletion of TrkB but not BDNF prevents epileptogenesis in the kindling model.** *Neuron* 2004, **43**:31-42.
45. Liu G, Gu B, He XP, Joshi RB, Wackerle HD, Rodriguiz RM, Wetsel WC, McNamara JO: **Transient inhibition of TrkB kinase after status epilepticus prevents development of temporal lobe epilepsy.** *Neuron* 2013, **79**:31-38.
46. He XP, Pan E, Sciarretta C, Minichiello L, McNamara JO: **Disruption of TrkB-mediated phospholipase Cgamma signaling inhibits limbic epileptogenesis.** *J Neurosci* 2010, **30**:6188-6196.
- On the basis of experiments on mice carrying a mutation (Y816F) that uncouples TrkB from PLCγ1, this study indicates that TrkB-dependent activation of PLCγ1 signaling is an important molecular mechanism of limbic epileptogenesis. Notably, this pathway has been implicated in regulation of KCC2 expression (cf. [1]).
47. Brunig I, Penschuck S, Berninger B, Benson J, Fritschy JM: **BDNF reduces miniature inhibitory postsynaptic currents by rapid downregulation of GABA(A) receptor surface expression.** *Eur J Neurosci* 2001, **13**:1320-1328.
48. Jovanovic JN, Thomas P, Kittler JT, Smart TG, Moss SJ: **Brain-derived neurotrophic factor modulates fast synaptic inhibition by regulating GABA(A) receptor phosphorylation, activity, and cell-surface stability.** *J Neurosci* 2004, **24**:522-530.
49. Rivera C, Voipio J, Thomas-Crusells J, Li H, Emri Z, Sipilä S, Payne JA, Minichiello L, Saarma M, Kaila K: **Mechanism of activity-dependent downregulation of the neuron-specific K-Cl cotransporter KCC2.** *J Neurosci* 2004, **24**:4683-4691.
50. Ferrini F, De Koninck Y: **Microglia control neuronal network excitability via BDNF signalling.** *Neural Plast* 2013, **2013**:429815.
51. Fabene PF, Navarro MG, Martinello M, Rossi B, Merigo F, Ottoboni L, Bach S, Angiari S, Benati D, Chakir A, Zanetti L, Schio F, Osculati A, Marzola P, Nicolato E, Homeister JW, Xia L, Lowe JB, McEver RP, Osculati F, Sbarbati A, Butcher EC, Constantin G: **A role for leukocyte-endothelial adhesion mechanisms in epilepsy.** *Nat Med* 2008, **14**:1377-1383.
52. Chamma I, Heubl M, Chevy Q, Renner M, Moutkine I, Eugene E, Ponce JC, Levi S: **Activity-dependent regulation of the K/Cl transporter KCC2 membrane diffusion, clustering, and function in hippocampal neurons.** *J Neurosci* 2013, **33**:15488-15503.
53. Baudry M, Chou MM, Bi X: **Targeting calpain in synaptic plasticity.** *Expert Opin Ther Targets* 2013, **17**:579-592.
54. Zhou HY, Chen SR, Byun HS, Chen H, Li L, Han HD, Lopez-Berestein G, Sood AK, Pan HL: **N-methyl-D-aspartate receptor- and calpain-mediated proteolytic cleavage of K⁺-Cl⁻ cotransporter-2 impairs spinal chloride homeostasis in neuropathic pain.** *J Biol Chem* 2012, **287**:33853-33864.
55. Feng ZH, Hao J, Ye L, Dayao C, Yan N, Yan Y, Chu L, Shi FD: **Overexpression of mu-calpain in the anterior temporal neocortex of patients with intractable epilepsy correlates with clinicopathological characteristics.** *Seizure* 2011, **20**:395-401.
56. Das A, Wallace GC, Holmes C, McDowell ML, Smith JA, Marshall JD, Bonilha L, Edwards JC, Glazier SS, Ray SK, Banik NL: **Hippocampal tissue of patients with refractory temporal lobe epilepsy is associated with astrocyte activation, inflammation, and altered expression of channels and receptors.** *Neuroscience* 2012, **220**:237-246.
57. Gorter JA, Van Vliet EA, Rauwerda H, Breit T, Stad R, van SL, Vreugdenhil E, Redeker S, Hendriksen E, Aronica E, Lopes da Silva FH, Wadman WJ: **Dynamic changes of proteases and protease inhibitors revealed by microarray analysis in CA3 and entorhinal cortex during epileptogenesis in the rat.** *Epilepsia* 2007, **48**(Suppl. 5):53-64.
58. Buddhala C, Suarez M, Modi J, Prentice H, Ma Z, Tao R, Wu JY: **Calpain cleavage of brain glutamic acid decarboxylase 65 is pathological and impairs GABA neurotransmission.** *PLoS ONE* 2012, **7**:e33002.
59. Gomes JR, Lobo AC, Melo CV, Inacio AR, Takano J, Iwata N, Saido TC, de Almeida LP, Wieloch T, Duarte CB: **Cleavage of the vesicular GABA transporter under excitotoxic conditions is followed by accumulation of the truncated transporter in nonsynaptic sites.** *J Neurosci* 2011, **31**:4622-4635.

60. Baliova M, Knab A, Franekova V, Jursky F: **Modification of the cytosolic regions of GABA transporter GAT1 by calpain.** *Neurochem Int* 2009, **55**:288-294.
61. Tyagarajan SK, Ghosh H, Yevenes GE, Imanishi SY, Zeilhofer HU, Gerrits B, Fritschy JM: **Extracellular signal-regulated kinase and glycogen synthase kinase 3 β regulate gephyrin postsynaptic aggregation and GABAergic synaptic function in a calpain-dependent mechanism.** *J Biol Chem* 2013, **288**:9634-9647.
62. O'Dell CM, Das A, Wallace G, Ray SK, Banik NL: **Understanding the basic mechanisms underlying seizures in mesial temporal lobe epilepsy and possible therapeutic targets: a review.** *J Neurosci Res* 2012, **90**:913-924.
63. Ben-Ari Y, Khalilov I, Kahle KT, Cherubini E: **The GABA excitatory/inhibitory shift in brain maturation and neurological disorders.** *Neuroscientist* 2012, **18**:467-486.
64. Ruusuvuori E, Li H, Huttu K, Palva JM, Smirnov S, Rivera C, Kaila K, Voipio J: **Carbonic anhydrase isoform VII acts as a molecular switch in the development of synchronous gamma-frequency firing of hippocampal CA1 pyramidal cells.** *J Neurosci* 2004, **24**:2699-2707.
65. Erecinska M, Cherian S, Silver IA: **Energy metabolism in mammalian brain during development.** *Prog Neurobiol* 2004, **73**:397-445.
66. Vanhatalo S, Palva JM, Andersson S, Rivera C, Voipio J, Kaila K: **Slow endogenous activity transients and developmental expression of K⁺-Cl⁻ cotransporter 2 in the immature human cortex.** *Eur J Neurosci* 2005, **22**:2799-2804.
67. Hyde TM, Lipska BK, Ali T, Mathew SV, Law AJ, Metitiri OE, Straub RE, Ye T, Colantuoni C, Herman MM, Bigelow LB, Weinberger DR, Kleinman JE: **Expression of GABA signaling molecules KCC2, NKCC1, and GAD1 in cortical development and schizophrenia.** *J Neurosci* 2011, **31**:11088-11095.
68. Overman JJ, Carmichael ST: **Plasticity in the injured brain: more than molecules matter.** *Neuroscientist* 2013 <http://dx.doi.org/10.1177/1073858413491146>. in press.
69. Pitkanen A, Narkilahti S, Bezvenyuk Z, Haapalinna A, Nissinen J: **Atipamezole alpha(2)-adrenoceptor antagonist, has disease modifying effects on epileptogenesis in rats.** *Epilepsy Res* 2004, **61**:119-140.
70. Echegoyen J, Armstrong C, Morgan RJ, Soltesz I: **Single application of a CB1 receptor antagonist rapidly following head injury prevents long-term hyperexcitability in a rat model.** *Epilepsy Res* 2009, **85**:123-127.
71. Cohen I, Navarro V, Le DC, Miles R: **Mesial temporal lobe epilepsy: a pathological replay of developmental mechanisms?** *Biol Cell* 2003, **95**:329-333.
72. Hansen AJ: **Effect of anoxia on ion distribution in the brain.** *Physiol Rev* 1985, **65**:101-148.
73. Laughlin SB, Sejnowski TJ: **Communication in neuronal networks.** *Science* 2003, **301**:1870-1874.
74. Ross ST, Soltesz I: **Selective depolarization of interneurons in the early posttraumatic dentate gyrus: involvement of the Na(+)/K(+)-ATPase.** *J Neurophysiol* 2000, **83**:2916-2930.
75. Ikeda K, Onimaru H, Yamada J, Inoue K, Ueno S, Onaka T, Toyoda H, Arata A, Ishikawa T, Taketo MM, Fukuda A, Kawakami K: **Malfunction of respiratory-related neuronal activity in Na⁺, K⁺-ATPase alpha 2 subunit-deficient mice is attributable to abnormal Cl⁻ homeostasis in brainstem neurons.** *J Neurosci* 2004, **24**:10693-10701.
76. Khirug S, Ahmad F, Puskarjov M, Afzalov R, Kaila K, Blaesse P: **A single seizure episode leads to rapid functional activation of KCC2 in the neonatal rat hippocampus.** *J Neurosci* 2010, **30**:12028-12035.
77. Di Lieto A, Rantamaki T, Vesa L, Yanpallewar S, Antila H, Lindholm J, Rios M, Tessarollo L, Castren E: **The responsiveness of TrkB to BDNF and antidepressant drugs is differentially regulated during mouse development.** *PLoS ONE* 2012, **7**:e32869.
78. Mizoguchi Y, Ishibashi H, Nabekura J: **The action of BDNF on GABA currents changes from potentiating to suppressing during maturation of rat hippocampal CA1 pyramidal neurons.** *J Physiol* 2003, **548**:703-709.
79. Löscher W, Klitgaard H, Twyman RE, Schmidt D: **New avenues for anti-epileptic drug discovery and development.** *Nat Rev Drug Discov* 2013, **12**:757-776.
80. Hines RM, Davies PA, Moss SJ, Maguire J: **Functional regulation of GABA receptors in nervous system pathologies.** *Curr Opin Neurobiol* 2012, **22**:552-558.
81. Grabenstatter HL, Russek SJ, Brooks-Kayal AR: **Molecular pathways controlling inhibitory receptor expression.** *Epilepsia* 2012, **53**(Suppl. 9):71-78.
82. Pavlov I, Walker MC: **Tonic GABA(A) receptor-mediated signalling in temporal lobe epilepsy.** *Neuropharmacology* 2013, **69**:55-61.
83. Madsen KK, White HS, Schousboe A: **Neuronal and non-neuronal GABA transporters as targets for antiepileptic drugs.** *Pharmacol Ther* 2010, **125**:394-401.
84. Brickley SG, Mody I: **Extrasynaptic GABA(A) receptors: their function in the CNS and implications for disease.** *Neuron* 2012, **73**:23-34.